

Polyoma Plaque Assay

11/9/67.

8 plates mouse embryo II^+ cultures.

Sucked off medium and washed 2x in ~5 ml Tris.

Virus stocks.

LP 148 A 1.4×10^8 pfu/ml rec'd from MV.

Diluted as follows 10 ml \rightarrow 5.0 ml Tris-5% HS = 2.8×10^8 pfu/ml. (SAVED)

Made additional dilutions as follows. 10 ml \rightarrow 5.0 (1:500); 10 ml \rightarrow 4.0 (1:400) \rightarrow 0.5 \rightarrow 2.5 (250 pfu/ml)
 \rightarrow 0.1 \rightarrow 2.0 (70 pfu/ml)

① 0.2 ml of diln cty \sim ~~70~~ 70 pfu/ml. (~14)

②

③ 0.2 ml of diln " \sim 250 pfu/ml. (~64)

④

P16/S4K $\sim 5 \times 10^{10}$ pfu/ml.

Diluted as follows. 10 ml \rightarrow 5.0 ml Tris-5% HS = 10^8 pfu/ml (SAVED)

then

2.5×10^5 pfu/ml
10 ml \rightarrow 4.0 (1:400); 10 ml \rightarrow 4.0 (1:400), 600 pfu \rightarrow 1:2 (300 pfu/ml)

\rightarrow 0.5 \rightarrow 2.5 (120 pfu/ml) \rightarrow 1:2 (60 pfu/ml)

⑤ 0.2 ml of diln cty 60 pfu/ml. (~12)

⑥

⑦ 0.2 ml of diln cty 300 pfu/ml (~60)

⑧

Adsorbed for 30' at 37° then overlaid with agar-Eagles-3.5% HS.
Incubated at 37° for 2 days then shifted to 33°

Polyoma Plaque Assay.

On 11/20 (the 12th day) overlaid with neutral-red agar ad pot at 33°

On 11/22 (14th day) could see clear plaques on the LP 148 A infected plates but none the P16 plates.

On 11/24 (16th day). nice clear plaques on LP but no easily visible plaques in P16.

On 11/27 (19th day) counted plaques

LP 148 A

| | | Au | Predicted from titre of stock |
|---|----|------|-------------------------------|
| ① | 9 | } 12 | ~ 14 |
| ② | 16 | | |
| ③ | 36 | } 33 | ~ 64 |
| ④ | 30 | | |

Only few plaques seen with P16 ∴ left at 33°

On 11/30/67

| | | Expected on basis of titre |
|---|------|----------------------------|
| ⑤ | 4 | } ~ 12 |
| ⑥ | lost | |
| ⑦ | 6 | } 60 |
| ⑧ | 4 | |

Either down by factor of 10 (stock is at 3rd pass cell) or plating efficiency at 33 is lower.

Attempt to make plaques on PY-6

12/2/67

8 ~~plates~~ trays of 6 plates each. Plated at ca 2×10^5 cells/plate on 11/30

Virus stock addition

LP148A 2×10^8 p.t.u.

10.4 \rightarrow 5.0 (1:10) \rightarrow 0.64 \rightarrow 5.0 (1:10) \rightarrow 0.64 \rightarrow 6.0 (1:10) 10^3 p.t.u.

Used 0.2 ml aliquots to infect

0.64 \rightarrow 6.0 (1:10) 10^2 p.t.u.

Plates washed in 25-64 Tris. Infected for 30' at 37° then overlaid with agar and different media then incubated at 35° or 37°

(see next page)

The first two of each set infected with 200 p.t.u. and the second set infected with 20 p.t.u.

38°

33°

* *
 (1) } 3.5% HTS
 (2) } + Hunter's
 (3) }
 (4) }

most cells dead - little staining - cells detached and floaty

* *
 (4) } 6.5% HTS
 (10) }
 (11) }
 (12) }

* *
 (17) } 0% HTS
 (18) }
 (19) }
 (20) }

* *
 (25) } 0% HTS
 (26) } 1:25
 (27) } Trypan
 (28) }

Well stained layer although much clumping: easily visible plaques: but only few/plate by 11 days

* *
 (33) } 0% HTS
 (34) } 1:100
 (35) } Trypan
 (36) }

Less well stained but clearly live cells in layer. Lack of clear areas but not certain there are plaques

* *
 (41) } 0% HTS
 (42) } 1:250
 (43) } Trypan
 (44) }

Still less well stained but perhaps some plaques

* *
 (6) } 3.5% HTS
 (7) } + Hunter's
 (8) }

Same as 38°

* *
 (13) } 6.5% HTS
 (14) }
 (15) }
 (16) }

* *
 (21) } 0% HTS
 (22) }
 (23) }
 (24) }

* *
 (29) } 2% HTS
 (30) } 1:25
 (31) } Trypan
 (32) }

Same as 38°

* *
 (37) } 0% HTS
 (38) } 1:100
 (39) } Trypan
 (40) }

Same as 38°

* *
 (45) } 6% HTS
 (46) } 1:250
 (47) } Trypan
 (48) }

Same as 38°

* Stained on 12/7
 all has stained on 12/9