

9/28

Started Second Exp with PY-6 to see if shorter X-ray exposures than 4' would be correct does for getting good plating cells for plaque assay with polyoma.

Took 2 large plates of PY-6.

Removed medium, washed in trypsin-TD and then incubated with ~4 ml of trypsin-TD at 37° for ~5'.

Cells come off glass easily.

Added 1 ml calf serum to stop trypsin action.

Removed cells and mixed the suspensions.

Distributed 5 ml to each of 2 small plates.

X-ray as before for 3' and 2'. (Allowed 5' between exposures).

Cells centrifuged and resuspended in 5 ml Eagle's serum.

Counted cells. - 6.3×10^6 cells/ml.

Took 2 ml of each suspension and diluted to 30 ml with Eagle's serum.

Distributed 5 ml to each of six plates and incubated at 37° overnight.

9/29

Cells had settled on plate and were "dividing".

At about 3 PM removed medium washed with ~5 ml Tris and infected in polyoma (LP140)

Titers was 2×10^8 - diluted in Tris as follows 1:50, 1:50, 1:40 to give 10^{-5} then another 1:10 to give 10^{-6} .

	<u>2'</u>	<u>3'</u>
0.1 ml of 10^{-5}	hi (3)	hi (3).
0.1 ml of 10^{-6}	lo (3)	lo (3).

After ~ 1.5 hours at 37° overlaid with 292r-Eagle's cells and incubate at 37°