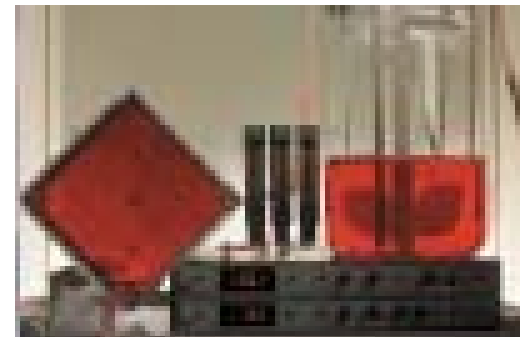
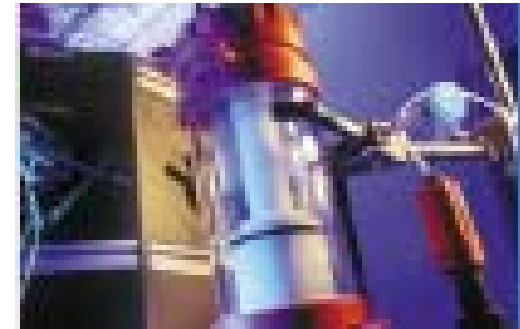


Assays for Bovine Viruses

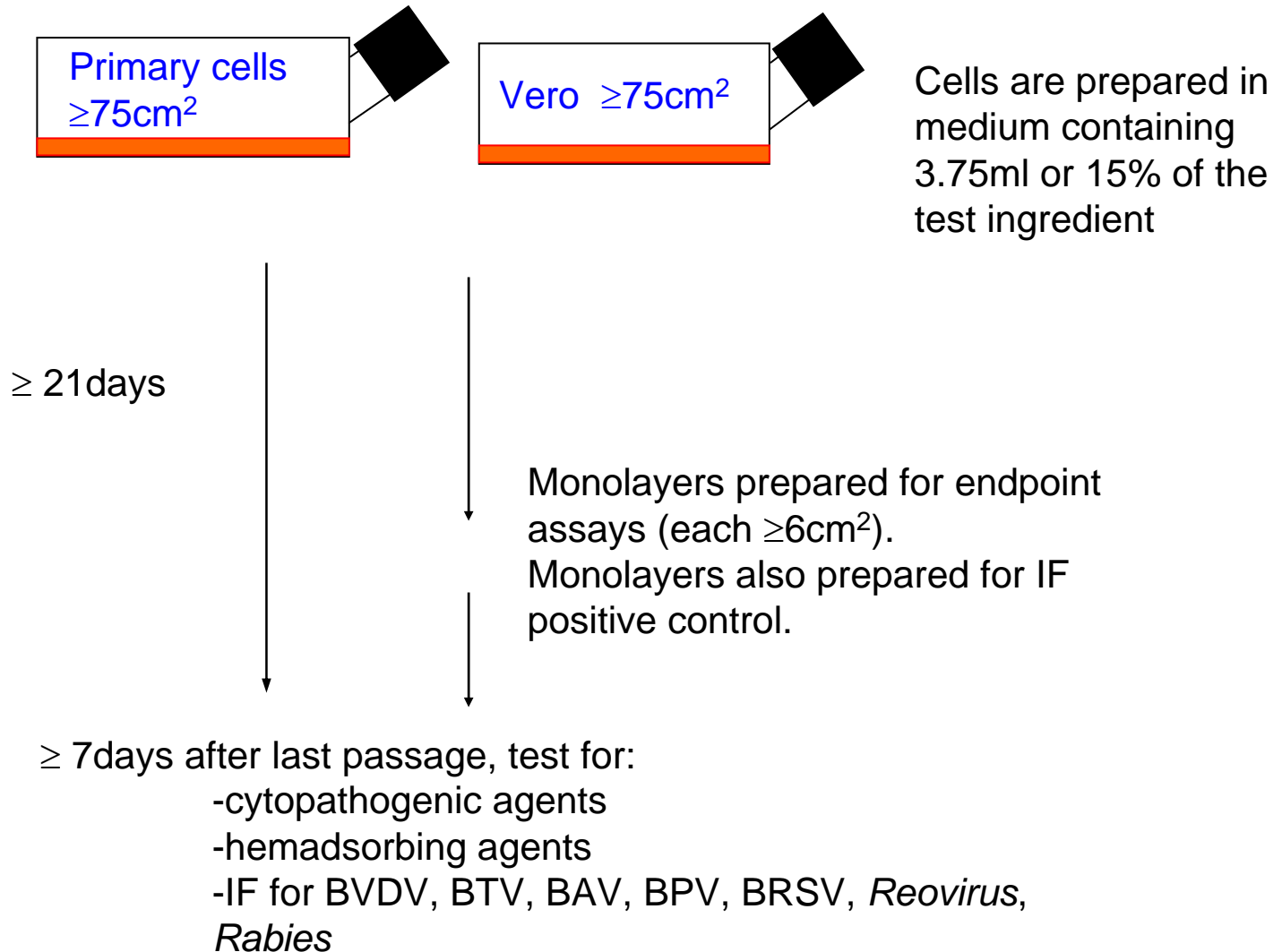
Dr Martin Wisher
Director for Market Development and
Regulatory Affairs



- US Code of Federal Regulations
 - 9CFR 113.53 Requirements for ingredients of animal origin used for production of biologics.
 - 9CFR 113.46 Detection of cytopathogenic and/or hemadsorbing agents
 - 9CFR 113.47 Detection of extraneous viruses by the fluorescent antibody technique
- EMEA CPMP Note for Guidance on the Use of Bovine Serum in the Manufacture of Human Biological Medicinal Products. CPMP/BWP/1793/02
- European Pharmacopoeia Draft Monograph 'Bovine Serum' (2003)

- General tests for:
 - Cytopathogenic agents
 - Haemadsorbing agents
- Specific test (fluorescent antibody) for:
 - Bovine viral diarrhoea virus (BVDV)
 - Blue tongue (BTV)
 - Bovine adenovirus (BAV)
 - Bovine parvovirus (BPV)
 - Bovine respiratory syncytial virus (BRSV)
 - Reovirus 3
 - Rabies

Testing according to 9CFR Part 113.53



- Tests for viral contaminants (prior to inactivation)
- General tests for
 - Haemadsorbing viruses (e.g., PI3)
 - Cytopathic agents (e.g., IBRV)
- Specific tests for:
BTV, BAV, BPV, BRSV, BVDV, Reo3, Rabies

CPMP Note for Guidance recommendations for BVDV

- Presence of BVDV in serum cannot be completely avoided
- Test for BVDV before inactivation, and if detected titrate virus. Test for BVDV again after inactivation when no infectious virus should be detected.

- Validated test for anti-BVDV antibodies should be conducted
- Serum should be free of BVDV antibodies or at a level that does not interfere with the detection of BVDV
- Comparative titration tests
 - prepare cell cultures in media with test serum
 - subculture at least 3 times using media supplemented with test serum
 - use cells in a titration of a reference control BVDV stock
- High antibody titres can be detected



The European Agency for the Evaluation of Medicinal Products
Evaluation of Medicines for Human Use

London, 18 June 2003
 CPMP/BWP/1793/02

**COMMITTEE FOR PROPRIETARY MEDICINAL PRODUCTS
 (CPMP)**

**NOTE FOR GUIDANCE ON THE USE OF BOVINE SERUM IN THE
 MANUFACTURE OF HUMAN BIOLOGICAL MEDICINAL PRODUCTS**

DISCUSSION IN THE BIOTECHNOLOGY WORKING PARTY	September 2001 – March 2002
TRANSMISSION TO CPMP	March 2002
RELEASE FOR CONSULTATION	April 2002
DEADLINE FOR COMMENTS	October 2002
DISCUSSION IN THE BIOTECHNOLOGY WORKING PARTY	November 2002- June 2003
TRANSMISSION TO CPMP	June 2003
ADOPTION BY CPMP	July 2003
DATE FOR COMING INTO OPERATION	1 October 2003

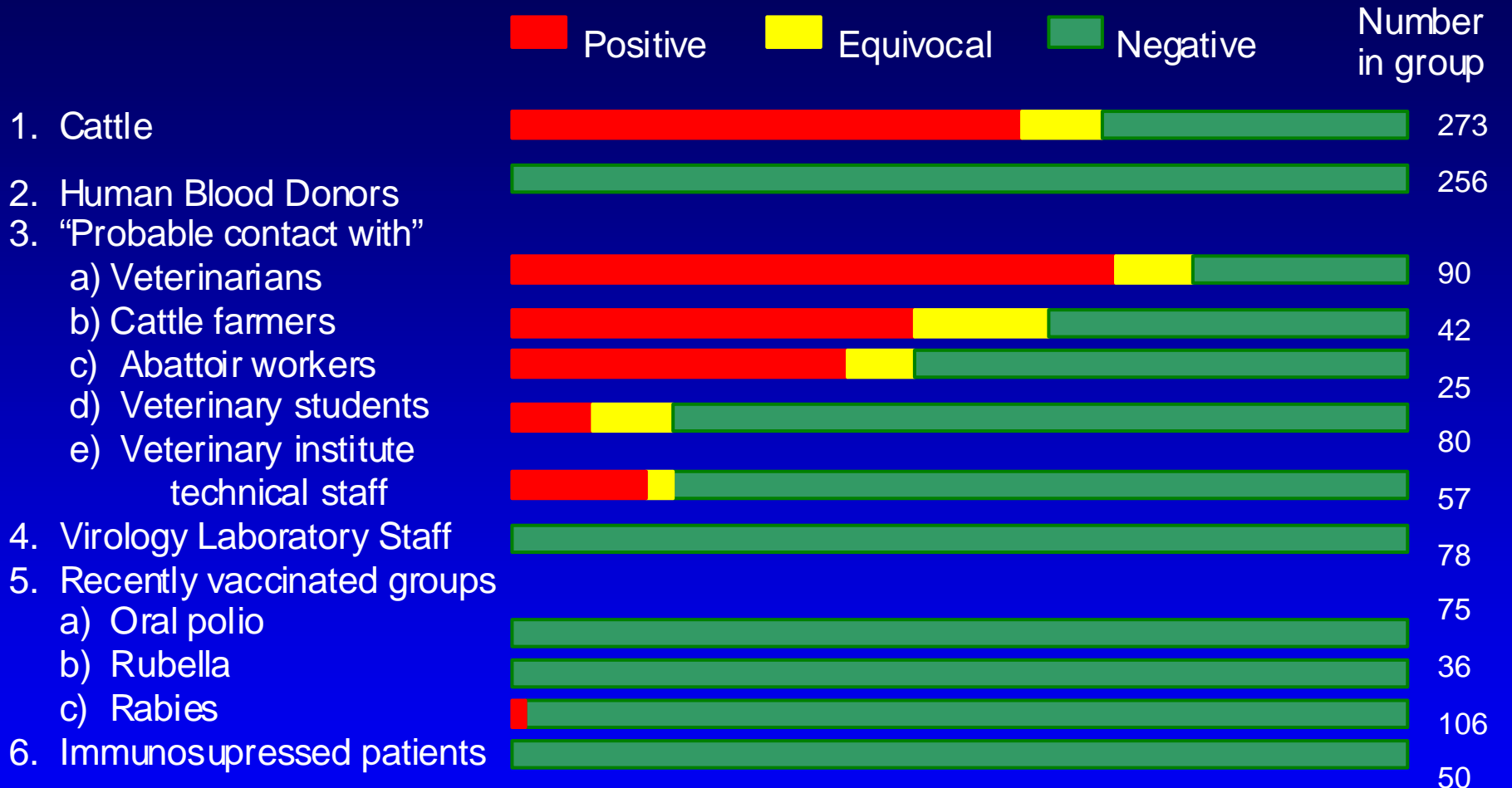
“Methods based on the detection of viral DNA suggest that Bovine Polyoma virus is a common contaminant of bovine serum.

Serum manufacturers and users are encouraged to apply infectivity assays for BPyV and to investigate methods for inactivation/removal of BPyV in order to limit or eliminate infectious virus from batches of serum”

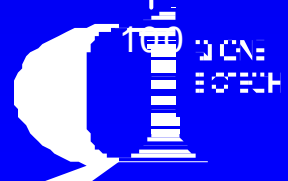
- Member of small DNA tumour virus family
 - ability to transform established cell lines
 - immortalise primary cell cultures
 - induce tumours in animals
- BPyV - small circular DNA genome of 4697bp (SV-40, 5243bp) (Schuurman *et al.*, 1990)
- Constructs containing early region of BPyV under control of the RSV promoter capable of inducing tumour formation in rodent cells following transfection (Schuurman *et al.*, 1992)

- Virus isolated from kidney cell cultures of stump-tailed macaque (Wognum *et al.*, 1984) and from foetal rhesus monkey kidney cell lines (Parry *et al.*, 1983)
- No viral antibodies present in monkey hosts
- High proportion of cattle shown to be positive for antibody to the virus
- Individuals who had been in close contact with cattle possessed antibodies to the virus -zoonotic potential of virus? (Parry & Gardner, 1986)

Antibody to BPyV in Humans



0 20 40 60 80
percentage of total specimens in each category
Parry & Gardner (1986)

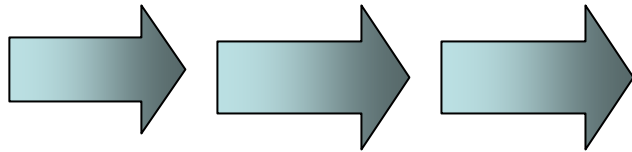


- Evidence suggested that virus was of bovine origin and that primate cultures had become infected from use of contaminated bovine serum in culture medium
- Studies have shown that approximately 70% of bovine serum batches are positive for BPyV by PCR (Schuurman *et al.*, 1991, Kappeler *et al.*, 1996)
- BioReliance in-house testing found 90% of serum samples to be positive by conventional PCR
 - 71 batches, 7 negative
 - USA, Australia, New Zealand, South America, Europe

- MDBK cells used as detector cells
- No visible cytopathic effect. Polyoma viruses are not released by cell lysis
 - Q-PCR endpoint required
- Cells inoculated with test article for 2-3 hours. Cells maintained in low serum concentrations with passage at least every 2 weeks
- Culture period 8 weeks. Test cell supernatants by Q-PCR at weeks 1 and 8

Cell Line	Copies Passage 1	Copies Passage 5
CHO-K1	10^7	0
Vero	10^7	0
BHK	10^7	0
FRHk	10^7	0
Primary Calf Kidney	10^7	10^5
MRC-5	10^7	0

Passage at 3-day intervals



No detectable virus producing cells
($<10^2$)

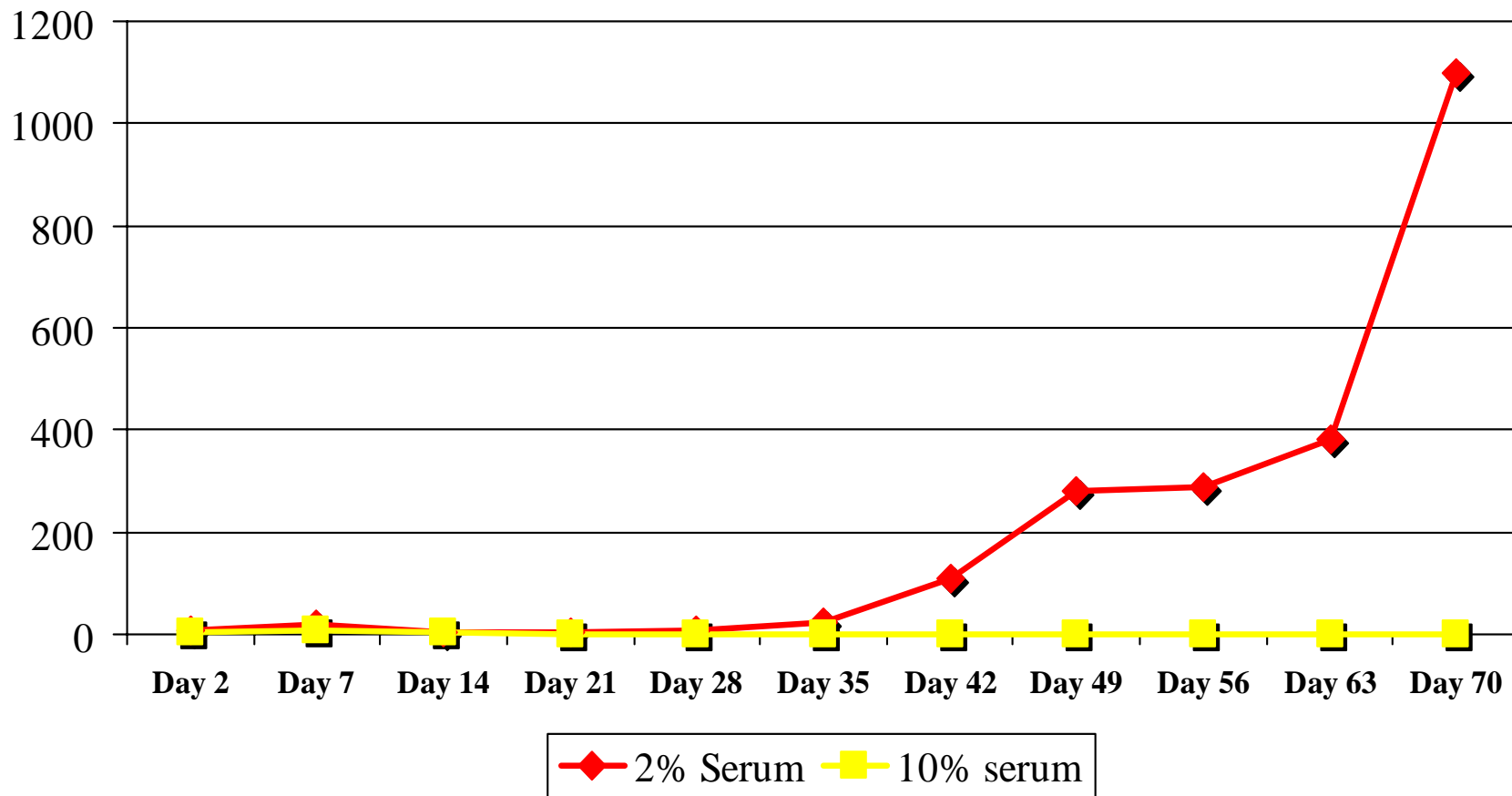
Passage at 2 week intervals

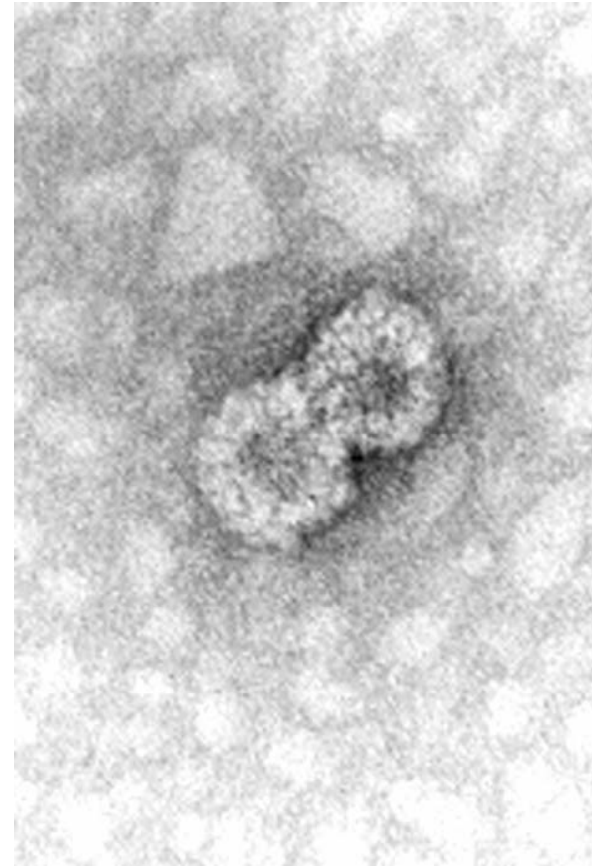
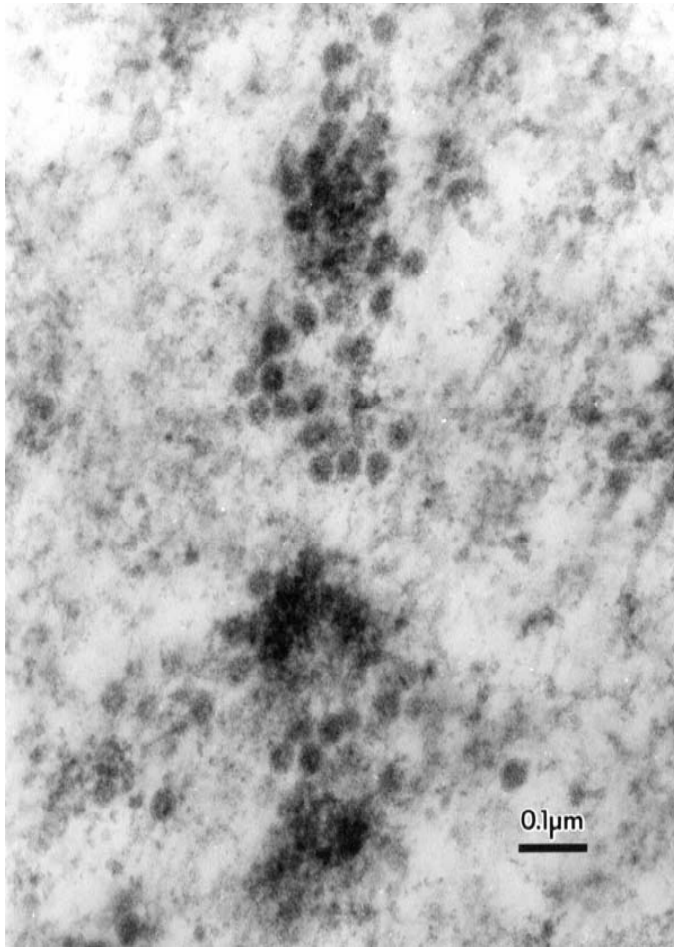


15% of cells virus producing

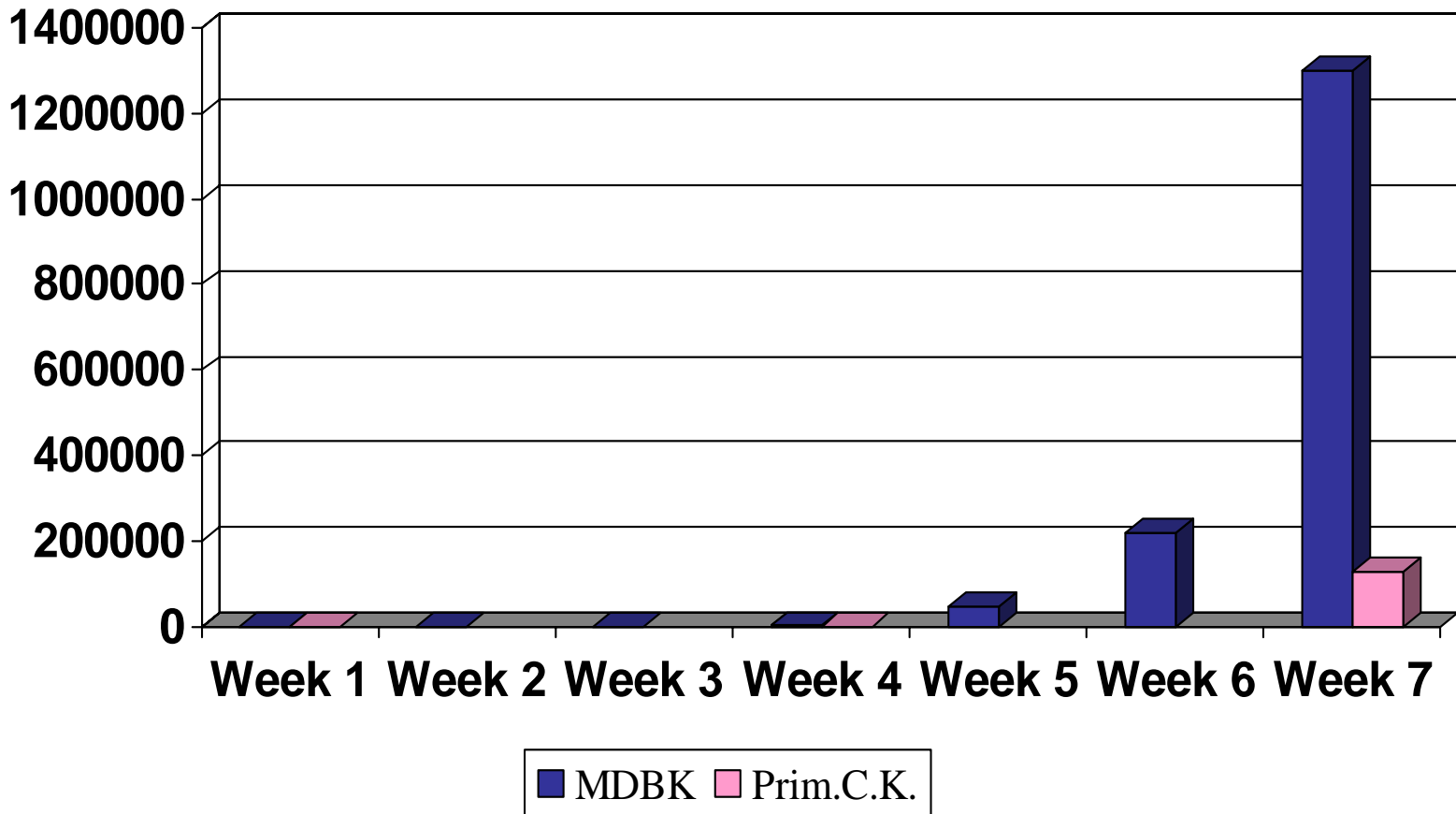
3 months of passage at 2 week intervals ~ 30% cells virus
producing 2×10^4 genomes per cell

Growth Dynamics of Bovine Polyoma virus

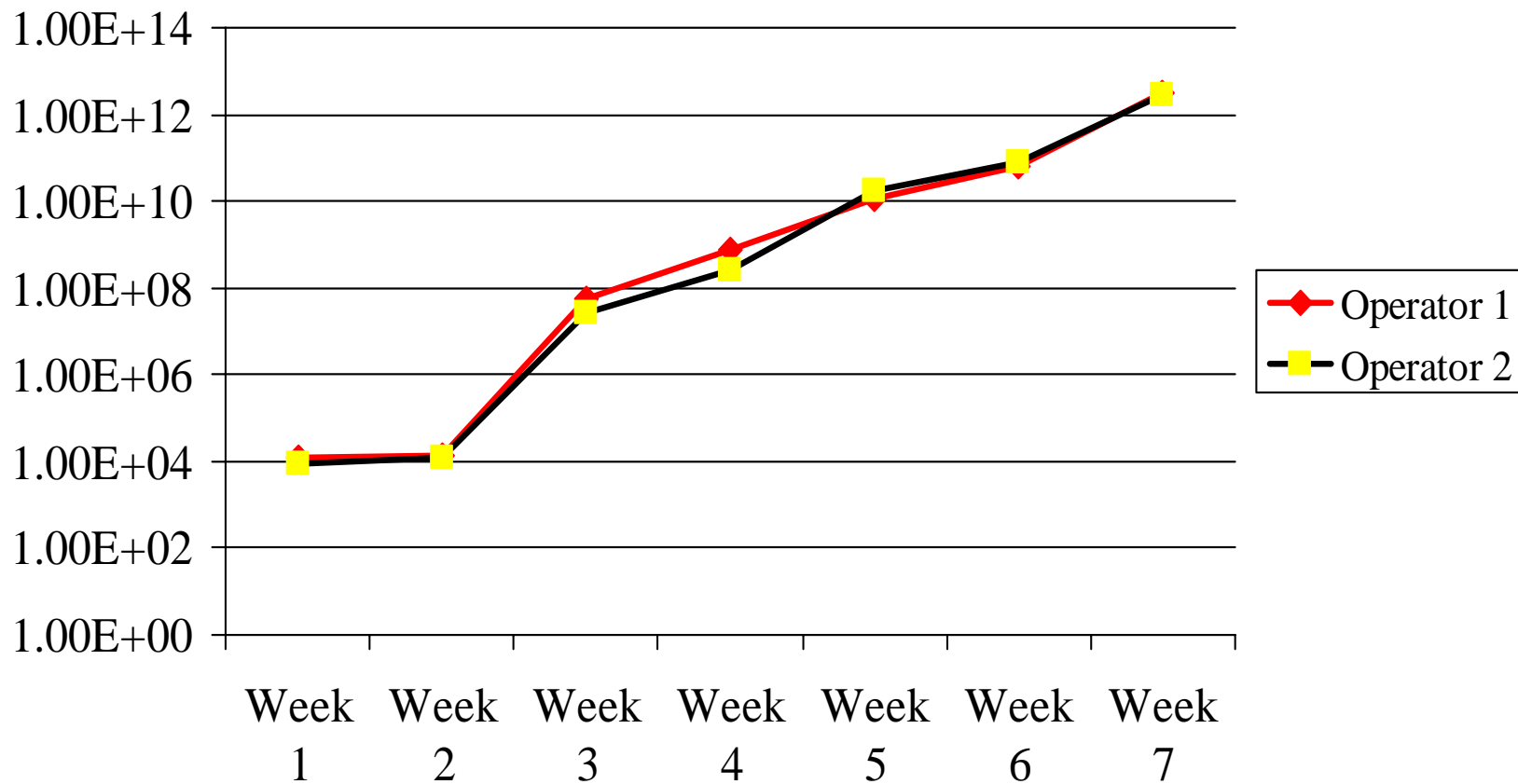




Photographs by Euan Milne



- Sensitivity and Reproducibility
 - Titration of virus stock from 10^7 to 10^{-1} carried out in duplicate on MDBK cells by 2 operators- cultured for 7 weeks
- Specificity
 - Virus (10^7) inoculated onto MRC-5, CHO-K1, 293 and MA104 cells- cultured for 7 weeks
 - Inactivated virus inoculated on MDBK
- Qualification of Test Material
 - Mock-test performed by inoculating bovine serum spiked with BPyV onto calf kidney cells to determine if virus would be neutralised by serum



- BPyV **not** detected by conventional PCR in:
 - Human: HEK 293, HeLa, MRC-5, A549, Hs27, RD, MCF
 - Monkey: CV-1, LLC-MK2, Vero
- BPyV **not** detected by Q-PCR in:
 - Human: HEK 293, HeLa, MRC-5
 - Monkey: Vero, Cos-7
 - Bovine: BT, MDCK
 - Rodent: CHO, BHK-21, NS0,
 - Avian: QT35
- BPyV **not** detected by infectivity assay in:
 - MRC-5, HEK 293
 - CHO

- 28 batches of FBS tested by both Q-PCR and infectivity assays for BPyV
- All batches of FBS tested by Q-PCR were positive
- Range 1.0×10^3 to 2.6×10^4 genome equivalents/ml
- No BPyV infectivity detected in any batch of FBS tested.

BVDV

- What is the risk of using high antibody titre bovine serum if the raw material is treated by a validated viral inactivation method?

BPyV

- Is there a significant risk from infectious BPyV in bovine sera?

- Daniel Galbraith
- Ian Forgie
- Archie Lovatt
- Karen McDonald
- Carron Nairn