NOSEMA (MICROSPORIDA: NOSEMATIDAE) SPECIES AS POTENTIAL BIOLOGICAL CONTROL AGENTS OF CACTOBLASTIS CACTORUM (LEPIDOPTERA: PYRALIDAE): SURVEYS FOR THE MICROSPORIDIA IN ARGENTINA AND SOUTH AFRICA

ROBERT W. PEMBERTON¹ AND HUGO A. CORDO²
¹USDA-ARS Invasive Plant Research Laboratory, Ft. Lauderdale, FL

²USDA-ARS South American Biological Control Laboratory, Hurlingham, Buenos Aires Province, Argentina

Abstract

Cactoblastis cactorum Berg is an invasive moth in North America where it damages and threatens many native Opuntia cacti. Nosema species of C. cactorum may have potential as biological control agents of the moth. Surveys for Nosema species were made in South Africa, where two of these Microsporidia were described from the moth and in Argentina where these pathogens may have originated. No Nosema were found in the C. cactorum larvae from South Africa and low levels of infection (0-6%) were found in the South American larvae. The low abundance of C. cactorum and the time of collection (austral summer) may be the reasons for the absence of or rarity of Nosema in these surveys. Winter collections of the larvae are suggested to obtain more abundant Nosema for evaluation as potential biocontrols of C. cactorum.

Key Words: biological control, surveys, insect pathogens, Opuntia

RESUMEN

Cactoblastis cactorum Berg es una polilla invasora en Norte America, donde daña y amenaza muchas especies nativas de Opuntia. Especies de Nosema que atacan a C. cactorum podrían tener potencial como agentes de control biológico. Se realizaron búsquedas de Nosema en Sudafrica donde dos de estos microsporidiosfueron descriptos y en Argentina donde estos patógenos podrían haberse originado. No se hallaron Nosema en larvas de C. cactorum de Sudafrica y se hallaron niveles muy bajos de infección (0-6%) en las larvas de Sud America. La baja abundancia de C. cactorum y el momento de las colecciones (verano austral) podrían ser las razones de la ausencia o rareza de Nosema en las búsquedas. Se sugieren colecciones de larvas durante el invierno para obtener mayor abundancia de Nosema para su evaluación como un agente potencial de control biológico de C. cactorum.

Cactoblastis cactorum Berg, is a famous biological control agent of weedy prickly pear cacti (Opuntia) in many parts of the world (Moran & Zimmermann 1984). This Argentine moth was introduced to Nevis in the Caribbean in 1957 for the control of native weedy Opuntia (Simmons & Bennett 1966). In 1989, C. cactorum was found in Florida where it attacks native Opuntia (Pemberton 1995; Johnson & Stiling 1996). The moth has the potential to spread to the western United States and Mexico where it could harm numerous native and economic Opuntia (Strong & Pemberton 2000; Zimmermann et al. 2000).

The potential and risk of using biological control as a possible solution to the *C. cactorum* threat was raised by Bennett and Habeck (1992) and is discussed in detail by Pemberton and Cordo (2001, this volume). Recorded parasitoids of the moth (Mann 1969; Zimmermmann et al. 1979) may have potential to control it, but they probably lack the host specificity needed to avoid use of native cactus moths and other Lepidoptera. Some insect patho-

gens such as *Nosema* species may be potent biological control agents of insects and also have enough host specificity to prevent non-target effects to native insects (R. Soper, pers. comm.). In many areas of the United States, *Nosema pyrausta* (Paillot) is the most important control agent of the European corn borer, *Ostrina nubialis* (Hübner) (Kogan et al. 1999). At times, *N. pyrausta* infects and kills 100% of the corn borers.

Two Nosema species have been recorded to attack Cactoblastis, both in South Africa (Fantham 1939). Nosema cactoblastis Fantham was described from C. cactorum which was originally imported for biological control from Australia. The second species, N. cactorum Fantham, was described from another Cactoblastis species, of uncertain identity, that was imported directly from Argentina. In South Africa, N. cactoblastis caused high mortality of C. cactorum larvae and pupae, and seriously hindered the ability of the moth to control weedy Opuntia (Petty 1948). Given this impact and the possibility that these Nosema

might have narrow host ranges, we decided to try to acquire them so their potential as biological control agents of the *C. cactorum* in the United States could be examined. They were sought in both South Africa and in Argentina where they may have originated.

MATERIALS AND METHODS

In South Africa, searches were made of *Opun*tia ficus-indica (L.) Miller plants growing at 15 sites in the eastern Cape during March 1996 by the senior author and P. Hulley of Grahamstown University. Most of the searches were on wild plants growing in the Uitenhage area, where high levels of N. cactoblastis infection of C. cactorum had been reported (Petty 1948). Additional searches were made at Fort Hare in two plantations of O. ficus-indica cultivated for fruit, and two wild populations. Plants varied greatly in size (from approximately 1-4 meters tall) and density (from scattered individual plants to dense thickets of intertwined plants, to rows of spaced plants in the plantations). At each site pads with characteristic damage (exit holes, frass, hollowed out, and/or with decay) and egg sticks were looked for, then cut open for verification of living larvae. A total of 429 living larvae, from 41 pads collected at nine sites, and six frass samples were shipped to Argentina for Nosema detection. Argentine importation permits enable the larvae to be shipped to the Instituto Nacional de Tecnología Agropecuaria quarantine facility in Buenos Aires. After confirmation of the identification, the larvae were transferred in secured containers to Carlos Lange, contracting pathologist at University of La Plata, for *Nosema* detection. Each living larva was squashed to create a smear on a glass slide. These smears were then examined with phasecontrast microscope to search for the characteristic spores of *Nosema*.

In Argentina, surveys were made in the northern half of the country, which is the largest part of

the native range of *C. cactorum*. In March 1995, we surveyed Opuntia species in the Argentine provinces of Entre Rios, Corrientes, Chaco, Formosa, and Santa Fe, and also along the eastern side of the Uruguay River in Uruguay. Additional surveys were made by the second author in Chaco and Formosa in Argentina and Paraguay during November-December 1995, and also in Buenos Aires, Cordoba, Santiago del Estero, Tucuman and Salta during December 1996. A total of 54 sites were surveyed in Argentina, three in Uruguay and one in Paraguay. A total of 867 plants were examined, 575 belonging to five identified Opuntia species and another 66 from Opuntia species that were not identified (Table 1). Cactoblastis cactorum was found in wild-growing O. aurantiaca Lindley, O. paraguayensis Schumann and O. ficus-indica at 14 sites. No larvae were found in either O. monacantha Haworth, O. quimilo Schumann or from unidentified Opuntia. A total of 528 larvae were collected for Nosema detection. *Opuntia* pads containing *C. cactorum* larvae were either transferred to La Plata for Nosema detection or reared in the laboratory to late instars to enhance detection and then transferred to La Plata for Nosema detection.

RESULTS AND DISCUSSION

Nosema was not detected in the 342 South African larvae or the six frass samples examined. Low levels of Nosema were detected in only 4 of the 528 larvae collected in Argentina. Nosema spores were found in 2 of the 34 (5.88%) larvae collected from O. paraguayensis at Tres Isletas in Chaco Province in March 1995. The other Nosema infections were in larvae from O. ficus-indica in Cordoba Province sites collected in December 1996. At Cruz del Eje, 1 of 68 (1.28%) were infected and at Dean Funes, 1 of 26 (3.85%) were infected. The low levels of Nosema infection found is consistent with the apparent health of collected larvae from both South Africa and Argentina. To-

TABLE 1. CACTOBLASTIS CACTORUM COLLECTION IN ARGENTINA AND DETECTED NOSEMA.

| Opuntia species | No. sites with the species ¹ | No. plants/ species | C. cactorum larvae | | | |
|------------------|---|------------------------|--------------------|---------------------------|--------------------------|--------------------|
| | | | Total | Mean no. plant/species | Mean no. species/site | No. with Nosema |
| O. aurantiaca | 15 | 124 | 135 | 1.08 | 9 | 0 |
| O. ficus-indica | 7 | 126 | 154 | 1.22 | 22 | 2 |
| O. monacantha | 2 | 11 | 0 | 0 | 0 | 0 |
| O. paraguayensis | 36 | 482 | 239 | 0.49 | 6.64 | 2 |
| O. quimilo | 7 | 58 | 0 | 0 | 0 | 0 |
| O. spp. | 11 | 66 | 0 | 0 | 0 | 0 |
| Total | 58 | 867 | 528 | | | 4 |

¹Twenty sites had more than one Opuntia species.

tal mortality was less than 5% (25 of 600) in the Argentine larvae and less than 25% (87 of 429) in the South African larvae. This mortality probably resulted from handling and shipping conditions. Due to the low number of larvae infected, no effort was made to culture the spores to characterize the *Nosema* species involved.

The low incidence of Nosema in South America material may relate to the low numbers of *C. cac*torum encountered. Forty-four of the 58 Argentine sites examined (75.9%) had no detectable populations of the moth. The mean number of C. cactorum at these 58 sites was 9.1 larvae. At the 14 sites, where the moth was present, the mean number of larvae was 37.7 per site. Although only three of five (or more) *Opuntia* spp. plants examined had C. cactorum larvae, all are known to be host of the moth (Dodd 1940). Opuntia ficus-indica and O. aurantiaca had an average of one larva per plant (1.09 and 1.22 respectively) (Table 1). The most common prickly pear encountered, O. paraguayensis, had less than one (0.49) larva per plant. Opuntia ficus-indica had more larvae per site, a mean of 22 compared to about 7 for O. paraguayensis and 9 for O. aurantiaca. This may be due to a greater number of *O. ficus-indica* per site (18 compared to 13.4 and 8 for the others).

The absence of *Nosema* samples from South Africa may also relate to the relatively low levels of *C. cactorum* in the areas surveyed. The moth was more common than in South America occurring at 60% (9/15) of the sites compared to 24% (14/58) in South America. However, at five of the South Africa sites, only a single infested pad was found. The other 4 sites infested had 6, 6, 9 and 15 infested pads despite the large numbers of plants at some sites.

The absence of Nosema in South Africa and the low levels of the disease found in South America probably also relate to the season of collections summer. In South Africa, Nosema cactoblastis infections of *C. cactorum* are known to be much less abundant in summer than in the winter (Petty 1948). For instance, at one South African site, N. cactoblastis was detected in 100% of the sampled larvae of the winter brood but in none of the larvae of the following summer brood. The infection rates of *Nosema* species we detected in Argentina (0.0, 1.28, 3.85, and 5.88%) are comparable to the infection rates (an average of 2%) at six coastal sites in South Africa during the summer (Petty 1948). These same sites had an average infection rate of 55.6% during the winter.

The absence of a formal biological control program for *C. cactorum* ended the research. Our efforts were opportunistic and done in conjunction with other research. We recognize that our collection times were not optimal for the detection of *Nosema*. The recent spread of the moth from Florida to Georgia and the increased concern that it will continue to spread and damage valued native

Opuntia in North America, suggests that acquisition and examination of Nosema species associated with the moth should be continued. The Nosema spores we found are frozen and may be able to be cultured and evaluated. Renewed efforts to obtain Nosema species from field populations of C. cactorum should be more successful if winter collections are made in South Africa and Argentina.

The South African experience with *N. cactoblastis* suggests that the disease (its occurrence in the more humid winter months and its greater impact in more humid coastal regions) would be more effective in humid areas such as Florida and in the Southeast, than in the drier areas of the West. The effectiveness of the *Nosema* in low populations of *C. cactorum* is expected to be less than in high populations of the moth.

ACKNOWLEDGMENTS

We thank Patrick Hulley, Rhodes University, Grahamstown, South Africa for helping locate and collect *C. cactorum* in South Africa, and Carlos Lange, University of La Plata, Argentina for the examination of larval samples for *Nosema*. Juan Briano, South American Biological Control Laboratory, USDA ARS, Hurlingham, Argentina, critically reviewed and improved the manuscript.

REFERENCES CITED

Bennett, F. D., and D. H. Habeck. 1992. *Cactoblastis cactorum*: a successful weed control agent in the Caribbean, now a pest in Florida?, pp. 21-26. *In* E. S. Delfosse & R. R. Scott [eds.]. Proc. 8th International Symposium on Biological Control of Weeds, Cantebury, New Zealand, CSIRO, Melbourne, Australia.

FANTHAM, B. 1939. Nosema cactoblastis, sp. n. and Nosema cactorum, sp. n. Microsporidian parasites of species of Cactoblastis destructive to prickly pear. Proc. Zool. Soc. London (B) 108: 688-705.

JOHNSON, D. M., AND P. D. STILING. 1996. Host specificity of *Cactoblastis cactorum* (Lepidoptera: Pyralidae), an exotic *Opuntia*-feeding moth, in Florida. Environ. Entomol. 25: 743-748.

KOGAN, M., D. GERLING, AND J. V. MADDOX. 1999. Enhancing biological control in annual agricultural environments, pp. 789-818. *In T. S. Bellows*, and T. W. Fisher [eds.]. Handbook of Biological Control. Academic Press, San Diego, CA.

MANN, J. 1969. Cactus feeding insects and mites. U.S. National Museum Bull. 256. 158 pp.

MORAN, V. C., AND H. G. ZIMMERMANN. 1984. The biological control of cactus weeds: achievements and prospects. Biocontrol News and Information 5: 297-320.

PEMBERTON, R. W. 1995. Cactoblastis cactorum (Lepidoptera: Pyralidae) in the United States: an immigrant biological control agent or an introduction of the nursery industry? Amer. Entomol. 41: 230-232.

PETTY, F. W. 1948. The biological control of prickly pears in South Africa. Union of South Africa Depart. of Agriculture, Entomology Series N. 22.

SIMMONS, F. J., AND F. D. BENNETT. 1966. Biological control of *Opuntia* sp. by *Cactoblastis cactorum* in the Leeward Islands (West Indies). Entomophaga 11: 183-189.

- STRONG, D. R., AND R. W. PEMBERTON. 2000. Biological control of invading species—risk and reform. Science 288: 1969-1970.
- ZIMMERMANN, H. G., R. E. McFadyen, and H. E. Erb. 1979. Annotated list of some cactus-feeding insects of South America. Acta Zoologica Lilloana 32: 101-112.
- ZIMMERMANN, H. G., V. C. MORAN, AND J. H. HOFFMANN. 2000. The renowned cactus moth, *Cactoblastis cactorum*: its natural history and threat to native *Opuntia* in Mexico and the United States of America. Diversity and Distributions 6: 259-269.