

**Numerical Simulation of the Anaerobic Transformation of Tetrachloroethene (PCE) to Ethene in a Continuous Flow Aquifer Column**

#### **ABSTRACT**

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Numerical modeling was conducted to simulate reactive transport with sequential anaerobic transformation processes	\n $\frac{\partial C_{rec}}{\partial t} = D \frac{\partial^2 C_{rec}}{\partial x^2} - v \frac{\partial C_{rec}}{\partial x} + \frac{K_{un,rec}x + C_{rec}}{K_{s,rec} + C_{rec}} - \frac{K_{un,rec}x + C_{rec}}{K_{u,rec}} + C_{rec}$ \n	\n $K_{u,rec} = \frac{1}{2} \left(1 + \frac{C_{rec}}{K_{u,rec}}\right) + C_{rec}\left(1 + \frac{C_{rec}}{K_{un,rec}}\right)$ \n	\n $0.14$ \n		
continuous-flow, and injection of PCE in synthetic groundwater. The column was packed with aquifer solids from the Hamiltonian. In the first continuous	\n $\frac{\partial S_{rec}}{\partial t} = K_{rec}(K_{a,rec} + C_{rec} - S_{rec})$ \n	\n $\frac{\partial S_{rec}}{\partial t} = K_{rec}(K_{a,rec} + C_{rec} - S_{rec})$ \n	\n $\frac{\partial C_{un,Dec}}{\partial t} = \frac{\partial^2 C_{un,Dec}}{\partial x} + \frac{K_{un,rec}x + C_{rec}}{K_{un,rec}} + C_{rec}$ \n	\n $\frac{\partial C_{un,Dec}}{\partial t} = K_{rec}(K_{a,rec} + C_{rec} - S_{rec})$ \n	\n $\frac{\partial C_{un,Dec}}{\partial t} = \frac{\partial^2 C_{un,Dec}}{\partial x} + \frac{K_{un,rec}x + C_{rec}}{K_{un,rec}} + \frac{K_{un,rec}x + C_{rec}}$

in continuous-flow columns and compared with experimental observations. The column studies were conducted with continuous flow and injection of PCE in synthetic groundwater. The column was packed with aquifer solids from the Hanford DOE site and bioaugmented with the Evanite (EV) dechlorinating enrichment culture. In the first continuous flow column experiment, enhanced desorption resulting from anaerobic reductive dechlorination of PCE was simulated and compared with experimental observations (Figure 1). The experiments showed that cis-dichlorethene (c-DCE) concentrations in the column effluent exceeded the influent PCE concentration. In the second continuousflow column experiment pre-reduced soil was used and <sup>a</sup> rapid reduction of PCE to ethene was observed. Simulating the second column using two microbial communities captured the observed CAHs breakthrough curves. Comparing model predictions with dehalogenation observations, reasonable agreement for CAHs transformation processes were found when using kinetics, inhibition constants and sorption parameters which were determined in batch laboratory experiments.



Figure 1: Simulation and experimental results of CAHs effluent concentration in continuous-

C: aqueous concentration, S: sorbed concentration, D: dispersion coefficient , V: velocity,  $K_{\text{max}}$ : the maximum specific utilization rate, X: the total biomass,  $K_s$ : the half-saturation coefficient, ρ : soil bulk density, θ: porosity, Vs: soil volume, VL : liquid volume ,  $\mathsf{K}_\mathsf{Cl}$  :  $\,$  Competitive inhibition constant.

## **OBJECTIVES**

Simulate reactive transport in continuous-flow column based on experimentally determined kinetics, inhibition constants and sorption parameters.

> An advection and dispersion transport was coupled with rate-limited desorption and reductive dechlorination (including competitive inhibition and microbial growth and decay) to describe the fate and transport processes occurring in continuous flow system fed PCE. The kinetics for the EV culture (Table-2) used in the model simulations were those determined by Yu & Semprini (2004). Sorption parameters (K $_{\text{\tiny d}}$  and K ) were determined by least square fit of the model equations (Eqn1 & 2) to batch experimental results (Table-1). Competitive inhibition constants  $(K<sub>CI</sub>)$ were set equal to their respective half-velocity coefficients  $(K_s)$  (Yu, 2004). Initial biomass distribution was assumed to be in the first 5 cm of the influent part of the columns.

the solid: liquid distribution coefficient  $(K_d)$  of PCE and its transformation products, and the first order mass transfer coefficient (K) between the aqueous and solid phase. All batch experiments were carried out in triplicate in 156 mL glass reactors. The reactors were filled with an approximately 1:1 solid liquid (w/w) ratio. Stock solutions containing the CAHs were spiked into each reactor. The following equations were used to model time dependent batch sorption experiments:

Model rate-limited enhanced desorption and biological transformation processes during one dimensional solute transport.



	soil water partitioning coefficient	mass transfer coefficient
	$K_{d}$ (L/K <sub>a</sub> )	(1/day) ĸ.
<b>PCE</b>	1.9	0.068
<b>TCE</b>	0.76	0.032
<b>CDCE</b>	0.23	0.021
<b>VC</b>	0.099	0.009
Eth	0.031	0.005

**Table 1:** Sorption Kinetic parameter values determined for Hanford soil and CAHs using batch experiments.

Develop <sup>a</sup> model for complete PCE reductive dechlorination to ethene in continuous flow column that incorporates inhibition between chlorinated ethenes and rate-limited sorption desorption.



**Future work will continue to evaluate electron donor limitations andcompetition for electron donor in continuous-flow column and incorporating** two microbial population in the modeling analysi

Where:

Develop <sup>a</sup> model with two microbial dehalogenating systems and simulate the biotransformation based on the spatial distribution and activity of the biomass**PCE\*** biomass. **<sup>12</sup> <sup>4</sup>**

### **METHOD and APPROACH**

The resulting sets of simultaneous equations were numerically solved by the finite

-M. Azizian, S. Behrens, A. Sabalowsky, P. Ruiz-Haas, J. D. Ingle, Jr., M. Dolan, A. Spormann, and L. Semprini Continuous-Flow Bioreactor Study of Reductive Dehalogenation of PCE Upon Bioaugmentation ith Evanite Enrichment Culture.. (submitted for publication)



Figure 2: Simulation and experimental results of CAHs effluent concentration after 6 cm in continuous flow column, using pre- reduced aquifer material ( Column experiment 2) .

### **MODEL DEVELOPMENT**

**Table 2:** Kinetic parameter values determined for the Evanite cultures. \* Yu and Semprini, (2004), \$ used for 2<sup>nd</sup> dehalogenating population.

### **DISCUSSION**

 **The simulations indicated that enhanced desorption responsible for the increase in effluent cDCE concentration above the influent PCE.**

# **CONCLUSIONS**

 **The simulations indicated that two dehalogenating communities are likely responsible for the transformation of PCE to ethene.**

Model simulations show higher c-DCE concentrations in the effluent compared to influent PCE concentration, consistent with the experimental results indicating enhanced desorption of PCE from the aquifer material.

> **Simulation of continuous-flow column using batch laboratory generated** microbial and sorption kinetics captured the overall response observed in **the column experiments.**

$$
\frac{\partial C_{PCE}}{\partial t} = \frac{-Vs}{VL} * \frac{\rho}{\theta} * K * (K_d * C - S) \tag{1}
$$

$$
\frac{\partial S_{PCE}}{\partial t} = K * (K_d * C - S) \tag{2}
$$

 Simulating columns 1 and 2 with biomass distributed spatially in the column influent where they were bioaugmented gives good agreement with the experimental results. The results indicated that most of thecolumn.

> **Insufficient mixing at the column influent likely resulted in deviations from 1-D transport and the tailing observed in Figure 2.**

 Successful simulation of the second column using two dehalogenating communities supports the hypothesis that at least two populations are responsible (Figure 2).

**Column Experiment 1:** Continuous-flow anaerobic **p** column experiment was performed using Hanford soil bioaugmented with EV culture, the column was fed lactate as <sup>a</sup> fermenting electron donor. The column is 30 cm long and 7 cm diameter.

**delingate 1 Column Experiment 2:** Using the same aquifer material *Experiment* 2: Using the same aquifer material and culture, the effect of the pre- reduction of aquifer material was evaluated. The aquifer material was pre-treated with 5 mM Na<sub>2</sub>S solution to reduce Fe  $^{\text{\tiny{\textsf{(III)}}}}$ to Fe  $^{\text{\tiny{\textsf{(II)}}}}$ 

Equations 3-9 were used for the simulation shown in Figure-1. The same equations were used for the simulation shown in Figure-2 by adding cDCE transformation to Eqn-7 and with similar additional equations for VC

Biodegradation rates of CAHs during flow in porous media often depends on the mixing between nutrients and CAHs . Experimental results from port one (Figure-2) shows tailing in breakthrough curves which may be due to insufficient mixing between nutrients and CAHs that are flowing through the aquifer material.

### **REFERENCES**



reduction of aquifer material all was pre-treated with 5 mM	$\frac{\partial C_{PCE}}{\partial t} = D \frac{\partial^2 C_{PCE}}{\partial x^2} - v \frac{\partial C_{PCE}}{\partial x} - \frac{K_{max, PCE} X * C_{PCE}}{K_{s, PCE} + C_{PCE}}$ \n\n <th>ACKNOWLEDGEMENTS This study was supported through: Cheter, sponsorted through: Center, sponsorted by the U.S. Environmental Protection Agency.</th> \n	ACKNOWLEDGEMENTS This study was supported through: Cheter, sponsorted through: Center, sponsorted by the U.S. Environmental Protection Agency.
\n $e^{(II)}$ \n	Alternative-Geograf	Wth Evanite Enrichment Culture. (submitted for publication YU, S. and L. Semprini, Kinetics and Modeling of Redu Coneentrations, Biotechnology and Bioengineering, vol. 88, r\n

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