

## ACETALDEHYDE by HPLC

3507

CH<sub>3</sub>CHO

MW: 44.05

CAS: 75-07-0

RTECS: AB1925000

METHOD: 3507, Issue 2

EVALUATION: FULL

Issue 1: 15 August 1987

Issue 2: 15 August 1994

**OSHA :** 200 ppm  
**NIOSH:** carcinogen; lowest feasible  
**ACGIH:** C 25 ppm; suspect carcinogen  
 (1 ppm = 1.801 mg/m<sup>3</sup> @ NTP)

**PROPERTIES:** liquid; d 0.78 g/mL @ 20 °C;  
 BP 20.4 °C; MP -123 °C;  
 VP 100 kPa (750 mm Hg; 99% v/v)  
 @ 20 °C; explosive range 4 to 60% in air

**SYNONYMS:** ethanal; acetic aldehyde.

SAMPLING		MEASUREMENT	
<b>SAMPLER:</b>	LIQUID IN BUBBLER (midget bubbler containing 15 mL Girard T solution @ pH 4.5)	<b>TECHNIQUE:</b>	HPLC, UV
<b>FLOW RATE:</b>	0.1 to 0.5 L/min	<b>ANALYTE:</b>	Girard T derivative
<b>VOL-MIN:</b>	6 L @ 200 ppm	<b>SAMPLE PREPARATION:</b>	dilute 5 mL sample to 100 mL with HPLC mobile phase
<b>-MAX:</b>	60 L	<b>INJECTION VOLUME:</b>	50 µL
<b>SHIPMENT:</b>	seal bubblers to prevent leakage before shipping; protect from light	<b>COLUMN:</b>	50 cm x 2-mm ID SS, Zipax SCX
<b>SAMPLE STABILITY:</b>	1 week @ 25 °C in dark [1]	<b>DETECTOR:</b>	UV @ 245 nm for acetaldehyde
<b>FIELD BLANKS:</b>	2 to 10 field blanks per set	<b>MOBILE PHASE:</b>	HPO <sub>4</sub> <sup>2-</sup> /HPO <sub>4</sub> <sup>-</sup> buffer, 0.75 mL/min
<b>ACCURACY</b>		<b>CALIBRATION:</b>	standard solutions of acetaldehyde in Girard T reagent
<b>RANGE STUDIED:</b>	170 to 670 mg/m <sup>3</sup> [1] (60-L samples)	<b>RANGE:</b>	2 to 60 mg per sample [1]
<b>BIAS:</b>	1.2%	<b>ESTIMATED LOD:</b>	0.1 mg per sample [1]
<b>OVERALL PRECISION (<math>\hat{S}_{r,T}</math>):</b>	0.053 [1]	<b>PRECISION (<math>\hat{S}_r</math>):</b>	0.024 @ 11 to 43 mg per sample [1]
<b>ACCURACY:</b>	± 14.4%		

**APPLICABILITY:** The working range is 18 to 372 ppm (33 to 670 mg/m<sup>3</sup>) for a 60-L air sample. The method is sensitive enough for short-term exposure sampling and can be used to measure lower concentrations by diluting samples to less than the recommended 100 mL.

**INTERFERENCES:** Other volatile aldehydes and ketones (e.g., acetone, acrolein, benzaldehyde, formaldehyde, furfural, methyl ethyl ketone, and propionaldehyde) compete for the Girard T reagent which should be kept at a two-fold molar excess over aldehyde concentration. Chromatographic conditions may be adjusted to resolve acetaldehyde from other aldehydes [1].

**OTHER METHODS:** This revises S345 [2]. Method 2538 is an adaptation of OSHA Method 68, which uses solid sorbent collection and GC analysis. Other reported methods for acetaldehyde use collection in 2,4-dinitrophenylhydrazine solution [3,4].

**REAGENTS:**

1. Acetaldehyde.\*
2. Citric acid.
3. Disodium hydrogen phosphate ( $\text{Na}_2\text{HPO}_4$ ).
4. Girard T reagent [(carboxymethyl)-trimethylammonium chloride hydrazide] recrystallized from 95% ethanol.
5. Water, distilled, deionized (DD).
6. Ethanol, 95%.
7. Sodium dihydrogen phosphate monohydrate ( $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$ ).
8. Girard T solution: 5.39 g citric acid, 6.63 g  $\text{Na}_2\text{HPO}_4$ , and 16.77 g Girard T reagent diluted to 500 mL with DD water. Store in annealed flask in the dark. Use within two weeks.
9. HPLC mobile phase: 0.22 M  $\text{Na}_2\text{HPO}_4 \cdot \text{H}_2\text{O}$ , 0.019 M  $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$  in 20% ethanol. Dissolve and dilute 31.2 g  $\text{Na}_2\text{HPO}_4$  and 26.2 g  $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$  to 1 L with DD water. Filter through 5- $\mu\text{m}$  PTFE filter and degas prior to use. Bubble helium through the solution to prevent bacterial growth.
10. Calibration stock solution, 4.32 mg/mL acetaldehyde in 0.2 M Girard T solution. Weigh 216 mg freshly-distilled acetaldehyde into 50-mL volumetric flask containing 49 mL Girard T solution. Make to volume with Girard T solution. Use within one day.
11. Helium.

\* See SPECIAL PRECAUTIONS.

**SAMPLING:**

1. Calibrate each personal sampling pump with a representative sampler and trap in line.
2. Add exactly 15 mL Girard T solution to each bubbler using a 15-mL pipet. Mark the initial liquid level in the bubbler with a glass marker. Make impinger-to-trap and trap-to-sampling pump connections with flexible inert tubing.
3. Sample at an accurately known flow rate between 0.1 and 0.5 L/min for a total sample size of 6 to 60 L.

NOTE: Higher flow rates will cause frothing of the collection medium. If amount of liquid condensed in the trap is greater than 1 mL, collection efficiency of bubbler may be reduced and sample may be invalid.

**SAMPLE PREPARATION:**

4. Tap bubbler stem lightly against bubbler body to drain contents into the body. If necessary, bring samples up to the 15-mL mark with distilled water. Swirl bubbler to mix contents well. Do not add solution collected in the trap to the sample.
5. Transfer a 5-mL aliquot to a 100-mL flask and bring to volume with HPLC mobile phase.

**EQUIPMENT:**

1. Sampler: bubbler, glass, midget, with fritted glass stems, annealed,\* with PTFE stoppers for shipping.
2. Personal sampling pump, 0.1 to 0.5 L/min, with trap made from midget bubbler with stem broken off and inert, flexible connecting tubing.
3. High pressure liquid chromatograph, with 245-nm UV detector, integrator, and column (page 3507-1) with 50- $\mu\text{L}$  injection loop or autosampler.
4. Syringe, 2-mL, Luer-lock.
5. Distillation apparatus for preparation of high purity acetaldehyde.
6. Flasks, volumetric, 1-L; 10-, 50-, and 100-mL; and 500-mL, annealed.\*
7. Pipets, 0.02- to 1-mL; 5-, 10-, and 15-mL.
8. Marker, glass.
9. Cylinder, graduated, 250-mL.
10. Filter, 5- $\mu\text{m}$ , PTFE, 37-mm, with holder for liquid filtration.
11. Balance, readable to 0.1 mg.

\* Heat in an oxidizing atmosphere at 580 °C.

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**SPECIAL PRECAUTIONS:** Acetaldehyde is extremely volatile and a fire hazard. Cool containers of acetaldehyde to ice bath temperature to reduce pressure buildup and open in an exhaust hood only.

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**CALIBRATION AND QUALITY CONTROL:**

6. Calibrate daily with at least six working standards over the range 0.007 to 4 mg acetaldehyde per mL (0.1 to 60 mg acetaldehyde per sample).
  - a. Add known amounts of calibration stock solution to Girard T solution in 10-mL volumetric flasks and dilute to the mark. Dilute 5 mL of each of these solutions to 100 mL with HPLC mobile phase. Prepare at least two blanks in the same manner.
  - b. Analyze together with samples and blanks (steps 8 and 9).
  - c. Prepare calibration graph (peak area vs. mg acetaldehyde per sample).
7. Analyze three quality control blind spikes and three analyst spikes to ensure that the calibration graph is in control.

**MEASUREMENT:**

8. Set liquid chromatograph according to manufacturer's recommendations and to conditions given on page 2507-1. Inject 50- $\mu$ L sample aliquot with injection loop or autosampler.  
NOTE: If peak area is above the linear range of the working standards, dilute with HPLC mobile phase, reanalyze, and apply the appropriate dilution factor in calculations.
9. Measure peak area.

**CALCULATIONS:**

10. Determine the mass, mg of acetaldehyde found in the sample (W), and in the average media blank (B).
11. Calculate concentration, C, of acetaldehyde in the air volume sampled, V (L):

$$C = \frac{(W - B) \cdot 10^3}{V}, \text{ mg/m}^3.$$

**EVALUATION OF METHOD:**

Method S345 was issued on March 16, 1979 [2], and validated over the range 170 to 670 mg/m<sup>3</sup> at 21 °C and 756 mm Hg using a 60-L sample [1,5]. Overall precision,  $\hat{S}_{rT}$ , was 0.053 with an average recovery of 101.2% representing a non-significant bias. The concentration of acetaldehyde was independently verified by calibrated gas chromatograph. Collection efficiency of a single bubbler was determined to be >0.998 when 61-L air samples were taken at 0.5 L/min in atmospheres containing 670 mg/m<sup>3</sup> acetaldehyde.

**REFERENCES:**

- [1] Backup Data Report, S345, Acetaldehyde, prepared under NIOSH Contract 210-76-0123 (unpublished).
- [2] NIOSH Manual of Analytical Methods, 2nd ed., Vol. 5, S345, U.S. Department of Health, Education, and Welfare, Publ. (NIOSH) 79-141 (1979).
- [3] Kuwata, K., M. Uebori and Y. Yamasaki. *J. Chromatog. Sci.*, **17**, 264-268 (1979).
- [4] Lipari, F. and S.J. Swarin. *J. Chromatog.*, **247**, 297-306 (1982).

- [5] NIOSH Research Report-Development and Validation of Methods for Sampling and Analysis of Workplace Toxic Substances, U.S. Department of Health and Human Services, Publ. (NIOSH) 80-133 (1980).

**METHOD REVISED BY:**

Eugene R. Kennedy, Ph.D., NIOSH/DPSE; Method S345 was validated under NIOSH Contract 210-76-0123.