

## **Evaluation of a Solubility Protocol for In Vitro Cytotoxicity Testing**

J Strickland<sup>1,2</sup>, M Paris<sup>1,2</sup>, W Stokes<sup>1</sup>, S Casati<sup>3</sup>, R Tice<sup>1,3</sup>, H Raabe<sup>4</sup>, C Cao<sup>5</sup>, R Clothier<sup>6</sup>, J Harbell<sup>5</sup>, G Mun<sup>4</sup>, A Sizemore<sup>4</sup>, G Moyer<sup>4</sup>, J Madren-Whalley<sup>5</sup>, C Krishna<sup>5</sup>, M Owen<sup>6</sup>, N Bourne<sup>6</sup>, M Wenk<sup>7</sup>, and M Vallant<sup>8</sup>

<sup>1</sup>NICEATM, RTP, NC, <sup>2</sup>ILS, Inc, RTP, NC, <sup>3</sup>ECVAM, JRC, Ispra, Italy, <sup>4</sup>IIVS, Gaithersburg, MD, <sup>5</sup>US Army ECBC, APG, MD, <sup>6</sup>Univ. of Nottingham, UK, <sup>7</sup>BioReliance Corp, Rockville, MD, and <sup>8</sup>NIEHS, RTP

#### Introduction

Solubility is an important determinant of toxicity in an *in vitro* assay system since it affects the availability of potential toxins to the cells. Solubility in any particular solvent is a specific property of the involved chemical structures while solubility determination, like any measurement, can vary from lab to lab with certain precision and accuracy. To eliminate solubility and solvent selection as potential sources of variation in multi-laboratory validation studies, the management teams often recommend an appropriate solvent for each test article and may also specify the concentrations to be tested (Knox et al. 1986; Spielmann et al. 1998; Clothier et al. 1999).

During a validation study to evaluate two neutral red uptake (NRU) *in vitro* cytotoxicity assays for estimating acute *in vivo* systemic toxicity, we evaluated a solubility protocol (NICEATM 2003) designed to identify the solvent that would provide the highest soluble concentration of a test chemical for *in vitro* testing. To avoid the use of different solvents by the labs when testing the same chemical, the Study Management Team (SMT) assigned the solvents used for *in vitro* testing. The objectives of this part of the validation study were to:

- Evaluate the utility and appropriateness of the solubility protocol
- Evaluate the concordance among labs in the solvent selected for each of the 72 chemicals tested in the validation study

### Methods

The study design was for three labs to test 72 coded chemicals in two *in vitro* basal cytotoxicity assays (for more information, see Poster 1628). While the three *in vitro* test labs used the protocol presented here, a fourth lab purchased and coded the chemicals, performed solubility testing using a different protocol, and distributed aliquots to the *in vitro* test labs.

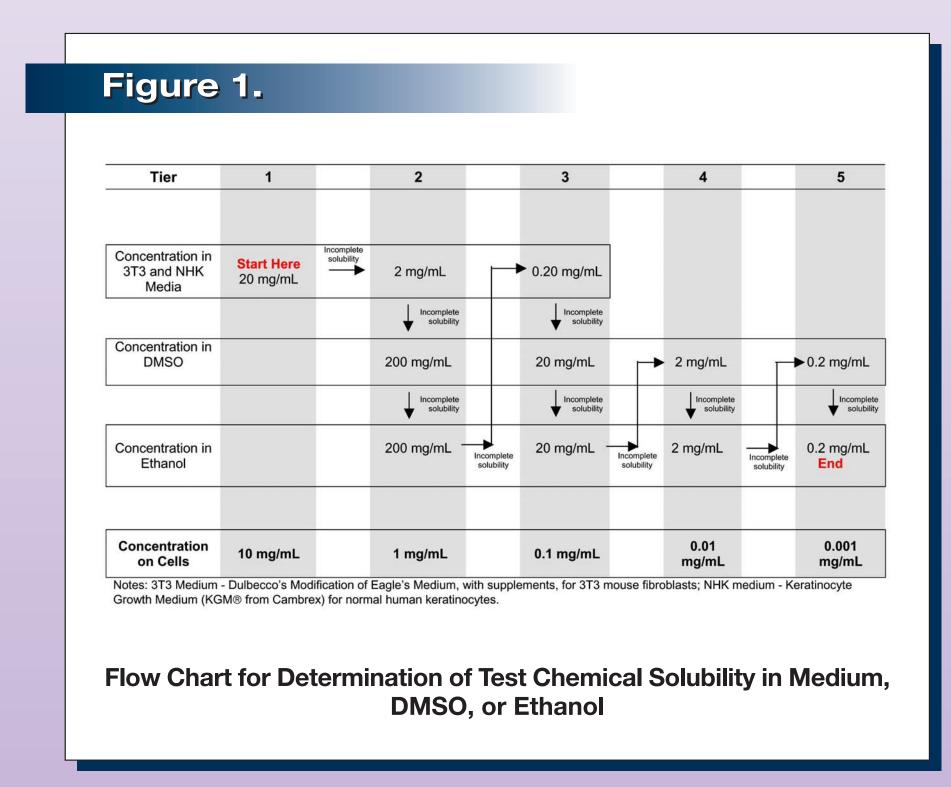
### Solubility Protocol

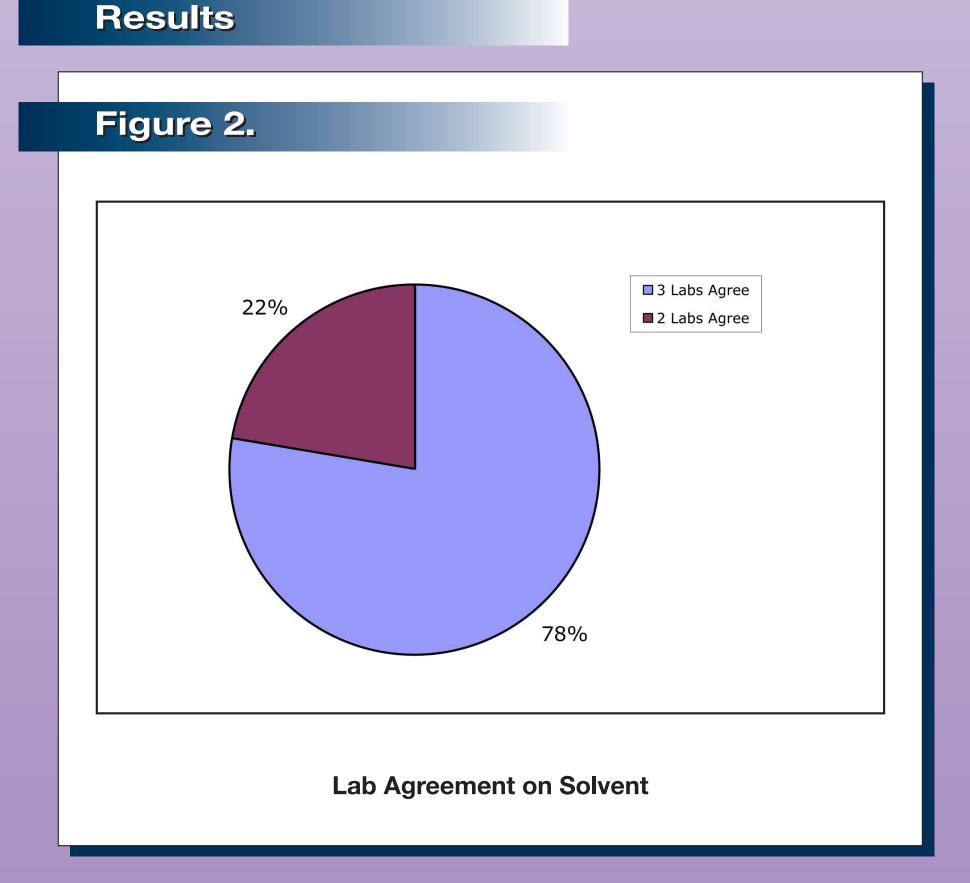
The order of preference for solvents was culture media, dimethyl sulfoxide (DMSO), and ethanol (ETOH). The protocol was based on a US Environmental Protection Agency guideline (US EPA 1998) and involved testing for solubility in a particular solvent, beginning at a relatively high concentration and proceeding to successively lower concentrations by adding more solvent as necessary for dissolution. At each concentration, mixing procedures (i.e., vortexing, sonicating, and heating to 37°C) were employed as necessary. Testing stopped when, upon visual observation, the procedure produced a clear solution with no cloudiness or precipitate. Chemicals considered insoluble in media were tested using DMSO and chemicals considered insoluble in DMSO were tested using ETOH. Practical limitations were the large volumes of solvent required for testing the more insoluble chemicals since approximately 50 mg test material would require >> 250 mL solvent.

The chemical purchaser/distributor was the first to evaluate the solubility of the test chemicals, first in media, then in DMSO, and then in ETOH at 400 and 200 mg/mL. Based on this experience, a solubility protocol for the *in vitro* labs was developed to test at lower test article concentrations and to test with the various solvents at concentrations that would be equivalent when applied to the cultures. The solubility flow chart in Figure 1 shows, for example, that 2 mg/mL medium and 200 mg/mL DMSO or ETOH are equivalent concentrations since they yield 1 mg/mL in cell culture. (Note: When applied to cultures, medium was diluted by one-half. DMSO and ETOH were diluted by 200 to achieve 0.5% [v/v] final concentrations.) At each concentration, the following mixing procedures were employed, as necessary to completely dissolve the test chemical, in this order:

- Gently mix. Vortex (1 –2 min).
- Sonicate for up to 5 min.
- Warm to 37°C for 5 60 min.

If test chemical was still undissolved, the next concentration/solvent was tested.







# Table 1. Solubility Results in mg/mL 200 NT NT DMSO <2 <2 200 NT <2 <2 200 NT 0.2 0.2 200 NT < 2 < 2 40 < 200 DMSO < 2 < 2 200 NT < 2 < 2 50<sup>6</sup> < 200 < 0.2 < 0.2 20 < 200</p> DMSO <2 <2 200 NT <0.2 <0.2 200 NT <2 <2 200 N <2 <2 500 NT DMSO <2 <2 200 NT <2 <2 200 NT <2 <2 <2 <2 </p> Medium 20 20 NT NT 20 20 NT NT 20 20 NT NT 2 2 200 < 200 DMSO 2 2 NT NT <2 <2 200 NT <2 <2 200 NT 40 40 400 400 Medium 20 20 NT NT 20 20 NT NT 20 20 NT NT Phenylthiourea 2 2 400 < 200 DMSO 2 < 2 200 NT 20 20 NT NT < 2 < 2 200 NT Physostigmine 2 2 400 200 DMSO 2 2 NT NT <2 <2 200 NT Medium 0.2 < 0.2 2 2 0.2 0.2 < 200 < 200 < 0.2 < 0.2 < 0.2 < 0.2 <2 <2 2 <20 DMSO 0.2 0.2 <200 <200 <0.2 <0.2 2 <2</p> Notes: 1 Used a different solubility protocol from the in vitro cytotoxicity labs. 2 Solvents selected by the SMT for cytotoxicity testing. Chemical distributor results were used to determine DMSO was selected. <sup>3</sup>Used protocol in Figure 1. <sup>4</sup>Dulbecco's Modification of Eagle's Medium. <sup>5</sup>Keratinocyte Growth Medium (KGM® from Cambrex). <sup>6</sup>Protocol deviation. In vitro labs agreed on solvent. Protocol did not provide enough information to select a solvent.

### Table 2. Differences in Media Solubility

Chemical	Lab 1	Lab 2	Lab 3
Lithium carbonate	3T3	3T3	3T3
Sodium oxalate	3T3		
Propranolol HCI	3T3		NHK
Aminopterin	NHK	3T3	
Copper sulfate	NII IIZ		
pentahydrate	NHK		
Valproic acid			NHK
Chloramphenicol	NHK		
Diethylphthalate		NHK	
Phenylthiourea	NHK		
Strychnine	NHK		
Xylene		NHK	
Total	8	4	3

Notes: <sup>1</sup>3T3 - solubility lower in 3T3 medium than in NHK medium, NHK – solubility lower in NHK medium than in 3T3 medium. No entry indicates no difference in solubility for the NHK and 3T3 media.

Although solubility in 3T3 medium was the same as that for NHK medium for 85% (61/72) of the chemicals, solubility was different for 11 chemicals in at least one lab. Two chemicals exhibited lower solubility in the 3T3 medium while seven chemicals had lower solubility in NHK medium in at least one lab. Two chemicals, propranolol HCl and aminopterin, had lower solubility in 3T3 medium in one lab and lower solubility in NHK medium in another lab. Although the differences for most chemicals were reported by one of three labs, medium solubility differences for lithium carbonate, propranolol HCl, and aminopterin were reported by at least two labs.

### Conclusions

- For in vitro cytotoxicity testing, culture medium was used for 38 chemicals, DMSO was used for 34 chemicals, and ETOH was not used
- Lab agreement was good for identifying the appropriate solvent. The in vitro labs selected the same solvent for 78% of the chemicals tested. Agreement was poor only with chemicals that exhibited low solubility, ~ 2 mg/mL, in medium. This was a limit concentration for determining whether to use medium or DMSO. If a chemical was insoluble in medium at 2 mg/mL, but soluble in DMSO at 200 mg/mL, then DMSO was selected as the solvent.
- Due to the 0.2 mg/mL lower limit for solubility testing, at least one in vitro lab failed to determine solubility for five chemicals (arsenic trioxide, sodium oxalate, strychnine, thallium sulfate, and triethylenemelamine). To determine the level of solubility for such compounds, future protocols should include testing at concentrations below 0.2 mg/mL. The chemical distributor used a protocol that required testing at concentrations as low as 0.05 mg/mL and never reported insolubility.
- Sixty-one of 72 chemicals were soluble in the 3T3 and NHK media to the same extent, despite differences in media composition.
- Solubility may be in the eye of the beholder. Differences in solubility among the labs may be attributed, in part, to the subjectivity of visual observations in judging whether the chemicals had fully dissolved.

### References

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