## 3 METHODS FOR MONITORING EXPOSURES

#### 3.1 EXPOSURE MONITORING

A worker's exposure to airborne acrylamide should be determined by using a personal sampling train consisting of a glass-fiber filter in a Swinnex cassette (13-mm) followed by a silica gel tube. Plastic cassettes (37-mm) yielded poor recoveries of acrylamide and are therefore unsuitable. Samples should be collected at a maximum flowrate of 1 liter/min for a minimum of 2 hr; the maximum air volume should be 120 liters. The silica gel tube should then be treated with methanol to extract the acrylamide. An important step in this method is the transfer of the glass-fiber filters to glass vials containing 1 ml of methanol immediately after sampling to avoid losses of acrylamide from the filter by evaporation. Analysis should be conducted by gas chromatography using a nitrogen/phosphorus detector. The limit of detection for this procedure is 1.3 parts per billion (ppb) (0.004 mg/m³). This method is described in Method 21 of the OSHA Analytical Methods Manual [OSHA 1985].

## 3.2 BIOLOGICAL MONITORING

The International Programme on Chemical Safety (IPCS) has recommended that a biological monitoring method for acrylamide be developed based on the determination of the adduct formed with hemoglobin [WHO 1985]. However, no biological monitoring test acceptable for routine use has yet been developed for acrylamide.

## 4 RELATIONSHIP BETWEEN EXPOSURE AND ADVERSE HEALTH EFFECTS

No data are available from studies in humans to establish an occupational exposure limit for acrylamide on the basis of neurotoxic, developmental, reproductive, or carcinogenic effects. However, many studies demonstrate a relationship between exposure and adverse health effects in animals. These studies are the bases for occupational exposure limits recommended by many organizations and government agencies. A few of these studies are described here.

EPA has used results from studies in animals to determine no-observable-effect levels (NOELs) and lowest-observable-effect levels (LOELs). With the use of the appropriate uncertainty factors, EPA has also recommended a "safe" concentration for human exposure (i.e., a concentration that is not expected to produce adverse health effects in exposed individuals) [EPA 1988b].

In 1988, EPA performed a detailed risk assessment of acrylamide based on animal studies of neurotoxicity, carcinogenicity, and reproductive effects [EPA 1988a]. Studies of neurotoxic effects in animals are summarized in Table 6. The NOEL was 0.2 to 2.0 mg/kg per day and the LOEL was 1.0 to 3.0 mg/kg per day. The reference dose (RfD) for acrylamide exposure (formerly acceptable daily intake, ADI) was calculated as 0.0002 mg/kg per day [EPA 1988b]. This value is based on an NOEL for neurotoxicity in a subchronic rat study of 0.2 mg/kg per day [Burek et al. 1980]. The exposure to 0.0002 mg/kg per day corresponds with a TWA concentration of 0.0014 mg/m³ (assuming an average 70-kg human breathing 10 m³ of air in an average working day with 100% absorption), which is approximately 20-fold lower than the OSHA PEL of 0.03 mg/m³. The RfD was obtained by dividing the NOEL by a factor of 1,000 to account for the use of animal data and subchronic exposure. This safety (uncertainty) factor was suggested by the National Academy of Science [NAS 1977].

A designation of B2 (probable human carcinogen) was proposed according to EPA cancer guidelines on the basis of data from studies of two different animal species [EPA 1988a]. Because risks at low exposures cannot be measured directly by experiments in animals or by epidemiologic studies, a number of mathematical models have been developed to extrapolate from high to low doses. To assess the cancer risk posed by acrylamide, EPA used a linear model (i.e., linearized multistage procedure) [EPA 1988a]. Data from the Johnson et al. [1986] study were used to estimate risk from acrylamide exposure. The EPA guidelines for cancer risk assessment recommend pooling tumor incidence data on the grounds that risk estimates derived from the incidence of site-specific tumors may not predict (and may in fact underestimate) whole-body risks that are determined with the pooled animal data. The dose-response curves for each sex are based on the pooled tumor incidence (benign and malignant) and comprise the data sets of choice for risk assessment. The most sensitive sex and species observed in this study (female rats) was chosen to represent possible human risk.

Table 6. Key animal studies of the neurotoxic effects of subchronic and chronic exposure to acrylamide\*,\*

Reference	Species (route)	Exposure duration	NOEL <sup>§</sup> (mg/kg)	LOEL <sup>§</sup> (mg/kg)	
Burek et al. 1980	Rats	90 days	0.2	1.0	
Johnson et al. 1986	Rats	2 y ars	0.5	2.0	
Hamblin 1956	Cats (i.v.)	180 days		1.0	
Kuperman 1958	Cats (i.p.)	125 days		1.0	
McCollister et al. 1964	Cats	1 year	0.3	1.0	
McCollister et al. 1964	Monkeys	1 year	1.0	3.0	
Spencer 1979	Monkeys	1 year	2.0	3.0	
Schaumburg et al. 1982	Monkeys	1.5 years		1.0	

Adapted from EPA [1988a].
 All studies here except the two noted used the oral route of administration.
 NOEL = no-observable-effect level; LOEL = lowest-observable-effect level.

The linearized multistage procedure was followed by EPA [1988a], with GLOBAL 86 as the computer program. Among the models that showed adequate fit with one to six stages, the model that gave the least q<sub>1</sub>\* (slope factor) was selected as the model with which to calculate carcinogenic risks using the lifetime average daily exposures provided by the exposure assessment for acrylamide. For the female rats with tumors of the thyroid, oral cavity, uterus, CNS, or mammary glands, this model had two stages. The cancer potency factor obtained for acrylamide by the linearized multistage procedure was 4.5 (mg/kg per day)<sup>1</sup>. The cancer potency factor describes the increased risk of developing cancer over a 70-year lifetime per unit of exposure where the unit of exposure is expressed as mg chemical/kg body weight per day. When based on an experimental animal study, the cancer potency factor is the 95% upper confidence limit slope of the dose-response relationship for a carcinogen as the dose approaches zero. Calculated upper-bound excess risks for individuals exposed to acrylamide are presented in Table 7. The highest risks were estimated for sewer repair workers, whose excess risks ranged from 10<sup>1</sup> to 10<sup>2</sup>. EPA calculated that exposure to airborne concentrations of 0.03 mg/m³ for a working lifetime of 40 years would result in an excess cancer risk of 2×10<sup>3</sup> [EPA 1988a].

The previous American Conference of Governmental Industrial Hygienists (ACGIH) threshold limit value (TLV®) of 0.3 mg/m³ [ACGIH 1971] was derived from a study of a small number of cats orally dosed with acrylamide and observed for neurotoxic effects [McCollister et al. 1964]. No effects were observed after feeding the cats acrylamide at the rate of 0.3 and 1 mg/kg per day, 5 days/week for 1 year. On the basis of these data, ACGIH recommended that human exposures not exceed a total intake of 0.05 mg/kg per day. The average 70-kg human who breathes 10 m³ of air in an average working day would not exceed this recommended exposure limit if he or she were exposed solely to airborne concentrations that did not exceed 0.3 mg/m³—the TLV adopted by ACGIH.

In 1989, OSHA adopted a new PEL for acrylamide—0.03 mg/m³ with a skin notation [29 CFR 1910.1000]. This new PEL was based on the increased incidence of cancer in laboratory animals and demonstrated dermal absorption of acrylamide [54 Fed. Reg. 2332]. NIOSH agreed with the proposed PEL of 0.03 mg/m³ and the supporting evidence of carcinogenicity [NIOSH 1988].

The World Health Organization reviewed cases of acrylamide poisoning in humans [WHO 1985]. Acute exposure to high doses of acrylamide appeared to affect the CNS, and long-term cumulative exposure to smaller doses produced peripheral neuropathy. Signs of peripheral neuropathy appeared after a latent period, which was dose-dependent and decreased with increasing dose.

On the basis of these neurotoxicity data, the World Health Organization recommended that the exposure not exceed a daily intake of 0.012 mg/kg body weight [WHO 1985]. For the average 70-kg human who breathes 10 m³ of air in an average working day, is occupationally exposed to airborne concentrations only, and has 100% absorption, this intake would result from breathing air with an acrylamide concentration of 0.094 mg/m³. This recommendation did not consider the risk of cancer or interference with reproduction.

Table 7. Estimates of excess cancer risk for individuals exposed to acrylamide over a lifetime\*

Exposure category	Upper-bound individual risk estimates		
Manufacturing/processing	10 <sup>-3</sup> - 10 <sup>-2</sup>		
Soil grouting (sewer workers)	10 <sup>-2</sup> - 10 <sup>-1</sup>		
Drinking water	10 <sup>-6</sup> ~ 10 <sup>-5+</sup>		

<sup>\*</sup> Adapted from EPA [1988a].
\* Worst case to typical case, based on residual acrylamide allowed.

## **5 RESEARCH NEEDS**

Very few studies address dermal absorption of acrylamide, although the dermal route may be the most significant one for acrylamide exposure in the workplace [He et al. 1989]. Therefore, quantitative studies should be performed to assess the absorption of acrylamide through the skin. Because dermal exposure appears to be a significant route of acrylamide uptake and it is difficult to monitor dermal exposures routinely, it is important to develop biomonitoring that can accurately reflect total exposure to acrylamide. A valid biomonitoring technique for acrylamide is presently unavailable. However, the literature on the toxicokinetics of acrylamide indicates that biomonitoring may be feasible either in urine or blood. More than 50% of the given dose is reported to be excreted in urine as the metabolite N-acetyl-S-(3-amino-3-oxy-propyl)cysteine [Miller et al. 1982]. By collecting the urine of workers whose exposure is monitored, it may be feasible to correlate the total exposure (that is, both inhalation and skin) to acrylamide with measured concentrations or total amounts of metabolites in 24-hr urine samples.

Another possibility for biomonitoring is measurement of acrylamide binding to red blood cells. Hashimoto and Aldridge [1970] observed that after rats received a single i.v. dose of <sup>14</sup>C acrylamide, the radioactivity in blood after 24 hr was entirely associated with red blood cells. Miller et al. [1982] found that the concentration of <sup>14</sup>C in whole blood reached a plateau at 12% of the total dose after 1 hr and remained constant throughout the time period examined (7 days). The binding to erythrocytes accounted for essentially all of the remaining radioactivity in the whole blood. In vitro studies showed that acrylamide was covalently bound to cysteine residues in protein and, on acidic hydrolysis, the adduct yielded a compound with chromatographic properties identified as S-(2-carboxyethyl)cysteine (CEC) [Hashimoto and Aldridge 1970; Bailey et al. 1986].

Bailey et al. [1986, 1987] used gas chromatography to measure the presence of CEC in red blood cells of rats dosed with less than 1 mg/kg. Research is needed to increase the sensitivity of the method (using high-performance liquid chromatography and ion chromotography) and to determine its applicability to human biomonitoring.

Another area that may require more research is elucidation of dose-response relationships for neurotoxic effects. The quantitative data on dose response were adequately addressed in only one species (rat) [Burek et al. 1980]. Better quantitative studies in other species (mice, cats, or rabbits) may be useful.

In light of the reported genotoxicity and carcinogenicity of acrylamide, information on the binding of this compound to DNA is of considerable interest [Moore et al. 1987, Bull et al. 1984a, 1984b; Johnson et al. 1986]. Studies addressing the mechanisms of genotoxicity (in vivo and in vitro DNA binding and effects) would be useful.

## Acrylamide

Acrylamide is widely used in research laboratories for making polyacrylamide gels. EPA estimated that 100,000 to 200,000 U.S. laboratory workers are potentially exposed [EPA 1988a]. Because exposure data are currently not available on this working population, a survey of potential exposure to acrylamide in research laboratories would be very useful.

## **6 DISCUSSION AND EVALUATION**

Studies in rats and mice indicate an association between the induction of cancer and exposure to acrylamide. Four types of response have been generally accepted as evidence of induction of neoplasms (tumors) [Williams and Weisburger 1986]: (1) the presence of tumors not observed in controls, (2) an increase in the incidence of a specific tumor type observed in controls, (3) the development of tumors earlier than those observed in controls, and (4) an increased number of tumors per animal. Acrylamide satisfies all these criteria. An increased incidence of tumors was observed in one strain of rats (female and male F344) and three strains of mice (male and female A/J, and female Sencar and Swiss-ICR). Increased incidences of lung adenoma and carcinoma occurred in ICR-Swiss female mice dosed by gavage six times over a period of 2 weeks. Similar increases in lung tumor incidence were observed in A/J male and female mice. In female and male rats, the increases in tumor incidence occurred at multiple sites (testes, thyroid, and adrenal gland in males and mammary gland, CNS, oral tissues, uterus, and clitoral gland in females). The incidence of tumors was dose-related both in rats and mice. In A/J mice, acrylamide increased the yield of lung tumors in both sexes in a dose-related manner. In addition, a highly significant doseresponse relationship existed for the time to occurrence of first tumors and for the number of tumors per animal in female Sencar mice.

Although carcinogenicity has not been demonstrated in workers occupationally exposed to acrylamide, the limitations of the epidemiologic studies preclude any conclusions regarding the association of exposure to acrylamide monomer and the risk of cancer. Prudent public health practice calls for regarding acrylamide as a potential occupational carcinogen.

On the basis of studies in animals, the International Agency for Research on Cancer (IARC) determined in 1986 that sufficient evidence existed to conclude that acrylamide was carcinogenic in animals and classified it as a 2B carcinogen (a possible human carcinogen) [IARC 1986]. Animal studies also indicate that reproduction is adversely affected by acrylamide exposure. Testosterone levels were depressed in rats [Ali et al. 1983], and decreased fertility was observed in male mice following oral exposure to acrylamide in drinking water [Sakamoto and Hashimoto 1986]. Degeneration of testicular epithelial tissue in male mice treated by gavage has been observed [Hashimoto 1981], as have dominant lethal effects in male rats exposed by drinking water [Smith et al. 1986] and mice exposed i.p. [Shelby et al. 1986]. Oral exposure of male or female mice and rats to acrylamide in drinking water has caused an increased resorption rate [Nalco Chemical Company 1987; Sakamoto and Hashimoto 1986].

## 7 SUMMARY

Acrylamide is an odorless, white, crystalline solid used as a monomer or as a raw material in the production of polyacrylamides. Workers potentially exposed to acrylamide monomer are employed in acrylamide manufacturing and processing, grouting operations, and research and analytical laboratories.

Only the acrylamide monomer is toxic; polyacrylamide products are generally nontoxic. Acrylamide monomer may be neurotoxic, carcinogenic, genotoxic, and hazardous to reproduction. Recent studies confirm that acrylamide exposures cause cancer and reproductive effects in animals, but epidemiologic studies have not demonstrated these effects in humans.

Key Words: Acrylamide, carcinogenicity, dermal exposures, grouting, neurotoxic effects, occupational exposure, polyacrylamide, reproductive effects.

## 8 REFERENCES

ACGIH [1971]. TLVs<sup>©</sup>: threshold limit values; documentation of the threshold limit values for substances in workroom air. 3rd ed. Cincinnati, OH: American Conference of Governmental Industrial Hygienists, pp. 5–6.

ACGIH [1986]. Documentation of the threshold limit values and biological exposure indices. 5th ed. Cincinnati, OH: American Conference of Governmental Industrial Hygienists, pp. 12-13.

ACGIH [1989]. TLVs<sup>®</sup>: threshold limit values and biological exposure indices for 1989–90. Cincinnati, OH: American Conference of Governmental Industrial Hygienists.

Agrawal AK, Squibb RE, Bondy SC [1981]. The effects of acrylamide treatment upon the dopamine receptor. Toxicol Appl Pharmacol 58(1):89–99.

Ali SF, Hong JS, Wilson W, Uphouse L, Bondy S [1983]. Effect of acrylamide on neurotransmitter metabolism and neuropeptide levels in several brain regions and upon circulating hormones. Arch Toxicol 52:35-43.

American Cyanamid Company [1969]. Chemistry of acrylamide. Wayne, NJ: American Cyanamid Company, Process Chemicals Department.

American Cyanamid Company [1980]. Toxic Substances Control Act (TSCA) FYI submission. A fetal toxicity study of acrylamide in rats. Washington, DC: U.S. Environmental Protection Agency, Office of Toxic Substances, EPA Document No. FYI-OTS-0680-0076. Unpublished report.

American Cyanamid Company [1983]. Toxic Substances Control Act (TSCA) 8(d) submission. CHO/HGPRT mammalian cell forward gene mutation assay. Acrylamide. Washington, DC: U.S. Environmental Protection Agency, Office of Toxic Substances, EPA Document No. 878211686.

American Cyanamid Company [1985]. Toxic Substances Control Act (TSCA) 8(d) submission. Drosophila sex-linked recessive assay of acrylamide. Washington, DC: U.S. Environmental Protection Agency, Office of Toxic Substances, EPA Document No. 878216232.

Backer LC, Dearfield KL, Erexson GL, Campbell JA, Westbrook-Collins B, Allen JW [1989]. The effect of acrylamide on mouse germ-line and somatic cell chromosomes. Environ Mol Mutagen 13:218–226.

Bailey E, Farmer PB, Bird I, Lamb JH, Peal JA [1986]. Monitoring exposure to acrylamide by the determination of S-(2-carboxyethyl)cysteine in hydrolyzed hemoglobin by gas chromatography-mass spectrometry. Anal Biochem 157:241–248.

Bailey E, Farmer PB, Shuker DEG [1987]. Estimation of exposure to alkylating carcinogens by the GC-MS determination of adducts to hemoglobins and nucleic acid bases in urine. Arch Toxicol 60:187–191.

Banerjee S, Segal A [1986]. In vitro transformation of C3H/10T 1/2 and NIH/3T3 cells by acrylonitrile and acrylamide. Cancer Lett 32:293-304.

Berti-Mattera LN, Lopachin RM, Schrama L, Lowery J, Eichberg J [1986]. Acrylamide alters axonal protein phosphorylation and polyphosphoinositide metabolism. 16th Annual Meeting of the Society for Neuroscience, Part 1, Washington, DC, Nov. 9–14, 1986. Soc Neurosci Abstr 12:(1)94.

Bisby MA, Redshaw JD [1987]. Acrylamide neuropathy: changes in the composition of proteins of fast axonal transport resemble those observed in regenerating axons. J Neurochem 48:924–928.

Brismar T, Hildebrand C, Tegner R [1987]. Nodes of ranvier in acrylamide neuropathy: voltage clamp and electron microscopic analysis of rat sciatic nerve fibres at proximal levels. Brain Res 423:135-143.

Bull RJ, Robinson M, Laurie RD, Stoner GD, Greisiger E, Meir JR, Stober JA [1984a]. Carcinogenic effects of acrylamide in sencar and A/J mice. Cancer Res 44:107-111.

Bull RJ, Robinson M, Stober JA [1984b]. Carcinogenic activity of acrylamide in the skin and lung of swiss-ICR mice. Cancer Lett 24:209-212.

Burek J, Albee R, Beyer J, Bell T, Carreon R, Morden D, Wade C, Hermann E, Gorzinski S [1980]. Subchronic toxicity of acrylamide administered to rats in the drinking water followed by up to 144 days of recovery. J Environ Pathol Toxicol 4:157–182.

Carlson GP, Weaver PM [1985]. Distribution and binding of [14C] acrylamide to macromolecules in sencar and BALB/c mice following oral and topical administration. Toxicol Appl Pharmacol 79:303-313.

29 CFR 1910.1000. Air contaminants. Occupational Safety and Health Administration. Washington, DC: U.S. Government Printing Office, Office of the Federal Register.

CMR [1985]. Chemical profile: acrylamide. Chemical Marketing Reporter, January 7, 1985.

Collins JJ, Swaen GMH, Marsch GM, Utidjian HMD, Caporossi JC, Lucas LJ [1989]. Mortality patterns among workers exposed to acrylamide. J Occup Med 31:614-617.

Cook WA [1987]. Occupational exposure limits—worldwide. Akron, OH: American Industrial Hygiene Association.

Davidson R, Volk H, Friedrick R [1980]. Polyacrylamides. Chapter 16. In: Handbook of water-soluble gums and resins. New York, NY: McGraw-Hill, pp. 1-19.

Dearfield KL, Abernathy CO, Ottley MS, Brantner JH, Hayes PF [1988]. Acrylamide: its metabolism, developmental, and reproductive effects, genotoxicity, and carcinogenicity. Mutat Res 195:45–77.

De Rojas TC, Goldstein BD [1987]. Primary afferent terminal function following acrylamide: alterations in the dorsal root potential and reflex. Toxicol Appl Pharmacol 88:175–182.

Dixit R, Mukhtar H, Seth PK, Murti CRK [1981]. Conjugation of acrylamide with glutathione catalyzed by glutathione-S-transferases of rat liver and brain. Biochem Pharmacol 30(13): 1739–1744.

Dixit R, Das M, Seth PK, Mukhtar H [1986]. Interaction of acrylamide with bovine serum albumin. Environ Res 40:365-371.

Eckert BS, Yeagle PL [1988]. Acrylamide treatment of PtK1 cells causes dephosphorylation of keratin polypeptides. Cell Motil Cytoskeleton 11:24-30.

Edwards PM [1975]. The distribution and metabolism of acrylamide and its neurotoxic analogues in rats. Biochem Pharmacol 24:1277–1282.

Edwards PM [1976]. The insensitivity of the developing rat foetus to the toxic effects of acrylamide. Chem Biol Interact 12:13–18.

EPA [1987]. Assessment of airborne exposure and dermal contact to acrylamide during chemical grouting operations. Washington, DC: U.S. Environmental Protection Agency, Office of Toxic Substances. EPA 560/5-87-009.

EPA [1988a]. Preliminary assessment of health risks from exposure to acrylamide. Washington, DC: U.S. Environmental Protection Agency, Office of Toxic Substances.

EPA [1988b]. Integrated risk information system (IRIS): Reference dose (RfD) for chronic oral exposure. Acrylamide. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Criteria and Assessment Office.

37 Fed. Reg. 329 [1972]. Food and Drug Administration. Food additives: acrylate-acrylamide resins.

54 Fed. Reg. 2332 [1989]. Occupational Safety and Health Administration: air contaminants; final rule. (To be codified at 29 CFR 1910.)

Frantz SW, Dryzga MD, Freshour NL, Watanabe PG [1985]. In vivo/in vitro determination of cutaneous penetration of residual monomer from polyacrylamides [Abstract]. Toxicologist 5:39.

Garland TO, Patterson MWH [1967]. Six cases of acrylamide poisoning. Br Med J 4:134–138.

GCA [1980]. Acrylamide technical control options analysis. Washington, DC: GCA Corporation. Prepared for EPA, Contract No. 68–01–5960, Report No. GCA-TR-80-75-G.

Gold BG [1987]. The pathophysiology of proximal neurofilamentous giant axonal swellings: implications for the pathogenesis of amyotrophic lateral sclerosis. Toxicology 46:125-39.

Goldstein BD, Fincher DR [1986]. Paradoxical changes in spinal cord reflexes following the acute administration of acrylamide. Toxicol Lett 31:93-99.

Hamblin D [1956]. The toxicity of acrylamide—a preliminary report. Hommage au Doyen René Fabre (Paris) Sede 3:195–199.

Hashimoto K, Aldridge WN [1970]. Biochemical studies on acrylamide, a neurotoxic agent. Biochem Pharmacol 19:2591–2604.

Hashimoto K, Sakamoto J, Tanii H [1981]. Neurotoxicity of acrylamide and related compounds and their effects on male gonads in mice. Arch Toxicol 47:179–189.

Hashimoto K, Tanii H [1985]. Mutagenicity of acrylamide and its analogues in Salmonella typhimurium. Mutat Res 158:129-133.

He F, Zhang S, Wang H, Li G, Zhang Z, Li F, Dong X, Hu F [1989]. Neurological and electroneuromyographic assessment of the adverse effects of acrylamide on occupationally exposed workers. Scand J Work Environ Health 15:125–129.

Hersch MI, McLeod JG, Satchell PM, Early RG, Sullivan CE [1989]. Breathing pattern, lung inflation reflex and airway tone in acrylamide neuropathy. Respir Physiol 76:257–276.

Hills BW, Greife AL [1986]. Evaluation of occupational acrylamide exposures. Appl Ind Hyg 13:148-152.

Howland RD, Ali P [1986]. Altered phosphorylation of rat neuronal cytoskeletal proteins in acrylamide induced neuropathy. Brain Res 363:333-339.

Hsie A, Recio L, Katz D, Lee C, Wagner M, Schenley R [1986]. Evidence for reactive oxygen species inducing mutation in mammalian cells. Proc Natl Acad Sci, USA 83:9616-9620.

Husain R, Srivastava S, Srivastava SP, Seth PK [1986]. Effect of acrylamide on energy-linked functions in rat brain. Bull Environ Contam Toxicol 37:427–432.

Husain R, Dixit R, Das M, Seth PK [1987]. Neurotoxicity of acrylamide in developing rat brain: changes in the levels of brain biogenic amines and activities of monoamine oxidase and acetylcholine esterase. Ind Health 25:19–28.

IARC [1986]. IARC monographs on the evaluation of the carcinogenic risk of chemicals to humans: acrylamide. Vol. 39. Lyon, France: World Health Organization, International Agency for Research on Cancer.

IARC [1987]. Overall evaluations of carcinogenicity: an updating of IARC monographs, Vols. 1 to 42. Supplement 7. Lyon, France: World Health Organization, International Agency for Research on Cancer, pp. 32 and 43.

IHE [1985]. Studies on the genetic toxicology of acrylamide monomer, 1985, FYI submission. Washington, DC: U.S. Environmental Protection Agency, Office of Toxic Substances; Brussels, Belgium: Institute of Hygiene and Epidemiology, EPA Doc. Control No. FYI-OTS-0885-044A.

Ikeda GJ, Miller E, Sapienza PP, Michel TC, King MT, Turner VA, Blumenthal H, Jackson WE, Levin S [1983]. Distribution of <sup>14</sup>C-labelled acrylamide and betaine in foetuses of rats, rabbits, beagle dogs, and miniature pigs. Food Chem Toxicol 21:49–58.

Ikeda GJ, Miller E, Sapienza PP, Michel TC, King MT, Sager AO [1985]. Maternal-foetal distribution studies in the late pregnancy. II. Distribution of [1-14C] acrylamide in tissues of beagle dogs and miniature pigs. Food Chem Toxicol 23:757-761.

Johnson KA, Gorzinski SJ, Bodner KM, Campbell RA, Wolf CH, Friedman MA, Mast RW [1986]. Chronic toxicity and oncogenicity study on acrylamide incorporated in the drinking water of Fischer 344 rats. Toxicol Appl Pharmacol 85:154–168.

Kaplan ML, Murphy SD, Gilles FH [1973]. Modification of acrylamide neuropathy in rats by selected factors. Toxicol Appl Pharmacol 24:564-579.

Khanna VK, Husain R, Seth PK [1988]. Low protein diet modifies acrylamide neurotoxicity. Toxicology 49:395-401.

Kuperman A [1958]. Effects of acrylamide on the central nervous system of the cat. J Pharmacol Exp Ther 123:180-192.

Lijinsky W, Andrews A [1980]. Mutagenicity of vinyl compounds in Salmonella typhimurium. Teratogenesis Carcinog Mutagen 1:259–267.

Marlowe C, Clark MJ, Mast RW, Friedman MA, Waddell WJ [1986]. The distribution of [14C] acrylamide in male and pregnant Swiss-Webster mice studied by whole-body autoradiography. Toxicol Appl Pharmacol 86:457–465.

McCollister DD, Oyen F, Rowe VK [1964]. Toxicology of acrylamide. Toxicol Appl Pharmacol 6(2):172-181.

McCollister DD, Hake CL, Sadek SE, Rowe VK [1965]. Toxicologic investigations of polyacrylamides. Toxicol Appl Pharmacol 7:639-651.

Merigan WH, Barkdoll E, Maurissen JPJ [1982]. Acrylamide-induced visual impairment in primates. Toxicol Appl Pharmacol 62:342-345.

Miller MJ, Carter D, Sipes I [1982]. Pharmacokinetics of acrylamide in Fischer 344 rats. Toxicol Appl Pharmacol 63:36-44.

Miller MS, Spencer PS [1985]. The mechanisms of acrylamide axonopathy. Ann Rev Pharmacol Toxicol 25:643-666.

Moore MM, Amtower A, Doerr C, Brock KH, Dearfield KL [1987]. Mutagenicity and clastogenicity of acrylamide in L5178Y mouse lymphoma cells. Environ Mutagen 9:261–267.

Nalco Chemical Company [1987]. Section 3e [Toxic Substances Control Act] combined two generation reproduction study and dominant lethal assay in Fischer 344 rats administered acrylamide in the drinking water. Washington, DC: U.S. Environmental Protection Agency, Office of Toxic Substances.

NAS [1977]. Drinking Water and Health. Vol. 3. Washington, DC: National Academy of Sciences, National Academy Press.

NIOSH [1976]. Criteria for a recommended standard: occupational exposure to acrylamide. Cincinnati, OH: U.S. Department of Health, Education, and Welfare, Public Health Service, Center for Disease Control, National Institute for Occupational Safety and Health, DHEW (NIOSH) Publication No. 77–112.

NIOSH [1983]. National occupational exposure survey (NOES), 1981–1983. Cincinnati, OH: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control, National Institute for Occupational Safety and Health. Unpublished data base; provisional data as of 1/1/90.

NIOSH [1988]. Testimony of the National Institute for Occupational Safety and Health on the Occupational Safety and Health Administration's proposed rule on air contaminants. Presented August 1, 1988, Washington, D.C. NIOSH policy statements. Cincinnati, OH: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control, National Institute for Occupational Safety and Health.

O'Donoughue JL, ed. [1985]. Acrylamide and related substances. In: Neurotoxicity of industrial and commercial chemicals. Vol. II. Boca Raton, FL: CRC Press, pp. 170-177.

OSHA [1985]. Analytical methods manual. Method 21. Salt Lake City, UT: U.S. Department of Labor, Occupational Safety and Health Administration, Analytical Laboratory.

Ott MG, Kolesar RC, Scharnweber HC, Schneider EJ, Venable JR [1980]. A mortality study of employees engaged in development or manufacture of styrene-based products. J Occup Med 22:444–460.

Ramsey JC, Young JD, Gorzinski SJ [1984]. Acrylamide: toxicodynamics in rats. Midland, MI: Dow Chemical Company. Unpublished report.

Sakamoto J, Hashimoto K [1986]. Reproductive toxicity of acrylamide and related compounds in mice—effects on fertility and sperm morphology. Arch Toxicol 59:201–205.

Satchell PM, McLoad JG [1981]. Megaoesophagus due to acrylamide neuropathy. J Neurol Neurosurg Psychiatry 44:906-913.

Schaumburg HH, Arezzo J, Spencer PS [1982]. Short-latency somatosensory evoked potentials in primates intoxicated with acrylamide: implications for toxic neuropathies in man. Presented at the 1982 meeting of the Society of Toxicology, Boston, MA. Toxicologist 2:139. Abstract No. 490.

Schaumburg HH, Spencer PS [1979]. Clinical and experimental studies of distal neuropathy—a frequent form of brain and nerve damage produced by environmental chemical hazards. Bronx, NY: Departments of Neurology, Neuroscience and Pathology (Neuropathology), Albert Einstein College of Medicine. Grant No. R01–OH–00535, NIOSHTIC, RN 00091771.

Shelby MD, Cain TK, Hughes L, Braden P, Generoso WM [1986]. Dominant lethal effects of acrylamide in male mice. Mutat Res 173:313-324.

Shelby MD, Cain TK, Cornett CV, Generoso WM [1987]. Acrylamide: induction of heritable translocations in male mice. Environ Mutagen 9:363–368.

Smith MK, Zenick H, Preston RJ, George EL, Long RE [1986]. Dominant lethal effects of subchronic acrylamide administration in the male Long-Evans rat. Mutat Res 173:273-277.

Sobel W, Bond GG, Parsons TW, Brenner FE [1986]. Acrylamide cohort mortality study. Br J Ind Med 43:785-788.

Solomon JJ, Fedyk J, Mukai F, Segal A [1985]. Direct alkylation of 2'-deoxynucleosides and DNA following in vitro reaction with acrylamide. Cancer Res 45:3465–3470.

Spencer PS [1979]. A neuropathologic study of acrylamide intoxication. Unpublished final report. Washington, DC: U.S. Department of Health, Education, and Welfare, Public Health Service, Center for Disease Control, National Institute for Occupational Safety and Health. Contract No. OH 00535.

Spencer PS, Schaumburg HH [1974]. A review of acrylamide neurotoxicity. Part I. Properties, uses and human exposure. Can J Neurol Sci 1:143-150.

Spencer PS, Schaumburg HH [1975]. Nervous system degeneration produced by acrylamide monomer. Environ Health Perspect 11:129–133.

Spencer PS, Schaumburg HH [1976]. Central-peripheral distal axonopathy—the pathology of dying-back polyneuropathies. Prog Neuropathol 3:253–295.

Spencer PS, Schaumburg HH [1977]. Ultrastructural studies of the dying-back process. IV. Differential vulnerability of PNS and CNS fibers in experimental central-peripheral distal axonopathies. J Neuropathol Exp Neurol 36(2):300-320.

Srivastava SP, Das M, Seth PK [1983]. Enhancement of lipid peroxidation in rat liver on acute exposure to styrene and acrylamide: a consequence of glutathion depletion. Chem Biol Interact 45:373–380.

Srivastava SP, Seth PK, Das M, Mukhtar H [1985]. Effects of mixed-function oxidase modifiers on neurotoxicity of acrylamide in rats. Biochem Pharmacol 34:1099-1102.

Sublet V, Smith MK, Randall J, Zenick H [1986]. Spermatogenic stages associated with acrylamide (ACR) induced dominant lethality [Abstract]. Toxicologist 6:292.

Takahashi M, O'Hara T, Hashimoto K [1971]. Electrophysiological study of nerve injuries in workers handling acrylamide. Int Arch Arbeitsmed 28:1-11.

Tilson HA, Spencer PA, Cabe PS [1979]. Acrylamide neurotoxicity in rats: a correlated neurobehavioral pathological study. Neurotoxicology 1:89-104.

Tilson HA [1981]. The neurotoxicity of acrylamide: an overview. Neurobehav Toxicol Teratol 3:445-461.

Waalkens D, Joosten H, Taalman R, Scheres J, Yih T, Hoekstra H [1981]. Sister-chromatid exchanges induced in vitro by cyclophosphamide without exogenous metabolic activation in lymphocytes from three mammalian species. Toxicol Lett 7:229–232.

Walden R, Squibb R, Schiller S [1981]. Effects of prenatal and lactational exposure to acrylamide on the development of intestinal enzymes in the rat. Toxicol Appl Pharmacol 58:363-369.

WHO [1985]. Environmental Health Criteria 49: acrylamide. Geneva, Switzerland: World Health Organization, International Programme on Chemical Safety.

Williams GM, Weisburger JH [1986]. Chemical Carcinogens. In: Klaasen CD, Andrew MO, Doull J, eds. Casarett and Doull's Toxicology. New York, NY: Macmillan Publishing Company, pp. 99–173.

Zenick H, Hope E, Smith M [1986]. Reproductive toxicity associated with acrylamide treatment in male and female rats. J Toxicol Environ Health 17:457-472.

# 9 APPENDIX. INTERNATIONAL STANDARDS FOR WORKPLACE EXPOSURES TO ACRYLAMIDE

This appendix lists occupational exposure limits for airborne acrylamide in various countries (Table A-1), and it contains a discussion of the bases for other recommendations that depart from the frequently cited limit of 0.3 mg/m<sup>3</sup>.

Table A-1. Occupational exposure limits for airborne acrylamide in various countries (mg/m³)

Country	Time-weighted average (TWA)		Short-term exposure limit (STEL)		Ceiling	
Austria		0.3				
Belgium	S <sup>+</sup>	0.3				
Denmark	Š	0.3				
Federal Republic	J					
of Germany	S	0.3				
Finland	_	0.3		0.9		
Hungary	S	0.3	S	1.5		
Indonesia	S	0.3	_			
Italy	S	0.3		•		
Japan		0.3		***		
Korea		0.3		0.6		
Mexico	S	0.3				
Netherlands	S	0.3				
Sweden	S	0.3	S	0.9		
Switzerland	S	0.3				
Taiwan		0.3				
United Kingdom	S	0.3	S	0.6		
United States (OSHA)	S	0.03				
Venezuela	S	0.3			S	0.6
Yugoslavia	S	0.3				

Adapted from Cook [1987].

<sup>+ &</sup>quot;S" denotes potential absorption into the body through the skin.

In 1989, the Occupational Safety and Health Administration (OSHA) changed its permissible exposure limit (PEL) from S 0.3 mg/m³ to S 0.03 mg/m³ (TWA) [29 CFR 1910.1000].

In 1976, the National Institute for Occupational Safety and Health (NIOSH) recommended an exposure limit (REL) for acrylamide of 0.3 mg/m³ (0.1 ppm) as a time-weighted average (TWA) for up to a 10-hr workshift (40 hr per week) [NIOSH 1976]. At that time, the available human and animal studies did not provide enough information to alter the previously established OSHA permissible exposure limit (PEL) of 0.3 mg/m³ as an 8-hr TWA. The original OSHA PEL was based on the 1968 American Conference of Governmental Industrial Hygienists (ACGIH) threshold limit value (TLV®) [ACGIH 1971], which was derived mainly from a study of a small number of cats orally dosed with acrylamide and observed for neurotoxic effects [McCollister et al. 1964]. No effects were observed after feeding the cats acrylamide at the rate of 0.3 and 1 mg/kg per day, 5 days/week for 1 year.

The current Swedish standard for acrylamide [TWA =  $0.3 \text{ mg/m}^3$ ] is the same as the previous ACGIH recommendation, with an added short-term exposure limit (STEL) of  $0.9 \text{ mg/m}^3$ .

Since 1968, ACGIH has designated acrylamide as an A2 substance (suspected human carcinogen) and assigned it a TLV of 0.03 mg/m³ (0.01 ppm) as an 8-hr TWA with a skin notation [ACGIH 1986]. The revised TLV was based on data indicating a carcinogenic response in rats exposed to acrylamide in drinking water [Johnson et al. 1986]. The skin notation was assigned because of the demonstrated dermal absorption of acrylamide.

On the basis of studies in animals, the International Agency for Research on Cancer (IARC) determined in 1986 that there was sufficient evidence to conclude that acrylamide was carcinogenic in animals [IARC 1986] and classified it as a 2B carcinogen (a possible human carcinogen) [IARC 1987].

The World Health Organization recommended that acrylamide exposure not exceed a daily intake of 0.012 mg/kg body weight [WHO 1985]. For the 70-kg human who breathes 10 m³ of air in an average workday, is occupationally exposed to airborne concentrations only, and has 100% absorption, this intake would result from breathing air with an acrylamide concentration of 0.094 mg/m³. It should be emphasized that this value is based solely on the neurotoxicity of acrylamide and does not take into account the risk of cancer or interference with reproduction.

In 1989, OSHA adopted a new PEL for acrylamide—0.03 mg/m³ as an 8-hr TWA with a skin notation [29 CFR 1910.1000]. This new PEL was based on the increased incidence of cancer in laboratory animals and demonstrated dermal absorption of acrylamide [54 Fed. Reg. 2332 (1989)]. OSHA considered evidence of carcinogenicity derived from the studies by Johnson et al. [1986] and Bull et al. [1984a, 1984b] and stated that the evidence was sufficient to conclude that acrylamide is a carcinogen. In addition, OSHA cited the ACGIH and IARC evaluations of acrylamide as a carcinogen. NIOSH agreed with the proposed PEL of 0.03 mg/m³ and the supporting evidence of carcinogenicity [NIOSH 1988].

In 1988, the U. S. Environmental Protection Agency (EPA) proposed a reference dose (RfD) for acrylamide exposure (formerly acceptable daily intake, ADI) of 0.0002 mg/kg per day [EPA 1988b]. The RfD is based on a no-observable-effect level (NOEL) in a subchronic rat study of 0.2 mg/kg per day [Burek et al. 1980]. The RfD was obtained by dividing the NOEL by a factor of 1,000 to account for the use of animal data and subchronic exposure. This safety (uncertainty) factor was suggested by the National Academy of Science [NAS 1977]. The designation of B2 (probable

human carcinogen) was proposed for acrylamide according to EPA cancer guidelines on the basis of data from studies of two different animal species [EPA 1988a].

The U.S. Food and Drug Administration has recommended that residual acrylamide monomer not exceed 0.05% in molasses and in beet and cane sugar [37 Fed. Reg. 329 (1972)].

