2,4- AND 2,6-TOLUENEDIAMINE (in the presence of isocyanates)

METHOD: 5516 Issue 2		N: PARTIAL	Issue 1: 15 May 1989
$CH_3C_6H_3(NH_2)_2$	MW: 122.17	95-80-7 823-40-5	RTECS: 2,4-: XS9625000 2,6-: XS9750000

METHOD: 5516, Issue 2	EVALUATION: PARTIAL	Issue 1: 15 May 1989 Issue 2: 15 August 1994	
OSHA : no PEL NIOSH: 2,4-: lowest feasible; carcinogen ACGIH: no TLV	PROPERTIES:	2,4-: solid; MP 99 °C 2,6-: solid; MP 106 °C	

SYNONYMS: 2,4-: 4-methyl-1,3-benzenediamine; 2,4-diaminotoluene 2,6-: 2-methyl-1,3-benzenediamine; 2,6-diaminotoluene

SAMPLING		MEASUREMENT	
SAMPLER:	IMPINGER (solution of 1-(2-methoxyphenyl)-	TECHNIQUE:	HPLC, UV DETECTION
	piperazine in toluene, 15 mL)	ANALYTES:	2,4- and 2,6-bisacetamidotoluene
FLOW RATE:	1 L/min	PREPARATION:	acetylate 4 hour, evaporate, redissolve in 1.5 mL methanol
VOL-MIN:	30 L @ 10 μg/m ³		
-MAX:	500 L	INJECTION VOLUME:	10 µL
SHIPMENT:	routine		· • F=
		MOBILE PHASE:	F 9
SAMPLE STABILITY:	at least 2 weeks @ 25 °C		acetonitrile/water at pH 6.0; 1.0 mL/min; ca. 20 °C
•	in the dark [1]		
FIELD BLANKS:	2 to 10 field blanks per set	COLUMN:	10 cm x 8-mm octadecylsilylated silica
	-	_	(C_{18}), 5-µm particle size, in Waters
	ACCURACY		RCM-100 radial compression module
RANGE STUDIED	: not studied	CALIBRATION:	standard solution of analytes in methanol
BIAS:	not determined	DANOT	
OVERALL PRECISION (Ŝ _{rτ}): not determined		RANGE:	0.3 to 3 µg per sample [1]
ACCURACY:	not determined	ESTIMATED LOD	: 0.1 µg per sample [1]
		PRECISION (Š,):	0.06 @ 0.74 to 0.89 µg per sample [1]

APPLICABILITY: The working range is 3 to 30 μ g/m³ for a 100-L air sample. This method, based on that of Warwick <u>et al.</u> for isocyanates [2], determines 2,4- and 2,6-toluenediamine in air in the presence of isocyanates. Samples from polyurethane foam plants were analyzed simultaneously for 2,4- and 2,6-toluenediamine and 2,4- and 2,6-toluene diisocyanate [1].

INTERFERENCES: <u>m</u>-Phenylenediamine interferes in the determination of 2,4-toluenediamine.

OTHER METHODS: Holdren <u>et al.</u> [3] reported a similar method using <u>N</u>-(4-nitrobenzyl)propylamine in toluene for sampling and HPLC with electrochemical detection. Other methods are: (a) absorb on Tenax GC, desorb in toluene, GC [4]; (b) absorb on silica gel, desorb in 2-butanone, GC [5]; (c) sample in aqueous acid, work up, GC of free amines [6] or bis(hepta-fluorobutyrl) amides [7,8]; (d) sample in ethanolic KOH, workup, LC of free amine [9,10] or GC of bis(pentafluoropropionyl) amides [11]; (e) sample with sulfuric acid-coated filter, work up, GC of bis(heptafluorobutyrl)amides [12]. Some of these methods [3,9-11] can b e used for the simultaneous determination of toluenediamines and toluene diisocyanates.

REAGENTS:

- 1. Toluene, reagent grade.
- 2. 1-(2-Methoxyphenyl)piperazine, purified (see APPENDIX A).
- Sampling medium: 43 µg/mL 1-(2methoxyphenyl)piperazine in toluene.
- 4. 2,4-Toluenediamine*, reagent grade.
- 5. 2,6-Toluenediamine*, reagent grade.
- 6. Acetic anhydride, reagent grade.
- 7. Methanol, reagent grade.
- Mobile phase A: Dissolve 60 mg anhydrous sodium acetate in 1 L of 12% acetonitrile in distilled water. Add 17% (V/V) aqueous acetic acid. Bring pH to 6.0.
- 9. Mobile phase B: Acetonitrile, chromatographic quality.
- 10. Water, distilled deionized.
- 11. Sodium acetate, anhydrous.
- 12. 2,4-Bisacetamidotoluene, See APPENDIX B).
- 2,6-Bisacetamidotoluene, (See APPENDIX B).
- Calibration stock solution, 0.5 μg/μL. Dissolve 5 mg each of 2,4- and 2,6bisacetamidotoluene in methanol. Dilute to 10 mL.
- 15. Nitrogen, prepurified.
- 16. Pentane*, purified.
 - * See SPECIAL PRECAUTIONS

SAMPLING:

EQUIPMENT:

- 1. Sampler: midget impinger, 25-mL.
- 2. Personal sampling pump, 1 L/min, with flexible connecting tubing.
- 3. Liquid chromatograph (HPLC) with 229-nm UV detector, recorder, integrator, and column (page 5516-1).
- 4. Vials, 4-mL and 20-mL, glass, with PTFE-lined caps.
- 5. Pipets, pasteur, 14.6-cm, glass disposable.
- 6. Volumetric flasks, 10-mL.
- 7. Syringes, 10- and 100-µL.
- 8. Pipets, 2-mL (graduated) and 15-mL, glass, with pipet bulb.
- 9. Water bath.
- 10. Hotplate, spark-free, 35 to 50 °C.
- 11. Evaporator, Mini-Vap, six-port, or equivalent.
- 12. pH meter.
- 13. Beakers, 250-mL.
- 14. Flask, filtration, 500-mL.
- 15. Funnel, Buchner, fritted glass, 100-mL.
- 16. Vacuum pump.
- 17. Vacuum desiccator.
- 18. Watchglass.

SPECIAL PRECAUTIONS: 2,4-Toluenediamine is a cancer suspect agent [13]; 2,6-Toluenediamine may be mutagenic [14]. Handle these chemicals carefully in a hood or glove box, and avoid working-surface contamination which

- 1. Calibrate each personal sampling pump with a representative sampler in line.
- 2. Transfer 15.0 mL sampling medium to an impinger.
- 3. Connect the assembled impinger to a sampling pump.
- 4. Sample 30 to 500 L of air at an accurately measured sampling rate of 1 L/min. When it is necessary to add solvent for proper impinger operation during sampling, add only toluene. NOTE: The reagent in the sampling medium reacts with isocyanates present to form ureas, thus preventing reaction of the isocyanate with the toluenediamines.

CH ₃ OC ₆ H ₄ N(CH ₂ CH ₂) ₂ NH	+	$\text{R-N=C=0} \rightarrow $	CH ₃ OC ₆ H ₄ N(CH ₂ CH ₂) ₂ NCONHR
1-2(methoxyphenyl)-		isocyanates	ureas
piperazine			

5. Transfer sample solution, including condensed water, to a 20-mL vial for shipment. Rinse impinger with 1 to 2 mL toluene. Add rinsings to sample solution.

SAMPLE PREPARATION:

 Add 25 μL acetic anhydride to acetylate the 2,4- and 2,6-toluenediamine and excess 1-(2methoxyphenyl)piperazine. Allow 4 hours for completion of reactions. Evaporate sample to dryness under a gentle stream of nitrogen while warming to 40 to 50 °C on a hotplate. Redissolve residue in 1.5 mL methanol.

NOTE: The acetylation reaction for 2,4-toluenediamine to produce 2,4-bisacetamidotoluene is:

 $\begin{array}{rrrr} \mathsf{CH}_3\mathsf{C}_6\mathsf{H}_3(\mathsf{NH}_2)_2 & + & 2\mathsf{CH}_3\mathsf{COOCOCH}_3 & \rightarrow & \mathsf{CH}_3\mathsf{C}_6\mathsf{H}_3(\mathsf{NHCOCH}_3)_2 & + & 2\,\mathsf{CH}_3\mathsf{COOH}.\\ \texttt{2,4-toluenediamine} & & \operatorname{acetic} \ \texttt{anhydride} & & \texttt{2,4-bisacetamidotoluene} & & \operatorname{acetic} \ \texttt{acetic} \ \texttt{$

CALIBRATION AND QUALITY CONTROL:

- 7. Calibrate daily with at least six working standards.
 - a. Using aliquots of calibration stock solution, prepare working standards of 2,4- and 2,6bisacetamidotoluene in methanol covering the range 0.1 to 3 µg/mL each.
 - b. Analyze these with the unknown and blank samples (steps 9 through 11).
 - c. Prepare calibration graphs (peak height vs. μg 2,4- and 2,6-toluenediamine per sample). Multiply the concentration (μg/mL) of bisacetamidotoluene by 0.889 mL to obtain the quantity (μg) of toluenediamine per sample. NOTE: The factor 0.889 includes the MW of toluenediamine (122.17), the MW of bisacetamidotoluene (206.24), and the 1.5-mL solution volume from step 6.
- 8. Prepare three quality control samples by adding known quantities of 2,4- and 2,6-toluenediamine to 15 mL of sampling medium and analyze (steps 9 through 11).

MEASUREMENT:

- 9. Set up the HPLC system according to the manufacturer's recommendations and to the conditions given on page 5516-1. The mobile phase program is:
 - a. Linear gradient 100% A to 90% A over t = 0 to 8 min.
 - b. 90% A to 40% A over t = 8 to 19 min following the convex gradient % A = 90 31 (t 8) $\frac{1}{5}$.
 - c. Hold at 40% A for 1 min, or as long as necessary to clear the column.
 - d. Return to 100% A and hold for 7 min before the next run.
 - NOTE: If only 2,4- and 2,6-toluenediamine are to be quantified, the mobile phase program may be modified to hasten elution of the ureas derived from the isocyanates.
- 10. Inject a 10-µL aliquot of solution from step 6 or step 7b.
- 11. Measure the peak heights. Adjusted retention times for some compounds of interest are:

2,6-bisacetamidotoluene	5.2 min
1,4-bisacetamidobenzene	7.8 min
1,3-bisacetamidobenzene	9.4 min
2,4-bisacetamidotoluene	9.7 min
1-acetyl-4-(2-methoxyphenyl)piperazine	14.3 min
urea derivative of 2,6-toluene diisocyanate	17.0 min
urea derivative of 2,4-toluene diisocyanate	18.3 min

CALCULATIONS:

- 12. Using the calibration graphs, determine the mass, μg, of 2,4- and of 2,6-toluenediamine in each sample (W) and in the average media blank (B).
- 13. Calculate the concentration, C, of 2,4- and of 2,6-toluenediamine in the air volume sample, V
- (L):

$$C = \frac{(W - B)}{V}, mg/m^3.$$

EVALUATION OF METHOD: [1]

The relationship of peak height and concentration of 2,4- and 2,6-bisacetamidotoluene in methanol was found to be essentially linear over the ranges 0.05 to 141 µg/mL and 0.14 to 84 µg/mL, respectively. The time required for completion of the acetylation reaction was determined using samples equivalent to 0.7 µg of 2,4-toluenediamine and 0.9 µg of 2,6-toluenediamine in 15 mL of sampling medium. Aliquots (2 mL) were treated with 10 µL of acetic anhydride and allowed to stand 0.25 to 6 hrs before further workup and analysis. The acetylation of 2,6-toluenediamine was the slower reaction, but it appeared complete after 4 h. Sample stability was studied using solutions of 2,4- and 2,6-toluenediamine in sampling medium at levels corresponding to 0.89 and 0.74 µg per sample, respectively. The recoveries, ranging from 97% to 106%, suggested the samples were stable under the conditions of storage -- 1, 7, and 14 days at room temperature in the dark. The potential for interference from isocyanates was investigated by drawing air containing 7.3 µg each of 2,4- and 2,6-toluene diisocyanate through samples of 0.9 µg each of 2,4- and 2,6-toluenediamine in 15 mL of sampling medium. The recoveries from these samples, averaging 95%, suggested that, when compared to identical samples not treated with toluene diisocyanate, a small but statistically significant negative bias was caused by the isocyanate. Using the data from all of the recovery experiments, the relative standard deviation (S ,) for 2,4-toluenediamine (0.74 and 0.83 μ g per sample) ranged from 0.01 to 0.08 with a pooled average (S,) of 0.05 and for 2,6toluenediamine (0.89 μ g per sample) ranged from 0.02 to 0.14 with a pooled average (\ddot{S}) of 0.06.

REFERENCES:

- [1] Arnold, J. E. and A. W. Teass. Unpublished research, NIOSH (1985).
- [2] Warwick, C. J., D. A. Bagon, and C. J. Purnell. "Application of Electrochemical Detection to the measurement of Free Monomeric Aromatic and Aliphatic isocyanates in Air by High-performance Liquid Chromatography," <u>Analyst</u>, <u>106</u>: 676-685 (1981).
- [3] Holdren, M. W., C. W. Spicer, and R. M., Riggin. "Gas Phase Reaction of Toluene Diisocyanate with Water Vapor," <u>Am. Ind. Hyg. Assoc. J.</u>, <u>45</u>, 626-633 (1984).
- [4] Menzies, K. T., J. Worthington and B. Belinky. "2,4-Dinitrotoluene, 2,4-Diaminotoluene," in "Short-Term Method Development and Chemical Analysis: Final Report," Contract No. 210-89-0099, National Institute for Occupational Safety and Health, Cincinnati, OH, 213-222 (June, 1982), available as Stock No. PB-84-181-817 from NTIS, Springfield, VA 22161.
- [5] Becher, G. "Glass Capillary Columns in the Gas Chromatographic Separation of Aromatic Amines. II. Application to Samples from Workplace Atmospheres using Nitrogen-Selective Detection," J. <u>Chromatogr., 211</u>, 103-110 (1981).
- [6] Audunsson, G. and L. Mathiasson. "Simultaneous Determination of Amines and Isocyanates in Working Atmospheres by Gas-Liquid Chromatography," <u>J. Chromatogr., 261</u>, 253-264 (1983).
- [7] Skarping, G., C. Sango, and B. E. F. Smith. "Trace Analysis of Isocyanates in Industrial Atmospheres Using Gas Chromatography and Electron-Capture Detection," <u>J. Chromatogr.</u>, 208, 313-321 (1981).
- [8] Bishop, R. W., T. A. Ayers, and G. G. Esposito. "A Gas Chromatographic Procedure for the Determination of Airborne MDI and TDI," <u>Am. Ind. Hyg. Assoc. J.</u>, <u>44</u>, 151-155 (1983).
- [9] Nieminen, E. H., L. H. Saarinen, and J. T. Laakso, "Simultaneous Determination of Aromatic Isocyanates and Some Carcinogenic Amines in the Work Atmosphere by Reversed-phase Highperformance Liquid Chromatography," <u>Journal of Liquid Chromatography</u>, <u>6</u>, 453-469 (1983).
- [10] Darlene, M., L. Mathiasson, G. Skarping, C. Sango, and J. F. Sandstrom, "Trace Analysis of Airborne Aromatic Isocyanates and Related Aminoisocyanates and Diamines Using High-Performance Liquid Chromatography with Ultraviolet and Electrochemical Detection," J. Chromatogr., <u>435</u>, 469-481 (1988).
- [11] Skarping, G., L. Renman, G. Sango, L. Mathiasson, and M. Darlene, "Capillary Gas Chromatographic Method for the Determination of Complex Mixture of Isocyanates and Amines,"

<u>J. Chromatogr.</u>, <u>346</u>, 191-204 (1985).

[12] Elskamp, C. J., "Benzidine; 3,3'-Dichlorobenzidine; 2,4-Toluenediamine; 2,6-Toluenediamine. Method 65," OSHA Analytical Laboratory, Salt lake City, Utah (1987).

- [13] NIOSH [1988], <u>Registry of Toxic Effects of Chemical Substances, data base: 2,4-</u> <u>Toluenediamine.</u> D.V. Sweet, Ed., U. S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, National Institute for Occupational Safety and Health, Division of Standards Development and Technology Transfer.
- [14] <u>Ibid</u>., 2,6-Toluenediamine.

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APPENDIX A:

PURIFICATION OF 1-(2-METHOXYPHENYL)PIPERAZINE:

Place 25 g 1-(2-methoxyphenyl)piperazine (yellowish white solid) in a 250-mL beaker. Add ca. 125 mL pentane. Bring to a boil (CAUTION: Pentane is FLAMMABLE) on a water bath and allow to boil until all but a small amount of yellow oil is in solution. The 1-(2-methoxyphenyl)piperazine will melt as it is warmed in the pentane. Decant the solution into a clean beaker, cover with a watchglass, and cool in the freezer for 2 to 3 h. Collect the resulting white needles in a Buchner funnel using suction filtration and dry them in a vacuum desiccator. The crystals are hygroscopic and melt at 26 to 29 °C. Store them in an airtight container in a refrigerator.

APPENDIX B:

SYNTHESIS OF 2,4- AND 2,6-BISACETAMIDOTOLUENE:

Place 0.5 g of 2,4- or 2,6-toluenediamine in a 250-mL beaker. Add ca. 100 mL of distilled water and warm to dissolve the compound. Filter the solution, if necessary. Chill the solution in an ice bath, then slowly add 5 mL acetic anhydride and stir. After keeping the mixture at least 1 h in the ice bath, collect the solid product by suction filtration. Recrystallize the product from water by dissolving it in boiling water, filtering the hot solution, chilling the filtrate in a refrigerator, and collecting the precipitate by suction filtration. Dry the precipitate in a vacuum desiccator.

2,4-Bisacetamidotoluene recrystallizes as white needles and melts at 230 °C.

2,6-Bisacetamidotoluene recrystallizes as brownish needles and melts at ca. 318 °C.