HC	DCH <sub>2</sub> CH <sub>2</sub> OH	MW: 62.07	CAS: 107-21-1	RTECS: KW2975000	
METHOD: 5500, Issue 2		EVA	LUATION: FULL	Issue 1: 15 February 1984 Issue 2: 15 August 1993	
OSHA : NIOSH: ACGIH:	no standard Group III Pesticide C 50 ppm (1 ppm = 2.54 mg/m <sup>3</sup> @	NTP)	PROPERTIES:	liquid; MP - 13 °C; BP 198 °C; d 1.1135 g/mL @ 20 °C; VP 0.007 kPa (0.20 mm Hg) @ 20°C; explosive limits 3.2 to 15.3% in air	

SYNONYMS: 1,2-ethanediol; 1,2-Dihydroxyethane; ethylene alcohol; ethylene dihydrate

SAMPLING			MEASUREMENT	
SAMPLER:	FILTER + SORBENT		TECHNIQUE:	GAS CHROMATOGRAPHY, FID
	(glass fiber filter + silica	a gel, 520 mg/260 mg)	ANALYTE:	ethylene glycol
FLOW RATE: 0.2 L/min				
VOL-MIN: -MAX:	0.3 L 60 L		SILICA GEL DESORPTION:	1 mL 2% (v/v) 2-propanol in $H_2O$ , sonicate for 15-30 min (5 min not enough to desorb analyte)
<b>SHIPMENT:</b> filter in glass vial with 1 mL 2% (v/v) 2-propanol/H <sub>2</sub> O directly after sampling; silica gel tube sealed with plastic caps		INJECTION VOLUME: 1 µL		
		TEMPERATURE-I	NJECTION: 250 °C	
SAMPLE STABILITY:	at least 15 days (s	ilica gel) @ 25 °C [1]	-C	DETECTOR: 300 °C -COLUMN: 165 °C
FIELD BLANKS: 2 to 10 field blanks per set			CARRIER GAS:	$N_2$ or He, 30 mL/min
ACCURACY			COLUMN:	glass, 1.9 m x 2-mm; 3% Carbowax 20M on 80/100 Chromosorb 101
RANGE STUDIED:		45 to 98 mg/m <sup>3</sup> [1]	CALIBRATION:	solutions of ethylene glycol in 2% (v/v)
BIAS:		3.3% [1]		2-propanol in H <sub>2</sub> O
OVERALL PRECISION $(\hat{S}_{rT})$ :		0.084 (aerosol); 0.087 (vapor) [1]	RANGE:	0.02 to 1 mg per sample
<b>ACCURACY:</b> ± 20.4%			ESTIMATED LOD: 4 µg per sample	
			PRECISION (Ŝ,):	0.060 (filters); 0.061 (silica gel) [1]

**APPLICABILITY:** The working range is 7 to 330 mg/m  $^3$  for a 3-L air sample.

**INTERFERENCES:** A ghosting phenomenon which was described by Spitz [2] caused little or no error in measurements when samples were analyzed with standards at similar concentrations. Improved separation of ethylene glycol and other glycols can be achieved using a 30-m-Rtx-35 (0.53 mm ID; 3.0 µm film) capillary column.

**OTHER METHODS:** This method originally was designated P&CAM 338 [3] and was evaluated with a reference method which involved sampling with bubblers containing water, oxidation of ethylene glycol to formaldehyde with periodic acid and co lorimetric analysis [1].

### **REAGENTS:**

- 1. Ethylene glycol.
- 2-Propanol/water, 2% (v/v). Add 2 volumes
  2-propanol to 98 volumes freshly distilled water.
- 3. Helium or nitrogen, purified.
- 4. Hydrogen, prepurified.
- 5. Air, filtered.
- 6. Calibration stock solution, 10 mg/mL, in 2% 2-propanol/water.

# EQUIPMENT:

- Sampler: 13-mm glass fiber filter, free of binders, in filter holder (Cat. No. SX00 013 00, Millipore Corp. or equivalent) followed by glass tube, 8 cm long, 8-mm OD, 6-mm ID; two sections of 20/40 mesh silica gel (d = 0.72 g/cm<sup>3</sup>; surface area, 720 to 760 m<sup>2</sup>/g) separated by 3-mm urethane foam plug (front = 520 mg; backup = 260 mg). Filter holder and glass tube connected with two short pieces of plastic tubing. One piece (7 mm long) fits tightly around outlet of filter holder; other piece fits over first piece and inlet of glass tube.
- 2. Personal sampling pump, 0.2 L/min, with flexible connecting tubing.
- 3. Gas chromatograph, FID, integrator and column (page 5500-1).
- 4. Vials, glass, 1-mL, rubber caps.
- 5. Syringes, 10- $\mu$ L, readable to 0.1  $\mu$ L.
- 6. U-tube, glass, 25 cm x 15-mm ID.
- 7. Constant temperature bath, 75 °C.
- 8. Volumetric flasks, 10-mL

### SPECIAL PRECAUTIONS: None.

### SAMPLING:

- 1. Calibrate each personal sampling pump with a representative sampler in line.
- 2. Sample at 0.2 L/min for a total sample size of 0.3 to 60 L.
- 3. Immediately after sampling, disassemble the sampler and transfer the filter to a vial containing 1 mL 2% 2-propanol/water. Seal the vial.
- 4. Seal the ends of the silica gel tubes with plastic caps. Pack securely for shipment.

### SAMPLE PREPARATION:

- 5. Transfer front and backup sections of silica gel to separate vials. Add 1 mL 2% 2-propanol/water to each vial.
- 6. Place samples in ultrasonic bath for 15 to 30 minutes.

## CALIBRATION AND QUALITY CONTROL:

- 7. Calibrate daily with at least six working standards over the range 0.001 to 1 mg ethylene glycol per sample.
  - a. Add known amounts of calibration stock solution to 2% 2-propanol/water in 10-mL volumetric flasks and dilute to the mark.
  - b. Analyze together with samples and blanks (steps 10 through 12).
  - c. Prepare calibration graph (peak area or height vs. mg ethylene glycol).
- 8. Determine recovery from filters in the calibration range (step 7). Prepare three filters at each of three levels plus three media blanks.
  - a. Place a filter in a clean vial.

- b. Add a known quantity of ethylene glycol in 5 µL water solution to the filter.
- c. Seal the vial; ultrasonicate for 15 to 30 minutes.
- d. Prepare and analyze the filters together with working standards (steps 5, 6, and 10 through 12).
- e. Calculate recovery (ethylene glycol recovered/ethylene glycol taken).
- 9. Determine DE from silica gel for each lot of silica gel in the range of interest. Prepare three tubes at each of five levels plus three media blanks.
  - a. Draw ethylene glycol vapor with a pump at 0.2 L/min into sampler tubes with a pump from a U-tube partly immersed in a constant temperature bath at 75 °C.
    - NOTE: Values of DE for liquid and vapor spikes may be different [3].
  - b. Desorb (steps 5 and 6) and analyze (steps 10 through 12) together with working standards.
  - c. Prepare a graph of DE vs. mg ethylene glycol recovered.

### MEASUREMENT:

- 10. Set gas chromatograph according to conditions given on page 5500-1. Set flow rates of hydrogen and air according to manufacturer's instructions.
- 11. Inject sample aliquot manually using solvent flush technique or with autosampler. t  $_{r}$  = 4 min for ethylene glycol under these conditions.
- 12. Measure peak area or peak height.

## CALCULATIONS:

- 13. Determine the mass, mg (corrected for recovery or DE) of ethylene glycol found on the filter (W) and in the sample tube front (W  $_{f}$ ) and back (W  $_{b}$ ) sorbent sections, and in the average media blank filter (B), and front (B  $_{f}$ ) and back (B  $_{b}$ ) sorbent sections.
- NOTE: If  $W_{b} > W_{f}/10$ , report breakthrough and possible sample loss.
- 14. Calculate concentration, C, of ethylene glycol in the air volume sampled, V (L):

$$C = \frac{(W + W_{f} + W_{b} - B - B_{f} - B_{b}) \cdot 10^{3}}{V} , mg/m^{3}.$$

## **EVALUATION OF METHOD:**

The method was tested with spiked samplers and atmospheres generated with a diffusion cell and verified with bubblers of water followed by colorimetric analysis for 54 to 98 mg/m <sup>3</sup>. S<sub>r</sub> = 0.084 (six samples) for 6-L samples at 14 mg/m <sup>3</sup> (aerosol);  $\bar{S}_r = 0.065$  (17 samples, pooled) for 6-L samples at 45 to 84 mg/m<sup>3</sup> (vapor). Breakthrough volume (79 mg/m <sup>3</sup>, 0.2 L/min) = 261 L; recovery from filters (20 to 900 mg per sample) = 1.02; DE from silica gel (20 to 3120 µg per sample) = 0.81 to 0.87; filter storage stability = 49% of 85 mg evaporated from filters during 4 h at 24 °C, 78 µg stable on silica gel for 15 days at 25 °C. Area samples were collected side by side for 4 h at a field location by this method and a reference method (bubblers and colorimetric analysis); concentrations by this method, 0.63, 0.59, and 0.23 mg/m<sup>3</sup>, were 1.4, 0.7 and 26.5% higher than corresponding concentrations by the reference method.

### **REFERENCES:**

- [1] Tucker, S. P., and G. J. Deye. <u>Anal. Lett.</u>, 14 (A12), 959-976 (1981).
- [2] Spitz, H. D. J. Pharm. Sci., 1339-1340 (1972).
- [3] NIOSH Manual of Analytical Methods, 2nd ed., V. 7, P&CAM 338, U.S. Department of Health and Human Services, Publ. (NIOSH) 82-100 (1981).

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