

METHOD #: 58

(Issue July 1986)

TITLE: Coal Tar Pitch Volatiles (CTPV) Coke Oven Emissions (COE)
Selected Polynuclear Aromatic Hydrocarbons (PAHs)

ANALYTE:	CAS #
Coal Tar Pitch Volatiles CTPV Coke Oven Emissions COE Selected Polynuclear Aromatic Hydrocarbons PAHs	65996-93-2

INSTRUMENTATION: HPLC

Matrix: Air

Procedure: Air samples are collected by drawing known amounts of air through cassettes containing glass fiber filters (GFF). The filters are analyzed by extracting with benzene and gravimetrically determining the benzene-soluble fraction (BSF). If the BSF exceeds the appropriate PEL, then the sample is analyzed by high performance liquid chromatography (HPLC) with a fluorescence (FL) or ultraviolet (UV) detector to determine the presence of selected polynuclear aromatic hydrocarbons (PAHs).

Recommended air volume
and sampling rate: 960 L at 2.0 L/min

Special requirements: Each GFF must be transferred to a separate scintillation vial after sampling and the vial sealed with a PTFE-lined cap. Samples must be protected from direct sunlight.

Status of method: Evaluated method. This method that has been subjected to the established evaluation procedures of the Organic Methods Evaluation Branch.

Target concentrations: 0.20 mg/m³ for Coal Tar Pitch Volatiles (PEL)
0.15 mg/m³ for Coke Oven Emissions (PEL)
8.88 ug/m³ (1.22 ppm) for phenanthrene
0.79 ug/m³ (0.11 ppm) for anthracene
9.00 ug/m³ (1.09 ppm) for pyrene
3.27 ug/m³ (0.35 ppm) for chrysene
2.49 ug/m³ (0.24 ppm) for benzo(u)pyrene

Detection limits of the
overall procedure: 0.006 mg/m³ for BSF

0.427 ug/m³ (59 ppb) for phenanthrene (PHEN)
0.028 ug/m³ (4 ppb) for anthracene (ANTH)
0.260 ug/m³ (31 ppb) for pyrene (PYR)
0.073 ug/m³ (8 ppb) for chrysene (CHRY)
0.045 ug/m³ (4 ppb) for benzo(a)pyrene (BaP)

Reliable quantitation

limits: 0.034 mg/m³ for BSF
0.740 ug/m³ (100 ppb) for PHEN
0.066 ug/m³ (9 ppb) for ANTH
1.13 ug/m³ (140 ppb) for PYR
0.273 ug/m³ (29 ppb) for CHRY
0.207 ug/m³ (20 ppb) for BaP

Standard errors of estimate at the target

concentration: 8.3% for BSF
(Section 4.6.) 6.0% for PHEN
6.8% for ANTH
6.7% for PYR
6.3% for CHRY
5.8% for BaP

1.0 General Discussion

1.1 Background

1.1.1 History

Coal tar pitch volatiles (CTPV) include the fused polycyclic hydrocarbons which volatilize from the distillation residues of coal, petroleum (excluding asphalt), wood, and other organic matter (Ref. 5.1.). Coke oven emissions (COE) are the benzene-soluble fraction (BSF) of total particulate matter present during the destructive distillation or carbonization of coal for the production of coke (Ref. 5.2.). Coal tar is obtained by the distillation of bituminous coal (Ref. 5.3.). Coal tar pitch is composed almost entirely of polynuclear aromatic compounds and constitutes 48-65% of the usual grades of coal tar (Ref. 5.3.).

The purpose of this work was to evaluate the sampling and analytical method routinely used by OSHA, and to make appropriate modifications if necessary. That method required samples be collected with glass fiber filters (GFF) in three-piece polystyrene cassettes. The sealed cassettes were shipped to the laboratory at ambient temperature and upon receipt were stored in a refrigerator until analyzed. The GFFs were placed in test tubes containing benzene and sonicated for 20 min. The resulting solutions were filtered with fine fritted glass filter funnels. The GFFs were then rinsed twice with benzene and the filtered rinses combined with the original extract. The benzene extracts were concentrated to 1 mL. A 0.5-mL aliquot of each sample was taken to dryness and the BSF was determined gravimetrically. The other half of each sample was saved to be analyzed by HPLC if the

BSF was over the PEL.

Alternate samplers were not considered because the O~HA standard defines CTPV and COE as a function of those components that collected on a GFF. However, the following modifications were made to the previous procedure to reduce costs and improve the sensitivity and precision:

1. Samples are collected closed-face with a two-piece cassette containing a GFF and a backup pad. A three-piece cassette is not necessary.
2. The GFF is removed from the cassette and placed in a glass vial which is sealed with a cap containing a polytetrafluoroethylene (PTFE) liner before shipment. This increases the recovery of the analytes over the old procedure.
3. The total extraction volume is reduced from 10 mL to 3 mL. This eliminates the concentration step of the old procedure (concentration to 1 mL) and greatly improves the recovery and precision.
4. The extracted samples are filtered through pure PTFE membrane filters instead of fritted-glass filter funnels. Blank corrections, which were 30-70 ug with the old procedure, are reduced to 5-20 ug.

The modified procedure resulting from this evaluation requires that the GFFs be removed from the polystyrene cassettes before shipment and placed in sealed vials. Three milliliters of benzene are added to the sample vials and then the vials are placed in a mechanical shaker and shaken for 1 h. The resulting solutions are filtered through pure PTFE membrane filters. One and one-half milliliters of the benzene extract are taken to dryness and the BSF is determined gravimetrically. The rest of the sample is saved to be analyzed by HPLC if the BSF is over the PEL.

The selected PAHs used in this evaluation are phenanthrene (PHEN), anthracene (ANTH), pyrene (PYR), chrysene (CHRY), and benzo(a)pyrene (BaP). These compounds are analyzed by HPLC and are marker compounds to indicate the presence of PAHs. The presence of BaP, identified by GC/MS, is used to confirm the presence of CTPV or COE when the BSF exceeds the appropriate PEL.

- 1.1.2 Toxic effects (This section is for information only and should not be taken as a basis for OSHA policy.)

The following information was reported in "Occupational Health Guidelines for Chemical Hazards". (Ref. 5.4.)

Coal tar pitch volatiles (CTPV) are products of the destructive distillation of bituminous coal and contain polynuclear aromatic hydrocarbons (PNA's). These hydrocarbons sublime readily, thereby increasing the amounts of carcinogenic compounds in the working areas. Epidemiologic evidence suggests that workers intimately exposed to the products of combustion or distillation of bituminous coal are at risk of cancer at many sites. These

include cancer of the respiratory tract, kidney, bladder, and skin. In a study of coke oven workers, the level of exposure to CTPV and the length of time exposed were related to the development of cancer. Coke oven workers with the highest risk of cancer were those employed exclusively at topside jobs for 5 or more years, for whom the increased risk of dying from lung cancer was 10-fold; all coke oven workers had a 7 1/2-fold increase in risk of dying from kidney cancer. Although the causative agent or agents of the cancer in coke oven workers is unidentified, it is suspected that several PNA's in the CTPV generated during the coking process are involved. Certain industrial populations exposed to coal tar products have a demonstrated risk of skin cancer. Substances containing PNA's which may produce skin cancer also produce contact dermatitis; examples are coal tar, pitch and cutting oils. Although allergic dermatitis is readily induced by PNA's in guinea pigs, it is only rarely reported in humans from occupational contact with PNA's; these have resulted largely from therapeutic use of coal tar preparations. Components of pitch and coal tar produces cutaneous photosensitization; skin eruptions are usually limited to areas exposed to the sun or ultraviolet light. Most of the phototoxic agents will induce hypermelanosis of the skin; if chronic photodermatitis is severe and prolonged, leukoderma may occur. Some oils containing PNA's have been associated with changes of follicular and sebaceous glands which commonly take the form of acne. There is evidence that exposure to emissions at coke ovens and gas retorts may be associated with an increased occurrence of chronic bronchitis. Coal tar pitch volatiles may be associated with benzene, an agent suspected of causing leukemia and known to cause aplastic anemia.

1.1.3 Operations where exposure may occur

In 1970, there were over 13,000 coke ovens in operation in the United States. It is estimated that approximately 10,000 persons are potentially exposed to COE. (Ref. 5.5.)

Coal tar pitch is used in metal and foundry operations, electrical equipment installations, pipe coating operations, and at construction sites. About 145,000 people are potentially exposed to CTPV. (Ref. 5.6.)

The PAHs that were studied in this evaluation have been found in many substances. These include coke oven emissions, coal tar pitch, creosote, exhaust of internal combustion engines, and cooked meats. Benzo(a)pyrene and chrysene have also been isolated from cigarette smoke. (Refs. 5.5.-5.7.)

1.1.4 Physical properties (Ref. 5.8.)

Phenanthrene
CAS no.: 85-01-8
MW: 178.22
bp: 340-C at 760 mm Hg
mp: 100-C
color: white crystals
structure: Figure 1.1.4.

Anthracene

CAS no.: 120-12-7
MW: 178.22
bp: 342-C at 760 mm Hg
mp: 218-C
color: colorless crystals
structure: Figure 1.1.4.

Pyrene

CAS no.: 129-00-0
MW: 202.24
bp: 404-C at 760 mm Hg
mp: 156-C
color: colorless crystals
synonyms: benzo(def)phenanthrene
structure: Figure 1.1.4.

Chrysene

CAS no.: 218-01-9
MW: 228.28
bp: 448-C at 760 mm Hg
mp: 254-C
color: white crystals
synonyms: 1,2-benzophenanthrene;
benzo(a)phenanthrene
structure: Figure 1.1.4.

Benzo(alpha)pyrene

CAS no.: 50-32-8
MW: 252.30
bp: 311-C at 10 mm Hg
mp: 179-C
color: yellow needles
synonyms: 3,4-benzopyrene; 6,7-benzopyrene
structure: Figure 1.1.4.

Benzene-soluble fraction (The sum of those components collected on a GFF and soluble in benzene.)

color: brownish-yellow to black tar

1.2 Limit defining parameters (The analyte air concentrations listed throughout this method are based on an air volume of 960 L and a solvent extraction volume of 3 mL. Air concentrations listed in ppm are referenced to 25-C and 760 mm Hg.)

1.2.1 Detection limits of the analytical procedure

1.2.1.1 Benzene-soluble fraction

The detection limit of the analytical procedure is 6 ug per sample and is based on the precision of the analytical balance used. This is the weight which corresponds to twice the standard deviation of the precision data for a 50-mg weight, which is the approximate weight of an average PTFE

cup. (Sections 4.1.1. and 4.4.1.) The detection limit also takes into account the dilution factor of 2.

1.2.1.2 Selected PAHs

The detection limits of the analytical procedure are listed below. These are the amounts of analyte which will give a peak whose height is about five times the height of the baseline noise. (Section 4.1.2.)

Table 1.2.1.2 Analytical Detection Limits

compound	ng/injection	detector*
PHEN	0.132	UV(254 nm)
PHEN	0.910	FL
ANTH	0.090	FL
PYR	0.960	FL
CHRY	0.386	FL
BoP	0.175	FL

* Fluorescence was more sensitive than UV for each PAH except PHEN

1.2.2 Detection limits of the overall procedure

The detection limits of the overall procedure are listed below. These are the amounts of analyte, determined from Figures 4.2.1.-4.2.6., which when spiked onto the sampling device would allow recovery of an amount of analyte equivalent to the detection limits of the analytical procedure. (Section 4.2.)

Table 1.2.2 Detection Limits of the Overall Procedure

	BSF	PHEN	ANTH	PYR	CHRY	BoP
ug/sample	6	0.41	0.027	0.25	0.070	0.043
ug/m3	6	0.43	0.028	0.26	0.073	0.045
ppb	--	59	4	31	8	4

1.2.3 Reliable quantitation limits

The reliable quantitation limits are listed below. These are the smallest amounts of analyte which can be quantitated within the requirements of a recovery of at least 75% and a precision (+/- 1.96 SD) of +/- 25% or better. (Section 4.3.)

Table 1.2.3 Reliable Quantitation Limits

	BSF	PHEN	ANTH	PYR	CHRY	BoP
ug/sample	33.1	0.71	0.064	1.08	0.262	0.199
ug/m3	34.5	0.74	0.066	1.13	0.273	0.207
ppb	--	100	9	140	29	20

The reliable quantitation limit and detection limits reported in

the method are based upon optimization of the instrument for the smallest possible amount of analyte. When the target concentration of an analyte is exceptionally higher than these limits, they may not be attainable at the routine operating parameters.

1.2.4 Sensitivities

The sensitivities of the analytical procedure over a concentration range representing about 0.5 to 2 times the target concentrations are listed below. These values were determined by the slope of the calibration curves. (Section 4.4.) The sensitivity will vary with the particular instrument used in the analysis. The values listed were obtained using an FL detector.

Table 1.2.4 Sensitivities of Selected PAHs

compound	area counts/(ug/mL)
PHEN	19000
ANTH	178000
PYR	21100
CHRY	58900
BoP	125000

1.2.5 Recoveries

The recovery of analytes from samples stored in vials used in the 15-day storage test remained above the percentages listed below. (Section 4.6.) The recovery of the analytes from the collection medium during storage must be 75% or greater.

Table 1.2.5 Recoveries from Ambient Storage

compound	% recovery
BSF	89.4
PHEN	92.2
ANTH	90.7
PYR	86.9
CHRY	96.2
BoP	99.9

1.2.6 Precisions (analytical procedure)

The pooled coefficients of variation obtained from replicate determinations of analytical standards at about 0.5 to 2 times the target concentration are shown below. The values were obtained using an FL detector. (Section 4.4.)

Table 1.2.6 Analytical Precision

compound	CV
PHEN	0.0092
ANTH	0.0051

PYR	0.0128
CHRY	0.0094
BoP	0.0150

1.2.7 Precisions (overall procedure)

The precisions at the 95% confidence level for the 15-day ambient storage tests are listed below. (Section 4.6.) These include an additional +/- 5% for sampling error. The overall procedure must provide results at the target concentration that are +/- 25% or better at the 95% confidence level.

Table 1.2.7 Precision of the Overall Procedure

compound	percent
BSF	16.2
PHEN	11.8
ANTH	13.4
PYR	13.0
CHRY	12.3
BoP	11.3

1.2.8 Reproducibilities

Six samples, spiked with coal tar by liquid injection, and a draft copy of this procedure were given to a chemist unassociated with this evaluation. The samples were analyzed after 21 days of storage at about 22-C. Another set of six samples, spiked with PAHs by liquid injection, and a draft copy of this procedure were given to another chemist unassociated with this evaluation. The samples were analyzed after 3 days of storage at about 22-C. The average recoveries are listed below. (Section 4.7.)

Table 1.2.8 Reproducibilities

compound	mean	SD
BSF	94.2	5.4
PHEN	98.0	3.4
ANTH	90.4	2.4
PYR	101.4	3.4
CHRY	98.7	2.7
BoP	100.6	3.0

1.3 Advantages

1.3.1 Recovery of the analytes is improved by placing the GFF in sealed glass vials before shipment.

1.3.2 The amount of benzene required for each sample is reduced from 10 mL to 3 mL per sample. This reduces the exposure to a suspected human carcinogen.

1.3.3 The reliable quantitation limits are much lower than those of

the previously used procedure.

1.3.4 The use of pure PTFE membrane filters, instead of fritted glass filter funnels, lowers the blank correction and provides much better precision.

1.3.5 The amount of time samples spend in the nitrogen evaporator for the previous procedure is eliminated, a savings of about 2 h.

1.4 Disadvantages

The GFF must be transferred from the cassette to a scintillation vial by the industrial hygienist.

2.0 Sampling Procedure

2.1 Apparatus

2.1.1 A personal sampling pump that can be calibrated to within +/- 5% of the recommended flow rate with the sampling device in line.

2.1.2 A two-piece cassette containing a glass fiber filter is the sampling device.

2.1.3 Forceps to transfer the GFF to a scintillation vial.

2.1.4 Scintillation vials with PTFE-lined caps.

2.1.5 Aluminum foil or an opaque container to protect collected samples from light.

2.2 Reagents

No sampling reagents are required.

2.3 Sampling technique

2.3.1 Attach the cassette to the sampling pump with flexible, plastic tubing so that the GFF in the sampling cassette is exposed directly to the atmosphere. Do not place any tubing in front of the sampler. The sampler should be attached vertically in the worker's breathing zone in such a manner that it does not impede work performance. The sampling device should be protected from direct sunlight (Ref. 5.9.).

2.3.2 After sampling for the appropriate time, remove the sampling device and install the two plastic plugs in the open ends of the cassette.

2.3.3 As soon as it is conveniently possible, but before the sample is shipped, fold the filter into quarters (sampling surface inside) and insert it into a scintillation vial (Figure 2.3.3.). Always handle the GFF with clean forceps. To avoid losing any particulate material, the inside of the cassette should be wiped with the folded filter. Install a cap that has a PTFE liner, not a Poly-seal cap. Wrap each vial in aluminum foil or place it in an opaque container to protect the sample from light.

2.3.4 Wrap each sample end-to-end with an OSHA seal (Form 21).

2.3.5 Submit at least one blank with each set of samples. The blank should be handled the same as the other samples except that no air is drawn through it.

2.4. Extraction efficiencies

The average extraction efficiencies of the analytes are listed below.

The target concentrations were used for this determination. (Section

4.5.)

Table 2.4. Extraction Efficiency from GFF

compound	percent
BSF	100.3
PHEN	105.9
ANTH	112.5
PYR	101.4
CHRY	107.5
BoP	108.7

2.5 Recommended air volume and sampling rate

2.5.1 The recommended air volume is 960 L.

2.5.2 The recommended air sampling rate is 2.0 L/min.

2.6 Interferences (sampling)

Suspected interferences should be reported to the laboratory with submitted samples.

2.7 Safety precautions (sampling)

The sampling equipment should be attached to the worker in such a manner that it will not interfere with work performance or safety.

3.0 Analytical Procedure

3.1 Apparatus

3.1.1 Benzene-soluble fraction

3.1.1.1 A calibrated microbalance capable of determining a weight to the nearest microgram. A Mettler M3-03 balance with a data transfer recorder was used in this evaluation.

3.1.1.2 Thirteen-millimeter stainless steel filter holder with a female Luer-Lok fitting.

3.1.1.3 Thirteen-millimeter pure PTFE membrane filters with 5-um pores.

3.1.1.4 Two-milliliter PTFE cups, Cahn Scientific.

3.1.1.5 Two-milliliter disposable pipets.

3.1.1.6 Ten-milliliter glass syringe barrels with male Luer-Lok fittings.

3.1.1.7 Disposable culture tubes (13 x 100 mm).

3.1.1.8 Vacuum oven.

3.1.1.9 Mechanical shaker.

3.1.1.10 Forceps.

3.1.2 Selected PAHs

3.1.2.1 High performance liquid chromatograph equipped with a fluorescence (FL) or an ultraviolet (UV) detector, manual or automatic injector, gradient flow programmer and chart recorder. A Waters M-6000A pump, Waters WISP 710B

autosampler, Waters 660 solvent programmer, Schoeffel 970 FL detector, Waters 440 UV detector, and a Houston dual pen recorder were used in this evaluation.

- 3.1.2.2 HPLC column capable of separating PAHs from any interferences. A 25-cm x 4.6-mm i.d. Dupont Zorbax ODS (6 μ m) column was used during this evaluation.
- 3.1.2.3 An electronic integrator, or some other suitable method of measuring detector response.
- 3.1.2.4 Vials, 4-mL with PTFE-lined caps.
- 3.1.2.5 Volumetric flasks, pipets, and syringes.

3.2 Reagents

- 3.2.1 Acetonitrile (ACN), HPLC grade.
- 3.2.2 Water, HPLC grade. A Millipore Milli-Q system was used to prepare the water for this evaluation.
- 3.2.3 Benzene, HPLC grade.
- 3.2.4 Nitrogen gas.
- 3.2.5 Phenanthrene (PHEN).
- 3.2.6 Anthracene (ANTH).
- 3.2.7 Pyrene (PYR).
- 3.2.8 Chrysene (CHRY).
- 3.2.9 Benzo(a)pyrene (BoP).
- 3.2.10 Tetrahydrofuran (THF), HPLC grade.

3.3 Standard preparation for selected PAHs

A stock standard solution is prepared by dissolving the PAHs in benzene. All dilutions of the stock solutions are made with benzene to arrive at the working range.

3.4 Sample preparation

- 3.4.1 Benzene-soluble fraction (CAUTION - All work with benzene must be done in a fume hood.)
 - 3.4.1.1 Clean the PTFE Cups by sonicating them in THF for a few minutes, and rinsing them twice with clean THF. Place the cups into a numbered holder. The cups are placed in a preheated oven (40-C under about 20 in. Hg vacuum) for 1 h. The cups are allowed to cool to room temperature and weighed to the nearest microgram. Handle the cups with clean, dry forceps.
 - 3.4.1.2 Pipet 3.0 mL of benzene to each scintillation vial containing the sample filter.
 - 3.4.1.3 Shake the vials for 60 min.
 - 3.4.1.4 Insert a 13-mm pure PTFE membrane filter (5- μ m) into the stainless steel holder and attach the holder to a syringe barrel. Add about 3 mL of benzene to the syringe and push the benzene through the filtering unit with nitrogen to check for leaks. A rubber stopper is used on the nitrogen line to pressurize the syringe barrel to 10 psig. Dry the filter by allowing the nitrogen to pass through the filter for 30 s.
 - 3.4.1.5 Transfer the benzene extract from the vial into the syringe barrel, one sample per syringe. If the vial contains

a considerable amount of particulate material, decant the extract into the syringe barrel. Push the benzene extract through the filters into the disposable culture tube (13 x 100 mm) with nitrogen gas.

3.4.1.6 Pipet 1.5 mL of the benzene extract to a tared PTFE cup.

3.4.1.7 Place the PTFE cups in a preheated oven (40-C under about 15 in. Hg vacuum). Provide some air flow in the oven to sweep benzene vapor out of the oven. Heat the cups for about 3 to 4 h. Close the vent valve for the last hour of the drying period.

3.4.1.8 Remove the PTFE cups from the oven and allow them to cool to room temperature. Weigh the cups to the nearest microgram.

3.4.2 Selected PAHs

Transfer the remaining benzene solution from the culture tube to a vial and seal with a PTFE-lined cap. This fraction of the sample will be analyzed for PAHs if the concentration of the BSF is equal to or greater than the PEL.

3.5 Analysis

3.5.1 Reverse phase HPLC conditions

column:	25-cm x 4.6-mm i.d. stainless steel column packed with 6-um DuPont Zorbax ODS
mobile phase:	85:15 ACN/water (v/v)
flow rate:	1.0 mL/min for 5 min, Curve 10 (flow program) for 5 min to 1.5 mL/min, then hold for 10 min
FL detector:	254 nm excitation 370 nm emission
UV detector:	254 nm
injection size:	10 uL
retention time:	7-18 min
chromatogram:	Figure 3.5.1.

3.5.2 An external standard procedure is used to prepare a calibration curve using at least 2 stock solutions from which dilutions are made. The calibration curve is prepared daily. The samples are bracketed with analytical standards.

3.6 Interferences (analytical)

3.6.1 Benzene-soluble fraction

3.6.1.1 Any compound that is soluble in benzene and is not normally found in coal tar pitch volatiles or coke oven emissions is an interference. Anything that falls into or adheres to the PTFE cups during the time between weighings will give high results.

3.6.1.2 It has been reported that mineral oil is an interference with the BSF determination in the aluminum industry (Ref. 5.10.). The problem of separating mineral oil from the BSF

was not addressed in this evaluation but a status report from the Aluminum Association Health Committee showed that the ANCAL IATROSCAN TH-10 could quantitate the amounts of oil and BSF separately. This is done by using a special thin-layer chromatography rod that is analyzed by a flame ionization detector after a separation has been completed with the rod. The report indicated good agreement between the total amount of oil and BSF by the Iatroscan method and weight of BSF determined gravimetrically. (Ref. 5.11.)

3.6.2 Selected PAHs

- 3.6.2.1 Any compound having a similar retention time as the PAHs is a potential interference. Generally, chromatographic conditions can be altered to separate an interference from the analyte.
- 3.6.2.2 Retention time on a single column is not proof of chemical identity. Analysis by an alternate HPLC column, absorbance response ratioing, and mass spectrometry are additional means of identification.

3.7 Calculations

3.7.1 Benzene-soluble fraction

The concentration in ug/m³ of BSF present in a sample is determined from the two weighings (in micrograms) of the PTFE cup. The factor "2" in the equation compensates for the fact that only 1/2 of the sample was used for the gravimetric procedure.

$$\text{ug/m}^3 = \frac{2[(\text{final wt} - \text{tare wt}) - (\text{blank final wt} - \text{tare wt})]}{(\text{air volume, m}^3)}$$

3.7.2 Selected PAHs

The concentration in ug/mL of the PAHs present in a sample is determined from the detector response of the analytes. Comparison of sample response with a least squares curve fit for standards allows the analyst to determine the concentration of the PAHs in ug/mL for the sample. Since the total sample volume was 3 mL, the results in ug/m³ of air are expressed by the following equation:

$$\text{ug/m}^3 = 3 \text{ mL}(\text{ug/mL})/[(\text{air vol., m}^3)(\text{extrac. effic.})]$$

This value can be converted to an equivalent concentration in parts per million with the following equation:

$$\text{ppm} = (\text{mg/m}^3)(24.46)/\text{MW}$$

where

24.46 = molar volume at 25-C and 760 mm Hg

MW = molecular weight of PAH

3.8 Safety precautions (analytical)

- 3.8.1 Avoid exposure to all standards.
- 3.8.2 Avoid skin contact with all solvents.
- 3.8.3 Wear safety glasses at all times.
- 3.8.4 All work with benzene is to be performed in a fume hood. Benzene is a suspected human carcinogen.

4.0 Backup Data

4.1 Detection limit of the analytical procedure

4.1.1 Benzene-soluble fraction

The detection limit of the analytical procedure is 6 ug per sample. This is the weight which corresponds to twice the standard deviation of the precision data for a 50-mg weight, the weight of an average PTFE cup. The data presented in Table 4.4.1. are applicable only to the Mettler M3-03 balance used in this evaluation. The detection limit also takes into account the dilution factor of 2.

4.1.2 Selected PAHs

The detection limits of the analytical procedure are listed below and were determined by injecting 10 uL of a standard. These amounts produced peaks whose heights were about 5 times the height of the baseline noise. The injection volume recommended in the analytical procedure (10 uL) was used in the determination of the detection limits for the analytical procedure. (Figures 4.1.2.1.-4.1.2.5. show chromatographs obtained with the FL detector.)

Table 4.1.2 Analytical Detection Limits

compound	ug/mL	ng/injection	detector
PHEN	0.0132	0.132	UV (254 nm)
PHEN	0.0910	0.910	FL
ANTH	0.0090	0.090	FL
PYR	0.0960	0.960	FL
CHRY	0.0386	0.386	FL
BoP	0.0175	0.175	FL

4.2 Detection limit of the overall procedure

The detection limits of the overall procedure are listed in Table 4.2. The values were determined graphically (Figures 4.2.1.-4.2.6.) by plotting amount spiked versus amount recovered and determining the amount that corresponds to the analytical detection limit. The values listed are based on an FL detector.

Table 4.2 Detection Limits of the Overall Procedure

	BSF	PHEN	ANTH	PYR	CHRY	BoP
ug/sample	6	0.41	0.027	0.25	0.070	0.043
ug/m3	6	0.43	0.028	0.26	0.073	0.045
ppb	--	59	4	31	8	4

4.3 Reliable quantitation limit

4.3.1 Benzene-soluble fraction

The reliable quantitation limit is 33.1 ug (34.5 ug/m³) of BSF per sample. Seven samples were prepared by injecting 4 uL of a coal tar solution (8.28 mg/mL) onto GFFs. The samples were analyzed the same day and the average results are reported in Table 4.3.1.

Table 4.3.1 Reliable Quantitation Limit

Sample	% Recovery
1	99.7
2	93.7
3	86.6
4	99.7
5	105.7
6	81.6
7	87.6

X = 93.5
SD = 8.7 1.96 SD = 17.1

4.3.2 Selected PAHs

The reliable quantitation limits are listed below. Six samples were prepared by injecting several microliters of a benzene solution containing PAHs onto GFFs. The samples were analyzed the same day.

Table 4.3.2 Reliable Quantitation Limit

analyte	PHEN	ANTH	PYR	CHRY	BoP
spike (ug)	0.71	0.064	1.08	0.262	0.199
(ug/m ³)	0.74	0.066	1.13	0.273	0.207
(ppb)	100	9	140	29	20

% recovery	94.7	90.1	91.1	93.3	97.0
	92.7	91.2	102.0	96.3	105.4
	91.1	89.4	92.2	97.5	102.5
	89.9	86.4	93.9	94.9	98.6
	91.0	87.0	82.3	93.8	99.3
	97.9	87.0	86.9	97.0	95.7
X	92.9	88.5	91.4	95.5	99.8
SD	3.0	2.0	6.7	1.7	3.6
1.96 SD	5.8	3.9	13.1	3.4	7.1

4.4 Sensitivity and precision (analytical method only) 4.4.1. Precision

data for the benzene-soluble fraction

The following data were obtained from multiple weighings of calibration weights that are approximately 0.5 to 2 times the nominal weight of a PTFE cup. This was done to establish the precision of the analytical balance.

Table 4.4.1 Precision Data

	25 mg	50 mg	100 mg			
	25.005	49.991	99.998			
	25.003	49.990	100.001			
	25.007	49.993	100.001	25.007	49.993	100.001
	25.005	49.993	100.002			
	25.006	49.994	100.002			
	25.005	49.992	100.000			
	25.006	49.992	100.000			
	25.008	49.994	100.000			
	25.006	49.994	100.003			
X	25.006	49.993	100.001			
SD	0.0014	0.0015	0.0013			
CV	0.00006	0.00003	0.00001			

4.4.2 Sensitivity and precision data for selected PAHs

The following data were obtained from multiple injections of analytical standards. This data was used to establish calibration curves for each analyte from which the sensitivity was determined. The data are also presented graphically in Figures 4.4.2.1.-4.4.2.5.

Table 4.4.2.1 Precision and Sensitivity Data Approximately 0.5x Target Concentration

analyte ug/mL	PHEN	ANTH	PYR	CHRY	BaP
	2.49	0.255	2.94	1.27	0.525
areas	45900.5	51249.6	62246.7	75163.5	66750.5
	47374.6	51970.1	65309.6	77086.5	69435.0
	47183.4	52000.0	64947.2	77164.0	68508.0
	46965.1	51575.7	65054.5	77073.2	68420.0
	46142.1	51108.6	63987.1	75900.5	67441.5
	46512.1	51627.2	64048.0	76050.7	67287.0
X	46679.6	51588.5	64265.5	76406.4	67973.7
SD	590.3	363.8	1129.4	825.6	987.0
CV	0.0126	0.0071	0.0176	0.0108	0.0145

Table 4.4.2.2 Precision and Sensitivity Data Approximately 1x Target Concentration

analyte	PHEN	ANTH	PYR	CHRY	BoP
ug/mL	4.98	0.51	5.88	2.54	1.05

areas	89773.1	103477	126795	151961	136383
	89874.4	103385	127081	152486	136615
	89365.4	103311	126379	151748	135617
	89247.6	103251	125730	150675	134451
	88542.6	102573	125370	149400	134111
	89070.5	103281	126281	150541	134593
X	89312.3	103213	126273	151135	135295
SD	487.1	324	630	1136	1061
CV	0.0055	0.0031	0.0051	0.0075	0.0078

Table 4.4.2.3 Precision and Sensitivity Data Approximately 2x Target Concentration

analyte	PHEN	ANTH	PYR	CHRY	BoP
ug/mL	9.71	0.99	11.76	5.08	2.10

areas	184607	182281	248424	299267	262309
	180064	184202	247112	297709	260233
	182561	183493	246260	297519	259748
	182924	183448	252779	302237	271134
	182992	183215	253143	303651	269743
	183198	184599	252296	303771	270109
X	182724	183540	250002	300692	265546
SD	1482	808	3088	2885	5329
CV	0.0081	0.0044	0.0124	0.0096	0.0201

Table 4.4.2.4 The Pooled Coefficients of Variation

PHEN	ANTH	PYR	CHRY	BoP

0.0092	0.0051	0.0128	0.0094	0.0150

4.5 Extraction efficiency

4.5.1 Benzene-soluble fraction

The following data represent the analysis of GFFs that were liquid spiked with coal tar solution prepared by the procedure in Section 4.8. at the target concentration (207 ug/GFF). These data only show that compounds derived from the specially prepared coal tar pitch solution can be extracted from a GFF. Since the BSF is a collection of many compounds, the extraction efficiency is not applied to the calculations. The PTFE cups were reweighed 24 h later and the results were still valid.

Table 4.5.1 Extraction Efficiency of Benzene-solubles

	first day	24 h later
percent recovered	98.1	102.9
	98.1	101.0
	100.0	100.0
	100.0	99.0
	97.1	99.0
	105.8	106.8
	102.9	107.7
X	100.3	102.3
SD	3.1	3.6

4.5.2 Selected PAHs

The data listed below represent the results of the analysis of GFFs that were liquid spiked with PAHs at the target concentration. These samples were allowed to dry and then extracted with benzene and analyzed the same day. The samples were reanalyzed 24 hr later and found to be stable (Table 4.5.2.2.).

Table 4.5.2.1 Extraction Efficiency of Selected PAHs

analyte ug/sample	PHEN 8.5	ANTH 0.76	PYR 8.6	CHRY 3.1	BaP 2.4
% recovery	107.6	117.7	106.3	111.6	110.9
	108.9	117.0	110.0	112.0	113.2
	104.8	110.0	100.9	105.2	105.9
	104.6	109.9	96.6	103.8	105.2
	106.8	112.0	98.7	107.9	109.3
	106.0	113.0	100.3	108.0	110.4
	104.4	111.5	101.0	106.7	108.2
	104.0	108.7	97.0	104.7	106.5
X	105.9	112.5	101.4	107.5	108.7
SD	1.8	3.3	4.6	3.1	2.8

Table 4.5.2.2 Extraction Efficiencies 24 Hours Later

analyte ug/sample	PHEN 8.5	ANTH 0.76	PYR 8.6	CHRY 3.1	BaP 2.4
% recovery	115.9	121.3	118.4	119.2	122.3
	114.5	119.0	117.7	119.7	120.3
	110.9	116.4	111.4	112.9	118.9
	111.5	117.8	110.3	115.0	119.5
	108.6	116.6	111.2	113.7	115.8
	106.3	109.5	102.3	106.9	112.5
	108.7	110.4	104.7	109.7	110.1
	107.3	111.3	102.9	107.6	111.7

X	110.5	115.3	109.9	113.1	116.4
SD	3.4	4.4	6.2	4.9	4.5

4.6 Storage data

Storage samples were generated by liquid spiking 36 GFFs with coal tar and another 36 GFFs with PAHs. All of the spiked GFFs were stored in sealed glass vials. One-half of the vials were stored in a freezer at -20-C and the other half were stored in a closed drawer at ambient temperature (about 22-C). The results (percent recovery versus storage time) are given in Tables 4.6.2.-4.6.3. and shown graphically in Figures 4.6.1.-4.6.12.

Table 4.6.1 Amount Spiked (ug/GFF)

BSF	PHEN	ANTH	PRY	CHRY	BaP
207	8.5	0.76	8.6	3.1	2.4

Table 4.6.2 Ambient Storage Test (% recovery)

day	BSF	PHEN	ANTH	PYR	CHRY	BaP
0	100.5	108.8	113.0	105.9	108.8	110.2
	89.9	107.8	113.4	105.4	108.3	109.5
	90.8	102.4	108.5	100.3	105.0	105.0
	91.8	103.7	108.5	102.6	105.3	110.2
	78.3	100.8	110.4	101.1	104.4	105.5
	88.9	102.9	109.6	100.1	102.8	107.3
3	90.8	98.5	104.3	91.3	97.9	101.4
	99.5	97.5	103.2	95.3	100.3	102.7
	73.4	98.3	100.2	91.5	98.8	100.6
6	87.4	99.6	99.3	99.4	102.8	105.3
	90.3	104.1	104.7	101.1	108.7	108.9
	87.4	103.0	105.9	105.3	110.5	108.4
9	86.5	94.2	94.6	90.4	102.0	104.0
	91.3	101.2	104.6	91.5	101.4	103.5
	90.3	97.5	98.4	91.5	99.2	101.5
12	100.5	88.1	84.3	82.5	92.4	97.3
	87.0	93.1	88.9	93.2	96.6	104.6
	93.7	91.7	88.7	89.6	96.7	101.0
15	91.8	92.0	93.1	85.5	94.4	97.3
	85.0	94.9	99.0	89.0	100.1	101.9
	85.0	95.2	95.3	88.8	95.3	100.0

Table 4.6.3 Refrigerated Storage Test (% recovery)

day	BSF	PHEN	ANTH	PYR	CHRY	BaP
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O	100.5	108.8	113.0	105.9	108.8	110.2
	89.9	107.8	113.4	105.4	108.3	109.5
	90.8	102.4	108.5	100.3	105.0	105.0
	91.8	103.7	108.5	102.6	105.3	110.2
	78.3	100.8	110.4	101.1	104.4	105.5
	88.9	102.9	109.6	100.1	102.8	107.3
3	86.0	98.9	103.3	92.7	100.7	104.5
	84.1	99.0	102.6	95.9	98.9	102.3
	92.8	98.3	101.3	92.8	99.0	104.1
6	90.3	105.0	110.6	100.2	105.0	107.5
	92.3	101.7	106.6	95.5	102.9	106.1
	98.1	99.3	104.4	95.6	102.2	105.6
9	88.4	97.6	100.3	91.3	99.3	101.0
	86.5	95.4	99.9	88.1	98.8	101.3
	93.2	95.4	97.5	87.0	96.1	98.6
12	94.7	95.8	96.7	89.6	96.0	107.6
	92.8	96.3	99.4	96.0	99.6	103.2
	108.2	98.3	99.0	90.1	95.4	101.8
15	90.8	96.5	100.6	94.8	100.6	97.7
	90.8	95.2	101.1	90.9	98.9	100.7
	84.1	101.6	105.2	98.5	102.8	107.9

4.7 Reproducibility data

Six samples, spiked with coal tar by liquid injection, and a draft copy of this procedure were given to a chemist unassociated with this evaluation. The samples were analyzed after 21 days of storage at about 22-C. Another set of six samples, spiked with PAHs by liquid injection, and a draft copy of this procedure were given to another chemist unassociated with this evaluation. The samples were analyzed after 3 days of storage at about 22-C. All the results are corrected for extraction efficiency except for the BSF data and are listed below.

Table 4.7 Reproducibility Results (percent of theoretical amount)

	BSF	PHEN	ANTH	PYR	CHRY	BaP
	101.4	99.1	91.7	105.4	102.2	105.0
	90.8	97.8	93.0	104.8	101.6	103.4
	87.0	91.8	86.1	96.5	95.5	97.5
	92.8	101.8	90.9	101.7	98.4	100.4
	99.5	99.8	90.9	100.4	98.1	99.2
	93.9	97.8	89.6	99.3	96.5	98.3
X	94.2	98.0	90.4	101.4	98.7	100.6
SD	5.4	3.4	2.4	3.4	2.7	3.0

4.8 Preparation of benzene-soluble standards used in evaluation

In this evaluation three different types of coal tar pitch were chosen at random from a collection of several confirmed coal tar pitch materials. Twenty grams of each pitch were placed in beakers containing 100 mL of benzene and sonicated for 1.5 h. The solutions were then combined and filtered twice with a fine fritted-glass filter funnel. The resultant solution was then passed through a glass fiber filter. The solution was concentrated with a stream of dry nitrogen and the gooey tar was placed in a heated oven (60-C under 20 in. Hg vacuum) for 4 h. A portion of the "dried" tar was used to prepare a stock solution in benzene. This was used to spike filters approximating a Coal Tar Pitch Volatile sample.

5.0 References

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