



# Assay Requirements

- Sensitive, quantitative, reproducible, high throughput and have correlative value
- Optimized and validated to meet GCLP requirements for human clinical trials
- Reagents need to be standardized and traceable



# Neutralizing Ab Assays

New technologies have allowed dramatic improvements in assay performance and validation

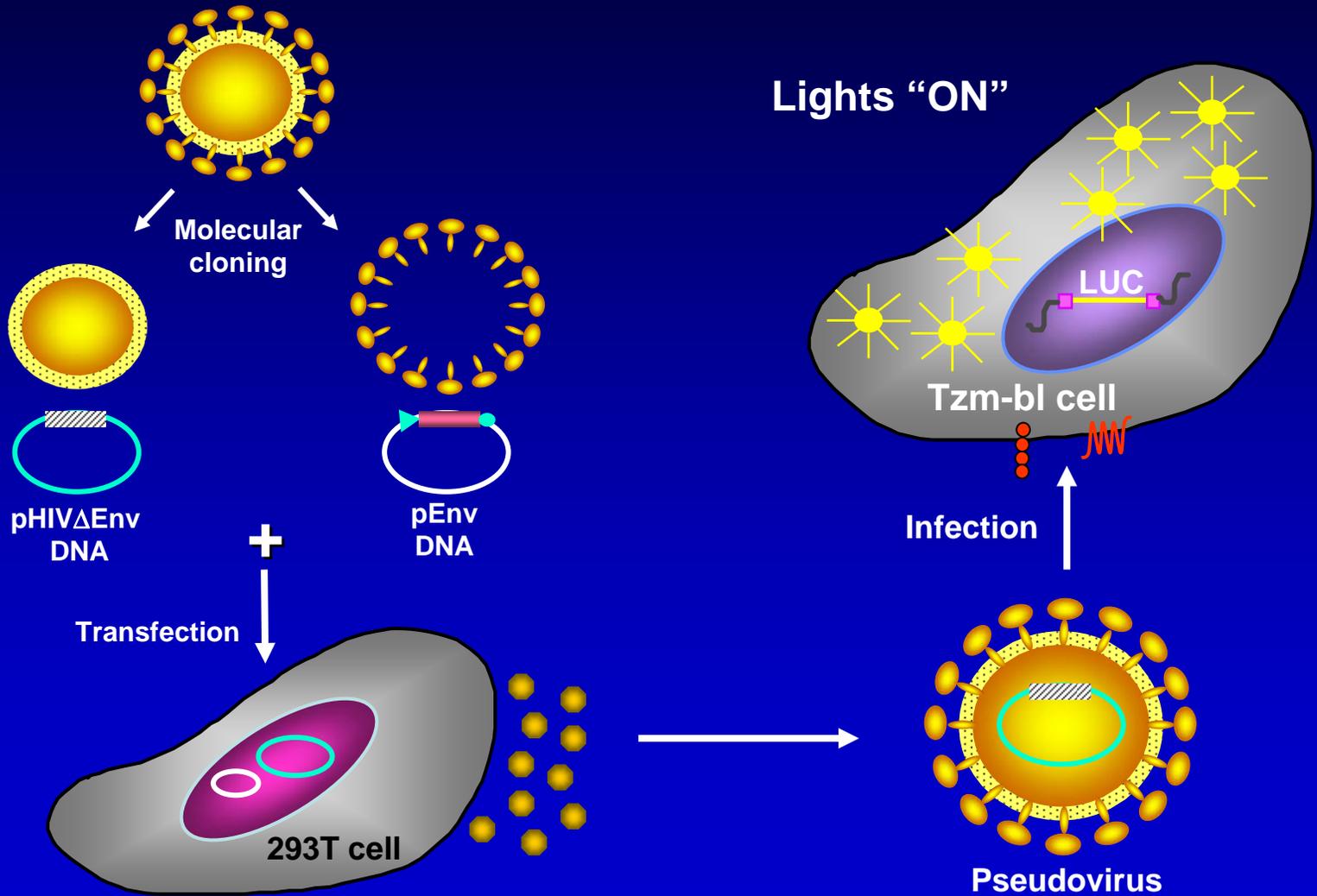


# Luciferase Reporter Gene Assay in TZM-bl Cells Based on Single-Round Infection with Molecularly Cloned Env-Pseudotyped Viruses

- TZM-bl (JC53-bl) is a genetically engineered HeLa cell line that expresses CD4, CXCR4 and CCR5 and contains Tat-inducible Luc and  $\beta$ -Gal reporter genes:
  - High success rate in single-round infections
  - Enhanced throughput capacity (2-day assay)
  - Increased precision (can measure 50% neutralization)
  - Improved level of standardization (e.g., stable cell line, clonal viruses)
  - **Recently optimized and validated**

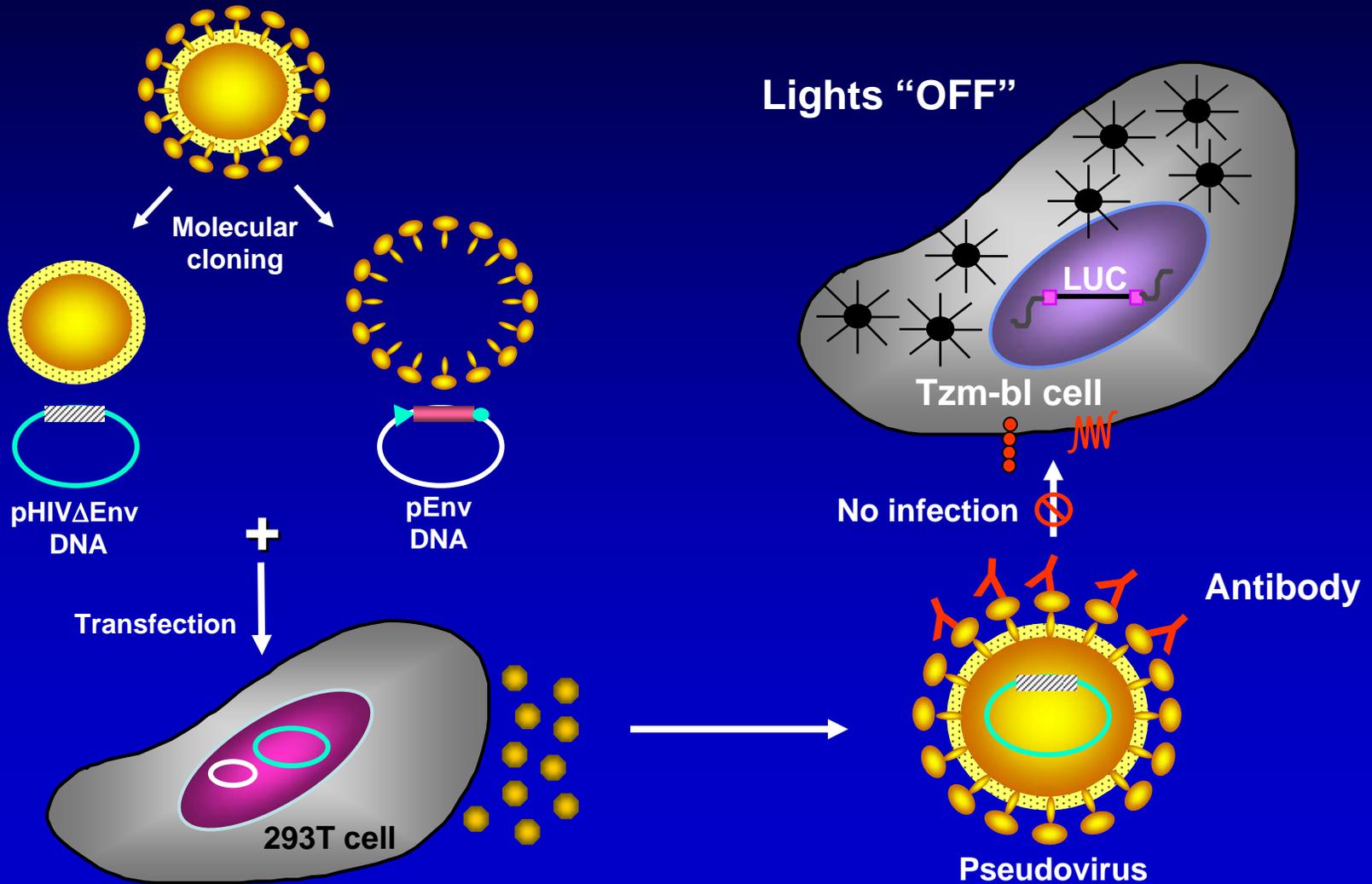


# SEQUENTIAL EVENTS IN DETECTING NEUTRALIZATION OF ENV-PSEUDOVIRUSES IN TZM-BL CELLS



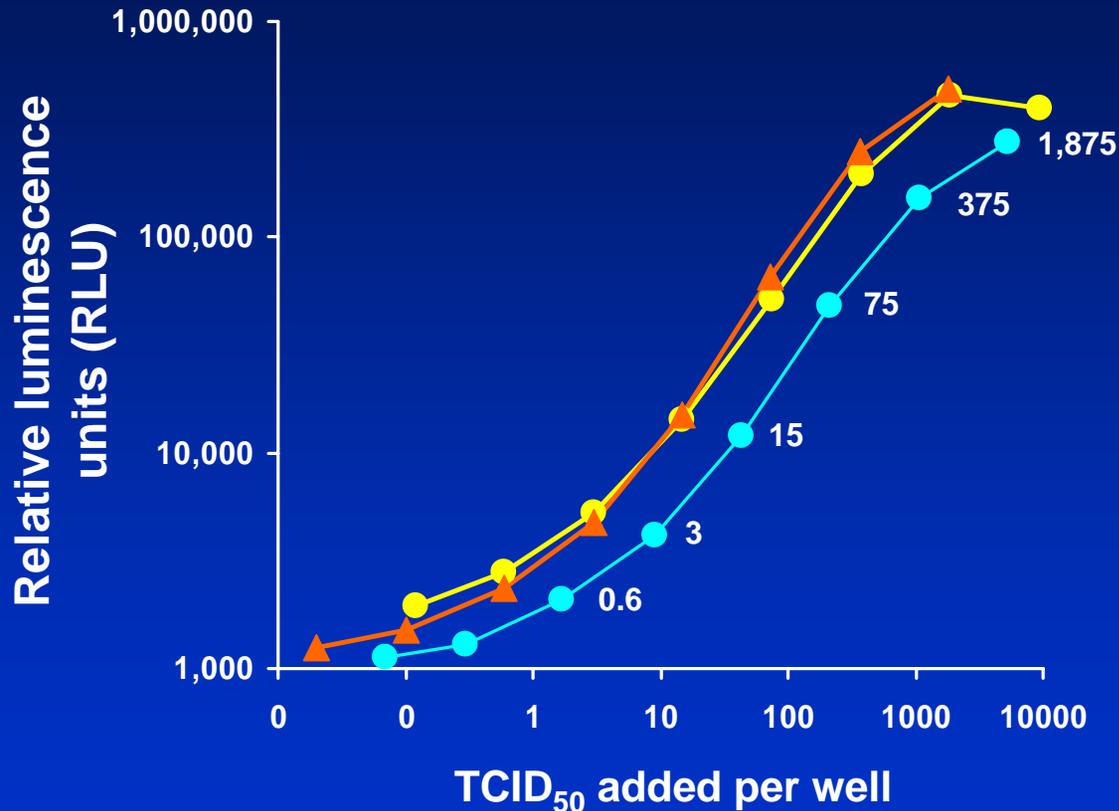


# SEQUENTIAL EVENTS IN DETECTING NEUTRALIZATION OF ENV-PSEUDOVIRUSES IN TZM-BL CELLS





# LINEAR RANGE OF INFECTION IN TZM-BL CELLS



Molecularly cloned  
Env pseudoviruses:

**SS1196.1**

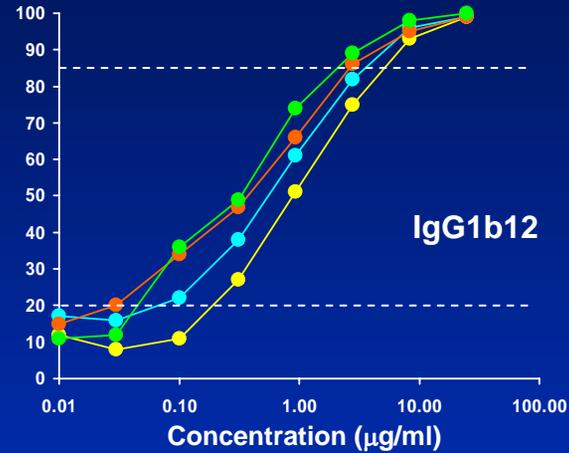
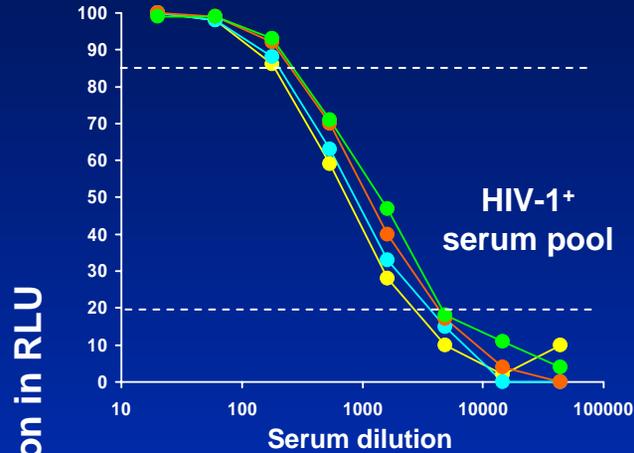
**QH0692.42**

**6101.10**

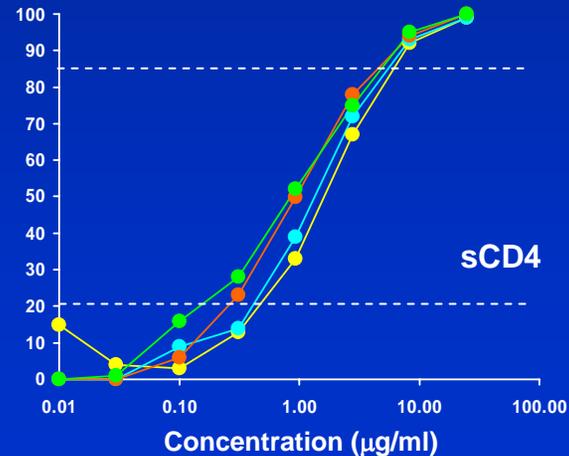
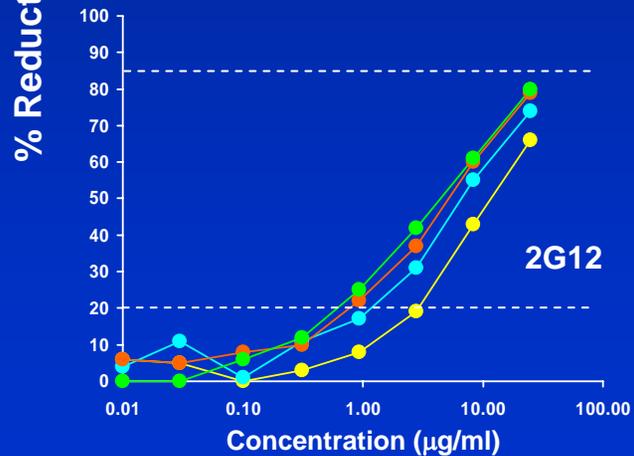
# LINEAR PORTION OF THE NEUTRALIZATION CURVE: 20% - 85%



Molecularly cloned pseudotype virus 1196.01,  
48 hr incubation



TCID50
50
200
1000
5000





# OPTIMIZATION OF THE TZM-BL ASSAY

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- Cell culture conditions
- Range of isolates that infect adequately
- Cell number
- Virus dose
- Incubation time
- Choice of 96-well plates for luminescence
- Luminescence readings
- DEAE-dextran
- Indinavir
- Unclassified vs cloned virus



# VALIDATION OF THE TZM-BL ASSAY

## Specificity:

- Background activity of normal human serum and plasma

## Accuracy:

- Comparisons have been made to other in-house assays and assays performed in other labs

## Precision:

- Well-to-well variability in cell control, virus control and test wells
- Intra- and inter-assay variability
- Intra- and inter-operator variability

## Limits of Quantitation:

- Upper and lower limits established

## Linearity & Range:

- Neutralization curves generated with positive serum samples and mAbs show a consistent pattern of linearity over a range of 20-85% reductions in RLU. Values in this range are directly proportional to the concentration of neutralizing antibodies in the sample.

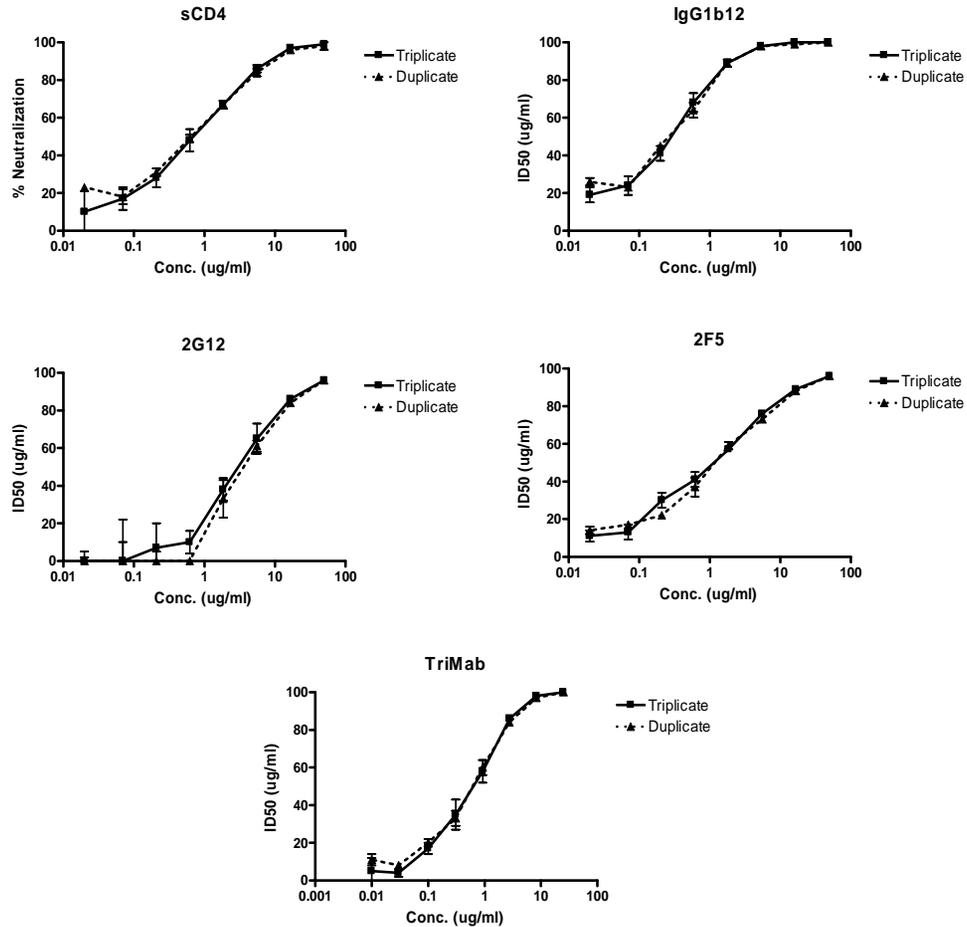
## Ruggedness & Robustness:

- Stability of CD4, CCR5 and CXCR4 expression
- Stability of TZM-bl infectivity after multiple passages
- Effect of DEAE-dextran on neutralizing antibody activity
- Effect of heat-inactivation on neutralizing antibody activity
- Serum vs plasma
- Uniformity of multiple luminometers



# Example of Intra-Assay Variation: Validation of Duplicate Assay Well Format

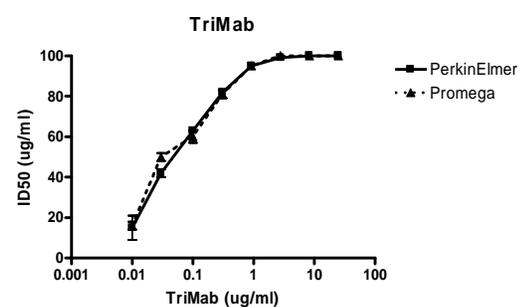
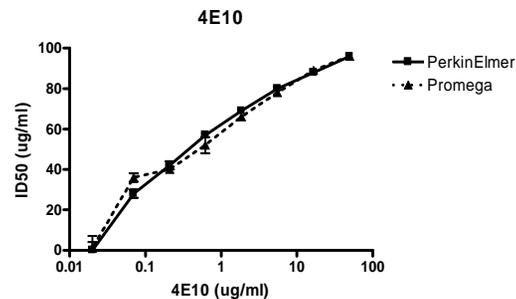
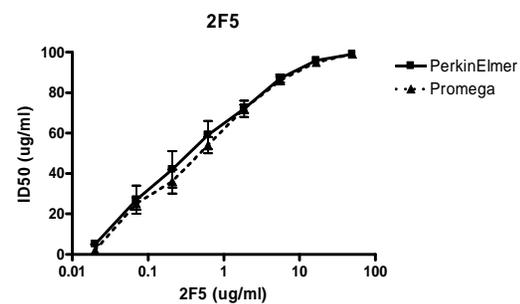
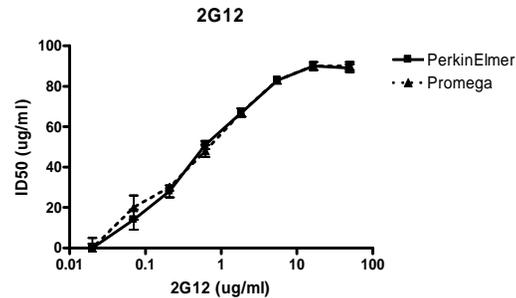
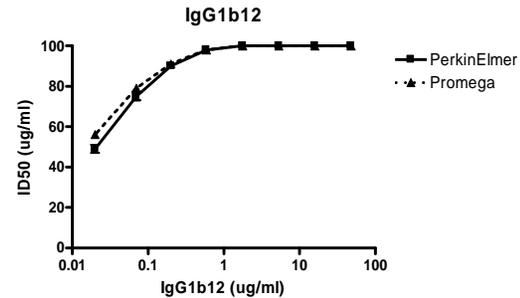
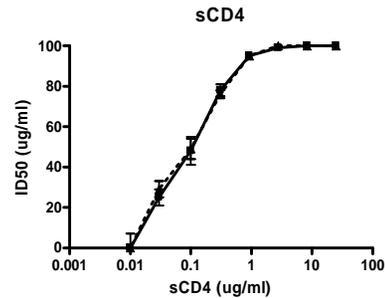
## Assays with QH0692.42





# Examples of Intra-Assay Variation: Comparison of Two Luciferase Kits (PerkinElmer vs Promega)

SF162.LS



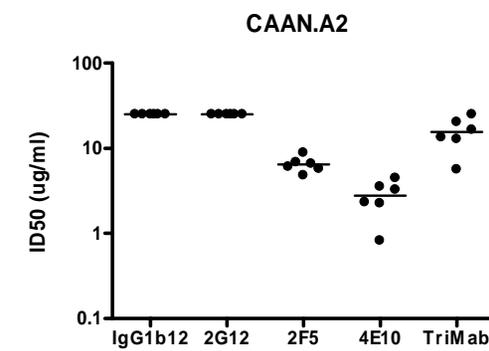
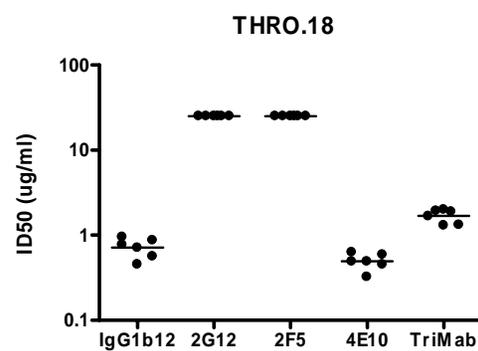
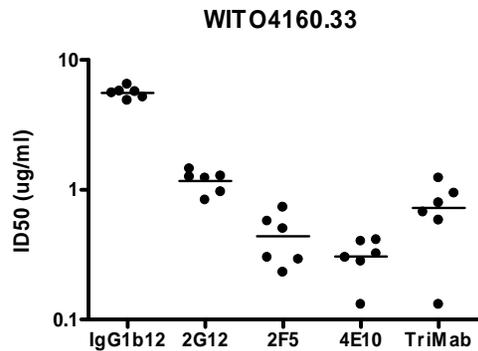
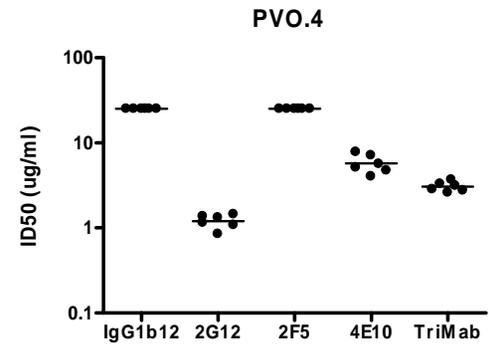
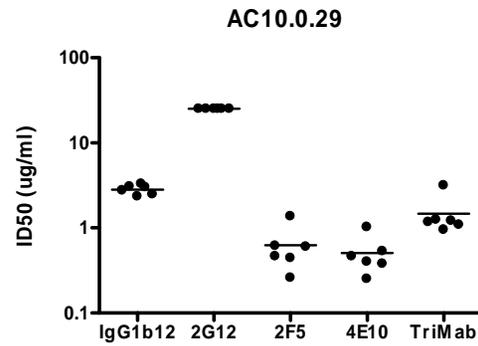
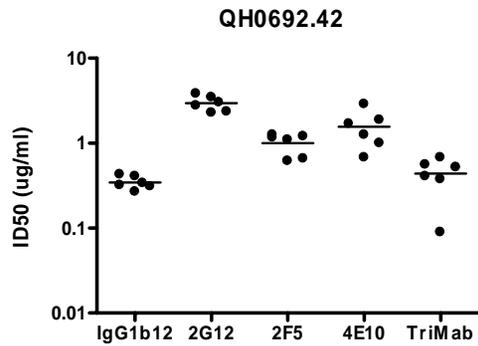


# **Inter-Operator Variation: Internal Proficiency Testing**

**Six operators assayed 7 positive serologic reagents against 6 reference strains of Env-pseudotyped HIV-1 in TZM-bl cells (SOP HVTN02-A0009).**

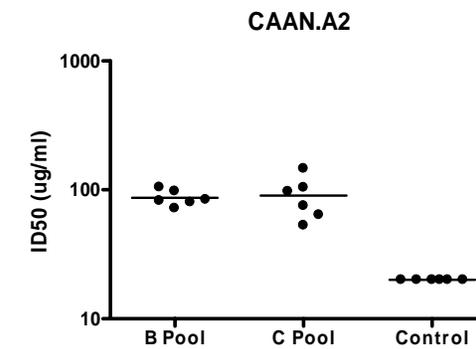
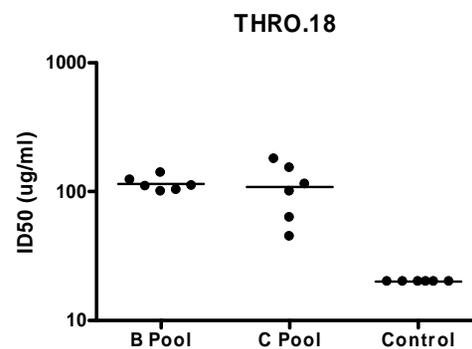
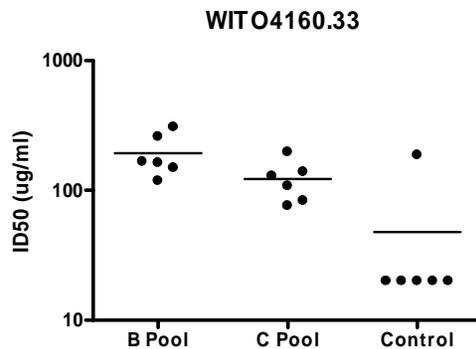
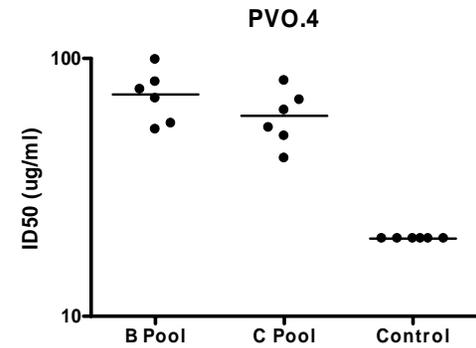
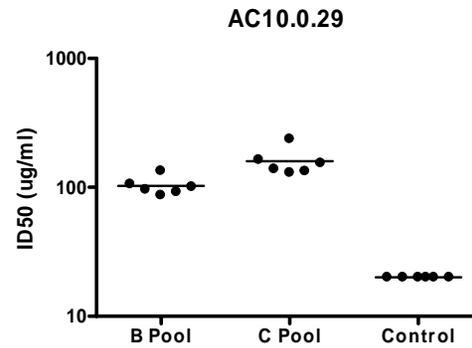
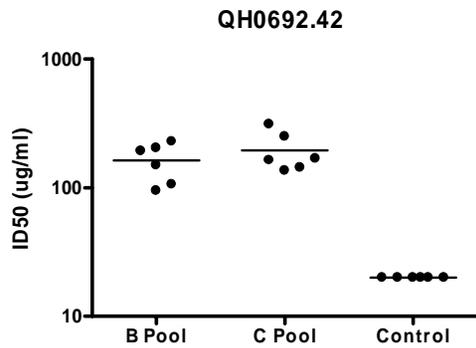


# Internal Proficiency Test Results with MAbs



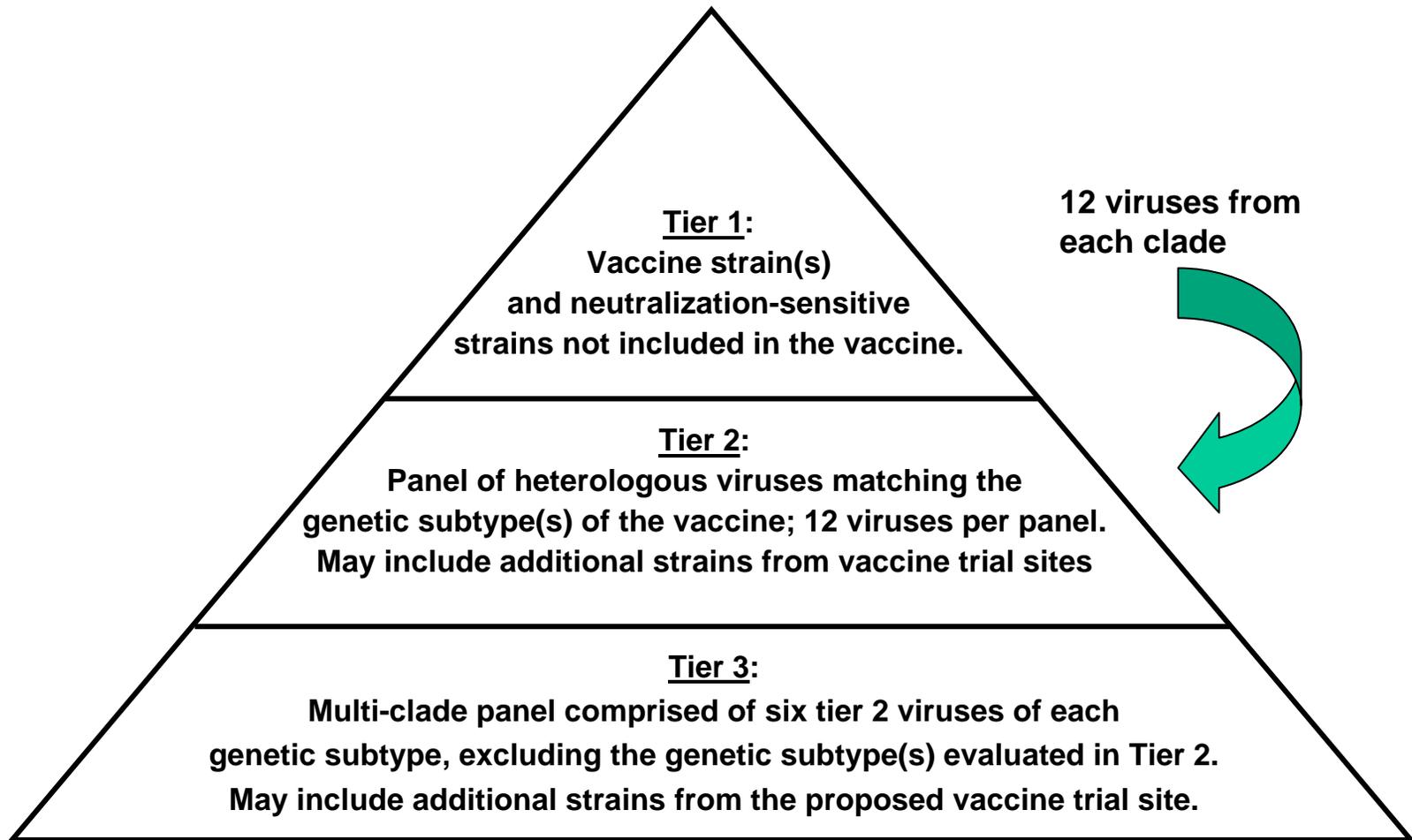


# Internal Proficiency Test Results with Serum Samples





# Tiered Approach to Assessing Vaccine-Elicited Neutralizing Responses





# Criteria for Selection: panel composition

- ❑ **Weighted in favor of recently collected viruses**
  - **Avoid potential genetic/antigenic drift over time**
- ❑ **Sexually transmitted viruses from acute/early infection**
- ❑ **Grouping by 6 major clades - 90% circulating HIV-1**
  - **Virus panels for A, B, C, D, E, A/G**
  - **Additional panels corresponding to vaccine trial sites**
- ❑ **Viruses with a representative distribution of neutralization phenotypes**
  - **HIV-1+ sera, mAbs, vaccine sera**
- ❑ **Use molecular clones for stability, reproducibility and epitope analysis**
  - **Non-replicating pseudoviruses**
  - **PBMC-grown viruses could also be made available**



# How many viruses in each panel?

## Statistical considerations - Peter Gilbert (SCHARP)

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**Estimate the # of viruses and vaccine recipients that would allow adequate power to differentiate immunogens.**



- Two vaccines in phase I - 20 subjects per arm
- Panel of 12 clade B viruses



**90% power to see a difference between vaccine that neutralized 10% of viruses vs. 35%**



# Advantage of Clonal Viruses

- ❑ **Reagent Characterization – precisely known Env sequence**
- ❑ **Stability – on regeneration of virus stock, same virus each time**
- ❑ **Easily transferable as plasmids**
- ❑ **Precision and reproducibility – clonal virus is the same in each assay**
- ❑ **Facilitate GCLP assay validation for clinical studies**
- ❑ **Facilitate the mapping of antibody specificities – relation to known Env sequence.**



## RECENT PROGRESS:

# Standard Panel of 12 Clade B HIV-1 Reference Strains

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- Full-length functional Env plasmids – **recently donated to NIAID AIDS repository**
- Clinically, demographically well characterized
- Acute/early sexually-acquired infection, R5 biologic phenotype
- Neutralization phenotype - representative primary isolates
- Genetically distinct and well-characterized, e.g. N-glycans, V-regions, MAb epitopes – **sequences deposited in GenBank**
- Gender diversity - most M-M, several F-M and one M-F transmissions



# Ongoing and Future Plans

- ❑ **Compose global panels for clades A, C, D, E, A/G**
- ❑ **Compose panels from international vaccine trial sites**
- ❑ **International networks - identify and donate reagents for non-B virus panels**
- ❑ **Address scientific questions identified by workshop**
- ❑ **Evaluate AIDSVAX phase III sera against the clade B panel (minimum bar)**
- ❑ **Design and implement an external proficiency testing program**



# Plans for an External Proficiency Testing Program: TZM-bl Neutralizing Antibody Assay

- ❑ **Initial round of testing**
  - **Assess inter-laboratory variation under conditions of relaxed standardization**
  
- ❑ **Subsequent rounds of testing**
  - **Confirm the key parameters that affect assay performance**
  - **Revise and validate the assay SOP**
  - **Develop an SOP for proficiency testing**
  - **Validate the proficiency testing SOP**

**First iteration of testing should begin in October 2005**



# Standard Neutralizing Antibody Potency Test for HIV-1 Vaccines?

- ❑ **Magnitude**: Geometric mean titer of neutralizing Abs against one or more Tier 1 viruses.
- ❑ **Breadth**: Frequency of positive neutralization against a standard panel of reference strains.
- ❑ **Consideration**: Protein boosting in the case of genetic vector vaccines.



# Acknowledgments

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