TESTING OF FIELD SPECIMENS FOR WEST NILE AND OTHER ARBOVIRUSES

Laura D. Kramer¹, Elizabeth B. Kauffman¹, Roger Nasci et al.² and the Arbovirus Laboratory¹

> ¹Wadsworth Center, NYSDOH and ²CDC, Arbovirus Diseases Branch

What is the question being asked?

Issues to consider in choice of assay

- Sensitivity
- Specificity
- Speed
- Staffing
- Safety
- \$\$

Specimens tested to monitor activity of West Nile virus -1

SERA for antibody detection



- Wild-caught avian species
- Captive birds
- Sentinels
- Cases equine, human





Diagnostic Assays for field and case specimens - 1

Serum, CSF

Serological Assays

> ELISA - indirect; competitive IgM IgG PRNT HI CF IFA Dipsticks Luminex



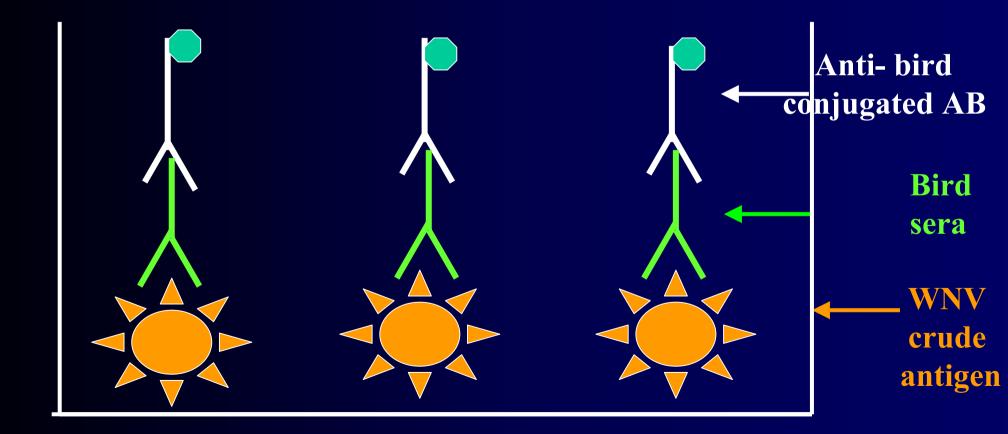
Serologic Assays

- Indirect ELISA using wild bird conjugate (Chiles and Reisen, 1998; Ebel, 2002)
 - Passeriformes (white-crowned sparrows)
 - Columbiformes (ringed turtle doves)
 - Galliformes (domestic chickens)
 - Anseriformes (muscovy ducks)

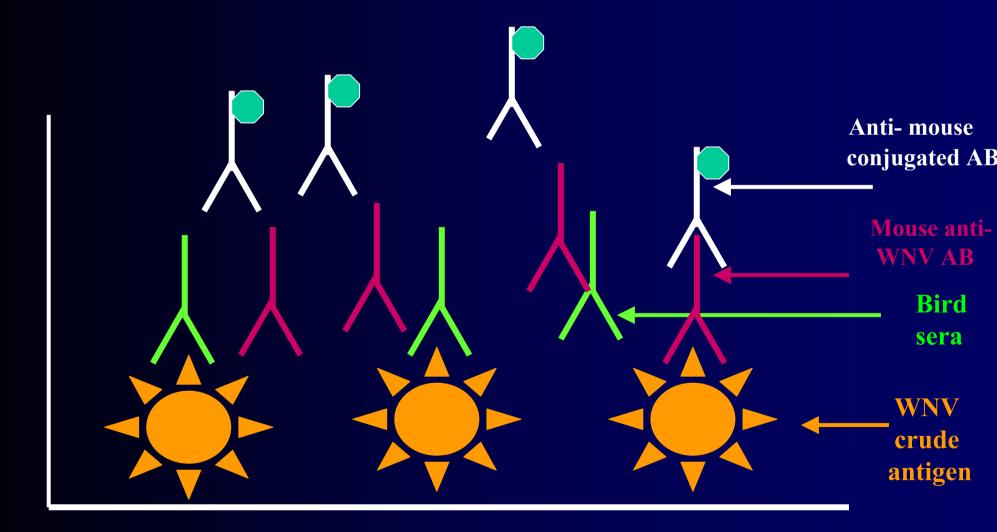
Competitive ELISA (Hall, 1994)

Confirmatory PRNT****

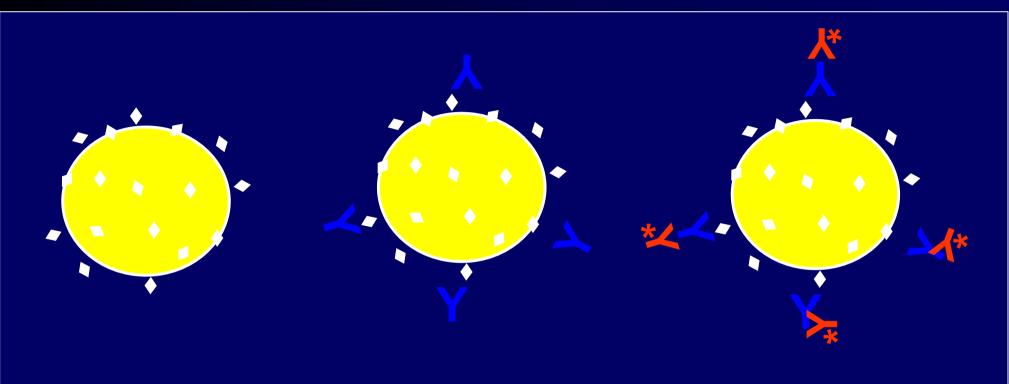
Indirect enzyme-linked immunosorbent assay



Competitive enzyme-linked immunosorbent assay



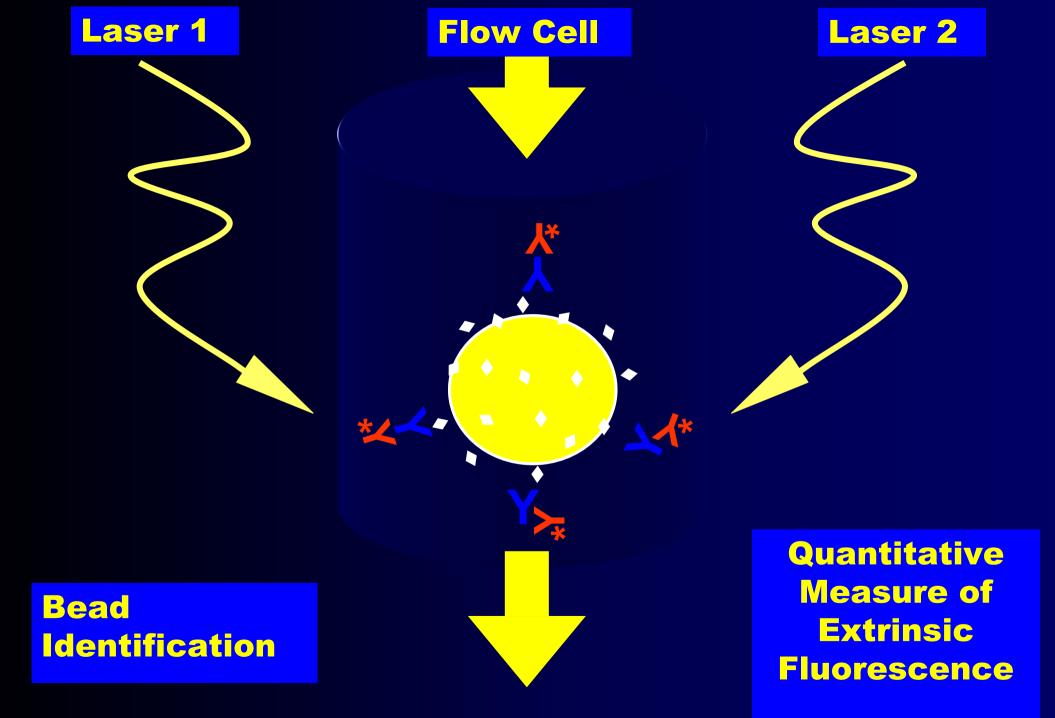
Suspension Array Technology Sequence of Events



Fluorescent Bead -Yellow Antigen - White

Patient's Antibody - Blue

PE Labeled Anti Antibody - Red



Specimens tested to monitor viral activity

TISSUES for virus isolation or detection

Mosquito pools



(50)

 Dead vertebrate tissues Oral/cloacal swabs



Living vertebrate - sera



Diagnostic Assays for Arboviruses - 2

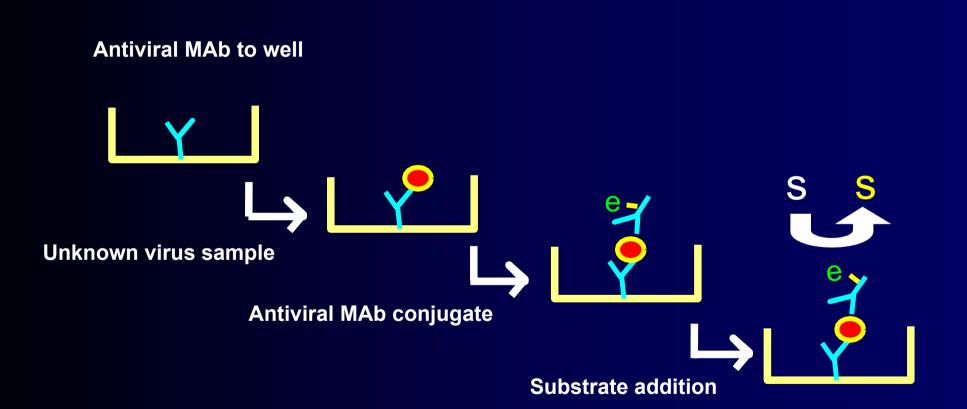
Mosquito pools, Tissues, Serum, CSF

Virus Detection Assays

> Virus isolation (cells, mice) IFA, IHC TaqMan RT-PCR Ang-capture ELISA RT-PCR Dipsticks NASBA

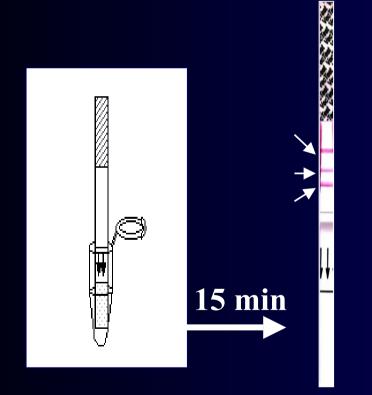


Antigen Capture Assay





Rapid assay for virus/antigen detection VecTestTM dip sticks



- Antigen-capture ELISA
- Mosquito homogenate
- Avian tissue
- Oral / cloacle swabs of avian carcasses
 - Crows & blue jays
 - 100% agreement with assay of brain tissues





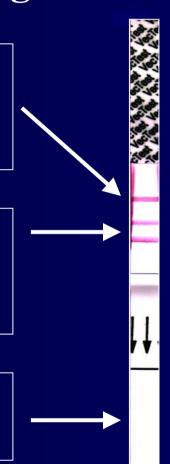
Procedure

• VecTest[™] assays for arbovirus antigen

Control Zone: captures unbound Ab-gold complex, confirms migration of sample through test zone.

Test Zone: virus-specific antibodies immobilized on membrane, SLE (top) and WN (bottom) in this assay.

Reagent Zone: virus-specific antibodies conjugated to gold particle.









Results

	Sensitivity Threshold in log ₁₀ PFU/ml				
	WN	SLE	EEE	WEE	
Plaque Assay	0.6 - 1.0				
TaqMan	0.1				
VecTest Assay	3.7	3.4	5.3	4.7	

Specificity: No evidence of cross reaction (false positives) in **TaqMan and VecTest** even at highest virus titer.





Results – Mosquito pools

34 TaqMan WNV-positive pools from Staten Island, NY 2000

TaqMan as standard



R Nasci

(No VecTest false positives)

Results – Oral swabs

Ontario, Canada (Barker et al.) Crows / oropharyngeal swabs 83% sensitivity 95% specificity

Oral swabs (Komar et al.)

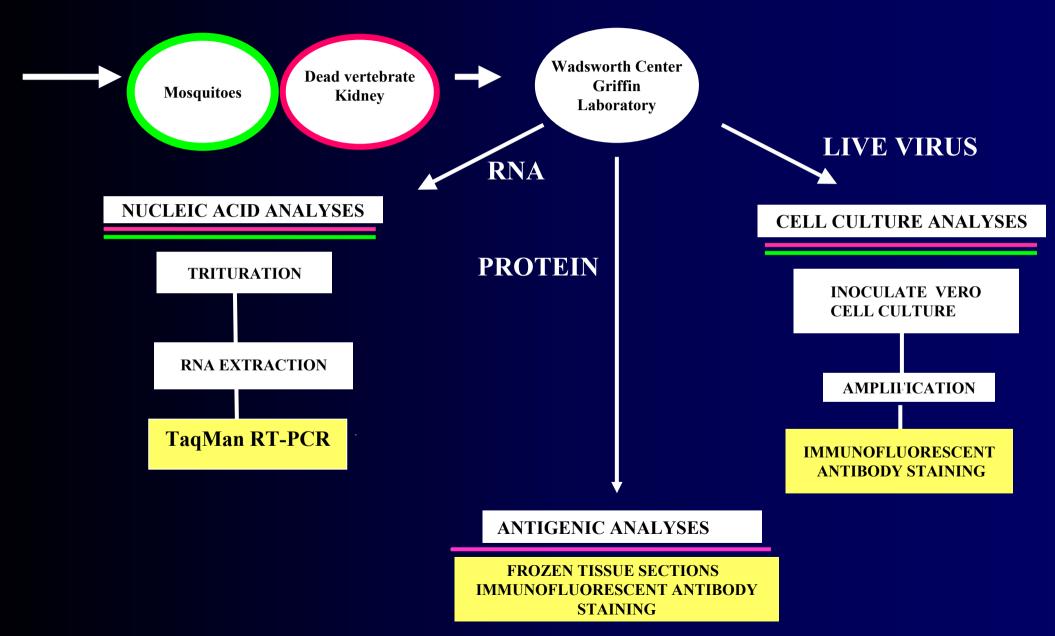
S

e

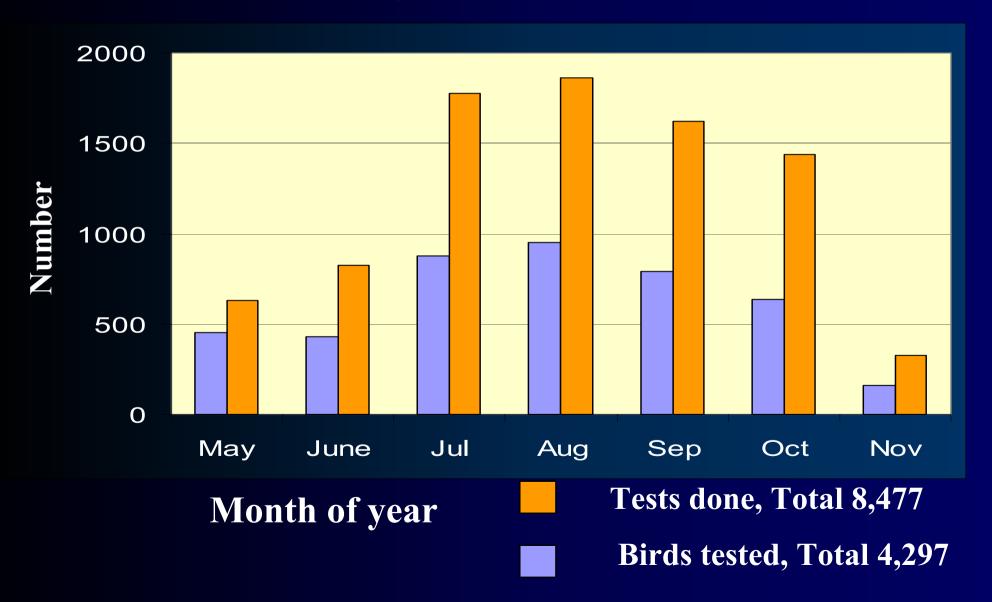
n

Sensitivity: 85% RT-PCR + 79% VecTest +

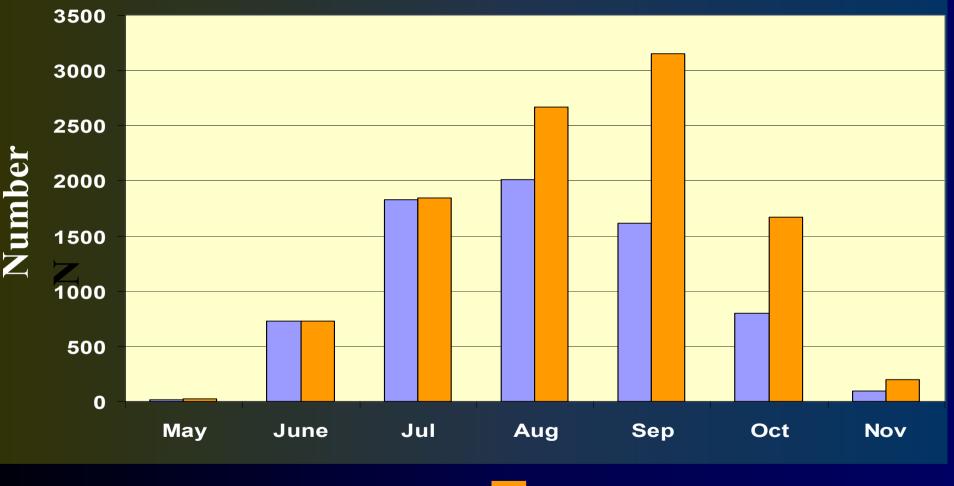
Flow chart for virologic testing of vertebrate and mosquito specimens



RT-PCR assays on avian tissue, 2002



RT-PCR assays on mosquito pools, 2002



Month of year

Tests done, Total 10,289

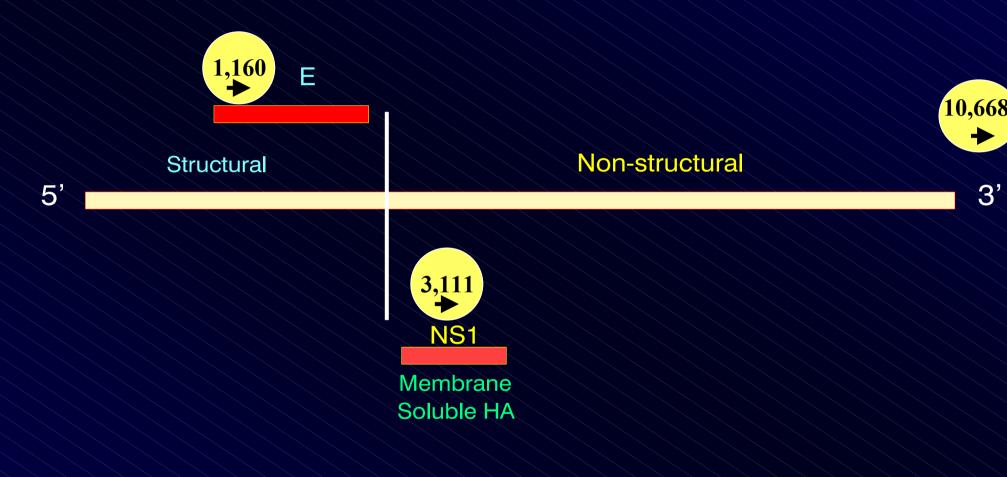
Mosq pools tested, Total 7,096

TRITURATION: Qiagen Mixer Mill MM 300



Disrupts 2 x 96 samples (1.2 ml) or 2 x 24 samples (2.0 ml) in 2-4 min

WNV Taqman primer / probe sets



Lanciotti 2000

High Throughput Testing

- Automated Nucleic Acid Workstation
 - Automates sample and reaction preparation for nucleic acid analysis
 - increase in productivity
 - cost efficient
 - high quality of product
 - decreased cross-contamination
 - consistency and reproducibility

ABI Prism 6700

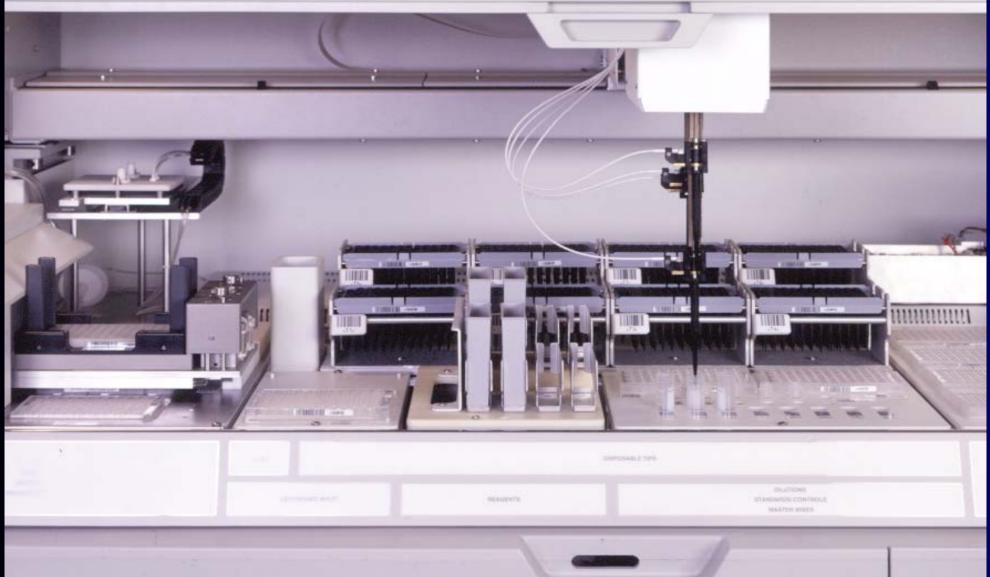
- Class II biosafety cabinet enclosure (HEPA filtered)
- Sets up dilutions and replicate samples in up to four 96-well output trays for TaqMan analysis
- Automatically seals output trays with a full cover optical blanket and holds them at 4C
- Completely compatible with TaqMan sequence detection system without additional manipulation
 - Software synergy
- One-step process from RNA purification to assay plate





mmmmmme	BERETER CONTRACTOR CONTRACTOR CONTRACTOR CONTRACTOR CONTRACTOR CONTRACTOR CONTRACTOR CONTRACTOR CONTRACTOR CONT	(~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~
		······

	***************************************	000000000000000000000000000000000000000



DESCRIPTION		ASSAY	PERSON HOURS
		COST	(Hands on labor)**
		per sample*	
Tissue sorting,	Birds/Mammals	\$0.75	2 hr per 96-well plate
excision,	Mosquitoes	\$0.50	3 hr per 96-well plate
homogenization	-		
Isolation of	RNeasy Method		
RNA	Birds/Mammals	\$3.25	5 h per 96-well plate
	Mosquitoes	\$3.25	5 h per 96-well plate
	ABI 6700 Robot		
	Birds/Mammals	\$2.00	1 h per 96-well plate
	Mosquitoes	\$2.75	1 h per 96-well plate
Real-time RT-	Manual setup	\$3.25	1 h per 96-well plate
PCR (TaqMan)	ABI 6700 Robot	\$3.85	30 m per 96-well plate

* Specific supplies only (no equipment, personnel, general supplies) **Robot run time: 85 m RNA extraction; 45 m TaqMan set-up

Comparison of TaqMan Ct values on RNA samples from naturally infected bird kidneys

Crow	ABI Prism 6700		RNeasy		Ratio
kidney	Mean C _t	SD	Mean C _t	SD	(6700:RNeasy)
A	14.97	0.8	16.56	0.7	0.90
B	17.85	0.6	18.45	0.6	0.97
C	14.59	0.6	16.93	0.5	0.86
D	14.71	0.7	16.78	0.6	0.88
E	17.57	0.4	21.95	1.3	0.80
F	30.13	1.5	29.54	5.5	1.02
G	40.00	0.0	40.00	0.0	1.00

Comparative assays on infected mosquito parts*

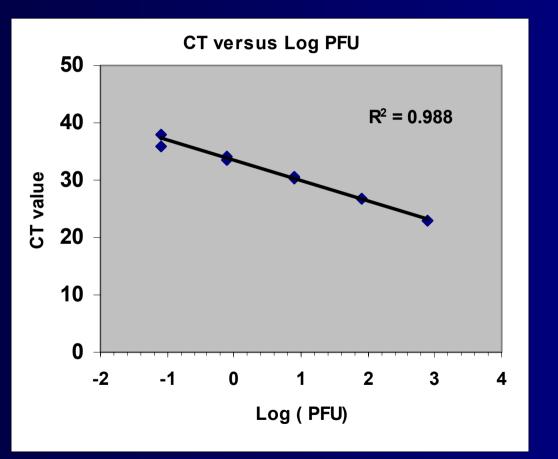
Infected	Ct Value		
Mosquito Part	Robot	RNeasy	
1 Leg	31.8	31.8	
2 Legs	29.9	30.6	
Abdomen	23.5	25.2	
Head	25.9	26.5	
Thorax	22.5	23.0	

* Added to pool of 50 uninfected mosquitoes

Summary of High Throughput Techniques

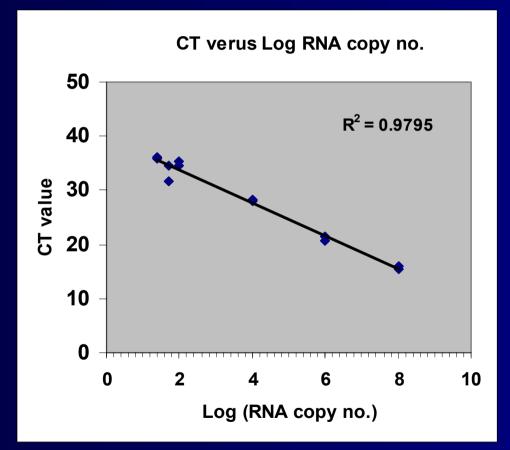
- Submission of sample data to laboratory on Excel spreadsheets
- High capacity mixer mill
- Robotic workstation for RNA extraction and real time RT-PCR setup
- Real time RT-PCR

Sensitivity: Log ₁₀ PFU by real time RT-PCR assay (primer - probe set to E gene)



SENSITIVITY 0.08 PFU in 5 ul

Sensitivity: Copy number by real time RT-PCR assay (primer - probe set to E gene)



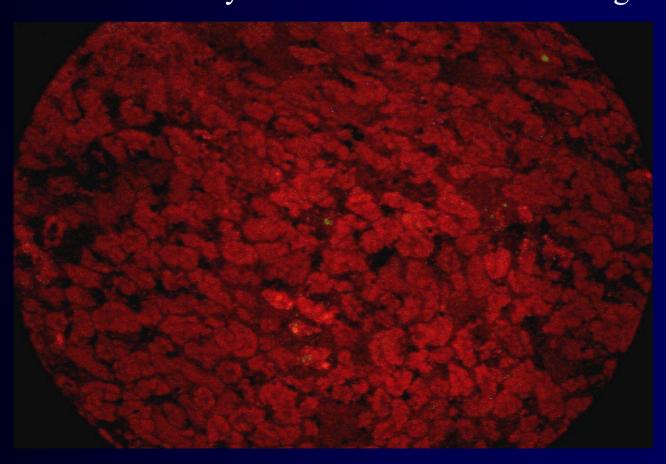
SENSITIVITY 40-60 copies in 5 ul

Sensitivity of multiplex real time RT-PCR using 2 sets of primer-probes for WNV

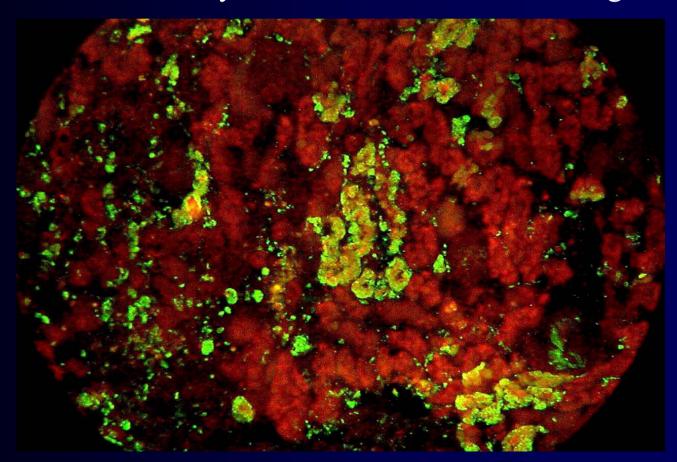
PFU	Avg. CT FAM	FAM SD.	Avg. CT VIC	VIC SD
8000	17.20	.15	17.54	.16
800	20.75	.08	21.07	.08
80	24.37	.12	24.62	.14
8	27.66	.12	28.34	.14
.8	30.55	.24	31.97	.24
──→ .08	40	00	40	00
.008	40	00	40	00

Output Sensitivity of single assay **RT-PCR**, 0.08 **PFU**

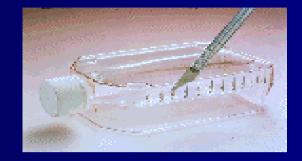
West Nile Virus Positive Kidney 107A Frozen Section, 200x, Evan's Blue counterstain First antibody = rabies glycoprotein-specific Mab (3D7) Second antibody = FITC labeled anti-mouse IgM



West Nile Virus Positive Crow Kidney 107A Frozen Section, 200x, Evan's Blue counterstain First antibody = West Nile Virus E protein-specific Mab (H5.46) Second antibody = FITC labeled anti-mouse IgM



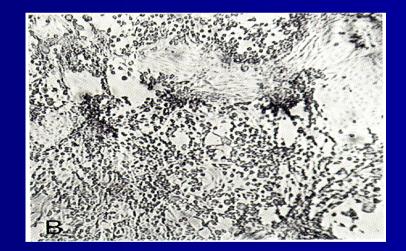
Cell culture inoculation



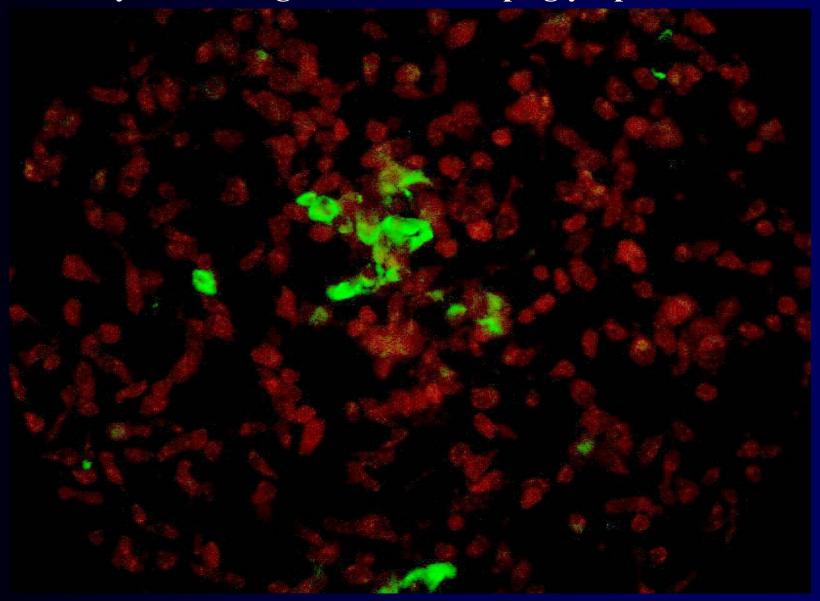
Vero cell cuture







WNV infected Vero cells stained with WNV specific monoclonal antibody directed against the envelope glycoprotein



Arbovirus Laboratory staff

