NHANES 2001–2002 Data Release May 2005 Documentation for Laboratory Results

Laboratory 28POC: Polychlorinated Dibenzo-*p*-Dioxins (PCDDs), Dibenzofurans (PCDFs), Non-Ortho-Substituted or Coplanar Polychorinated Biphenyls (cPCBs), other Polychlorinated Biphenyls (PCBs), Persistent Chlorinated Pesticides, and Selected Pesticide Metabolites

(1) Documentation File Date – July 2007

(2) Documentation File Name – Laboratory 28POC: Polychlorinated Dibenzo-p-Dioxins (PCDDs), Dibenzofurans (PCDFs), Non-Ortho-Substituted or Coplanar Polychorinated Biphenyls (cPCBs), other Polychlorinated Biphenyls (PCBs), Persistent Chlorinated Pesticides, and Selected Pesticide Metabolites

(3) Survey Years Included in this File Release – 2001–2002

(4) Component Description

Polychlorinated Dibenzo-*p*-Dioxins (PCDDs), Dibenzofurans (PCDFs), Non-Ortho-Substituted or Coplanar Polychorinated Biphenyls (cPCBs), other Polychlorinated Biphenyls (PCBs), Persistent Chlorinated Pesticides, and Selected Pesticide Metabolites

Organochlorines are diverse, synthetic chemicals that are persistent in the environment and tend to bioaccumulate. Most of these chemicals are banned in the U.S.

Assessment of exposure to persistent organochlorines in a representative sample of the U.S. population is needed to determine current prevalence and level of exposure and the potential for human health threat from exposure to these chemicals.

(5) Sample Description

Eligible Sample

One third subsample of participants aged 12 years and older.

(6) Description of the Laboratory Methodology

Substituted or Coplanar Polychlorinated Biphenyls (cPCBs), other Polychlorinated Biphenyls (PCBs), Persistent Chlorinated Pesticides, and Selected Pesticide Metabolites

These analytes are measured in serum by high-resolution gas chromatography/ isotope-dilution high-resolution mass spectrometry (HRGS/ID-HRMS). Serum samples are spiked with ¹³C₁₂-labeled internal standards and the analytes of interest are isolated using either a C₁₈ solid-phase extraction (SPE) or liquid-liquid extraction procedure followed by a multi-column automated cleanup and enrichment procedure. The analytes are chromatographed on a DB-5 ms capillary column (30m x 0.25 mm x 0.25 µm film thickness) using a Hewlett-Packard 6890 gas chromatograph and selected analytes quantified by ID-HRMS using selected ion monitoring (SIM) at 10,000 resolving power using either a Micromass AutoSpec ULTIMA or Finnigan MAT95 mass spectrometer in the EI mode. The concentration of each analyte is calculated from an individual standard linear calibration. Each analytical run is conducted blinded and consists of three unknown serum samples, a method blank, and a quality control sample. Detection limits, on a whole-weight and lipid-adjusted basis, are reported for each sample, corrected for sample weight and analyte recovery.

(7) Laboratory Quality Control and Monitoring

Serum specimens are processed, stored, and shipped to the Division of Environmental Health Laboratory Sciences, National Center for Environmental Health, Centers for Disease Control and Prevention for analysis.

(8) Data Processing and Editing

Detailed specimen collection and processing instructions are discussed in the NHANES Laboratory/Medical Technologists Procedures Manual (LPM). Vials are stored under appropriate frozen (–20°C) conditions until they are shipped to National Center for Environmental Health for testing.

(9) Data Access

All data are publicly available.

(10) Analytic Notes for Data Users:

Measures of polychlorinated dibenzo-*p*-dioxins (PCDDs), dibenzofurans (PCDFs), nonortho-substituted or coplanar polychorinated biphenyls (cPCBs), other polychlorinated biphenyls (PCBs), persistent chlorinated pesticides, and selected pesticide metabolites are assessed in participants aged 12 years and older on a one-third subsample. Use the special weights included in this data file when analyzing data. Read the "Special Sample Weights for this Dataset" information provided below before beginning analysis.

Detection limits

The detection limit was variable for all of the analytes in the data set. Two variables are provided for each of these analytes. The variable named LBD_LC indicates whether the results were below the limit of detection. There are two values: "0" and "1"; "1" indicates that the result was below the limit of detection. The other variable named LBX_ provides the analytic result for that analyte. In cases, where the result was below the limit of detection, the value for that variable is the detection limit divided by the square root of two.

The analysis of NHANES 2001–2002 laboratory data must be conducted with the key survey design and basic demographic variables. The NHANES 2001–2002 Household Questionnaire Data Files contain demographic data, health indicators, and other related information collected during household interviews. The Phlebotomy Examination file includes auxiliary information on duration of fasting, the time of day of the venipuncture, and the conditions precluding venipuncture. The Household Questionnaire and Phlebotomy Exam and other data files may be linked to the laboratory data file using the unique survey participant identifier SEQN.

(11) Special Sample Weights for this Dataset:

Special sample weights are required to analyze these data properly.

Measures of this urinary multi-analyte profile are assessed in participants aged 6 years and over on a randomly selected subsample. Specific sample weights for this subsample are included in this data file and should be used when analyzing these data.

The dataset includes 2-year and 4-year subsample weights. The 4-year weights should be used if these 2001–2002 data are combined with 1999–2000 data. The 1999–2000 data files have been updated to include the subsample 4-year weights. The recommended procedure for variance estimation requires use of stratum and PSU variables (SDMVSTRA and SDMVPSU, respectively), which are included in the demographic data file for each data release. For further information, see the NHANES Analytic Guidelines.

(12) References

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Quality Control and Limit of Detection

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Toxic Equivalency Factors (TEFs)

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