

FOOD AND DRUG ADMINISTRATION  
CENTER FOR DRUG EVALUATION AND RESEARCH

CLINICAL PHARMACOLOGY SUBCOMMITTEE  
OF THE  
ADVISORY COMMITTEE FOR PHARMACEUTICAL SCIENCE

8:34 a.m.

Tuesday, April 22, 2003

Conference Room  
5630 Fishers Lane  
Food and Drug Administration  
Rockville, Maryland 20857

ASSOCIATED REPORTERS OF WASHINGTON  
1523 North Carolina Avenue, N.E.  
Washington, D.C. 20002  
(202) 543-4809

## ATTENDEES

## SUBCOMMITTEE MEMBERS:

JURGEN VENITZ, M.D., PH.D., Acting Chair  
Associate Professor  
Department of Pharmaceutics  
Virginia Commonwealth University  
School of Pharmacy  
Box 980533  
410 North 12th Street  
Richmond, Virginia 23298-0533

KATHLEEN REEDY, R.D.H., M.S., Executive Secretary  
Advisors and Consultants Staff (HFD-21)  
Center for Drug Evaluation and Research  
Food and Drug Administration  
5600 Fishers Lane  
Rockville, Maryland 20857

EDMUND V. CAPPARELLI, PHARM.D.  
Associate Clinical Professor  
University of California, San Diego  
9500 Gillman Drive  
La Jolla, California 92093

HARTMUT DERENDORF, PH.D.  
Professor, Department of Pharmaceutics  
University of Florida College of Pharmacy  
P.O. Box 100494, Health Science Center  
Gainesville, Florida 32610-0494

DAVID FLOCKHART, M.D., PH.D.  
Professor, Departments of Pharmacology  
and Medicine  
Indiana University School of Medicine  
Division of Clinical Pharmacology  
Wishard Memorial Hospital WP OPW 320  
1001 West 10th Street  
Indianapolis, Indiana 46202

WILLIAM J. JUSKO, PH.D.  
Professor, Department of Pharmaceutics  
State University of New York at Buffalo  
School of Pharmacy  
Buffalo, New York 14260

## ATTENDEES (Continued)

## SUBCOMMITTEE MEMBERS: (Continued)

GREGORY L. KEARNS, PHARM.D.  
Professor and Division Chief  
Pharmacology and Toxicology  
Children's Mercy Hospital  
2401 Gillham Road  
Kansas City, Missouri 64143

MARY V. RELING, PHARM.D.  
Member, Pharmaceutical Sciences  
St. Jude Children's Research Hospital  
332 North Lauderdale, Room D-1052  
Memphis, Tennessee 38105

WOLFGANG SADEE, DR.RER.NAT.  
Chair, Department of Pharmacology  
College of Medicine and Public Health  
Ohio State University  
5072 Graves Hall, 333 West 10th Avenue  
Columbus, Ohio 43210

LEWIS B. SHEINER, M.D.  
Professor, Laboratory Medicine  
University of California, San Francisco  
Box 0626, C-255  
San Francisco, California 94143

MARC SWADENER, ED.D., Consumer Representative  
2235 Dartmouth Avenue  
Boulder, Colorado 80305-5207

## GUEST SPEAKER:

MATS KARLSSON, PH.D.  
Uppsala University  
Sweden

## ATTENDEES (Continued)

## FOOD AND DRUG ADMINISTRATION STAFF:

PETER LEE, PH.D.  
LARRY LESKO, PH.D.  
NHI NGUYEN, PH.D.  
HE SUN, PH.D.  
GENE WILLIAMS, PH.D.  
JENNY J. ZHENG, PH.D.

## C O N T E N T S

| AGENDA ITEM   | PAGE |
|---|------|
| MEETING STATEMENT<br>by Ms. Kathleen Reedy  | 7    |
| INTRODUCTION TO THE MEETING<br>by Dr. Larry Lesko   | 10   |
| TOPIC 1: QUANTITATIVE RISK ANALYSIS USING<br>EXPOSURE-RESPONSE FOR DETERMINING DOSE<br>ADJUSTMENT FOR SPECIAL POPULATIONS |      |
| Introduction - by Dr. Peter Lee   | 19   |
| Example 1 - by Dr. Nhi Nguyen   | 26   |
| Example 2 - by Dr. Jenny Zheng  | 42   |
| Committee Discussion  | 61   |
| Example 3 - by Dr. He Sun   | 82   |
| Committee Discussion  | 93   |
| OPEN PUBLIC HEARING   | 116  |
| Example 4 - by Dr. Mats Karlsson  | 118  |
| Committee Discussion  | 141  |
| TOPIC 2: PEDIATRIC POPULATION PHARMACOKINETICS<br>STUDY DESIGN TEMPLATE AND ANALYSES OF THE FDA<br>PEDIATRIC DATABASE     |      |
| Introduction - by Dr. Larry Lesko   | 150  |
| Pediatric PPK Template - by Dr. Peter Lee   | 158  |
| Committee Discussion  | 165  |
| Pediatric Database Analysis - Dr. Gene Williams   | 188  |
| Committee Discussion  | 203  |

## P R O C E E D I N G S

(8:34 a.m.)

1  
2  
3 DR. VENITZ: I'd like to call the meeting to  
4 order, please.

5 Welcome, everybody. This is the Clinical  
6 Pharmacology Subcommittee meeting. We have a full agenda,  
7 as you can tell.

8 I'd like to open the meeting by introducing the  
9 individuals around the table, maybe starting with Dr.  
10 Derendorf, please.

11 DR. DERENDORF: Hartmut Derendorf, University  
12 of Florida.

13 DR. CAPPARELLI: Edmund Capparelli, University  
14 of California, San Diego.

15 DR. FLOCKHART: Dave Flockhart from Indiana  
16 University.

17 DR. SHEINER: Lewis Sheiner, University of  
18 California, San Francisco.

19 DR. SWADENER: Marc Swadener, Boulder,  
20 Colorado.

21 MS. REEDY: Kathleen Reedy, Food and Drug  
22 Administration.

23 DR. VENITZ: Jurgen Venitz, Virginia  
24 Commonwealth University.

25 DR. JUSKO: William Jusko, University at

1 Buffalo.

2 DR. KEARNS: Greg Kearns, University of  
3 Missouri, Kansas City.

4 DR. RELLING: Mary Relling, St. Jude Children's  
5 Research Hospital in Memphis.

6 DR. SADEE: Wolfgang Sadee, Ohio State  
7 University.

8 DR. LESKO: Larry Lesko, Office of Clinical  
9 Pharmacology and Biopharmaceutics at FDA.

10 DR. LEE: Peter Lee, FDA.

11 DR. VENITZ: Thank you.

12 Our next order of business is the conflict of  
13 interest statement. Kathleen Reedy will read the conflict  
14 of interest statement.

15 MS. REEDY: Acknowledgement related to general  
16 matters waivers, Clinical Pharmacology Subcommittee of the  
17 Advisory Committee for Pharmaceutical Science, April 22,  
18 2003, an open session.

19 The following announcement addresses the issue  
20 of conflict of interest with respect to this meeting and is  
21 made a part of the record to preclude even the appearance  
22 of such at this meeting.

23 The topics of this meeting are issues of broad  
24 applicability. Unlike issues before a committee in which a  
25 particular product is discussed, issues of broad

1 applicability involve many industrial sponsors and academic  
2 institutions.

3 All special government employees have been  
4 screened for their financial interests as they may apply to  
5 the general topics at hand. Because they have reported  
6 interests in pharmaceutical companies, the Food and Drug  
7 Administration has granted general matters waivers to the  
8 following SGEs which permits them to participate in these  
9 discussions: Dr. Edmund Capparelli, Dr. William Jusko, Dr.  
10 Gregory Kearns, Dr. Howard McLeod, Dr. Wolfgang Sadee, Dr.  
11 Lewis Sheiner.

12 A copy of the waiver statements may be obtained  
13 by submitting a written request to the agency's Freedom of  
14 Information Office, room 12A-30 of the Parklawn Building.

15 In addition, Dr. Hartmut Derendorf, Dr. David  
16 Flockhart, Dr. Mary Relling, and Dr. Marc Swadener do not  
17 require special matters waivers because they do not have  
18 any personal or imputed financial interests in any  
19 pharmaceutical firms.

20 Because general topics impact so many  
21 institutions, it is not prudent to recite all potential  
22 conflicts of interest as they apply to each member and  
23 consultant.

24 FDA acknowledges that there may be potential  
25 conflicts of interest, but because of the general nature of

1 the discussion before the committee, these potential  
2 conflicts are mitigated."

3 With respect to FDA's invited guest speaker,  
4 Dr. Mats Karlsson reports that he has contracts and/or  
5 grants with AstraZeneca, Oasmia, Pfizer, and Servier. He  
6 also receives consulting fees from AstraZeneca, Ferring,  
7 Lilly, Pfizer, and Roche; and speaker fees from Johnson &  
8 Johnson and NovaNordisk.

9 In the event that the discussions involve any  
10 other products or firms not already on the agenda for which  
11 FDA participants have a financial interest, the  
12 participants' involvement and their exclusion will be noted  
13 for the record.

14 With respect to all other participants, we ask  
15 in the interest of fairness that they address any current  
16 or previous financial involvement with any firm whose  
17 product they may wish to comment upon.

18 DR. VENITZ: Thank you, Kathleen.

19 Before we proceed to the official business of  
20 today, I'd like to welcome a few new committee members.  
21 Dr. Swadener to my right is the consumer representative who  
22 also serves on the Advisory Committee for Pharmaceutical  
23 Science. We've got Dr. Shek who couldn't make it today who  
24 is the industry representative, also serving on the  
25 Advisory Committee for Pharmaceutical Science. And Drs.

1 D'Argenio and Davidian who couldn't make it today.

2 I would also like to thank the two outgoing  
3 members of the committee, Dr. Lalonde and Dr. Hale, as well  
4 as Dr. Jusko to my left, for chairing the previous  
5 committee meeting.

6 With that said, I'd like to turn over the  
7 meeting to Dr. Lesko who is going to introduce the topics  
8 for the next day and a half. Larry.

9 DR. LESKO: Thank you, Jurgen.

10 Well, good morning, everybody and again welcome  
11 back to Rockville. This is the second Clin Pharm  
12 Subcommittee meeting. We had our first back six months  
13 ago, on October 22-23, and as reflect back on that meeting,  
14 it was an extremely productive meeting for us. The advice  
15 we received at that time was excellent, and we've been  
16 thinking about it since then as we move forward with the  
17 initiatives we introduced at that first meeting.

18 I do want to say thanks again. As I look  
19 around the room, I recognize everyone here as very busy in  
20 their own work, and taking time to come to Washington and  
21 participate with us is extremely important. I think you'll  
22 find that the mission that we have for this committee is a  
23 noble one relating to some exciting times in the area of  
24 drug development generally and clinical pharmacology  
25 specifically.

1           Let me talk a little bit about today's meeting.  
2           Let me start with what's new since we last met in October,  
3           and there are really three exciting things that are new  
4           that really impact the topics that we'll talk about today.

5           The first is an FDA-wide announcement that our  
6           Commissioner made back in January called Improving  
7           Innovation in Medical Technology Beyond 2002. What this  
8           initiative entails is quite lengthy. There are several  
9           goals, but basically it revolves around improving the  
10          process, including the drug development process, for  
11          bringing medical innovations, treatments, and devices to  
12          the marketplace as quickly as possible that would benefit  
13          public health.

14          A second part of that, however, is improving  
15          the review process at FDA through a quality systems  
16          approach. This is the goal that we are going to sort of  
17          use to couch today's topics because a quality systems  
18          approach, if I think of it in terms of goals, has the goals  
19          I have on the slide basically.

20          Application of advances in science. Well, what  
21          are those advances in science? They're clinical trial  
22          design. They're clinical pharmacology study designs,  
23          statistical approaches, modeling and simulation, use of  
24          dose response in PK/PD in interesting ways.

25          Use of new technology. Use of new technology

1 | embraces things like the quantitative methods we're going  
2 | to talk about today. It embraces the integration of  
3 | pharmacogenetics into drug development and review.

4 |           Rigorous analytic reviewer tools. This is the  
5 | how to do it. What are the tools that we can make  
6 | available to our reviewers to achieve this quality systems  
7 | approach? We'll be talking about one of those in the first  
8 | topic.

9 |           And finally, the overall goal of this  
10 | initiative is to provide for high quality reviews. That's  
11 | translated into effective reviews, efficient reviews, and  
12 | consistent reviews.

13 |           Initiative number two that's occurred since the  
14 | last time we met is an initiative under our Prescription  
15 | Drug User Fee Act, PDUFA. It's the premarketing risk  
16 | assessment initiative. We only recently had our first  
17 | public meeting having to do with the risk assessment  
18 | initiative, and Bob Meyer, our ODE II director, defined  
19 | risk assessment as the process of identifying, estimating,  
20 | and evaluating the nature and severity of risk of a drug  
21 | product. You'll see some links between this and the first  
22 | topic in today's presentations.

23 |           In that meeting on April 9th, Bob Temple  
24 | described the ideal safety database that we ought to be  
25 | striving for, and that included a complete characterization

1 of the clinical exposure-response relationship certainly  
2 for efficacy, but also for drug safety. And he made the  
3 point that this is important for making decisions about  
4 dosing adjustments, particularly in many of the clinical  
5 pharmacology studies when exposure goes up for a variety of  
6 reasons.

7 He recommended a good search for  
8 individualization factors. This obviously involves  
9 studying individual plasma drug levels, and he made the  
10 point that it's always necessary to assess polymorphic drug  
11 metabolism, and you'll see that this relates to one of our  
12 topics in this meeting.

13 Finally, he touched upon pediatrics, an  
14 important topic, and made the point that these pose special  
15 issues related to dose, PK, and PD, and we'll be touching  
16 upon that as well today.

17 The third initiative that's been launched since  
18 October is the one that relates to our FDA Science Board.  
19 We had this meeting last week on the same day as our risk  
20 assessment meeting. The theme of the Science Board meeting  
21 was integrating scientific advances into regulation with  
22 the emphasis on pharmacogenetics. Dr. Woodcock made a  
23 presentation at the Science Board and stated that genetic  
24 contributions to variability in toxicity include  
25 differences in drug metabolism, for example, thiopurine

1 methyltransferase, which was a topic of our last meeting,  
2 but more broadly recommended that we look at the use of  
3 genetic tests for metabolizer status to predict dosing.

4 So think of those three broad initiatives, and  
5 I hope it gives you a context for the discussion that we're  
6 going to have today and tomorrow.

7 We have four proposals, four topics on the  
8 agenda. The first is a proposal that we initiated our  
9 discussion in October and we've refined the proposal and  
10 we'll present with examples today the idea of a  
11 standardized approach to quantitate the impact that changes  
12 in exposure, related to efficacy or safety, result from  
13 changes in PK caused by a variety of intrinsic and  
14 extrinsic factors.

15 What we're trying to accomplish with this  
16 proposal and standardized approach relates to what I  
17 mentioned about the Commissioner's initiative, quality  
18 systems and review. We want to achieve a rational  
19 scientific basis for dosing adjustment that's quantitative  
20 and that links to the assessment of risk.

21 We have a goal of identifying individualization  
22 factors, and through our examples today you'll see some of  
23 those factors.

24 And finally at the end of the day, we hope to  
25 develop a standardized method that would rely on many

1 different tools, but a standardized thinking process that  
2 we hope would bring consistency to the label  
3 recommendations that we make in terms of dosing  
4 adjustments.

5 Our topic number two is pediatrics, and again  
6 going back to October, we opened the discussion of  
7 pediatrics very briefly the last time. Today we'll be  
8 making a proposal. The proposal will relate to a pediatric  
9 population PK design template. We'd like to recommend the  
10 use of this template for getting information about  
11 pediatrics during drug development. We feel that this  
12 approach is efficient in many cases. We feel it's under-  
13 utilized for a variety of reasons relating to perhaps a  
14 lack of understanding, perhaps related to a concern that  
15 FDA will not accept this type of study approach. But we'd  
16 like your input on the template that we'll be presenting.

17 Related to that, at the last meeting I had  
18 mentioned that we have this database at FDA related to  
19 pediatric studies. These are studies that were done under  
20 our pediatric rule. We felt this database is loaded with  
21 information that we could capitalize on by studying it,  
22 looking for trends, and learning something about the  
23 pediatric clinical pharmacology situation. We'll update  
24 you on our progress. It's been slow. It's been difficult  
25 because we don't have access to an electronic database, and

1 much of our time is simply gathering and assembling data.  
2 But nevertheless, we'd like to share with you some of the  
3 things that we've learned so far, but more importantly what  
4 we'd like to do going forward and look for input on  
5 designing the studies of this database.

6           The third topic, which we'll talk about  
7 tomorrow morning, is what I'll call a work in progress.  
8 We'll all familiar with the human genome. We've been  
9 bombarded with information about it, particularly this  
10 month on the 50th anniversary of identifying the double  
11 helix structure for DNA. Certainly the dream of genomics  
12 is to develop new and better treatments for disease states.  
13 But we feel there's a lot to be gained. There's a lot of  
14 substantial improvement that could be made by integrating  
15 pharmacogenetics into the treatments that we now have and  
16 using this science as it matures for identifying more  
17 optimal doses for subsets of the population.

18           We'll continue to talk about this tomorrow.  
19 What we're going to emphasize is moving forward with the  
20 knowledge that we have on polymorphic drug-metabolizing  
21 enzymes that influence variability in drug response,  
22 especially toxicity. It's a challenging area. Many  
23 questions come up in the context of this, things like how  
24 much variability will genetics explain, how much of an  
25 effect on drug dose will genetics explain, how important is

1 it.

2 But I think more importantly where we're  
3 heading is to create a general construct for looking at  
4 improvement in existing therapies, existing therapies being  
5 approved drugs, to determine what criteria we ought to be  
6 thinking about that would warrant updating labels for  
7 products to optimize drug dosing using genetic information.

8 Last time we talked about specifically  
9 thiopurine, TPMT. Tomorrow we'll touch upon that, but  
10 we'll also be looking for a broad way to best program in  
11 this area and what we need to be thinking about in  
12 assessing data, assessing evidence to update labels. So a  
13 rational scientific basis.

14 Now, our fourth topic today is going to be a  
15 new topic. We'll actually talk about it tomorrow. We've  
16 been working pretty much over the last year in the area of  
17 drug-drug interactions. It continues to be a major problem  
18 if you read the current literature in JAMA and the New  
19 England Journal of Medicine about adverse drug reactions  
20 and the high fraction of those that are related to drug  
21 interactions. We have some ideas on revising our guidance  
22 on drug-drug interactions. We have some questions on  
23 transporter based drug-drug interactions. As was stated in  
24 our risk assessment workshop, some matters always need  
25 assessment in regulatory review, including new interactions

1 that we may not have paid as much attention to in the past,  
2 such as interaction involving glucuronidation and  
3 transporter interactions like P-gp.

4 So we're going to bring this topic forward  
5 tomorrow with some issues and questions. We'll talk about  
6 the use and extension of a classification system for 3A4  
7 inhibition for single and multiple drug interactions. This  
8 is a classification system that we can see moving towards  
9 label language for bringing some consistency to how we  
10 report drug interactions in the label.

11 And a big question that we frequently get from  
12 sponsors during the drug development process and we ask  
13 ourselves is when and how should the role of P-glycoprotein  
14 in drug interactions be investigated. This is an emerging  
15 area and we're beginning to see clinical evidence that this  
16 important and we'd like to develop a path forward that's  
17 reasonable and rational.

18 Each of the presenters is going to have some  
19 specific questions on the topic, but I'd like you to think  
20 about some of the broader questions that we have for the  
21 session. For example, you'll hear many proposals during  
22 the next day and a half. Aside from the specific questions  
23 about the subtopics of today's meeting, think about the  
24 rationality of these proposals. Are they reasonable? Are  
25 they feasible? Overall, do you think these will enhance

1 the quality of drug development and regulatory review? Are  
2 these the priorities that we should be looking at in our  
3 clinical pharmacology program?

4 We have works in progress, topics number 3 and  
5 number 4. That means we need input as we move forward with  
6 these programs and some advice on whether these objectives  
7 are worthwhile. And in particular, what is the best way to  
8 integrate new science and technology, whether it's  
9 genetics, whether it's P-gp transporter information, into  
10 therapeutics and regulatory review?

11 Well, that's the introduction to the lineup for  
12 today. I look forward to the discussion. It was a  
13 terrific discussion in October. Again, as we look around  
14 the table, the expertise of this committee is really  
15 substantial, and we're looking forward to defining and  
16 expanding upon the proposals that we're going to make  
17 during this meeting. Thanks.

18 DR. VENITZ: Thank you, Larry.

19 Now we're moving to our first topic. As Larry  
20 indicated, we're going to talk about exposure response as a  
21 way of justifying dose adjustments. The introduction will  
22 be given by Peter Lee. He's Associate Director of the  
23 Office of Clinical Pharmacology and Biopharmaceutics.

24 DR. LEE: Good morning. The first topic we're  
25 going to discuss today is quantitative risk analysis using

1 exposure response for determining dose adjustment for  
2 special populations.

3 This topic has been discussed in our previous  
4 meeting back in October 2002. In the last meeting we  
5 talked about three main topics. We had proposed a  
6 standardized approach to estimate the probability of  
7 adverse events in special populations using exposure-  
8 response information. We also discussed a regulatory  
9 decision tree for recommending dose adjustment in special  
10 populations. And lastly, we also discussed the potential  
11 application of utility functions for risk and benefit  
12 assessment.

13 So today we're going to present several  
14 examples to illustrate what we talked about in the last  
15 meeting. We're going to present examples of a standardized  
16 approach for using exposure-response information to adjust  
17 dose in special populations. We also are going to present  
18 one example of using population analysis to obtain PK/PD  
19 information from the large clinical trials. And in the  
20 last example, we're going to present a methodology to  
21 applying utility function for an optimal dosing strategy.

22 Just to give a little bit of background  
23 information, as you know, many of the NDAs may contain up  
24 to 20 or more clinical pharmacology studies, and in these  
25 studies different intrinsic and extrinsic factors may be

1 studied and these factors may influence the  
2 pharmacokinetics of the drug in these special populations.  
3 Therefore, we need a consistent approach to determine the  
4 dosing adjustment requirement in these populations.

5 Here's one example. In this particular  
6 example, we have about 11 factors that have been studied in  
7 the NDA. As you can see, the area under the curve of the  
8 drug may change depending on what factors from 0 percent,  
9 which is no change, to a 60 percent increase in the special  
10 populations.

11 So the question is, how do we make the dose  
12 adjustment according to the pharmacokinetic results? Where  
13 is the cutoff? Do we adjust the dose at 30 percent  
14 increase of AUC, or do we adjust the dose at 60 percent  
15 increase of AUC?

16 So the answer is that we had to look at PK/PD  
17 information and determine what is the clinical significance  
18 of this AUC change.

19 So there are some issues related to the dosing  
20 adjustment in drug labels of NDA submissions. Quite often  
21 we have seen inconsistency in dosing adjustment  
22 recommendations in the initial label of NDA submissions.  
23 Exposure-response information, as is required to interpret  
24 the pharmacokinetic change, is now always available in the  
25 NDA submission. The FDA reviewer had to conduct additional

1 exposure-response analyses in order to interpret the AUC  
2 change. Therefore, we feel that a standard for analyzing  
3 and interpreting the exposure-response information will be  
4 critical and beneficial to regulatory decision making in  
5 terms of dose adjustment in special populations.

6 So to improve the current status, in the last  
7 meeting we had proposed to develop and evaluate a  
8 standardized approach for the reviewer and possibly for  
9 industry to quantitatively assess the impact of exposure  
10 change on either safety or efficacy that results from a  
11 change in pharmacokinetics due to intrinsic and extrinsic  
12 factors.

13 This is the standardized approach that we had  
14 proposed in the last meeting. In this example, basically  
15 we have seen an increase of exposure of the test population  
16 compared to the reference. Using exposure-response  
17 information, we can estimate the distribution of response  
18 in both reference and test populations. If we could  
19 determine what is the critical value of response, which is  
20 considered clinical significance, which is the vertical  
21 line here, then we can calculate the probability of a  
22 clinically significant response based on the PK change, as  
23 well as the PK/PD relationship.

24 So in order to interpret the clinical  
25 significance of pharmacokinetic change, we need to have

1 | observed data in pharmacokinetics in special populations.  
2 | We also need data of the PK/PD relationship. With this  
3 | information, then we can estimate the probability of  
4 | adverse events in the special populations with the response  
5 | that is greater than the clinically significant critical  
6 | value.

7 |           However, in order to determine the clinically  
8 | significant critical values, we need to base it on a risk  
9 | and benefit assessment of the drug therapy. Currently  
10 | we're doing that on a case-by-case basis through a  
11 | discussion between clinical pharmacology and the medical  
12 | reviewer. But in the last meeting with the committee, we  
13 | also proposed that we can use a utility function to assess  
14 | the risk and benefit of pharmacokinetic change in the  
15 | special populations.

16 |           In the last meeting, we also discussed a  
17 | decision tree for dosing adjustment recommendations. Since  
18 | the last meeting, based on the recommendation from this  
19 | committee, we have made some modifications of the decision  
20 | tree, and this is the current decision tree. Basically we  
21 | ask a number of questions.

22 |           First, according to our current guidance, we  
23 | ask whether the 90 percent confidence interval of test over  
24 | reference is within the default no-effect boundary. A no-  
25 | effect boundary could be, for example, 80 to 125. Now, if

1 | the answer is yes -- we think it is the no-effect boundary  
2 | -- then there's no dose adjustment required for the special  
3 | populations. But if the answer is no, which means the 90  
4 | confidence interval is outside the boundary, then we ask  
5 | the next question, whether we have a PK/PD relationship.

6 |           If we do have a PK/PD relationship, then we  
7 | will take the standardized approach to estimate the  
8 | probability of adverse events and probability of  
9 | effectiveness in the special populations and ask the  
10 | question whether that's a clinically significant change  
11 | from the typical population. If it is considered  
12 | clinically significant, then we will recommend a dose  
13 | adjustment or precaution or warning in the drug label.

14 |           As I mentioned, there will be several examples  
15 | discussed in today's meeting. The first two examples will  
16 | be used to illustrate the use of the proposed standardized  
17 | approach for estimating the probability of toxicity in  
18 | special populations using exposure-response information.

19 |           And the next example will be used to illustrate  
20 | the potential utility of population analyses to obtain  
21 | exposure-response information from large clinical trials.  
22 | We think this is a very important topic because the large  
23 | clinical trials represent a unique opportunity to obtain  
24 | exposure-response information from the studies.

25 |           The last example will be used to demonstrate a

1 method of applying utility functions to optimize a dosing  
2 strategy.

3 Today we have four speakers to present the  
4 examples. The first speaker is Nhi Nguyen from DPE I, and  
5 she will present the first example. The second speaker is  
6 Dr. Jenny Zheng from DPE III. She'll present a second  
7 example of the standardized approach. And the third  
8 presenter will be Dr. He Sun from DPE II, and he will  
9 present the population PK/PD approach. The last speaker,  
10 our guest speaker, is Dr. Mats Karlsson from Uppsala  
11 University, and he will present an example illustrating the  
12 utility functions.

13 After each presentation, we're going to ask one  
14 or two questions to the committee. I'd like to present the  
15 questions now so that hopefully the committee can keep  
16 those questions in mind when listening to the  
17 presentations.

18 The first two questions relate to the  
19 standardized approach. We would like to ask, under what  
20 treatment circumstances, for example, intrinsic or  
21 extrinsic factors or therapeutic areas, would this  
22 standardized approach not be applicable? We also ask a  
23 second question. Does the exposure response differ between  
24 special populations and typical populations? If so, how  
25 can the differences be detected?

1           The next questions will be related to the  
2 population PK/PD analysis, and there are two questions  
3 related to that topic. The first question is, what are the  
4 utility and general limitations of linking pharmacokinetics  
5 obtained from the population analysis to the response  
6 endpoints? And the second question is what are the general  
7 considerations in using exposure response for dose  
8 adjustment in special populations, especially using the  
9 population approach to obtain the exposure response?

10           The last question is related to the utility  
11 functions, and the question will be, can the presented  
12 approach of utility function be generalized to other  
13 scenarios?

14           So with that, I want to introduce our first  
15 speaker of the examples, Dr. Nhi Nguyen. She will present  
16 the first example of the standardized approach.

17           DR. NGUYEN: Good morning. This morning you  
18 will hear a presentation on a method of analysis and how it  
19 was applied in regulatory decision making.

20           This slide will summarize how we did the  
21 analysis for this NDA.

22           The first step is to develop your exposure-  
23 response models. The exposure-response models should be  
24 based on large, randomized clinical trials, trials that  
25 explore a wide exposure range and include a large number of

1 people and are of the longest duration.

2           The second step is to have an expectation for  
3 your target window of exposure. How much benefit does one  
4 need and how much risk is one willing to assume?

5           The third step is to example what happens to  
6 exposure response when various intrinsic and extrinsic  
7 factors are introduced. Typically studies in special  
8 populations include pharmacokinetic information and are  
9 underpowered to provide good response data. So with the  
10 appropriate assumptions about exposure response in these  
11 special populations, we took the data from the special  
12 population studies, the individual data, not just the mean  
13 data, and integrated it into the exposure-response models  
14 and then determined the probability of effectiveness and  
15 safety.

16           So probability is on the y axis here and the  
17 sum of these bars equals 100 percent. So you can see that  
18 we not only have a feel for the maximum likelihood of  
19 benefit or risk, but we also have a feel for the tails of  
20 the distribution.

21           This slide summarizes how we did the analysis  
22 for this review.

23           I will take one more slide to further explain  
24 the last step, determining the probabilities. And for this  
25 example, I've chosen the QTc interval which is a surrogate

1 for torsade, a fatal ventricular arrhythmia. A clinical  
2 trial that examines QTc prolongation may have a  
3 distribution of baseline QTc intervals that look like this.  
4 Modeling the data may result in concentration QTc slope  
5 distributions that look like this. So if we want to see  
6 what happens when an intrinsic or extrinsic factor is  
7 introduced, we took the results from the PK study, and for  
8 this example I'm illustrating Cmax and overlaid it into the  
9 known concentration-QTc relationship.

10 So, in essence, we sampled from each of these  
11 distributions and created a virtual patient, and we did  
12 this 1,000 times to determine empirically what happens with  
13 a concentration and achieving a specific QTc. So by doing  
14 these simulations, we were able to test combinations of the  
15 tails of the distributions that were untested.

16 So that leads me to our objective which was  
17 only to quantitate the risk-benefit of a drug. A decision  
18 about what to do about the risk-benefit should be made with  
19 the whole review team or the domain experts.

20 In developing the exposure-effectiveness model,  
21 for the primary endpoint, we chose the largest clinical  
22 trial which was also of the longest duration. The primary  
23 effectiveness endpoint and pharmacokinetics were collected  
24 at baseline, week 2, and week 4. This study also explored  
25 the largest exposure, and these doses have been changed for

1 | the purpose of this presentation, but let's just say that 1  
2 | milligram a day is the sponsor's recommended starting dose,  
3 | with titration to 2 milligrams a day. So this study  
4 | explored a dose greater than and a dose less than the  
5 | sponsor's recommended dose.

6 |           Next we developed the exposure-safety or  
7 | exposure-risk models. For these models, we used the  
8 | adverse event data from all the pivotal clinical trials.  
9 | Now, you can imagine that large clinical trials may have  
10 | hundreds of adverse events. So after discussions with  
11 | other members of the review team, we prioritized these  
12 | adverse events and focused on these six: dizziness, edema,  
13 | liver toxicity, palpitations, tachycardia, and vertigo.

14 |           We also analyzed QTc prolongation because of  
15 | drug properties suggestive of QTc prolongation. For this  
16 | analysis, we chose the study that had the most information  
17 | on the time course of QTc prolongation. You will note that  
18 | this was a drug-drug interaction study, and the sponsor  
19 | used half the recommended starting dose. ECGs were only  
20 | measured up to 4 hours post dose, so there were some study  
21 | design limitations. And the study contained 24 hours of  
22 | drug concentration data.

23 |           So now that you've seen the exposure  
24 | effectiveness and the exposure risks that were assessed,  
25 | let's take a look at the models.

1                   This is the exposure-primary effectiveness  
2 model, and the asterisk in the following slides will  
3 indicate the mean Cmax of the sponsor's recommended  
4 starting dose of 1 milligram. These lines indicate the  
5 mean Cmax for the 1, 2, and 4 milligram dose, and you will  
6 note that the increase in concentration is more than dose  
7 proportional. The maximum effect was 7.6 and the  
8 concentration that produced half the maximal effect was  
9 about .2 units.

10                   In the following slides, I show a mean Cmax  
11 line to keep the slide clean, but really we are considering  
12 the entire distribution of individual Cmax's. So it's  
13 something that may look like this with some overlap between  
14 the 1 and 2 milligram dose. So that's the exposure-  
15 effectiveness model.

16                   When you look at the risks, each blue line on  
17 this slide indicates one individual's concentration/QTc  
18 prolongation relationship. The QTc corrections shown here  
19 are individual corrections obtained by nonlinear mixed  
20 effects modeling.

21                   There was a statistically significant  
22 relationship between concentration and QTc prolongation.  
23 However, you can see that there is a lot of variability.  
24 The starting dose in some patients results in no QTc  
25 prolongation. However, in other patients, it results in

1 substantial QTc prolongation. You will also note that we  
2 have very little data around the concentration of the mean  
3 Cmax for the 2 milligram dose.

4 For our analysis of other adverse events, we  
5 found three adverse events to be statistically significant  
6 and that was tachycardia, palpitations, and edema.  
7 However, since the analysis of all these adverse events was  
8 similar, I will only present one for the sake of time.

9 This slide shows the probability of tachycardia  
10 by the effective dose, and the effective dose is an  
11 adjustment of the actual dose to account for the saturable  
12 first pass of the drug. So, 1, 2, and 6 really correspond  
13 to 1, 2, and 4 milligrams of drug.

14 The probability of tachycardia was dependent on  
15 weight and dose. So in a 70-kilogram patient, you can see  
16 that there is a very small probability of tachycardia, and  
17 this probability does not increase with a six-fold  
18 effective dose increase. Whereas, in a 50-kilogram  
19 patient, there is about a 5 percent probability of  
20 tachycardia, and then this increases about 1 percent with a  
21 six-fold effective dose increase.

22 So now you've seen the exposure-effectiveness  
23 model, the exposure and QTc model, and then the probability  
24 of tachycardia by effective dose. So now we're equipped  
25 with the models necessary to interpret results of the

1 special population studies.

2 This slide shows results of two special  
3 population studies presented in terms of changes in AUC and  
4 Cmax. Ketoconazole with a half a milligram of drug  
5 resulted in a 13-fold increase in AUC and a 7-fold increase  
6 in Cmax, and grapefruit juice with 1 milligram of drug  
7 resulted in a 7-fold increase in AUC and a 6-fold increase  
8 in Cmax.

9 So the next step is to see how these data  
10 integrate into the known exposure-response relationship.  
11 This is the same figure you saw earlier, only it's smaller,  
12 of the concentration effectiveness. When a half milligram  
13 of drug is given, you would expect to see an effectiveness  
14 around 3. Taking ketoconazole with a half milligram of  
15 drug increases concentrations about 7-fold, reaching the  
16 Emax of about 7.6. Taking 1 milligram of drug with  
17 grapefruit juice results in about a 6-fold increase in  
18 concentration, and no additional effectiveness. Now,  
19 hypothetically if ketoconazole were given with 1 milligram  
20 of drug, we might expect to see a similar increase in  
21 concentration.

22 If we look at the concentration/QTc  
23 relationship, a half milligram of drug would result in  
24 about this amount of QTc prolongation. Taking ketoconazole  
25 with a half milligram of drug pushes patients from this

1 amount of QTc prolongation over to this amount. Taking 1  
2 milligram of drug with grapefruit juice results in about a  
3 6-fold increase in concentration, and the concentrations  
4 are then off the figure and we do not have QTc data there.  
5 And then hypothetically again, if ketoconazole were given  
6 with 1 milligram of drug, we might expect to see a similar  
7 response. So in this situation, we would not be able to  
8 make any conclusions about what happens at these higher  
9 concentrations on QTc prolongation because we do not have  
10 data there.

11 Now, I also want to remind you again that there  
12 is a distribution of Cmax's and slopes. So really we are  
13 looking at data that looks like this. So if we want to  
14 consider the worst case scenario, we have to consider both  
15 of these distributions, and the results of some of those  
16 simulations will be presented in the next slide.

17 Now, if we look at the probability of  
18 tachycardia, taking a half milligram of drug with  
19 ketoconazole in a 50-kilogram patient barely increases the  
20 probability of tachycardia, whereas taking grapefruit juice  
21 with 1 milligram of drug increases the probability of  
22 tachycardia about 1 percent in the 50-kilogram person. In  
23 a 70-kilogram person, you can see that this slope is pretty  
24 much a straight line and there is not much of an effect on  
25 the probability of tachycardia. Then again if ketoconazole

1 | were given with 1 milligram of drug, we might expect to see  
2 | a similar response as that with grapefruit juice.

3 |           So to summarize the data integration,  
4 | ketoconazole with a half milligram of drug results in a 13-  
5 | fold increase in AUC and a 7-fold increase in Cmax. And  
6 | this translated into a 4-unit effect, and we did  
7 | simulations to determine that 5 percent of the population  
8 | may have a prolonged QTc greater than 32 milliseconds.  
9 | Realize that these simulations are determined from the data  
10 | in the ketoconazole study. So we could present this data  
11 | in other terms, such as change from baseline or percent or  
12 | the time above a certain threshold QTc.

13 |           And then the probability of tachycardia with a  
14 | half milligram of drug and ketoconazole was barely  
15 | increased or affected. Grapefruit juice with 1 milligram  
16 | of drug resulted in a 7-fold increase in AUC and a 6-fold  
17 | increase in Cmax, and this translated into no additional  
18 | effectiveness, and we were unable to conclude anything  
19 | about the effect on QTc because we did not have data at  
20 | those higher concentrations. And then the probability of  
21 | tachycardia increased about 1 percent in the 50-kilogram  
22 | patient, and there was negligible effect in the 70-kilogram  
23 | patient. And then again if ketoconazole were given with 1  
24 | milligram of drug, we may expect to see a similar response  
25 | as that seen with grapefruit juice.

1           So at this point we would present the table to  
2 the review team and weigh the effectiveness and risks and  
3 realize the assumptions of our models. One is that it is  
4 the higher drug concentrations, not the intrinsic or  
5 extrinsic factor itself, that alters response. We  
6 recommended to conduct an appropriate QT study, one that  
7 explores a wider concentration range and one that collects  
8 QT data over 24 hours.

9           DR. VENITZ: Thank you, Nhi.

10           We have time for questions. Go ahead.

11           DR. SHEINER: I gather that the various parts  
12 of the various models were gathered from different data  
13 sets sometimes.

14           DR. NGUYEN: For the effectiveness, we used the  
15 largest clinical trial, and then for the safety and risks,  
16 we used the largest clinical trial and the other pivotal  
17 trials.

18           DR. SHEINER: I guess the question is this.  
19 You've got several models that are translating from A to B  
20 and then B to C and so on.

21           DR. NGUYEN: That's correct.

22           DR. SHEINER: And the question is, were enough  
23 of them gotten from the same set of people so that you  
24 could look at things like correlations? Does it turn out,  
25 for example -- not that it should -- that the people with

1 | the high concentrations essentially -- in other words, your  
2 | model is kind of assuming that this association that you  
3 | see, there are no correlations. So it's not necessarily  
4 | true that somebody who has, let's say, a raise in level and  
5 | doesn't have toxicity will also not have efficacy or  
6 | something like that.

7 |           You've got a set of relationships that  
8 | translates from concentrations before you add ketoconazole,  
9 | let's say, to afterwards, and then you map from  
10 | concentration to effect, but there is no part of this thing  
11 | that says, well, when you raise the concentration to the  
12 | ketoconazole, maybe the relationship of concentration to  
13 | effect is not the same. I'm not saying that it is, but you  
14 | don't have any evidence to say one way or another. Is that  
15 | right?

16 |           DR. NGUYEN: Yes.

17 |           DR. VENITZ: Any other questions?

18 |           DR. FLOCKHART: One thing directly to that  
19 | point. There is some data -- but this might be addressable  
20 | -- to suggest that ketoconazole itself can affect cardiac  
21 | repolarization.

22 |           DR. NGUYEN: That's right.

23 |           DR. FLOCKHART: You might have data on that  
24 | from the control arms of the smaller trials. If they show  
25 | no effect, that would be reassuring. It doesn't completely

1 get to Lew's point because it's still possible that the  
2 concentration-effect relationship is different in the  
3 presence of ketoconazole.

4 DR. CAPPARELLI: Just a clarification so that I  
5 understand the terminology. When you say probability of  
6 tachycardia, you're speaking of sinus tachycardia in this  
7 case, not torsade de pointes?

8 DR. NGUYEN: That's correct, sinus tachycardia.

9 DR. CAPPARELLI: I just wanted to be clear  
10 because I think one question that I had is, did you take a  
11 similar approach to heart rate that you did to QTc?  
12 Because your sinus tachycardia is going to be relative to  
13 where you start from, at least the risk.

14 The one thing that I think was brought up as a  
15 question is, are there extrinsic factors that we need to  
16 think about in these models? Clearly, strictly from a PK  
17 standpoint, I wouldn't expect a 50-kilogram person to have  
18 a different response based on weight. So I think there  
19 clearly is an extrinsic factor that's linked to the 50-kilo  
20 patient rather than the 70-kilo patient. But I'm not  
21 certain that it's gotten here. So I have some questions  
22 about the classification scheme based strictly on weight,  
23 and maybe linking to where their baseline heart rates would  
24 be of help from that standpoint.

25 DR. NGUYEN: Actually for the tachycardia

1 analysis, we would have preferred to analyze it by heart  
2 rate, but the data were collected like that. So it was  
3 sinus tachycardia, yes or no. So we did a logistic  
4 regression.

5 DR. CAPPARELLI: Was there an age effect or  
6 other disease effects that you looked at in terms of heart  
7 failure? It's kind of difficult to look at this and see  
8 where you expect a large change in concentration such that  
9 the 70-kilo person at the highest dose is going to have  
10 much higher concentrations than the 50-kilo person at the  
11 lowest dose. And yet, you're seeing this differential PD  
12 response. Without understanding what's causing that, I  
13 think it becomes very difficult to extrapolate from this  
14 aspect of the analysis.

15 DR. NGUYEN: Probably the 50-kilogram person  
16 did receive more of a dose, milligram per kilo, than the  
17 70-kilogram person. But the analysis -- they were given  
18 the same dose. So we didn't have concentration data to  
19 analyze data by concentration and probability of  
20 tachycardia. We only had dose data.

21 DR. SHEINER: Just a comment. I think we're  
22 getting at the fundamental problem that what you want to do  
23 is extrapolate to new circumstances, people having these  
24 other co-factors. You want to get some reasonable guess as  
25 to what's going to happen, what's dangerous and what isn't.

1 Yet, you're extrapolating from observational data based  
2 models, which is the hardest thing to do because you don't  
3 know where causality resides in those models.

4 One of the solutions in the past is to just not  
5 do it, and I don't think that's the right solution. But I  
6 do think there is no easy solution, and we have to be quite  
7 careful about things and recognize that we're talking about  
8 outer boundaries and recognize that we're talking about  
9 sort of worst case scenarios or maybe even best case  
10 scenarios. We can't be sure. We have to somehow get  
11 comfortable with the increased degree of uncertainty that  
12 arises in this activity. But as I say, I think we should  
13 do it because the alternative is even greater uncertainty.

14 DR. DERENDORF: There are a lot of straight  
15 lines in your concentration-effect and dose-effect  
16 relationships. Is there enough evidence that you have  
17 linear relationships between those parameters, particularly  
18 when you use them to extrapolate?

19 DR. NGUYEN: Which one are you referring to?

20 DR. DERENDORF: The concentration-QTc  
21 prolongation plot and then also the one below the dose-  
22 tachycardia plot. You just have straight lines there that  
23 suggest that concentration and effect are linked that way.  
24 Do you know that?

25 DR. NGUYEN: No. I mean, that was the data

1 that we had. So like for the QTc, they measured ECGs at 0,  
2 1, 2, and 4 hours post dose, and they had 24 hours of  
3 concentration data. So that straight line is the  
4 relationship between the concentration and QTc.

5 DR. DERENDORF: You think it is or you know it  
6 is?

7 DR. NGUYEN: Well, that's what it was in that  
8 population. They could have gone with a higher dose range,  
9 and then we could have seen what type of model the  
10 relationship is. So I don't know.

11 DR. DERENDORF: Then the other question that's  
12 related to the previous question that I really am puzzled  
13 with is this 50- versus 70-kilo situation where you have a  
14 6-fold dose and a 70-kilogram person doesn't have  
15 probability versus a one-sixth of a dose in a 50-kilogram  
16 person. So there would be quite different exposures and  
17 very different risks.

18 DR. FLOCKHART: It just means to me that the  
19 relationship between dose and weight isn't a simple linear  
20 relationship. You're right. There may be something else  
21 involved, but that's not uncommon.

22 DR. SADEE: I have a comment also on the  
23 variability from one patient to the next. You're looking  
24 at two interactions. It's maybe one of the classic  
25 examples in pharmacogenetics where you have a variety of

1 different genetic variance from one patient to the next,  
2 and actually that could affect the interaction between the  
3 two drugs so that you may have specific cases in the single  
4 patients that are totally different in their exposure than  
5 others. So there would be no way of extrapolating from  
6 that because you're disturbing the very relationship with  
7 the dose response that you're looking at.

8 DR. FLOCKHART: My difficulty here is the FDA  
9 is faced with the problem of trying to make a rational  
10 prediction. We can sit as academics and ding them all over  
11 the place for it. You know, you can't do this and you  
12 can't do that. But the reality is you have to try. And I  
13 think Lew's point is salient. We have to try and include  
14 the error.

15 DR. KEARNS: And that's the point that I think  
16 I want to make. Back to your ECG slide. It's not to be  
17 critical. It's quite exciting to see 20 percent of people  
18 have a response that way and then one outlier at the top  
19 who really had one.

20 But I was struck by the recommendation that you  
21 showed on your slide which was, okay, we did this. Now we  
22 go back and recommend a trial with more concentrations,  
23 wider range. And as Dr. Sheiner was kind of getting at,  
24 there's a lot that's riding on this extrapolation. I think  
25 from a public side, there's always the question of how much

1 additional time is that going to take. From a medical  
2 side, there's always the question about that one or two  
3 outliers. Are those the people who die in that trial  
4 because they have some hERG channel defect that's not  
5 recognized at the outset?

6 So I think trying to go back and do the  
7 diligent thing is to wire up the model as best you can.  
8 For instance, if that was a pediatric population and you  
9 looked at baseline QTc's, you'd see quite a different  
10 dispersion based on age for no treatment than you would in  
11 an older free-living population.

12 So is the applicability of the model approach  
13 -- can we go across populations? It depends I think. But  
14 to jump to the study and to add the patients, to add the  
15 concentrations, to maybe add the risk until you've taken  
16 all the flies that you can out of the ointment could be  
17 premature.

18 DR. VENITZ: Thank you, Nhi.

19 Let's move to the next presentation. Dr. Jenny  
20 Zheng. She is a pharmacometrics reviewer in the Division  
21 of Pharmaceutical Evaluation III. She's going to give us a  
22 second example.

23 DR. ZHENG: Good morning. Today I'm going to  
24 present another example to illustrate how the dose-response  
25 relationship was used for recommending dose adjustment.

1           In our review process, it's very often to see  
2 the pharmacokinetics of a drug is influenced by intrinsic  
3 factors such as age, gender, impaired renal and hepatic  
4 function and extrinsic factors such as drug-drug  
5 interactions. In this situation, we have to ask the  
6 question what is the clinical significant of the changes in  
7 concentrations.

8           Currently the decision will be made based on  
9 the clinical assessment based on the clinical experience  
10 and totality of the evidence. But the disadvantage of that  
11 approach is it's not a quantitative and standardized  
12 approach. The assessment could be pretty subjective. In  
13 other words, the decision may not be the same based on who  
14 makes the assessment and from what perspective. Therefore,  
15 we propose from a clinical pharmacology perspective to use  
16 the exposure-response relationship to bridge the response  
17 and exposure data to quantitate the influence after changes  
18 in the exposure.

19           The example I'm going to present will focus on  
20 the drug concentration increase and the safety assessment.  
21 This is drug Z. It's a noncardiac drug. From both  
22 preclinical and phase I studies, it shows that the drug  
23 caused QT prolongation and this QT prolongation is  
24 concentration dependent.

25           The phase I PK studies showed three factors

1 increased the drug concentration. In an age study, it  
2 shows that the mean steady state maximum concentration was  
3 100 percent higher in elderly subjects as compared with  
4 Cmax in young subjects. The renal study demonstrates  
5 steady state Cmax in severely renally impaired subjects was  
6 50 percent higher as compared with healthy subjects. And  
7 drug interaction studies showed ketoconazole increased the  
8 steady state Cmax by 60 percent.

9           Knowing the concentration increase in this  
10 situation, the question raised is, should dose be adjusted  
11 in elderly, renally impaired subjects or when co-  
12 administered with ketoconazole? To answer that question,  
13 we need to understand the effect of increase in drug Z  
14 concentration on the QT prolongation which will rely on the  
15 exposure-response relationship.

16           The exposure-response relationship was obtained  
17 from several phase I studies. They were all placebo-  
18 controlled crossover studies. The doses included in the  
19 study were a clinical dose and two times of the clinical  
20 dose and three times of the clinical dose. The higher dose  
21 is important here to provide the wide range of the  
22 concentration which is the key for obtaining exposure-  
23 response relationship. From all these phase I studies,  
24 blood samples were collected for drug measurement. Also  
25 the QTs were measured.

1           The results of the analysis are shown in this  
2 slide. The QT prolongation is represented as delta QTc,  
3 which is the QT change from the baseline. So the  
4 association between the delta QTc with the concentration  
5 was described by a simple linear regression model. The  
6 linear mixed effect model was used for analyzing these  
7 data. The dashed lines represent individual regression  
8 lines. The solid line represent the population regression  
9 line. The wider band of the lines indicates that the  
10 inter-subject variability is quite high. The estimated  
11 slope ranged from 1.5 to 7.6, indicating that for some of  
12 the subjects, the delta QTc change is sensitive to the  
13 concentration change. In some of the subjects, the change  
14 is not quite as sensitive.

15           An outlier analysis is a very important part of  
16 QT assessment. We want to know how many subjects would  
17 experience the delta QTc longer, for example, 10  
18 milliseconds, 20 milliseconds, 30 milliseconds, or 40  
19 milliseconds. Unfortunately, the phase I study usually  
20 included a limited number of subjects which limits its  
21 ability for that type of analysis.

22           For the phase III study, even though hundreds  
23 of subjects may be included in that analysis, the QT  
24 measurement is not as intensive as the phase I study. So  
25 it's difficult sometimes to capture the outlier from the

1 phase III study. On the other hand, if you're interested  
2 in the special population, even a phase III study may not  
3 provide sufficient number, for example, severely renally  
4 impaired subjects.

5 So in order to make an outlier comparison  
6 between the population, a simulation exercise was conducted  
7 here. Most specifically, the phase I data, the  
8 concentration data was modeled assuming the logarithmic  
9 distribution in the PK parameter. Using that model, 2000  
10 maximum concentration was simulated for young subjects,  
11 elderly subjects, for renally impaired subjects. The same  
12 approach is used to simulate 2000 Cmax for when  
13 ketoconazole is co-administered with the drug Z. So we  
14 have 2000 concentration in each special population, special  
15 situation. Then we used the exposure-response relationship  
16 as described in the previous slide to predict delta QTc.

17 The results of the age effect are presented in  
18 this slide. The data is presented as the percent of the  
19 subjects who would have the delta QTc longer than 10  
20 milliseconds, 20 milliseconds, 30 milliseconds, 40  
21 milliseconds. These results indicated that about 2 percent  
22 of young subjects would have delta QTc longer than 40  
23 milliseconds and for the elderly, the percentage will  
24 increase to 7.3 percent. So for delta QTc longer than 30  
25 milliseconds, in young subjects it's about 8 percent; in

1 | the elderly, it's about 19 percent. So a similar trend  
2 | could be seen for a delta QTc longer than 20 milliseconds  
3 | and 10 milliseconds.

4 |           This slide presents the results for renal  
5 | function. As you can see, most subjects with severe renal  
6 | impairment would have longer QT prolongation than the  
7 | normal subjects. For example, for the normal renal  
8 | function subjects, 2 percent would have delta QTc longer  
9 | than 40 milliseconds, but if you have severely renally  
10 | impaired function due to the concentration increase, there  
11 | will be 5 percent of the subjects who would have a delta  
12 | QTc longer than 40 milliseconds.

13 |           The results in this slide show ketoconazole  
14 | increased the percent of subjects who experienced a certain  
15 | extent of delta QTc. Like if you're taking the drug alone,  
16 | 2 percent of subjects would experience delta QTc longer  
17 | than 40 milliseconds. But if you take drug Z with the  
18 | ketoconazole, the percentage will increase to 6.2 percent.

19 |           The percent of subjects with delta QTc longer  
20 | than 40 milliseconds is summarized in this slide. You can  
21 | see that the risk of having a delta QTc longer than 40  
22 | milliseconds is higher in elderly subjects, in severely  
23 | renally impaired subjects, and when the drug is co-  
24 | administered with ketoconazole.

25 |           Examination of the creatinine clearance

1 indicated that the age effect might be partially attributed  
2 by reduced renal function. So in the age study, creatinine  
3 clearance was 50 percent lower in elderly subjects as  
4 compared with the young subjects. So it's believed that  
5 age effect would be reduced if the renal function effect  
6 was corrected by dose reduction.

7           Since the consequence of the worst event could  
8 be very severe, the question was asked in the review team,  
9 what would be the effect of ketoconazole in subjects with  
10 severe renal impairment? Not many subjects would belong to  
11 this group, even from a phase III study. So in order to  
12 make that assessment, a simulation was conducted.

13           First, steady state Cmax in severely renally  
14 impaired subjects was simulated as I described earlier. In  
15 the second step, the Cmax ratio of drug Z at presence and  
16 absence of ketoconazole was obtained from the crossover  
17 study so that the ratio actually characterized the  
18 ketoconazole effect. From that study the ratio ranged from  
19 1 to 4. So the combined effect for both factors was  
20 simulated by just randomly multiplying the maximum  
21 concentration from step 1, which is the maximum  
22 concentration for severely renally impaired subjects, and  
23 the ratio from step 2 which characterized the ketoconazole  
24 effect.

25           The results are shown in this slide. As you

1 | can see, 19 percent of subjects who are severely renally  
2 | impaired would experience a delta QTc longer than 40  
3 | milliseconds when co-administered with ketoconazole.

4 |           This slide just simply summarizes all of the  
5 | factors, the effects. It summarizes young subjects. It's  
6 | the percent of subjects with a delta QTc longer than 40  
7 | milliseconds. For young subjects, it's about 2 percent.  
8 | In the elderly, it's almost triple the percentage, up to  
9 | 7.3 percent, and more than double that percentage in  
10 | severely renally impaired subjects. And when drug Z is co-  
11 | administered with ketoconazole, the effect could be very  
12 | dramatic if the two factors are combined.

13 |           That analysis leads to the conclusion that the  
14 | increase of concentration by age, severe renal function,  
15 | and co-administration with ketoconazole resulted in  
16 | increased number of subjects with a delta QTc longer than  
17 | 40 milliseconds. The effect is more significant when two  
18 | factors are combined.

19 |           Based on that analysis and the consideration of  
20 | the nature of an adverse event, a dose reduction was  
21 | recommended in severely renally impaired subjects. A dose  
22 | reduction was also recommended when drug Z is co-  
23 | administered with ketoconazole.

24 |           DR. VENITZ: Thank you, Jenny. We have about 5  
25 | minutes for questions.

1 DR. DERENDORF: I think it's the same issue as  
2 in the last case. You're assuming that the exposure--  
3 response relationship that you got from your phase I study  
4 is a constant and it doesn't change in elderly or in severe  
5 renal impairment. So the calculations that you're making  
6 are all focused on exposure, and then at the very end, you  
7 convert that into expected --

8 DR. ZHENG: I don't quite understand your  
9 point. Actually the delta QTc versus concentration, that  
10 is the relationship between the effect versus  
11 concentration. I don't think we make any assumption with a  
12 constant concentration.

13 DR. DERENDORF: No, not constant concentration.  
14 But the relationship between the exposure that you have in  
15 your different cases and the outcome -- you take that  
16 linear relationship that you have from your phase I study  
17 where you have concentration versus change of QTc and apply  
18 that to all of these cases assuming that this relationship  
19 holds true for all of these.

20 DR. ZHENG: Okay. So you have the problem with  
21 the extrapolation from the young healthy subjects to the  
22 elderly population.

23 DR. DERENDORF: I don't see any evidence that  
24 it holds.

25 DR. ZHENG: In one of the phase I studies, they

1 included not only the young subjects but the elderly. So  
2 we do look at the relationship there. We don't see much  
3 difference in terms of the slope, the relationship. So  
4 that's one of the information we could have.

5 In terms of the drug-drug interaction and the  
6 severely renally impaired subjects, yes, we don't have data  
7 to show that they are going to have the same relationship.  
8 But you are right. That's the assumption we have to make  
9 for these type analyses.

10 DR. CAPPARELLI: Just as a follow-up on that,  
11 because this is a recurrent issue -- I mean, this  
12 particular QT prolongation looking at drug concentration  
13 effects -- has there been a systematic look at several of  
14 these drugs looking at especially, say, with renal  
15 impairment where you're going to have changes in  
16 electrolyte abnormalities and looking really at sort of a  
17 population dynamic model to identify the covariates?

18 So I think as Hartmut was saying, we're going  
19 forward with the assumption that the exposure-response  
20 relationship is totally uncorrelated with the changes in --  
21 the disease states that are causing the changes in PK. I  
22 think this is actually a great example where maybe in some  
23 across-study evaluations, one could actually look at some  
24 of these populations not only at the variability in  
25 response in subpopulations, but maybe the electrolyte

1 differences in your renal failure patients may change that  
2 slope entirely. So adding these effects as we go along in  
3 the chain, it's nice along the way to test some of these  
4 assumptions.

5 DR. ZHENG: I think if we could have enough  
6 information, definitely that's a good thing to do. But I  
7 think here, unfortunately we just don't have that much  
8 information. So the focus here is simply the effect of  
9 concentration on the delta QT.

10 DR. SHEINER: Of course, the answer is get more  
11 information. But the answer to the problem when you don't  
12 have enough information is not that you have to make some  
13 assumption and go with it, but that you have very carefully  
14 display. It is sort of what I was indicating before. You  
15 have to carefully display the limits of your knowledge.

16 So if I look at, for example, the last page and  
17 the last several slides you showed, you've got these bars  
18 that are just heights, the amount of change with the  
19 elderly or renal failure, and there are no uncertainty  
20 intervals on them. Yet, this is exactly what you need to  
21 pay a lot of attention to, it seems to me, in this kind of  
22 a situation so that you can have a rational dialogue with  
23 other people.

24 And where do the uncertainties come from? And  
25 we have techniques whereby you say, well, look, I have this

1 | linear relationship between QTc change and the  
2 | concentration, but I could put in some uncertainties about,  
3 | let's say, whether it applies to other populations. Then I  
4 | can actually build that into my projections, and I can see  
5 | that instead of having 30 percent of people above QTc of  
6 | 40, it will be anywhere from 10 to 50 percent, or whatever  
7 | the numbers are.

8 |           That's the point. You've got the computers to  
9 | do it. It doesn't cost money. And that's the way I think  
10 | to deal with the problem that there are so many assumptions  
11 | that have to be made, sensitivity analyses and honest  
12 | uncertainty intervals which involve model uncertainty as  
13 | well as data uncertainty. And then everybody is talking  
14 | about the same thing. It may well be that the conclusion  
15 | stays the same.

16 |           DR. LEE: Dr. Sheiner?

17 |           DR. SHEINER: Yes.

18 |           DR. LEE: To follow up, if we don't know the  
19 | true relationship in different populations, how do we build  
20 | into the model the uncertainty due to population  
21 | difference?

22 |           DR. SHEINER: Well, let's say we'll talk about  
23 | the average slope. Let's say you're willing to assume it's  
24 | linear, but it's the average slope that's different in  
25 | different populations. So then you just talk to a bunch of

1 | people and you say, how big do you think it could be, and  
2 | you just build that uncertainty in. Now it spreads out all  
3 | of your predictions, and it means there's a larger fraction  
4 | of people who have low values, but there's a larger  
5 | fraction of people who have high values.

6 |           So it's a matter of assessing the risk. It's  
7 | what's the probability based on everything we know,  
8 | including all the uncertainty, that the value will be  
9 | greater than this. And that will be your most educated  
10 | guess.

11 |           The point is it will be everybody's most  
12 | educated guess, and anybody who says I don't think that  
13 | will happen, you'll say, well, you're pointing to the 40  
14 | percent that's still below the line because we have  
15 | uncertainty. And I understand you're betting on that 40  
16 | percent, but we're worried about the 30 percent. So that's  
17 | the way we're going to go.

18 |           The point is you're never going to get the  
19 | answer from doing the numerical calculations. All you get  
20 | is an honest statement of what you know and that everybody  
21 | can agree on. I think that's the big thing, is that  
22 | everybody can agree this is the state of our knowledge.  
23 | Therefore, if you're going this way and I'm going that way,  
24 | it's because we're valuing different outcomes differently,  
25 | and so the expected value comes out differently.

1 Another example here of a place for an  
2 opportunity for this is in the discussion -- well,  
3 actually, I'll let it go. But I think you get the idea.

4 DR. VENITZ: Let me just follow up to that,  
5 Jenny. Whenever you do an outlier analysis, which is  
6 really what you're trying to do, worst case scenario, what  
7 are the few that have a large change in QTc, distribution  
8 assumptions are very important in terms of what your final  
9 outcomes are. I look at your simulation slide. You're  
10 talking about a logarithmic distribution. I'm assuming you  
11 mean a log normal distribution.

12 DR. ZHENG: Right.

13 DR. VENITZ: How did you then actually simulate  
14 the changes due to the disease states or the drug-drug  
15 interaction? Did you just change the mean or did you  
16 change variances as well?

17 DR. ZHENG: Actually I just changed the mean  
18 because the model is fitted to the raw data. For example,  
19 the young subjects -- we modeled that. So we know the mean  
20 for that group.

21 DR. VENITZ: But what about the variance? I  
22 guess what I'm worried about, whenever you look at outliers  
23 and you have a change in variance, in other words your  
24 elderly or your renal population are probably more variable  
25 than your young reference population even in terms of

1 kinetics.

2 DR. ZHENG: We used the same model to model the  
3 data for young subjects and the data for elderly. I mean,  
4 the same compartment model.

5 DR. VENITZ: But in terms of your parameter  
6 variability, did you use the same variance in your --

7 DR. ZHENG: No. The data --

8 DR. VENITZ: So you used the actual data  
9 variance.

10 DR. ZHENG: Yes. I used the real data  
11 variance.

12 DR. VENITZ: Just to follow up on that, I'm  
13 assuming when you looked at your slopes, you assumed that  
14 the slopes followed normal distribution or log normal  
15 distribution?

16 DR. ZHENG: I did that analysis using NONMEM.  
17 So it's an additive model. So it's normal distribution.

18 DR. VENITZ: Well, based on what I've seen or  
19 based on the previous example, that may not be a good  
20 assumption. It could be that you just have a few outliers  
21 and have very steep slopes, but the rest of them have a  
22 fairly shallow slope. Whenever you do an outlier analysis,  
23 just as a general rule, the distribution assumption of the  
24 variances that you make really determine what your final  
25 outcome is.

1                   In addition to that, I would reinforce what Dr.  
2 Sheiner said, and that is, I was missing the fact that you  
3 didn't really give us an idea of the uncertainty --

4                   DR. ZHENG: Right. I think that's something I  
5 should have included. Probably I don't have enough  
6 information to speak to severely renally impaired subjects,  
7 what the relationship will be. But I do have the  
8 information about uncertainty of the parameter estimate.  
9 So I agree 100 percent.

10                  DR. VENITZ: Just look at your three slides  
11 where you tell us what happens for the young individuals.  
12 Let's say the QTC of less than 10 is 41, 44, and 4 and  
13 42.7.

14                  DR. ZHENG: Right.

15                  DR. VENITZ: Those are three different  
16 simulations. So you just do that a couple times and you  
17 know how much --

18                  DR. ZHENG: Right, yes. The uncertainty of  
19 that estimate should have taken into that exercise. I  
20 agree.

21                  DR. VENITZ: I think we have one more question.

22                  DR. JUSKO: I imagine you're using the best  
23 available metrics on evaluating the exposure-response  
24 relationships. But you might consider using the  
25 availability of these data to examine additional

1 possibilities. For example, if you look at absolute  
2 changes in QTc, there might be the possibility that a  
3 change of 10 or 20 in the elderly is a bigger problem than  
4 a change of 10 or 20 in the young subjects. And perhaps  
5 something in relation to baseline values should be  
6 considered.

7 Secondly, you're using Cmax as the exposure  
8 index, and it would seem to me that in addition to that,  
9 the duration of time that a person has an abnormal QTc  
10 interval could be an additional hazard that could be  
11 factored in in exploring bigger sets of data as you may be  
12 doing.

13 DR. VENITZ: Okay. Thank you. Thank you,  
14 Jenny.

15 Before we go on a break, just an announcement.  
16 For those of you on the committee who haven't handed in  
17 your lunch orders, now is the time to do it or you're going  
18 to starve.

19 With that, we're going to reconvene at 10:10.

20 (Recess.)

21 DR. VENITZ: I'd like to reconvene the meeting  
22 please.

23 All right. While Peter is posting the  
24 questions that the FDA is asking the committee, are there  
25 any additional specific questions to the two presenters,

1 | Dr. Zheng and Dr. Nguyen?

2 |           DR. KARLSSON: Yes. Just a question regarding  
3 | this last presentation. The elderly showed quite a change  
4 | when you looked at the distribution; 7.3 percent would be  
5 | above 40 milliseconds. Maybe that would be mitigated by  
6 | the renal impairment dose adjustment, although I guess even  
7 | in the elderly population, it wouldn't be that many that's  
8 | below 30 mls per minute in the elderly population. But I  
9 | guess you could look at that through simulations as well.

10 |           But another question is, when looking at the  
11 | percentage of a particular subpopulation that's outside, is  
12 | it only the percentage within the population you're looking  
13 | at, not the size of the population at all? Because I guess  
14 | the elderly population is very large in absolute numbers  
15 | compared to maybe severe renal impairment or ketoconazole.

16 |           Did I make myself clear?

17 |           DR. ZHENG: Actually could you repeat your  
18 | second question?

19 |           DR. KARLSSON: Well, if we're looking at dose  
20 | recommendations, is it only the percentage within the  
21 | population that's interesting? Is it not also the size of  
22 | the population as such?

23 |           DR. ZHENG: The simulation I did is 2,000  
24 | subjects. So it may change if you change the sample size  
25 | to --

1 DR. KARLSSON: No. In essence, what I mean is  
2 that the elderly population is maybe like 80 million people  
3 in the U.S. and the severe renal impairment population is  
4 maybe 1 million. I don't know. 2 million. So that would  
5 come into play as well when making dosing recommendations.

6 DR. ZHENG: Okay. Yes, I think at the time we  
7 make a decision, we should consider the population who use  
8 the drug, the impact.

9 DR. LESKO: Mats, I'm not clear how you would  
10 consider it, though. Would you consider it in the context  
11 of saying that equal changes in a population that's larger  
12 number would get more weight in a dose adjustment scheme?  
13 Or how would you think about it as far as that issue goes?

14 It's like saying because the elderly population  
15 is so large, there's a greater overall risk to public  
16 health than there would be with patients with severe renal  
17 function. But it would seem in labeling a product, I'm not  
18 sure that would be taken into account for dose adjustment.  
19 Or if it is, I'm not sure how it would be.

20 DR. SHEINER: It would be if you were thinking  
21 about what dose sizes to make, for example.

22 DR. LESKO: Okay, from a manufacturer's  
23 standpoint.

24 DR. SHEINER: It's more convenience. If 90  
25 percent of people are going to use this to make them safe

1 and 10 percent -- you know. They're the ones who are going  
2 to get out their pocket knives and hack the thing in half.

3 DR. VENITZ: Peter, do you want to review the  
4 questions for the committee one more time?

5 So those are the questions that we are asked to  
6 discuss with regard to the approach that we just two  
7 examples of using exposure-response information as a way of  
8 predicting probabilities of, in this case, adverse events  
9 as a way of deciding about dosing adjustments or not.

10 Lew?

11 DR. SHEINER: Did you want discussion on that?

12 DR. VENITZ: Yes.

13 DR. SHEINER: Well, I think we're back to where  
14 we were sort of in the very beginning. It's a good thing  
15 to do, but if it's not done with a little extra care, then  
16 maybe it's not a good thing to do. So I think it really  
17 comes down to that.

18 As I was saying to Jenny at the break, if you  
19 do a well-designed, even clinical experiment in which you  
20 know exactly the question you want to ask, you've got  
21 adequate data by good design, and you analyze it, in a  
22 funny way the statistics are relatively unimportant. The  
23 signal to noise is usually pretty high, and it's usually  
24 pretty clear what the result is. Yet, that's where most of  
25 the statistics that most of us have seen have been applied,

1 in making sure that type I error is controlled. And  
2 there's nothing the matter with that. It's a good idea.  
3 But it's not really where you need it. And it's not that  
4 you need sure inference here because you can't get sure  
5 inference when you're this uncertain.

6 But what we need is we need to have a good way  
7 of displaying what we know so that everybody is looking at  
8 the same thing and understands the uncertainties. It seems  
9 to me what we didn't see were two ways in which I feel that  
10 that needs to be done.

11 One, as soon as you generate a simulation  
12 model, you have to show me that that simulation model can  
13 simulate the data it was derived from. There are lots of  
14 different ways of going about convincing me of that. Some  
15 of them treat the data it was derived from as though they  
16 were new by leaving it out and then making the thing and  
17 remaking. There are many, many different techniques. But  
18 the fundamental idea is show me that the sorts of  
19 conclusions that you want me to draw about extrapolations  
20 are at least verified on the data that you built the thing  
21 from when you apply them to those data. So that's number  
22 one. I want to see a lot of that.

23 And then I want to see a real honest  
24 uncertainty in my simulation. We all understand we're not  
25 talking about anything that's sure here. But I want to

1 know how big the uncertainty is and I want to have some way  
2 of knowing where it came from. I personally really want to  
3 see model uncertainty as well as data uncertainty. In  
4 fact, I'm more concerned about model uncertainty than data  
5 uncertainty.

6           What do I mean by that? I mean if you have 100  
7 patients from whom you've generated the data set, I  
8 understand the next 100 patients are going to have somewhat  
9 different numbers, and so you're going to get somewhat  
10 different conclusions. And we all estimate that  
11 uncertainty, and it's not that tough to estimate. And  
12 sometimes it isn't that large because we have a fair number  
13 of patients.

14           What's really uncertain is whether or not the  
15 relationship we discovered on this population is going to  
16 apply to that population. There we have no data if we  
17 haven't studied that population, if we're extrapolating to  
18 it. So there we need just some reasonable guesses. How  
19 different have populations been with respect to this kind  
20 of thing in the past with similar sorts of things? This is  
21 where the science comes in. This is where the judgment  
22 comes in. But you can build those model uncertainties in,  
23 and I can get to see how big they are. That's kind of like  
24 a robustness test. It's kind of like a way of saying how  
25 much will conclusions vary if I vary my assumptions.

1                   Assumptions there will always be. I'm not  
2                   against assumptions. What I'm against is making  
3                   assumptions look like facts. It turns out that the things  
4                   we don't know anything about we put the least uncertainty  
5                   on, and that's very peculiar. We choose a form of a model.  
6                   So it's a bi-exponential. And then we say, boom, that's  
7                   it. No questions about that. And that's the thing we know  
8                   the least well. What we do know well is the data we  
9                   observed, and that we say, aha, that's got noise. So it's  
10                  kind of like backwards. I want to see the model  
11                  uncertainty.

12                  This is not to be critical. I believe in  
13                  modeling and I believe in trying to be quantitative about  
14                  conclusions. But without that kind of thing, you won't  
15                  ever get people around a table to agree on what you know,  
16                  and if you can't do that, they won't agree on where you  
17                  ought to go.

18                  DR. VENITZ: Any other comments to the first  
19                  question? Can we think of any specific examples or  
20                  circumstances, therapeutic areas where this approach may  
21                  not be applicable?

22                  DR. KEARNS: Yes. I think one glaring one --  
23                  and Dr. Sheiner again speaks of model uncertainty. As I  
24                  might understand it, it would be in the context of the  
25                  facts of an experiment that we saw examples of. But as Dr.

1 Flockhart mentioned, for QTc studies where a patient may  
2 ingest a medicine that can have effects on its own, that's  
3 not necessarily part of model uncertainty. I don't know  
4 that you could predict the rate of co-ingestion of those  
5 drugs. And I would argue that with some combinations that  
6 are available, the relationships that you so nicely shared  
7 with us could look quite different.

8           So are there treatment circumstances that the  
9 approach might not be applicable as it was presented? Yes,  
10 and I think that's one example.

11           DR. VENITZ: What about the second question? I  
12 think that's something that we talked about last time.  
13 Differences in the exposure-response relationship between  
14 the typical and special populations.

15           DR. DERENDORF: Yes. I think that there are  
16 some examples in the literature where there are clearly  
17 differences in exposure-response relationship. If you  
18 think of benzodiazepines, for example, with the sensitivity  
19 and all the patient changes, so for the same concentration,  
20 you'll get a different response. But there's actually very  
21 little hard data available in the literature because it's  
22 hard to study. If you want to do it right, you have to do  
23 a complete PK/PD study, and just focusing on exposure is  
24 simply easier and therefore it's done more frequently. But  
25 I think that's what we need: more clean PK/PD studies in

1 different populations to see how much variability and how  
2 much systematic change we have in the exposure-response  
3 relationship.

4 DR. VENITZ: Larry.

5 DR. LESKO: There are actually two levels of  
6 uncertainty that we're dealing with. The first -- and I  
7 think it was the first example. We were talking about  
8 exposure-response relationship across various special  
9 populations. The second example illustrated a different  
10 problem and that was the assumption of exposure-response  
11 relationships between healthy volunteers and then patients.  
12 That's just the fact of the way, at least currently, drugs  
13 are developed. So we have to find ways to think about  
14 that, and it would seem there are two things I thought  
15 about.

16 One was in the pediatric decision tree or in  
17 the pediatric rule, we make the assumptions, or at least we  
18 ask the questions, about disease progression being the same  
19 in adults and kids and whether or not the mechanism of  
20 action in the exposure response is the same in adults and  
21 kids, and then we proceed down a path of logic that  
22 requires perhaps a dose being changed based on simply  
23 pharmacokinetic differences to achieve the same type of  
24 exposure.

25 It gets to the question, though, is my base

1 | assumption that exposure response is similar in the absence  
2 | of hard data, and then I look for reasons, perhaps  
3 | mechanistic reasons, why it wouldn't be, or do I look and  
4 | say, well, let me assume it's different and find  
5 | mechanistic reasons that it should be the same? For  
6 | example, in the pediatric adult area, you might ask the  
7 | question, does a beta receptor's either density or  
8 | sensitivity change and is it safe to assume that with a  
9 | beta blocker I'm going to have similar exposure-response  
10 | relationships?

11 |           It just seems to me that there's a way to think  
12 | about it mechanistically if one understands the way the  
13 | drug is working and the changes that are occurring in the  
14 | special population. Like in the QTc, for example, if renal  
15 | patients have altered potassium levels, then we know that  
16 | affects sensitivity in terms of drug effects on QTc, and  
17 | that could be kind of a rationale for including some  
18 | assumption about heightened sensitivity or something like  
19 | that or a change in the exposure-response curve. But in  
20 | the absence of that kind of mechanistic information, it  
21 | would seem we have to go with the assumption that these  
22 | exposure-response relationships are the same.

23 |           I mean, does that line of thinking make sense?

24 |           DR. VENITZ: That would be the way I think. My  
25 | default position is there is no difference between my

1 | typical population and the special population unless I have  
2 | either hard data to show that it is, which is rare, or I  
3 | have mechanistic reasons based on the pathophysiology of  
4 | the disease of that special population and/or the mechanism  
5 | of action of the drug to suspect that it is. Then I either  
6 | have to question the need for additional studies to show  
7 | whether it exists or not or build it in as an uncertainty  
8 | in my model.

9 |           DR. KEARNS: And Larry, I think another answer  
10 | to your question that you posed is it depends on the  
11 | surrogate chosen to assess effect. For example, if we look  
12 | at studying a proton pump inhibitor in a child, there's  
13 | convincing physiologic evidence that the maturation of the  
14 | proton pump occurs very early and that the children respond  
15 | to those medicines in ways that are very similar to adults.

16 |           But if you go pick a surrogate far from the  
17 | tree of effect or drug action and you apply it and say, is  
18 | gastroesophageal reflux in a 6-month-old the same as in a  
19 | 46-year-old, and then try to make arguments about bridging,  
20 | you'll find that the pillars that you've constructed the  
21 | bridge out of are not worth traversing. So it depends on  
22 | how close your surrogate is to where the medicine works.

23 |           DR. VENITZ: Something else I think we're going  
24 | to talk about in a minute that I would also consider -- and  
25 | I'm pretty sure you do that in your briefings with the

1 | medical reviewers -- is what are the consequences of being  
2 | wrong. In other words, what's the utility of whatever  
3 | assumptions you may not be very certain about? Sometimes  
4 | that severity or that consequence may be relatively  
5 | inconsequential, and then it really doesn't make a  
6 | difference. Forget the fact that you have statistical  
7 | uncertainty associated with it.

8 | DR. LESKO: There are many ways these kind of  
9 | data are handled for purposes of dosing adjustment, and  
10 | maybe that's one of the reasons we're trying to arrive at a  
11 | standardized approach to doing it.

12 | It would seem the safest way of doing it is to  
13 | simply adjust the dose based on an area under curve change.  
14 | The question then becomes what is the threshold level for  
15 | that area under curve change to trigger that. And that's  
16 | where the difference of opinion occurs because you don't  
17 | have a method on the table that allows one, as people have  
18 | said, to discuss this in a quantitative way.

19 | So there may be, in essence, a lot of  
20 | likelihood of not optimal dosing by doing it that way,  
21 | either making dose adjustments when you don't need them or  
22 | not making them when you should based on people's  
23 | interpretation of the data without a methodology to  
24 | discuss.

25 | DR. VENITZ: But in the examples that you've

1 | shown, the endpoints, as far as I can understand them --  
2 | QTc. That's a surrogate of fatal arrhythmia. So you're  
3 | worried about a potential fatal consequence. There's a  
4 | high, in my terminology, negative utility associated with  
5 | it. On the other hand, things tachycardia or palpitations  
6 | would rank much lower on the totem pole of my concerns.  
7 | But I'm not sure how you quantitatively incorporate that  
8 | short of using utility functions.

9 |           DR. FLOCKHART: This is an editing, small  
10 | point. If you're going to talk about changes in AUC of a  
11 | compound, I think particularly when you're talking about  
12 | the QT -- but this may be representative of other things --  
13 | the area under the exposure curve is not necessarily the  
14 | main thing. The time of exposure to a drug is not the  
15 | trick. Parameters like the rate of rise to Cmax can be  
16 | very important and the QTc max at a given dose can be very  
17 | important. There are dis-relationships, blocks between the  
18 | time-effect curve so the time of the concentration Cmax is  
19 | absolutely not necessarily the time of the effect Cmax. It  
20 | can be later. So it's possible there would be situations  
21 | where a parameter other than a change in the PK AUC would  
22 | be the appropriate parameter. It could still be a PK  
23 | parameter, but it might be Cmax itself or it might be the  
24 | rate of rise to Cmax. And that would be a drug-specific  
25 | question.

1           If, for example, you looked at quinidine,  
2 quinidine has a very poor relationship to the QT interval.  
3 If you were able to talk about torsade, what really matters  
4 there is the rate of rise, how quickly you get to Cmax.  
5 And if you get to a very nasty Cmax very slowly, it's not a  
6 terribly dangerous thing it looks like, but if you get to  
7 the same Cmax very quickly, then it can be a very dangerous  
8 thing.

9           That's a drug-specific question, and I would  
10 caution about always using AUC. I mean, you can think of  
11 examples related to Greg's example too. Above a certain  
12 point, changes in the AUC of a proton pump inhibitor do  
13 nothing. They're meaningless. I guess that just  
14 emphasizes the point that you need to know the  
15 pharmacodynamic relationship first.

16           DR. SHEINER: True as what you say is, I quake  
17 at the notion that things as uncertain as area under the  
18 curves, which are essentially integrals and consequently  
19 smooth out error, and how you're telling us we're going to  
20 have to take derivatives, which augment error --

21           DR. FLOCKHART: Well, it's taking a smaller  
22 part of the data.

23           DR. SHEINER: We may never be able in sort of a  
24 naturalistic setting to estimate a derivative with any kind  
25 of accuracy.

1                   It would work at the level of a preparation.  
2    If you had a preparation that was rapidly absorbed and one  
3    that wasn't, and that was pretty consistent, then you'd  
4    know that you'd have more danger from one than another in  
5    that sort of circumstance. But that we'll ever discover  
6    who are the people who absorb more rapidly by sort of  
7    surveying the world and then trying to put it together  
8    across several models -- and I'm the mad modeler.

9                   DR. FLOCKHART: But, Dr. Sheiner, shouldn't  
10   that come out of some models? In other words, you would be  
11   able to see in a large population study whether people who  
12   get fast absorption get a QT longer.

13                  DR. SHEINER: Well, I don't know the rate of  
14   rise because I don't know when their level was drawn. I  
15   put it on the graph at a certain point because that's what  
16   they told me, but I can have two levels that are 10 minutes  
17   apart and they're really 2 hours apart.

18                  I guess what I'm trying to say is that -- and I  
19   was just being facetious, but I think we do need to temper  
20   the kinds of conclusions we hope people to draw from sort  
21   of messy clinical data. I'm just mentioning that  
22   derivatives are really hard.

23                  DR. VENITZ: But that's the empiricist talking.  
24   As a pharmacologist, I say maybe I can understand something  
25   about what's responsible for orthostatic tachycardia and it

1 | may well be that my rate of change in concentration or my  
2 | Cmax is much more physiologically important, and I know  
3 | that without having to do an empirical study.

4 |           DR. SHEINER: Right. What I'm saying is the  
5 | implications of what I'm saying, to be serious rather than  
6 | just making trouble, would be if that's the kind of thing  
7 | you want to know, if you're not sure about it, then you  
8 | need to do a very well-controlled experiment. You're not  
9 | going to be able to learn that from the same kind of data  
10 | you might be able to learn that area under the curve was  
11 | the determinant.

12 |           DR. VENITZ: Either that or you have some  
13 | mechanistic understanding how the drug concentration leads  
14 | to a response. That's the point that I'm making.

15 |           Hartmut.

16 |           DR. DERENDORF: Well, I think this discussion  
17 | shows that it's really impossible to even try to have a  
18 | standardized approach in terms of a parameter like a  
19 | bioequivalence approach, that you have a single criterion  
20 | that would summarize it all up. I think each drug, each  
21 | class of drug is different, and each situation is  
22 | different. I'm not sure if we can find a standardized  
23 | approach as we're asked to.

24 |           DR. LEE: I guess by standardized approach, we  
25 | mean a standardized conceptual approach, which means we

1 always like to calculate the probability of an adverse  
2 event, rather than saying that we're going to standardized  
3 an Emax model as the method to be used or the magical AUC  
4 or Cmax. So, again, we're trying to standardize the  
5 conceptual approach.

6 DR. SHEINER: I think it's really worthwhile  
7 focusing on the positive here, which is this is a difficult  
8 problem and the very fact that people involved in  
9 regulation are acknowledging that it's worthwhile to try to  
10 be quantitative about things that are extremely uncertain  
11 and to try to come up with a better way to be more  
12 quantitative about a problem where there will never cease  
13 to be disagreement about any particular case because you'll  
14 never nail anything down close enough -- you'll be saying  
15 this is what we think we ought to do in terms of dosage  
16 recommendations. And it will be based upon an information  
17 base which would allow a rational person to say, no, you  
18 don't need to do that. That's where we're going to wind  
19 up, and to wind up anywhere else would be so prohibitively  
20 expensive that it would not justify the effort.

21 So I'm extremely encouraged. It's not as it  
22 has been in the past, hands being thrown up and we can't do  
23 this well enough, so we won't do it at all. That's not the  
24 right attitude. And that you're seeking advice on how to  
25 do this difficult thing I think is a very good thing.

1                   But I do think that some kind of  
2                   standardization, for example, about turning things into  
3                   probabilities and utilities in an honest way so that  
4                   everybody can be on the same page -- they can all  
5                   understand what you know and what values you're applying.

6                   DR. LESKO: A lot of the context for the  
7                   discussion in the case studies so far have been what we've  
8                   seen, what has come in in an NDA, but is there a way to  
9                   translate the methodology we're talking about, let's say,  
10                  to a drug development program in order to get studies  
11                  designed that would provide for information that would  
12                  reduce some of the uncertainty that we work with in the  
13                  absence of some formal recommendation to do studies a  
14                  certain way?

15                  In other words, let's say a standardized method  
16                  evolves over time, and let's say that that could perhaps  
17                  evolve into a guidance on study design that would provide  
18                  for information that would be better apt to provide the  
19                  information that we're asking here in terms of dose  
20                  adjustment and quantitation of risk. Is that a logical  
21                  follow-through on the path we're on in the minds of people?

22                  DR. VENITZ: But is the uncertainty and the  
23                  consequence of the uncertainty that we currently have so  
24                  large that we really need to do a whole lot more  
25                  experimental work, short of what you're doing right now,

1 | which is on a case-by-case basis, evaluate whether the  
2 | information is sufficient for you to assess the risk-  
3 | benefit, and then as in Jenny's case, recommend to the  
4 | sponsor that they would have to do a larger study to look  
5 | at high exposures?

6 |           DR. LESKO: I guess I was sort of asking the  
7 | question -- in Jenny's case, for example, QTc was obtained  
8 | for the first 4 hours. Would a study design that looked at  
9 | a longer period of time -- wait a minute. Was that your  
10 | case? Well, it was one case. Sorry, Nhi. I should know  
11 | these data.

12 |           There was one case where the QTc was obtained  
13 | for only 4 hours and blood levels were obtained for a  
14 | longer period of time. And would a different study design  
15 | have provided a better basis to make the recommendations  
16 | that people were trying to make with the analysis of the  
17 | data? That's sort of where I'm heading with that.

18 |           DR. FLOCKHART: I think the answer to that is  
19 | obviously it depends. 4 hours might have been long enough  
20 | for that drug, but there are other drugs -- haloperidol  
21 | comes to mind -- where that would not have been enough.

22 |           To go back to my point, I think actually it is,  
23 | Dr. Lee, very generalizable. I think there is a  
24 | generalizable conceptual approach. My point about bringing  
25 | up just sticking to the AUC was just to be educated about

1 that. I suspect that the AUC would very often be a  
2 valuable parameter, but you have to be open to using other  
3 things when that's biologically and pharmacologically  
4 appropriate.

5 DR. VENITZ: The only thing I would add is, as  
6 you've heard the committee talk about last time, as well as  
7 this time, I think there's a lot of favorable sentiment.  
8 As Dr. Sheiner likes to point out, it's better than what we  
9 currently have. It beats the competition.

10 The one thing that I would reinforce, though,  
11 is that it's very important to communicate it  
12 appropriately, and that has to do with all the assumptions  
13 that are being made. Are they verifiable to some extent or  
14 not? Do you want to err on the conservative side or on the  
15 more liberal side? So that the people that deal with the  
16 clinical pharmacology reviews interact with the medical  
17 reviewers. They may not understand the technical side of  
18 it, but they're the domain experts and they can follow  
19 those kind of thoughts. So it's really a matter of risk  
20 communication in my mind more than it is the actual  
21 process.

22 DR. KEARNS: And to pick up on a point that Dr.  
23 Flockhart mentioned too, it has to be driven by biological  
24 or pharmacological plausibility. To use an approach, a  
25 guidance across the board can create information that is

1 | not factual.

2 |           For example, I had the occasion to look at a  
3 | new molecule just last week with a sponsor to talk about a  
4 | study design. Of course, they had received some input  
5 | about that study design, which included multiple ECGs over  
6 | time that was coincident with the sampling time for the  
7 | pharmacokinetics. When I inquired as to the preclinical  
8 | data about the ability of the molecule to prolong QT, about  
9 | the only way that I could be convinced it could happen is  
10 | if the structure could be inserted in the chart of the  
11 | alphabet and somehow got between the letters Q and T.

12 |           (Laughter.)

13 |           DR. KEARNS: So what will we see when we do the  
14 | trial? We do multiple ECGs, in this case, on children.  
15 | What happens if we see a relationship come out of that that  
16 | can be described by a host of models with all the  
17 | appropriate variability nested in? Will we have proven  
18 | something that wasn't shown by prudent preclinical testing,  
19 | or will we be finding ourselves in the midst of yet another  
20 | epi phenomenon that has implications about how the drug  
21 | might be used?

22 |           So I think one has to use caution in making  
23 | sure that when we do these things, we have good reason to  
24 | do it based upon what we know. I'm not saying that we will  
25 | always know everything up front. We clearly, clearly

1 don't. It's an imperfect science in an imperfect world.  
2 But to just cast it out there as indiscriminate use of an  
3 approach carries with it some liability that might not  
4 serve the public at the end of the day.

5 DR. DERENDORF: Just a follow-up comment to  
6 what Dr. Sheiner said. I fully agree that it is worthwhile  
7 doing it and it is a good thing to do it. But the  
8 standardization -- really my point was that it stops at the  
9 point where we say each drug is different and the more you  
10 know about the exposure-response relationship for that  
11 particular drug, the more we can use it to make predictions  
12 and the better they will be. That's a trivial conclusion,  
13 but I think that's where the standardization ends. Then  
14 from there on, really each case is different and needs to  
15 be dealt with individually.

16 DR. VENITZ: Would it be helpful, as far as the  
17 internal workings are concerned, to come up with a list of  
18 questions that you typically consider when you go through  
19 this process and for the committee to have a look at them?  
20 I'm not sure whether the approach is something that can be  
21 unified, but maybe the kind of questions that should be  
22 asked every time you do this can be found consensus on.  
23 Does that make sense to the committee? So maybe at a  
24 future meeting, the questions that you would ask, what  
25 surrogate markers do you have, what relationships do you

1 | have, do you use areas or Cmax, those kinds of things that  
2 | you go through every time that you have to review an NDA  
3 | based on your experience.

4 |           DR. SHEINER: I think you can go a little  
5 | further. I think there are sort of best practices. Maybe  
6 | that's the way to think about it in doing this kind of  
7 | thing. Since I don't know anything about anything in  
8 | particular, I've been dwelling on the generals of showing  
9 | clearly what you know and what you don't know and somehow  
10 | checking your models against your current data and so on.  
11 | So I think there are best practices in this and I think  
12 | there are some things you can say in general, although I  
13 | agree with you, when you get down to putting the labels on  
14 | the x axis and the y axis, then suddenly you're in the  
15 | domain area and you've got to talk to the right folks.

16 |           DR. VENITZ: Any further comments by the  
17 | committee or any further questions from the FDA staff?  
18 | Larry.

19 |           DR. LESKO: Yes. Maybe this is a deeper  
20 | question and there isn't time to discuss it, but it does  
21 | lead us down the path if we develop a standardized  
22 | approach, the question that I have, in terms of labeling,  
23 | comes into my mind. Right now we put in the label  
24 | descriptive information, for example, in the clinical  
25 | pharmacology section about a change in an area that

1 describes the, let's say, drug interaction or a special  
2 population change, and then if it warrants, a change in the  
3 dosage and administration section as to what to do about  
4 it. But if you have more data in hand, i.e., the  
5 likelihood of a risk or the probability of a risk or the  
6 probability of an increase in risk or other things that  
7 might come out of a standardized approach, the question  
8 would be to what extent would this information be helpful  
9 to prescribers or would it be a distraction to the  
10 prescribers and how can we enhance labels. Because we now  
11 know there are certain pieces of information that go into  
12 labels that at least the consumers, public and physicians  
13 tell us are not helpful to them and they can't interpret,  
14 and drug interaction seems to be one of those areas we  
15 frequently hear about.

16 So we're thinking of ways of improving labels  
17 in terms of consistent language, the scope of information  
18 that goes into it, and with this kind of standardized  
19 approach, it could lead to some interesting ways of  
20 revising labels to convey different information to  
21 prescribers and patients.

22 DR. VENITZ: Any comments?

23 (No response.)

24 DR. VENITZ: Okay. Then let's move to the  
25 third example for today, which is going to be presented by

1 | Dr. He Sun. He is a pharmacometrics reviewer in the  
2 | Division of Pharmaceutical Evaluation II.

3 | DR. SUN: Good morning. I will try to discuss  
4 | some general questions in my discussion, and we may switch  
5 | the specific detail in numbers to general issues.

6 | These will be the questions I'm going to ask  
7 | after the presentation, but I just put it up front to get  
8 | some initial feelings.

9 | The first question is, if we get adverse  
10 | reaction data from clinical studies, these data can be  
11 | treated as either a continuous variable or a categorical  
12 | variable. Then, what should we do? Do you have any  
13 | preference, and why? I will show some examples to  
14 | illustrate it further for this.

15 | The second question is, in phase III clinical  
16 | trials, lots of subjects, but we may not have observations  
17 | in special subjects. Therefore, the population PK approach  
18 | may give us the opportunity to either simulate or predict  
19 | the exposure parameter for the population who don't have  
20 | exposure observation but do have response observation. So  
21 | what's the limitation and utility of this approach?

22 | Now, if we have a PK model based on the above  
23 | approach, we get some kind of conclusion on side effect  
24 | versus drug concentration relationship. How do we make a  
25 | dose adjustment recommendation for subpopulations?

1           There's some limited information here to  
2 present what data distribution pretty much looks like. On  
3 the slide here, in this corner it shows what the data  
4 distribution may look like. It can be dense data from  
5 phase II trials or it can be sparse data from phase III  
6 clinical trials, or some kind of a combination with an  
7 unbalanced situation.

8           Now, the safety information can be either a  
9 single critical key adverse reaction parameter which is a  
10 continuous variable, like QTc variable or high blood  
11 pressure and so on. But it can also be a categorical  
12 variable like pain or "yes or no" for liver toxicity and so  
13 on.

14           But these two actually are switchable. Let's  
15 say blood pressure. You can set up a cutoff marker that  
16 says if above such and such, it is abnormal, below such and  
17 such, it is normal. So the continuous variable actually  
18 can be changed to a categorical variable.

19           Now, a categorical variable, although it can be  
20 a "yes or no" situation, but for the group with "yes," you  
21 can also give a score of 1, 2, 3, 4, 5 or from 0 to 10. So  
22 it becomes some kind of a continuous variable.

23           So these two actually have no clear cut.  
24 That's why I ask this question in this presentation. If  
25 you have this situation, which one do you prefer? Of

1 course, this will also change your data analysis process.

2 So you can also have a combination of both with  
3 multiple ADR observations. But for phase III trials,  
4 pretty much we have this kind of situation: mixed types,  
5 multiple ADRs, and unbalanced. Then that is why population  
6 approach can play here.

7 Let me first show some data sets. Then we go  
8 back to see what analysis process we can apply. This data  
9 set is used just to illustrate the question or the process  
10 I mentioned before. Forget the exact numbers and the true  
11 terms. Sometimes I have to modify this.

12 Let's say we have two clinical trials, very big  
13 size, 1,500 evaluable treatment patients. And we also have  
14 multiple dose levels from X to 4 times higher. And the  
15 patient plasma drug concentration was measured, although  
16 for some are dense and for some are sparse. Therefore, the  
17 total data set is kind of unbalanced.

18 Then we have endpoints for safety measurement.  
19 This can be some blood chemistry variables which usually is  
20 a continuous variable at the beginning, but a clinician can  
21 define some value as a cutoff point shows this variable as  
22 normal/abnormal to claim at such situation there's no ADR  
23 and others will be ADR.

24 The ADR can be also present as a "yes or no"  
25 situation for some, like headache, liver toxicity,

1 phototoxicity values. And this variable again can be  
2 changed to different scores for the extent of headache,  
3 like mild, moderate, or severe.

4 Now, PK results. Let's say drug-drug  
5 interaction causes AUC to increase by almost 300 percent.  
6 The Cmax changes by 150 percent. AUC and Cmax may also be  
7 changed by age, gender, or so on and so forth, even between  
8 ethnic groups.

9 The safety results. We will not focus on  
10 efficacy in the presentation. We will only focus on safety  
11 parameters. Safety parameters usually are very, very small  
12 in percentage and very sparse. So there are some cases  
13 where you never have any so-called "maximal effect" for  
14 side effect terms.

15 The efficacy results. Let's make this  
16 discussion a little simple. We see efficacy has no such  
17 exposure-efficacy relationship detected although we see  
18 there's a demonstration of clinical efficacy in total.

19 Now, with these data sets, let's come back and  
20 see what process we usually can do. First of all, of  
21 course, there are managing/editing data processes we can  
22 use. For this part, we start to have a question: shall we  
23 treat the data as a continuous variable or should we treat  
24 the data as a categorical variable?

25 Then we can conduct a population PK analysis

1 based on exposure data such as building a base model, add  
2 variability, add covariate, and so on and so forth. Now,  
3 there's one problem: if we cannot find a significant  
4 covariate in the PK model, then the next step for  
5 predicting E for the new population or new individuals will  
6 have a little problem. But let's say we have the model  
7 built and the model validated. Then we can go to the next  
8 step and determine individual exposure or subpopulation  
9 exposure parameters. There are two parts here. We can do  
10 post hoc for the subjects who are already included in the  
11 study, or we can do a simulation trial to determine  
12 exposure parameter for the population who was not really in  
13 the trial or the observation was not available in the  
14 particular patient.

15 The next step will be to derive secondary  
16 exposure parameters, such as AUC, Tmic, effect compartment  
17 concentration and so on. And another important factor here  
18 I want to emphasize is that the accumulative exposure  
19 parameter can be estimated and determined, like say what if  
20 after multiple dose or long exposure situation.

21 With these exposure parameters available  
22 through the above processes, we can determine exposure-  
23 response relationship for individuals or for special  
24 populations. We have lots of methods here. We can use  
25 classification method. I will give you some examples later

1 on. We can do classification or regression tree analysis,  
2 logistic regression for binary data, and so on.

3           Then we consider the accumulative exposure  
4 time, then perform statistical analysis on response data.  
5 Now, this again correlates with what do we do with the  
6 data? If our data is a continuous variable and we have  
7 odds ratios with uncertainty built in, we can do  
8 statistics. But sometimes if the variable purely is  
9 categorical and is divided to either above the mean or  
10 below the mean, the statistics will be hard.

11           Now, with all this situation, the next step,  
12 finally we will make a dose adjustment. What are we going  
13 to do especially if we have multiple variables? Let's say  
14 the exposure-response depends on age, gender, body weight,  
15 and blah, blah, so on. If we take all of this condition  
16 together for making a dose adjustment, it may be too  
17 complicated in drug labeling. Shall we only consider the  
18 one which is critical, or shall we consider the one which  
19 has most frequently occurred, or some other method? I want  
20 to hear some discussion on this.

21           Let's see some results. If we're dealing with  
22 this process, what result can we get?

23           Classification. The first part we can see is  
24 to divide the whole population into some equal populated  
25 segments. Let's say every 25 percent subjects from low

1 exposure to high exposure. Or we can divide this whole  
2 population into equally distanced segments, the percentile;  
3 that is, the first 25 percent in concentration, the second  
4 25 percent, and so on and so forth. Then we count what's  
5 the frequency of ADR. For example, the results can be  
6 presented as total ADR is 18 percent if AUC is greater than  
7 the mean and only 5 percent if AUC is less or equal to the  
8 mean value. That makes the whole discussion for this kind  
9 of classification results. Of course, there are lots of  
10 pros and cons. I really want to hear a discussion later  
11 on.

12 Now, the second one is that we can do a  
13 classification analysis based on severity. Let's say  
14 we reclassified R values, the response values, as different  
15 class or different scores, as severe, moderate, or mild,  
16 and so on. Then we see the example. Severe ADR occurs if  
17 Cmax is greater than 10 but only mild ADR is apparent if  
18 Cmax is less than 2, although the frequency probably  
19 between these two has no significant difference. But in  
20 this situation, we see it looks like 10 is some kind of  
21 cutoff value to avoid severe ADRs.

22 Then we can throw this data into a computer to  
23 search for the best maximum split, maximum split  
24 distinctions between R values as by a classification tree  
25 or regression tree. One result I present here, for

1 | example, the first split on ADR frequency was at AUC equals  
2 | to 871. So when the split occurs on ADR, it was 23 percent  
3 | if AUC is greater than 871 and would be 2.5 percent if AUC  
4 | is less than 871.

5 |           Next, we will do modeling work. We can do  
6 | modeling work for the same data set for different ADR or  
7 | the same ADR parameters. When we do modeling work, there  
8 | are several ways. I do not want to discuss further on this  
9 | part. I only want to show the possible ways. We can base  
10 | on purely statistical models to do the modeling work, or  
11 | base on some kind of physiological-based, meaningful  
12 | models. There's a lot of discussion on the pros and cons  
13 | for each. But let's go to the next one. Our model can be  
14 | a linear model or a nonlinear model. Of course, there are  
15 | uncertainty parts when building nonlinear models, adding  
16 | fixed effects and random effects, again, with this model.

17 |           So two examples. We can do a simple regression  
18 | analysis based on so-called logistic regressions. So, for  
19 | example, we can get a result with even a 95 percent  
20 | confidence interval for the logistic regression results for  
21 | odds ratios. For example, we can see the odds ratio for  
22 | acute tissue rejection increases 23 percent if AUC 0-24  
23 | decreases by 10 percent. That's one way to present this  
24 | data.

25 |           Then we can treat all the data as a continuous

1 variable, do as the next few examples for modeling work.  
2 We can get an equation that says the HDL drops below normal  
3 on day 95 if the concentration, average concentration, is  
4 greater than 10 micrograms per ml on day 95 for patients  
5 with high body weight and low initial HDL at baseline. So  
6 there's a covariate effect built in and it also has this  
7 kind of a drug concentration curve profile.

8           The back pain we treat as 0 to 10 scores, some  
9 kind of semi-categorical variable. It's nonlinearly  
10 correlated with plasma drug daily AUC and the number of  
11 treatment days and the dose regimen. So three factors.  
12 The result here found was b.i.d. actually has less  
13 incidence of back pain. T.i.d. will have more, although  
14 the total daily doses are equivalent. And the number of  
15 treatment days significantly correlate with or predict back  
16 pain scores.

17           QTc prolongation. Now, we can find some  
18 models. The relationship between QTc and the drug  
19 concentration can be described by an Emax model. Then you  
20 can find some E-R parameter like E0, Emax, ED50, and so on.  
21 But these parameters can be, as we discussed before,  
22 correlated with either gender or age or some other  
23 subpopulation variables.

24           Phototoxicity. Now, we only have two  
25 variables, yes or no. We get a "yes" value on day 10 if C

1 average on day 10 is greater than 8. This will occur only  
2 in female subjects. So this can be due to either data  
3 limitation or this really is a true result that female  
4 subjects are more sensitive to the drug in terms of having  
5 phototoxicity occurring.

6           Liver toxicity. We can get some kind of  
7 frequency linearly correlated with C in the initial few  
8 hours of time of exposure. It may not correlate exactly  
9 with the Cmax, but it correlated with the initial range of  
10 concentration average.

11           Blood chemistry. Now, blood chemistry actually  
12 is a continuous variable, but we can treat the blood  
13 chemistry variable as a categorical "yes or no," normal or  
14 abnormal, or give them a score from 0 to 10. Now, if we  
15 have a score from 0 to 10, we can get some kind of  
16 correlation with plasma concentration, either AUC or number  
17 of treatment days.

18           So these are examples I want to show. So one  
19 single variable can be treated by different ways, but in  
20 one study we have lots of different variables, and  
21 different variables may be treated by different ways, and  
22 use different data sets as different base for information.

23           And there are others, probably we never find  
24 any relationship, never can find a cutoff point.  
25 Descriptive. Here I give some examples. Some we just can

1 | have a definition of normal/abnormal values, but may not  
2 | see any difference between different subgroups like these  
3 | four examples I show here.

4 |           So now we come to the end. We have all the  
5 | information, by all different methods, with different  
6 | bases, from different data sets, so on. We definitely  
7 | already used population PK, did it two times. One is  
8 | predicting exposure parameter for subjects who do not have  
9 | observation in exposure but do have a response measurement,  
10 | and in the second part, use a population approach,  
11 | nonlinear mixed effect modeling, to show whether the E-R  
12 | relationship depends on some other co-variables.

13 |           So with all this population of subjects and  
14 | information, now we will make a dose adjustment. See, for  
15 | example, we can do this: the average upper therapeutic  
16 | limit is probably around 10 for Cmax and 871 for AUC.  
17 | Remember, these two variables are gathered from previous  
18 | toxicity analysis. And the female subjects seemed the most  
19 | sensitive to phototoxicity, and the concentration average  
20 | should be less than 8. Then we see a b.i.d. dose regimen  
21 | is preferred because it reduced one of the particular  
22 | toxicity results.

23 |           Now let's go back to my questions with all the  
24 | data we have seen in the examples. First is what are the  
25 | utility and general limitations of linking PK obtained from

1 population analysis to response endpoints? And what are  
2 the general considerations in E-R based dose adjustment for  
3 special populations? And should we treat this data as a  
4 continuous or categorical variable? What's the preference?

5 Again, I really want to hear a discussion more  
6 focused on the general concept and ideas based on the  
7 experience you have and let us know the pros and cons for  
8 each situation rather than focusing on the numbers because  
9 I have modified the values somewhat to make the  
10 presentation smooth. Thanks.

11 DR. VENITZ: Thank you, He.

12 Any questions about Dr. Sun's presentation  
13 before we delve into his proposed questions?

14 On one of your slides, you mentioned Cint. Can  
15 you tell me what that meant?

16 DR. SUN: This is the initial concentration  
17 exposure.

18 DR. VENITZ: Oh, initial concentration. Okay.

19 Do you want to pose the questions and then let  
20 the committee bat it around?

21 DR. SUN: Okay.

22 DR. VENITZ: The first question regarding the  
23 limitations of linking PK from a population analysis to  
24 response endpoints. Do you want to elaborate on that  
25 question what specifically you had in mind?

1 DR. SUN: Okay. This question was the utility  
2 and general limitations of using population PK for  
3 population for this analysis. As I mentioned, we have two  
4 places we can use nonlinear mixed effect modeling work for  
5 doing data analysis for this data. The first part is in  
6 clinical phase III trials, we may not have a concentration  
7 exposure measure for every subject, but we do have a  
8 response measure for every subject. In this situation, if  
9 we have sufficient information, use the population PK, get  
10 the model, then we can predict or estimate concentration or  
11 other exposure parameter for the population we see in the  
12 phase III trial. Or in some situation, patients only have  
13 one or two trough measures and we want to determine the  
14 total exposure and the time of exposure. So this is one  
15 place population PK can be used.

16 The second part is after we have E data and R  
17 data, either categorical or continuous variable, now we can  
18 use mixed effect modeling to see whether these two  
19 variables are related to each other based on some covariate  
20 factors.

21 So that's my question. What are the utility  
22 and general limitations on this if we do it this way?

23 DR. SHEINER: I can't decide if what you're  
24 asking is, is there a manual for how you treat any given  
25 set of data to come up with the conclusions that you're

1 going to find most believable. We can't address that. So  
2 when I look at that first question, your description of  
3 what you might do sounded sort of like something I might  
4 do.

5 The only thing I can say, the only serious  
6 general limitation about which even good data analysis  
7 cannot help you is the problem of confounding. Both the PK  
8 and the responses are endpoints, are outcomes, and whenever  
9 you try to relate outcomes to outcomes, you have the  
10 problem that you can't tell which way causality runs. And  
11 you base that conclusion, if you do, on external  
12 information in the way of science or other things. You  
13 can't tell it from your data. So that's the limitation.

14 Now, that doesn't mean we don't proceed every  
15 day, based on observational data, to make the most  
16 important decisions in our lives. We do. But we have to  
17 understand that in a regulatory context, there are other  
18 forces operating. You want to be cautious in certain ways.  
19 And that's the serious problem.

20 Most of the other stuff, it seems to me, that  
21 you brought up were technical issues. And I'm not sure  
22 that we really want to spend -- even though I'm a real  
23 techno-wonk when it comes to model-based analyses, I don't  
24 think you want to hear me dilate on that.

25 I think the basic thing there is if you get

1 | different conclusions when you treat your data as  
2 | categorical versus continuous or when you use one kind of a  
3 | model or another, then there's something wrong. So you  
4 | ought to get all the same conclusions. What happens as you  
5 | turn data from continuous, if it's got a lot of  
6 | information, to categorical, you're losing information. So  
7 | some things will drop out. Some things will appear no  
8 | longer to have relationships that did appear before to have  
9 | relationships. That's got to happen as you limit the  
10 | information in your data.

11 |           But other than that, I think you want to  
12 | basically use the data representation that's most relevant  
13 | to the people who are going to use it and that keeps the  
14 | information, et cetera, all the good rules of modeling.  
15 | But I don't think we can get into too many details.

16 |           So maybe if you have particular instances where  
17 | you think, looking at data in different ways, the same data  
18 | in different ways led you to very different conclusions, I  
19 | think that's something that I might be interested in  
20 | hearing about. Otherwise, I don't know what we can say in  
21 | general.

22 |           DR. LEE: Can I rephrase the question a little  
23 | bit? The reason we're making this presentation is because  
24 | phase III studies actually present a unique opportunity for  
25 | us to look at exposure and response, especially the safety

1 | endpoint that we don't frequently observe in a phase II  
2 | study which is too small to capture a rare adverse event.  
3 | That's why we wanted to ask the committee whether a  
4 | population approach would be a good approach to look at  
5 | exposure response in the phase III studies.

6 |           However, there may be some limitation in terms  
7 | of study design. For example, we may not have enough  
8 | plasma samples or maybe the sampling time between the PK  
9 | and the pharmacodynamic endpoints is different.

10 |           So this is the type of question we're trying to  
11 | ask the committee, whether internal study design, whether  
12 | there's any limitation to conduct such type of a population  
13 | PK analysis. And if there are certain limitations, can we  
14 | recommend to the sponsor to design the study differently in  
15 | the future so that we can get a better quality PK/PD  
16 | relationship out of the phase III studies?

17 |           DR. SHEINER: Let me just say one more thing  
18 | about that. So you're talking about wanting to use this  
19 | confirmatory study for learning purposes. And there are  
20 | certain kinds of learning data elements that don't  
21 | interfere with your design that would make your life a lot  
22 | easier. Whether they're worth it or not, you can only say  
23 | afterwards. But measuring things serially, whether it be  
24 | toxicity or efficacy or both, rather than just the 6-month  
25 | and 1-year endpoint, or whatever it is that was the primary

1 | thing; measuring compliance, that is to say, what drug did  
2 | they actually take; measuring PK.

3 |           I think, by the way, my guess is that adherence  
4 | is more important an influence on outcome than PK is for  
5 | most cases. But when they say measuring PK, you want to  
6 | measure that in the case where adherence is assured so that  
7 | you have those as two separate variables. And so on.

8 |           Basically the idea is measuring biomarkers,  
9 | whether they be adherence or chemicals, you know, along the  
10 | causal path from the prescription to the effect, and  
11 | measuring them serially over time. That's the best you can  
12 | hope for without changing the design radically. And if you  
13 | want to be able to do these kinds of analyses, that's the  
14 | kind of data that you need.

15 |           But techniques for dealing with missing data,  
16 | techniques for dealing with other problems that arise, with  
17 | mixed kinds of data, both continuous and categorical, and  
18 | so on, those I don't think are essential. They exist.  
19 | They make your life a little tougher or a more interesting,  
20 | depending on where you come from. But they're there and  
21 | you should be able to get the information out of the data.

22 |           DR. SUN: He was asking whether I have an  
23 | example regarding how the data can be treated either  
24 | continuous and categorical.

25 |           DR. SHEINER: And reach different conclusions.

1 DR. SUN: Yes, reach a different conclusion.

2 Let's say this situation. If you treat data as  
3 a "yes or no" situation and you divide the concentration  
4 distribution to be above the mean and below the mean, do  
5 you will find the only conclusion you can get is the  
6 frequency of "yes" when concentration or AUC above the mean  
7 is 23 percent. If lower than the mean, it will be 5  
8 percent. That's all you can get.

9 If you treat it as a continuous variable, you  
10 can get some kind of sigmoid models, correlation between  
11 scores of adverse reaction versus the concentration. Then  
12 from the curve, you can pick up -- say you want to limit  
13 less than 10 percent of subjects has a score less than 2 --  
14 a concentration. So this becomes a different decision.  
15 And the curve becomes a smooth curve. You pick up a point  
16 at which you limit two factors. Percent of subjects reach  
17 a score of XYZ. Compared with the first one, you only can  
18 get a result if above the mean will be such and such, if  
19 below the mean will be such and such.

20 Then in labeling, it will be different. In  
21 labeling, when make a dose adjustment, say due to drug-drug  
22 interaction, if the population Cmax change, still somewhat  
23 below the mean values, for the overall population, or you  
24 can get a feeling what the frequency of side effects due to  
25 drug-drug interaction will be. But in the second

1 | situation, if you have a continuous curve, you can estimate  
2 | when concentrations switch a kind of 10 percent, what's the  
3 | percentage of patients will have a score of 2 or 3 increase  
4 | by such and such.

5 |           So when we recommended these two suggestions to  
6 | clinical or to the labeling committee for NDA review, these  
7 | two really makes different. That's why the question comes.  
8 | Any parameter really we can treat as one of, or we can  
9 | switch between the two. And what really we want to do?

10 |           DR. DERENDORF: I think as was said earlier,  
11 | whenever you move from continuous to categorical values,  
12 | you throw away information. I think it comes down to a  
13 | compromise that you have to make with the information that  
14 | you have and how you want to communicate it. If you make  
15 | it too complicated and include everything you know, nobody  
16 | is going to use it. So you have to find a way to focus on  
17 | the important things, but still come up with an accurate  
18 | conclusion.

19 |           You have a great example that you can overdo it  
20 | and make it too simple. One of the conclusions you have  
21 | here, the total ADR is 18 percent when the AUC is above  
22 | 1,200 and 5 percent when it's below. That may be true for  
23 | the data set that you have, but it's totally useless for  
24 | someone who wants to extrapolate it for a certain situation  
25 | because obviously you can have very, very low AUCs that