APPENDIX H
OHIO WATER MICROBIOLOGY LABORATORY
mEI AGAR PREPARATION

BASAL MEDIUM (mE AGAR)

*Don't need to mix—use manufactured mE agar

Peptone	10.0 g/L
Sodium chloride	15.0 g/L
Yeast extract	30.0 g/L
Esculin	1.0 g/L
Actidione	0.05 g/L
Sodium azide	0.15 g/L
Agar	15.0 g/L

- Add 71.2 g of pre-mixed dehydrated basal medium plus 0.75 g of indoxyl-β-D glucoside to 1 L of reagent water in a Wheaton bottle.
- Heat to dissolve (to boiling).
- Autoclave the Wheaton bottle and 10 empty dilution bottles with caps for 15 minutes.
- Mix 0.24 g nalidixic acid in 5 mL sterile reagent water (add a few drops of 0.1N NaOH to dissolve if needed). Add this solution to the tempered medium.
- Add 0.02 g triphenyl tetrazolium chloride to the medium and mix.
- Pour medium into the sterilized 100-mL dilution bottles.
- Store at 4°C for up to 6 months.

PREPARATION OF AGAR PLATES

- Melt the basal medium using a beaker with water on a hot plate. DO NOT AUTOCLAVE!
- Pour the plates after the agar is tempered (50-60°C).
- Store the plates at 4°C for up to 2 weeks in a tightly sealed container.