



Toxic Hydrology Program

Results of the Chemical and Isotopic Analyses of Sediment and Ground Water from Alluvium of the Canadian River Near a Closed Municipal Landfill, Norman, Oklahoma, Part 2

By George N. Breit, Michele L.W. Tuttle, Isabelle M. Cozzarelli, Cyrus J. Berry, Scott C. Christenson, and Jeanne B. Jaeschke

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Contents

Conversion Factors	v
Chemical and Isotopic Symbols.....	vii
Abstract.....	1
Introduction	2
Methods	2
Sample Site Selection.....	2
Sediment Sampling.....	4
Water Sampling and Field Analyses	4
Sediment Analyses.....	5
Water Laboratory Analyses.....	6
Results	7
Sediment Composition	7
Water Composition.....	12
References Cited	17

Figures

1. Location map of the IC sample sites	3
2. Cumulative abundance of size fractions measured by sieve analysis of sediment samples collected at IC 35, IC 54, and IC South.....	7
3. Depth variation in total iron concentration extracted using 0.5 N HCl and Ti^{3+} -EDTA on sediment samples collected at IC 36, IC 54, and IC South.....	8
4. Depth variation in the ratio of ferrous to total iron extracted with 0.5 N HCl from sediment samples collected at IC 36, IC 54, and IC South.....	9
5. Abundance of iron extracted from sediment samples with Ti^{3+} -EDTA relative to the abundance of particles smaller than 100 μm	10
6. Depth variation in the abundance of acid-volatile sulfide and disulfide extracted from the sediment samples collected at IC 36, IC 54, and IC South.....	11
7. Box plot of the sulfur isotope composition of sulfur species extracted from the sediment samples collected at IC 36, IC 54 and IC South.....	12
8. Depth variation of chloride concentration of ground water from the wells at IC 36, IC 54, and IC South in October 2004 and April 2005.....	14
9. Depth variation of iron concentration of ground water from the wells at IC 36, IC 54, and IC South in October 2004 and April 2005.....	14

10. Depth variation of sulfate concentration of ground water from the wells at IC 36, IC 54, and IC South in October 2004 and April 2005.....	15
11. Piper diagram of major ion composition of ground water samples from wells at IC 36, IC 54, and IC South.....	16

Tables

1. Location of sample sites.....	3
2. Summary statistics of water composition from IC 36, IC 54, and IC South	13

Appendices

1. Depth intervals of the wells sampled at IC 36, IC 54, and IC South	19
2. Elevations and descriptions of cored sediment samples.....	20
3. Results of grain-size analysis of sediment samples	22
4. Concentration of iron sequentially extracted from sediment samples using 0.5 N HCl and Ti^{3+} -EDTA	24
5. Concentrations of sulfur species in sediment samples and their sulfur isotopic composition	26
6. Results of chemical and isotopic analyses of ground water samples from wells at IC 36, IC 54, and IC South.....	28

Conversion Factors

Inch/Pound to SI

Multiply	By	To obtain
Length		
inch (in.)	2.54	centimeter (cm)
inch (in.)	25.4	millimeter (mm)
foot (ft)	0.3048	meter (m)
mile (mi)	1.609	kilometer (km)
mile, nautical (nmi)	1.852	kilometer (km)
yard (yd)	0.9144	meter (m)
Volume		
ounce, fluid (fl. oz)	0.02957	liter (L)
pint (pt)	0.4732	liter (L)
quart (qt)	0.9464	liter (L)
gallon (gal)	3.785	liter (L)
cubic inch (in ³)	0.01639	liter (L)
Mass		
ounce, avoirdupois (oz)	28.35	gram (g)
pound, avoirdupois (lb)	0.4536	kilogram (kg)

Temperature in degrees Celsius (°C) may be converted to degrees Fahrenheit (°F) as follows:

$$^{\circ}\text{F}=(1.8\times^{\circ}\text{C})+32$$

Temperature in degrees Fahrenheit (°F) may be converted to degrees Celsius (°C) as follows:

$$^{\circ}\text{C}=(^{\circ}\text{F}-32)/1.8$$

Vertical coordinate information is referenced to the North American Vertical Datum of 1988 (NAVD 88).

Horizontal coordinate information is referenced to the North American Datum of 1983 (NAD 83).

Elevation, as used in this report, refers to distance above the vertical datum.

SI to Inch/Pound

Multiply	By	To obtain
Length		
centimeter (cm)	0.3937	inch (in.)
millimeter (mm)	0.03937	inch (in.)
meter (m)	3.281	foot (ft)
meter (m)	1.094	yard (yd)
Volume		
liter (L)	33.82	ounce, fluid (fl. oz)
liter (L)	2.113	pint (pt)
liter (L)	1.057	quart (qt)
liter (L)	0.2642	gallon (gal)
cubic meter (m ³)	264.2	gallon (gal)
cubic decimeter (dm ³)	0.2642	gallon (gal)
liter (L)	61.02	cubic inch (in ³)
Mass		
gram (g)	0.03527	ounce, avoirdupois (oz)
kilogram (kg)	2.205	pound avoirdupois (lb)

Abbreviated Units Chemical concentration and water temperature are given only in metric units. Chemical concentration in water is given as milligrams per liter (mg/L). Chemical concentrations in solid phase samples are given as micromoles per gram ($\mu\text{moles g}^{-1}$). Grain size abundances are listed in weight percent (wt.%). Specific conductance is given in microsiemens per centimeter at 25 degrees Celsius ($\mu\text{S/cm}$ at 25 °C). Stable-isotope ratios are reported relative to internationally agreed-upon standards. Deviations from standards are expressed in per mil using the delta (δ) notation. The delta symbol in this report is followed by the chemical symbol for the heavier isotope measured during isotopic analysis.

Chemical and Isotopic Symbols

Ag_2S	silver sulfide
AVS	acid-volatile sulfide
BaCl_2	barium chloride
BaSO_4	barium sulfate
Ca	calcium
Cl^-	chloride
DI	disulfide sulfur
EDTA	ethylenediaminetetraacetic acid
Fe	iron
Fe^{2+}	ferrous iron
FeS_2	iron disulfide
Fe_{total}	sum of ferrous and ferric iron
HCl	hydrochloric acid
HCO_3^-	bicarbonate ion
H_2S	hydrogen sulfide
K	potassium
Mg	magnesium
N	normal (concentration)
Na	sodium
NH_4^+	ammonium
S	sulfur
SO_4^{2-}	sulfate
Ti	titanium
$\delta^2\text{H}_{\text{H}_2\text{O}}$	hydrogen isotope composition of water
$\delta^{18}\text{O}_{\text{H}_2\text{O}}$	oxygen isotope composition of water
$\delta^{18}\text{O}_{\text{SO}_4}$	oxygen isotope composition of sulfate
$\delta^{34}\text{S}_{\text{AVS}}$	sulfur isotope composition of acid-volatile sulfide
$\delta^{34}\text{S}_{\text{DI}}$	sulfur isotope composition of disulfide (reducible) sulfur
$\delta^{34}\text{S}_{\text{sulfate-HCl}}$	sulfur isotope composition of sulfate extracted with 6N HCl

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Abstract

Analytical results on sediment and associated ground water from the Canadian River alluvium collected subsequent to those described in Breit and others (2005) are presented in this report. The data presented herein were collected primarily to evaluate the iron and sulfur species within the sediment at well sites IC 36, IC 54, and IC South located at the USGS Norman Landfill study site. Cored sediment and water samples were collected during October 2004 and April 2005. The 52 sediment samples collected by coring were analyzed to determine grain size, the abundance of extractable iron species, and the abundance of sulfur forms and their isotopic compositions. Ground water was collected from cluster wells that sampled ground water from 11 to 15 screened intervals at each of the three sites. The depth range of the wells overlapped the interval of cored sediment. Concentrations of major ions, dissolved organic carbon (DOC), ammonium, and iron are reported with pH, specific conductance, and the isotopic composition of the water for the 75 water samples analyzed. Dissolved sulfate in selected water samples was analyzed to determine its sulfur and oxygen isotope composition.

Introduction

Landfills were sited adjacent to rivers prior to the passage of regulations intended to restrict the impact of solid wastes and leachates produced by interaction of ground and surface water with those wastes (Steiner and others, 1994). The U.S. Geological Survey (USGS) Toxic Hydrology Program is supporting the study of a closed municipal landfill located on Canadian River alluvium south of the city of Norman, Oklahoma, to evaluate the environmental impact of one of these waste repositories. The closed Norman municipal landfill operated in varying forms beginning in the 1920s until 1985 (fig. 1) (Schlottmann, 2001). Initially the wastes were dispersed along the north bank of the river. Beginning in 1960, the operators excavated trenches to the water table and buried wastes beneath 15 cm of sand. Trenches were abandoned in 1971, and wastes were deposited on lifts of sand 0.6 meters above the water table. Wastes were buried daily with sand. The landfill was closed in 1985 and covered with a low-permeability cap, leaving two mounds rising about 12 meters above the surrounding land surface (Schlottmann, 2001).

Movement of ground water through the waste produces a leachate distinguished by high concentrations of several dissolved constituents including organic carbon, iron, methane, ammonium, and boron (Cozzarelli and others, 2000; Schlottmann, 2001). Chemical and biological processes that degrade organic contaminants as the leachate moves through the Canadian River alluvium are the primary emphasis of the project. Collectively these attenuation processes may be important as a means of natural remediation. Bacterial oxidation of the contaminant organic compounds is supported by the presence of electron acceptors such as dissolved oxygen, nitrate, and sulfate, as well as solid manganese and ferric oxyhydroxides and oxides. This report presents data on the abundance of solid forms of iron and sulfur species in sediment within 2.8 meters of the ground surface as an addition to the previously published results of Breit and others (2005). In addition, ground water proximal to the sediment samples was chemically and isotopically analyzed. The depth range evaluated in this study is subject to seasonal fluctuations in water level (Becker, 2002; Scholl and others, 2005), with the shallowest ground water originating largely from infiltration of precipitation into the sediment (Scholl and others, 2005; 2006). The annual changes in water level are anticipated to affect the redox environment of the sediment and therefore the abundance of electron acceptors. A greater abundance of reduced forms of iron and sulfur are predicted following high water levels that are characteristic of spring, and the converse behavior is anticipated in the fall because of low water levels in late summer (Scholl and others, 2006). Such an annual fluctuation may impact the availability of electron acceptors necessary for natural attenuation processes. To test this hypothesis, samples of sediment and ground water were collected in October 2004 and April 2005 to represent seasons of generally low and generally high water levels, respectively. Results of sediment and water analyses are presented in this report.

Methods

Sample Site Selection

Previous studies by Scholl and others (2005, 2006) explored the annual variations in water composition at sites of three cluster wells, IC 36, IC 54, and IC South. Because of the known variation in water composition at these sites, they were the focus of this investigation. The IC sites are south of the landfill mounds (fig. 1; table 1), which is down the hydrologic gradient (Scholl and Christenson, 1998). The locations of the sites are: IC 36, 35 m from the toe of the landfill mound; IC 54, 7 m south of the slough, which is a former channel of the Canadian River; IC South, 85 m south of the slough and located in an area disturbed by major floods and channel avulsion during the 1980s (Schlottmann, 2001).

Variations in the chemical and isotopic composition of water recovered from the IC sites reflect the distinct histories of water collected at the various sampled depths and locations (Scholl and others, 2005; 2006). All sites produced water in the shallowest intervals, which is attributed to dilute recharge by precipitation. Deeper water at IC 36 is considered to contain a component of leachate from the landfill. Water in the lower wells at IC 54 is consistent with recharge from the slough and possibly leachate. At IC South the deepest wells sampled may produce water originally recharged from the slough or from the Canadian River (Scholl and others, 2005; 2006).

Table 1. Location of the IC sample sites and water levels at the time of sampling. Horizontal coordinates relative to NAD83 datum. Elevation relative to NADV88 datum.

Sample Site	Latitude	Longitude	Land Surface Elevation (meters)	Water Elevation October 2004 (meters)	Water Elevation April 2005 (meters)
IC 36	35.167496° N	97.446614° W	330.96	330.11	330.28
IC 54	35.166582° N	97.447231° W	330.56	329.85	329.90
IC South	35.165146° N	97.446614° W	330.31	329.62	329.59

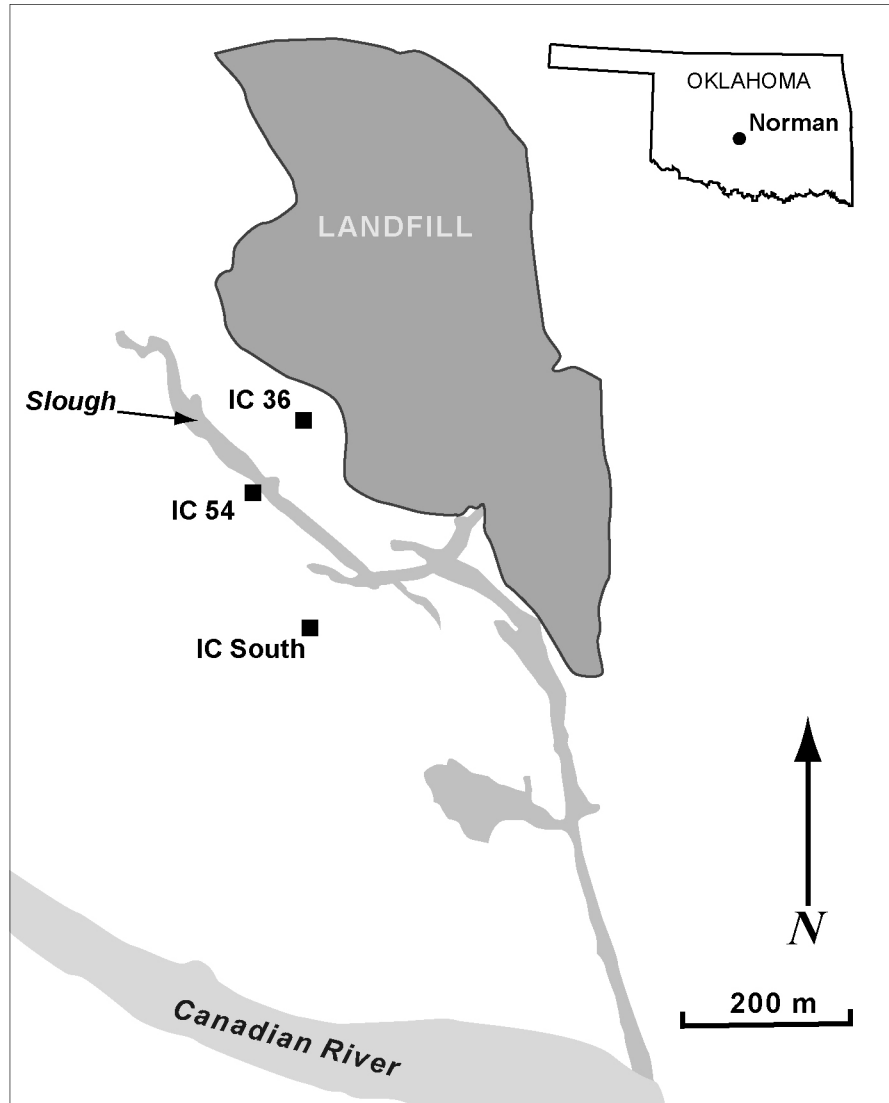


Figure 1. Location map of the IC sample sites (■). Topographic and hydrologic gradients slope to the south. Outline of landfill indicates the extent of the two capped landfill mounds.

Sediment Sampling

Cores of the alluvium were recovered within a few meters laterally of the three IC cluster well sites using a Geoprobe direct-push coring device. At each of the three sites sediment was collected from the ground surface to a depth ranging from 2.1 to 2.8 m. Recovery of the sandy sediment was incomplete in the October 2004 sampling because of problems retaining the flowing sediment in the core barrel. The missing depth range was the focus of recovery during the April 2005 core collection. During coring, sediment was captured in transparent acrylic liners that were 4.1 cm in diameter. The recovered core was cut into 0.1 to 0.4 m lengths, frozen on site with dry ice and shipped with dry ice to USGS laboratories in Denver. Upon arrival in Denver, the cores were transferred to a freezer and stored at -10 °C to inhibit redox or microbial changes to the sediment until they were prepared for analysis.

Within one month of sample collection, the frozen core was cut into lengths that approximated the depth range of each well in the cluster. While frozen, the core was extruded and split lengthwise into four approximately equivalent fractions using a hammer and chisel. One quarter of the sample was weighed and transferred to the glass reaction vessel for sulfur speciation as described below. The second quarter was transferred to a dish and allowed to air dry for the determination of moisture content. The third quarter of each sample collected during the October 2004 collection was weighed and transferred to a polyethylene bottle for extraction of water-soluble sulfate. This extraction was omitted during analysis of the April 2005 samples. The remaining sample was allowed to thaw slightly and, while still cold, was homogenized by manual mixing. Two aliquots of the homogenized sediment were transferred to glass vials equipped with butyl rubber septa. One vial was used for the iron speciation analyses, and the other was archived after freeze-drying.

Water Sampling and Field Analyses

Water was collected from existing wells (appendix 1) following the procedures outlined by Scholl and others (2005). Water was drawn slowly from each well in the cluster using a peristaltic pump. Specific conductance and pH were measured on site using calibrated meters. Ammonium content was measured spectrophotometrically on unfiltered samples that were diluted to fit the reporting range of the Nessler CHEMETS kit. Water samples for determination of alkalinity were filtered to <0.4 micrometer (μm) in the field, and transported with ice to the laboratory. An aliquot of water was filtered (<0.1 μm) and acidified with high-purity concentrated nitric acid to a pH less than 2 for analysis of cations and trace elements in the laboratory. Another aliquot was filtered in the field to <0.2 μm and was chilled with ice and transported to the laboratory for measurement of anions. Water passed through a 0.45 μm silver filter to remove bacteria and was stored in a baked glass bottle and acidified with phosphoric acid to a pH <2 for determination of the dissolved organic carbon content. Unfiltered water was also collected in a glass bottle sealed tightly with polyseal caps to prevent evaporation and refrigerated for later determination of the hydrogen ($\delta^2\text{H}$) and oxygen ($\delta^{18}\text{O}$) isotope composition of the water.

Sediment Analyses

Sediment samples were analyzed to determine the grain size distribution, the abundance of extractable iron species, and the abundance and isotopic composition of extractable sulfur species.

Stainless steel sieves were used to divide the air-dried sediment into five fractions: >500 μm , 500-250 μm , 250-100 μm , 100-62 μm , and <62 μm . The sediment and sieves were shaken manually for 5 minutes and the resulting sized fractions were weighed to the nearest 0.01 gram. Although large intraclasts of mud and silt were visually recognized, they were gently disaggregated using a mortar and pestle for the size analysis. Disaggregation is compatible with the emphasis of this study on the reactivity of the sediment, which is reasonably evaluated by the dispersed particles.

Chemical extractions for iron speciation were modified slightly from those of Heron and others (1994). Approximately 5 to 10 grams of sediment was combined with 40 mL of 0.5 normal (N) hydrochloric acid (HCl) in an amber glass serum vial that was purged with nitrogen gas ($\text{N}_2(\text{g})$). The sediment was reacted for 24 hours with intermittent agitation to suspend the sediment. Upon completion of the reaction, an aliquot of the 0.5N HCl was removed using a needle and syringe. The withdrawn solution was immediately filtered through a 0.2- μm syringe filter and reacted with orthophenanthroline according to the procedure of Clesceri and others (1998) to determine ferrous and total dissolved iron concentrations. The remaining sediment was subsequently recovered by filtration, rinsed thoroughly with distilled water, and air-dried overnight.

The air-dried sediment residue of the 0.5N HCl was weighed and then split using a mechanical splitter into a fraction weighing between 1.5 to 3.0 grams. The weighed fraction was combined with 40 mL of 0.008 molar titanous chloride ($\text{Ti}^{3+}\text{Cl}_3$) in 0.05 molar disodium ethylenediaminetetraacetic acid (EDTA) with pH of the combined solution adjusted to between 6 and 6.5 using sodium hydroxide. The Ti^{3+} -EDTA solution was prepared and introduced into an amber glass serum vial under $\text{N}_2(\text{g})$. The serum vial was sealed, purged with $\text{N}_2(\text{g})$ for 3 minutes, wrapped with foil, and shaken intermittently over the 24 hour reaction period. Following reaction, 30 mL of solution was removed through the septum using a needle and syringe and then filtered through a 0.2- μm syringe filter. The Ti^{3+} -EDTA solution has a medium-intensity magenta color prior to reaction with ferric iron. This color was apparent, although less intense, at the completion of the reaction indicating that the Ti^{3+} was not completely consumed during reaction with the sediment. The filtered solution was then analyzed for total iron using the orthophenanthroline procedure (Clesceri and others, 1998). Although the sediment was dried in air prior to evaluation of reducible iron, the readily reactive iron phases were removed by the 0.5 N HCl prior to air exposure, and any remaining reduced iron phases such as pyrite are slowly reactive and contribute little to the Ti^{3+} -EDTA extraction (Heron and others, 1994).

Water-soluble sulfate was determined by combining 40 to 80 grams of cold sediment with 200 milliliters of deionized water. The sample and water were agitated for 1 hour. The resulting solution was separated from the sediment by filtration, acidified to a pH between 3 and 4 with HCl, heated to approximately 90 $^\circ\text{C}$, and combined with 5 mL of barium chloride (BaCl_2). After several hours, the heating was discontinued, and the solution was allowed to cool overnight. The resulting barium sulfate (BaSO_4) precipitate was recovered by filtration, air-dried, and weighed.

The abundances of acid-volatile sulfide (AVS), acid-soluble sulfate, and total reducible-disulfide (DI) were determined following the extraction scheme of Tuttle and others (1986). Approximately 40 to 80 grams of frozen sediment was transferred under a nitrogen atmosphere into a glass reaction vessel. Under flowing $\text{N}_2(\text{g})$ the sample was combined with 50 mL of 6 N HCl and 1 gram of stannous chloride and heated to evolve hydrogen sulfide (H_2S) from AVS phases. The stannous chloride was added to reduce ferric iron present in hematite grain coatings that might oxidize

evolved H₂S (Rice and others, 1993). The H₂S was then carried in flowing nitrogen through a reservoir of pH 4 buffer to remove HCl vapor and subsequently bubbled through a trap containing silver nitrate, which resulted in the formation of silver sulfide (Ag₂S). The Ag₂S was collected by filtration, dried, and weighed to determine the mass of recovered sulfide.

Following completion of release of H₂S, the acid solution was separated from the remaining sediment by filtration. The acid solution was reacted with 10 mL of 10 weight percent BaCl₂ to precipitate dissolved sulfate as BaSO₄, following the procedure outlined for water-extracted sulfate. The precipitated sulfate is considered acid-soluble sulfate.

The residual solid remaining after the 6 N HCl extraction was air-dried, weighed and placed in a reaction vessel under flowing nitrogen, where it was combined with 1M chromous chloride (CrCl₂) in 4 N HCl with 10 mL of ethanol. Chromium(II) reduces sulfur in FeS₂ phases, known to be present in the sediment (Breit and others, 2005), to produce H₂S. As in the AVS analysis, evolved H₂S was carried through a pH 4 buffer to strip HCl vapors and then bubbled through a silver nitrate solution that precipitated the H₂S as Ag₂S. The resulting Ag₂S was collected by filtration, air-dried, and weighed to determine the amount of contained disulfide sulfur. Replicate analyses produced concentrations of sulfide phases within ±10 weight percent.

The sulfur isotope composition of Ag₂S produced by the AVS and DI extractions, as well as the precipitated BaSO₄, were measured by direct combustion (Kester and others, 2001) at the USGS Isotope Laboratory in Denver. Sulfur isotope data are reported relative to Vienna Canon Diablo Troilite (VCDT) in standard per mil values. Precision of the measurement of δ³⁴S values is within ± 0.3 per mil.

Water Laboratory Analyses

Major ion, trace element, dissolved organic carbon, alkalinity, and isotope composition were determined on selected water samples in USGS laboratories in Reston, Virginia (Baedecker and Cozzarelli, 1992). Measurement of alkalinity was completed by titration using a Titra Lab Tim 900 autotitrator with 0.0254 N HCl. Alkalinity is reported as bicarbonate ion (Baedecker and Cozzarelli, 1992). Direct-current plasma spectrometry determined the cation and trace element abundances in of the water samples. Anions were measured using a Dionex 120 ion chromatograph equipped with an Ion Pac AS14 column. Dissolved organic carbon (DOC) was measured by the method of Qian and Mopper (1996). Addition of dissolved barium chloride to selected water samples precipitated dissolved sulfate for isotope analyses, using the same procedure for precipitation of sulfate that was extracted by water from the sediment samples. The pH of the water samples was adjusted to be between 3 and 4 with addition of HCl to remove dissolved carbonate species while minimizing exchange of oxygen bound to sulfate. The isotopic composition of recovered sulfate and water were determined at the USGS Reston Stable Isotope Laboratory. Sulfur isotopes and oxygen isotope composition of sulfate were determined by continuous flow with precisions of ± 0.2 and ±0.3 per mil respectively. Sulfate sulfur is reported relative to Vienna Canon Diablo Troilite (VCDT) and oxygen is reported relative to Vienna Standard Mean Ocean Water (VSMOW) as per mil values. The δ²H composition of water was determined by equilibration with gaseous hydrogen; resulting precision of this analysis was ± 1 per mil. δ¹⁸O of the water was determined through equilibration with CO₂; resulting precision was ± 0.1 per mil. Both hydrogen and oxygen isotope compositions are reported as per mil relative to

VSMOW and normalized such that the $\delta^2\text{H}$ and $\delta^{18}\text{O}$ values of Standard Light Antarctic Precipitation (SLAP) are -5.55 per mil and -4.28 per mil respectively.

Results

Sediment Composition

The alluvium deposited by the Canadian River contains sand, silt, clay, and aggregates of fine-grained sediment in intraclasts (Breit and others, 2005). Most of the samples considered in this study are red-brown as the result of grain-coating hematite that was eroded from Permian red beds within the drainage basin. Exceptions to this pervasive coloration are the samples from deeper than approximately 330 m elevation at the IC 36 well site, which are distinguished by their gray-brown color. Bedforms observed in the IC cores vary from finely laminated to massive (appendix 2). Sedimentary structures are readily recognized in sediment collected above the water table; water-saturated sediment was likely liquefied during the coring process, resulting in disruption of primary sedimentary structures to produce the massive appearance.

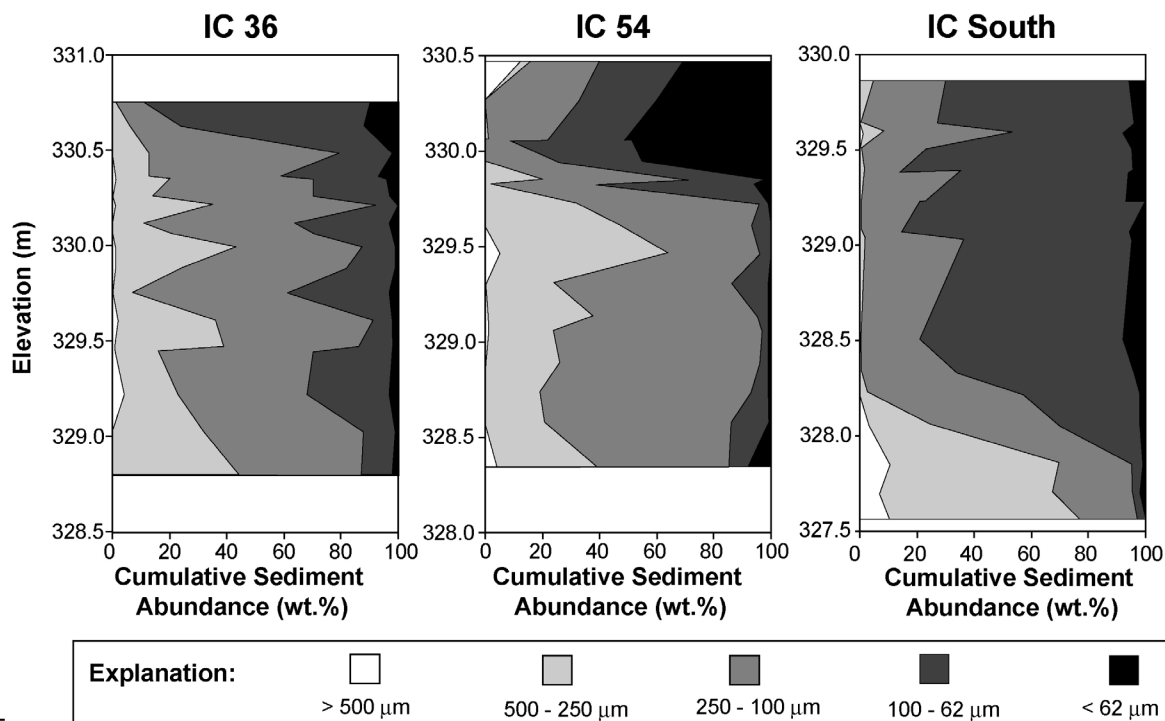


Figure 2. Cumulative abundance of size fractions measured by sieve analysis of sediment samples recovered at IC 36, IC 54, and IC South.

Results of the sieve analysis of the sediment samples are presented in figure 2 and listed in appendix 3. Sediment recovered from all cores is mainly medium to very fine sand. The deepest sediment from IC South is composed of relatively coarse-grained sediment consistent with the presence of pebble-sized intraclasts recovered in this interval. Sediment recovered from the upper

0.5 m of IC 54 is notable for the greater abundance of particles less than 62 μm compared to other cored intervals.

The amounts of iron extracted with the 0.5 N HCl and Ti^{3+} -EDTA extractions are presented in figure 3 and listed in appendix 4. The amount of total iron extracted by 0.5 N HCl ranged from 1.2 to 18.8 micromoles per gram ($\mu\text{moles g}^{-1}$). For all samples, Ti^{3+} -EDTA released greater amounts of iron (7.9 to 48 $\mu\text{moles g}^{-1}$) than the 0.5 N HCl extraction. At all sites the greatest amounts of Ti^{3+} -EDTA-extracted iron were from samples collected above the position of the water table at the time of sample collection. The abundance of ferrous iron relative to total iron extracted with 0.5 N HCl generally varied directly with position of the sample relative to the water table (fig. 4). Sediment samples collected above the water table, with the exception of one sample from IC 54, have $\text{Fe}^{2+}/\text{Fe}_{\text{total}}$ ratios <0.25 . Below the water table there is a general increase in the ratio, with values approaching 1.0 near the bottom of the cored intervals. Results of the iron extractions on samples collected in October 2004 and April 2005 are broadly consistent with similar chemical conditions in the sediment during both sampling events.

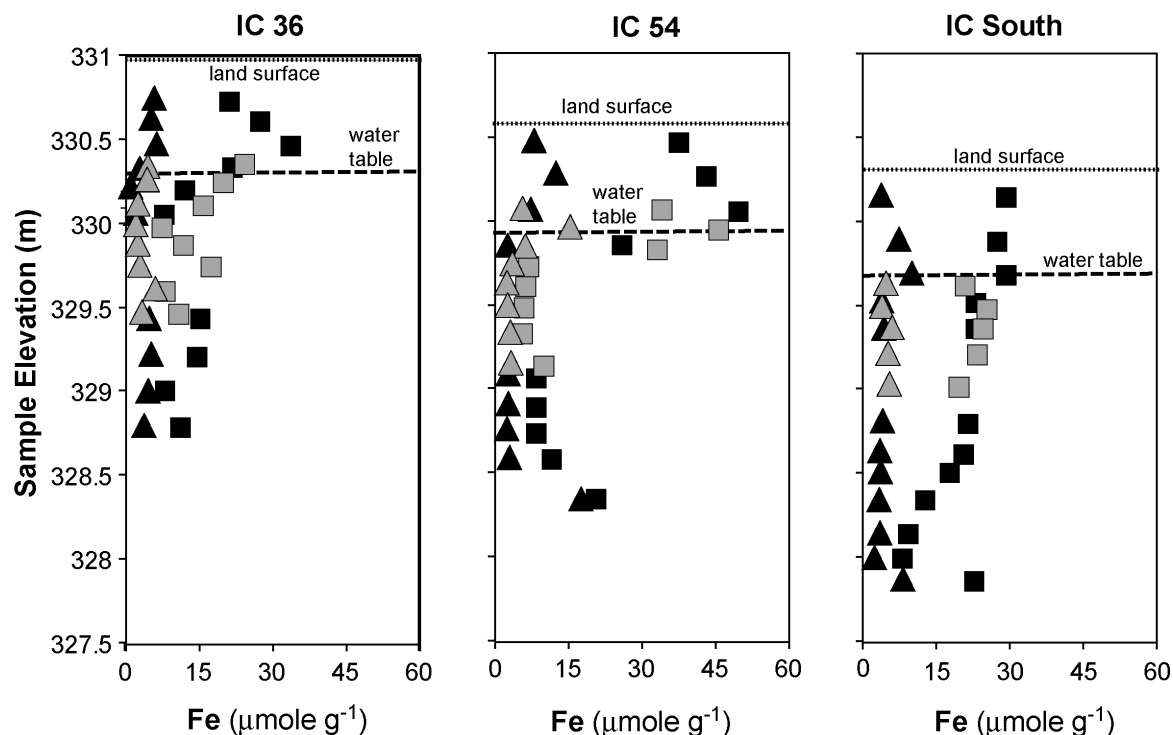


Figure 3. Depth variation in the total iron extracted using 0.5N HCl (\blacktriangle) and Ti^{3+} -EDTA (\blacksquare) on sediment samples collected at IC 36, IC 54, and IC South. Elevation of water table during the April 2005 sample collection is shown. (black, collected in October 2004; gray, collected in April 2005).

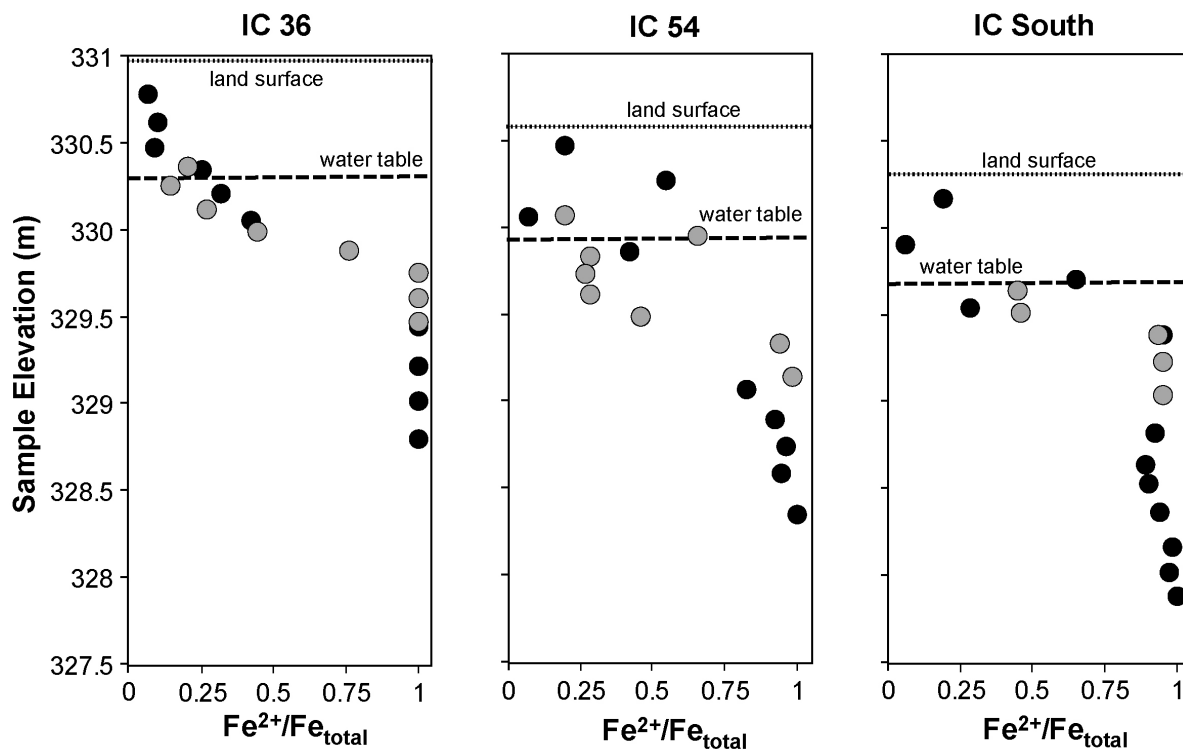


Figure 4. Depth variation in the ratio of ferrous (Fe^{2+}) to total iron (Fe_{total}) extracted with 0.5 N HCl from sediment samples collected at IC 36, IC 54, and IC South. Elevation of water table during the April 2005 sample collection is shown as a dashed line. (black, collected in October 2004; gray, collected in April 2005).

The amount of iron extracted was also evaluated relative to the grain size of the samples. No systematic distribution of 0.5 N HCl-extractable iron with grain size distribution of the sediment was recognized. In contrast, the amount of Ti^{3+} -EDTA-extractable iron did increase as the fraction of sediment smaller than 100 μm increased (figure 5). For samples with similar contents of particles smaller than 100 μm , those collected above the water table generally contain markedly greater amounts of Ti^{3+} -EDTA-extractable iron than samples from the saturated zone.

The abundances of water-soluble sulfate, acid-soluble sulfate, acid-volatile sulfide, and disulfide sulfur recovered from the sediment are listed in appendix 5 along with the $\delta^{34}S$ values of these phases. Results of the water-soluble sulfate extraction on sediment collected in October 2004 yielded only small amounts of sulfate (less than 0.5 $\mu moles\ g^{-1}$ sulfate sulfur); therefore, the extraction was not applied to the April 2005 sample set. The amounts of acid-soluble sulfate recovered vary from less than 0.1 to 2.4 $\mu moles\ g^{-1}$. Acid-volatile sulfide concentrations range from less than 0.1 to 3.8 $\mu moles\ g^{-1}$, and disulfide sulfur varies from less than 0.1 to 15 $\mu moles\ g^{-1}$. Most samples contain greater amounts of acid-volatile sulfide than disulfide sulfur.

Depth variations in the abundance of acid-volatile sulfide and disulfide sulfur are plotted on figure 6. The vertical variations in the concentrations of acid-volatile sulfide and disulfide sulfur are similar at each site, but the profiles are distinct among the IC sites. Sediment above the water table at IC 36 lacks detectable sulfide, whereas large amounts were recovered from the same depth range

at IC 54 and minor amounts at IC South. Sediment a few tens of centimeters below the water table at all sites has lower sulfide concentrations than deeper sediment. Sulfide concentrations at IC 36 and IC 54 increase sharply in samples below the water table, while the profile for IC South portrays a more gradual increase with depth.

The range of sulfur isotope compositions of acid-soluble sulfate (4 to 20 per mil) is distinct from the values measured for acid-volatile sulfide (-0.5 to -21.5 per mil) and disulfide sulfur (-4 to -25.1 per mil) (fig. 7). The $\delta^{34}\text{S}$ values of acid-soluble sulfate collected at IC South are consistently smaller than those measured at IC 36 and IC 54 (fig. 7). Sulfur isotope values of acid-volatile sulfide among the three cores are generally similar, while the $\delta^{34}\text{S}$ of disulfide sulfur measured in IC 54 core is greater than at the other sites. The typically negative values for $\delta^{34}\text{S}$ of the sulfide phases relative to the values of sulfate are consistent with formation of sulfide by bacterial sulfate reduction (Breit and others, 2007).

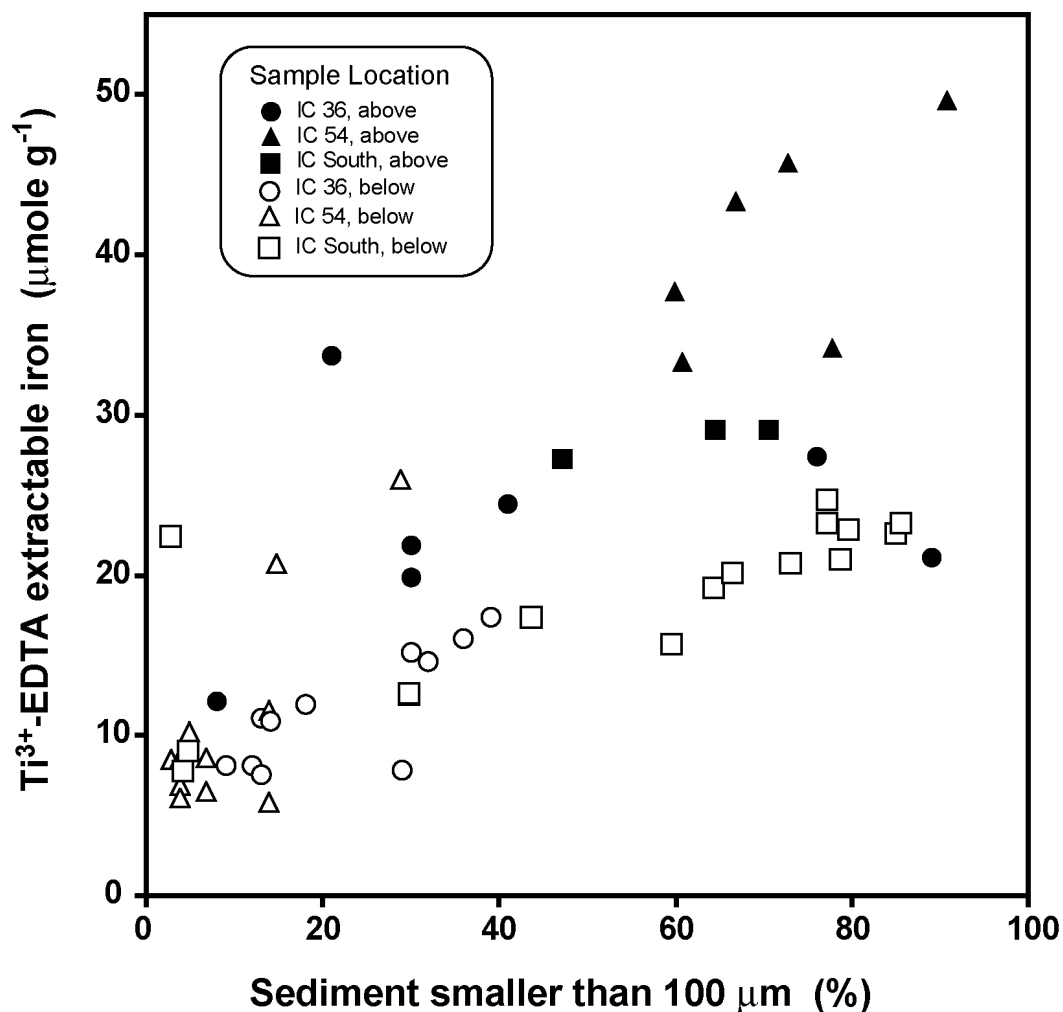


Figure 5. Abundance of iron extracted from sediment samples with Ti^{3+} -EDTA relative to the abundance of particles smaller than $100\ \mu\text{m}$. ('above', samples collected above the saturated zone; 'below', samples collected within the saturated zone).

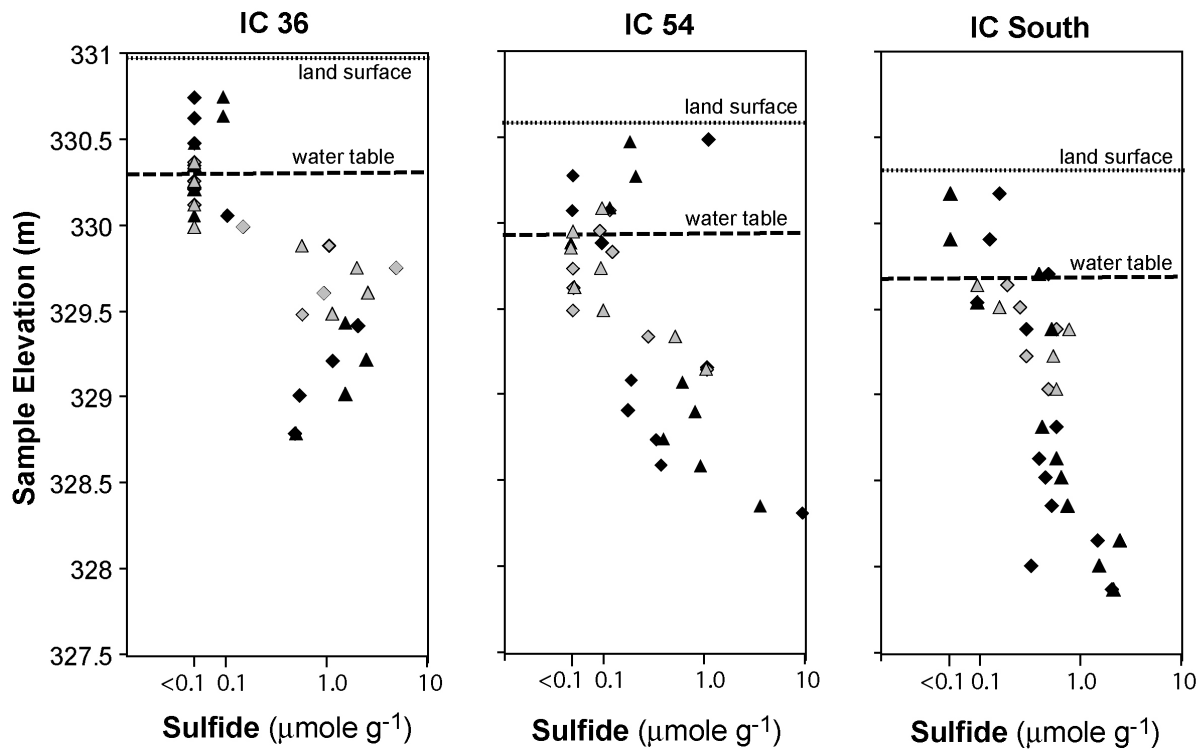


Figure 6. Depth variation in the abundance of acid-volatile sulfide (▲) and disulfide (◆) extracted from sediment collected at IC 36, IC 54, and IC South. Elevation of water table during the April 2005 sample collection is shown by the dashed line. (Black indicates samples collected in October 2004; gray indicates samples collected in April 2005).

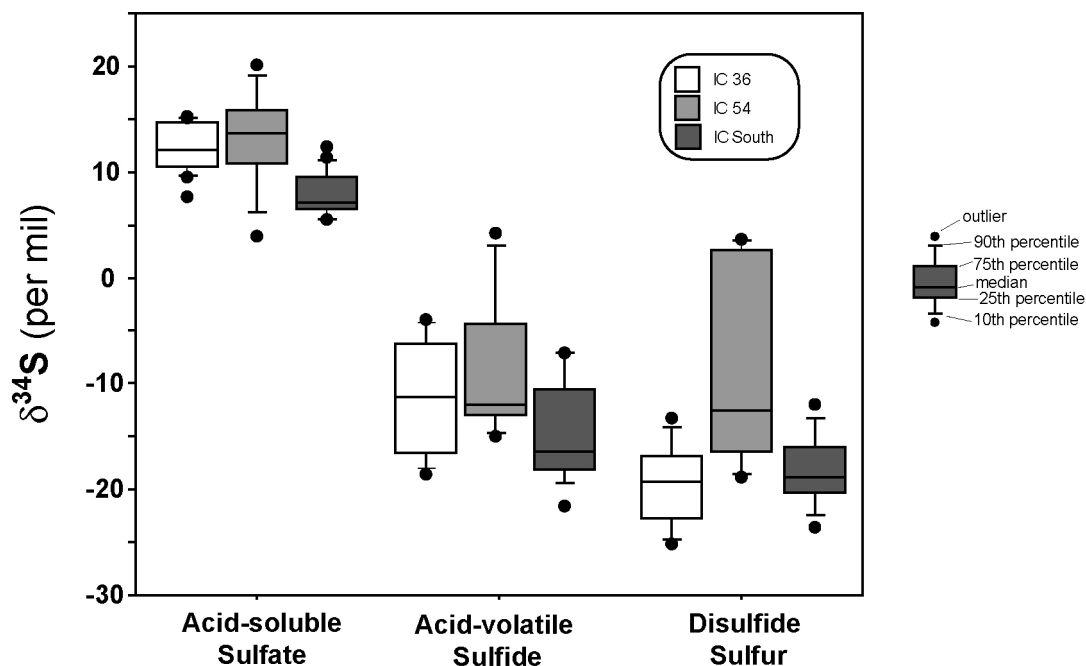


Figure 7. Box plot of the sulfur isotope composition of sulfur species extracted from the sediment samples collected at IC 36, IC 54 and IC South.

Water Composition

The composition of water samples is presented in appendix 6 and summarized in table 2. In general, the concentration of dissolved constituents at the IC site follows the vertical trends described by Scholl and others (2005, 2006). Low concentrations of most measured constituents were detected at shallow depths at all three sites. This tendency is indicated in the profiles of chloride concentration at IC 36 and IC South (fig. 8).

Dissolved iron concentrations generally increase with depth, but the trends vary at each IC site (fig. 9). Sample preservation problems limited the iron analyses of water samples from IC 36 that were collected in October, but the April samples suggest two maxima in iron content.

Samples from IC 54 have low dissolved iron contents without a clear maximum for samples gathered in October, whereas the April samples define a maximum 1.0 m below the ground surface and an increase in iron concentration in the two deepest wells. The profile for IC South is similar for both the October and April sampling, with maxima at approximately 1.5 m depth.

The concentration of dissolved sulfate increases with depth to a maximum value and then declines to concentrations near the limit of detection at all sites (fig. 10). The depth of the maxima is similar in both the October and April sampling, although the concentrations that define the maxima are greater in October.

Table 2. Statistical summary of parameters measured on ground water samples collected from IC wells in October 2004 and April 2005.

[Sp. Cond. – specific conductance; $\mu\text{S}/\text{cm}$, microSiemens per centimeter; DOC, dissolved organic carbon; mg/L, milligrams per liter].

	IC 36	IC 54	IC South
pH			
Median	7.07	7.24	7.10
Minimum	6.78	6.75	6.07
Maximum	7.49	7.70	7.80
Sp. Cond. ($\mu\text{S}/\text{cm}$)			
Median	3600	1400	920
Minimum	600	1100	620
Maximum	4800	2400	1500
Calcium (mg/L)			
Median	99	96	160
Minimum	67	50	100
Maximum	240	170	200
Magnesium (mg/L)			
Median	55	35	19
Minimum	7.5	22	8.0
Maximum	95	59	45
Potassium (mg/L)			
Median	150	6.4	4.2
Minimum	0.6	4.5	1.3
Maximum	230	9.1	5.2
Sodium (mg/L)			
Median	350	190	14
Minimum	0.6	77	5.4
Maximum	530	310	110

	IC 36	IC 54	IC South
Iron (mg/L)			
Median	7.8	1.9	3.6
Minimum	0.1	0.3	0.02
Maximum	14	7.8	8.6
Ammonium (mg/L)			
Median	65	2.8	1.0
Minimum	<0.7	<0.7	<0.7
Maximum	430	8.6	4.1
DOC (mg/L)			
Median	57	20	5.8
Minimum	6.0	2.1	1.1
Maximum	92	32	11
Bicarbonate (mg/L)			
Median	1850	864	578
Minimum	337	765	278
Maximum	2330	1150	830
Chloride (mg/L)			
Median	255	81	7.2
Minimum	8.9	41	0.3
Maximum	396	256	134
Sulfate (mg/L)			
Median	18	21	27
Minimum	0.2	0.2	4.7
Maximum	78	120	130

The major ion characteristics of water samples are depicted on a Piper plot (fig. 11) in which the bicarbonate concentration is multiplied by 0.1 to enhance contrast in anion composition among the water samples. Water samples from IC 36 and IC 54 tend to have consistently greater proportions of chloride than most samples from IC South. Similarly, IC 36 and IC 54 have a greater relative content of sodium than ground water sampled from IC South.

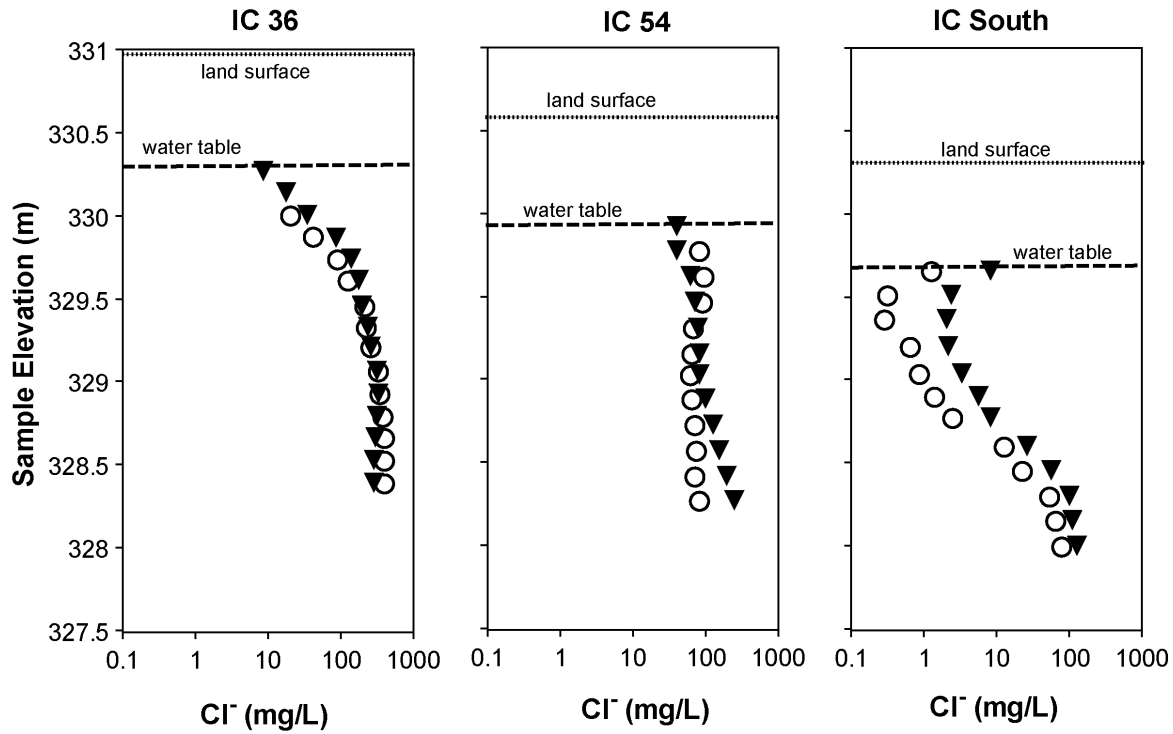


Figure 8. Depth variation of chloride concentration of ground water sampled from the wells at IC 36, IC 54, and IC South in October 2004 (○) and April 2005 (▼).

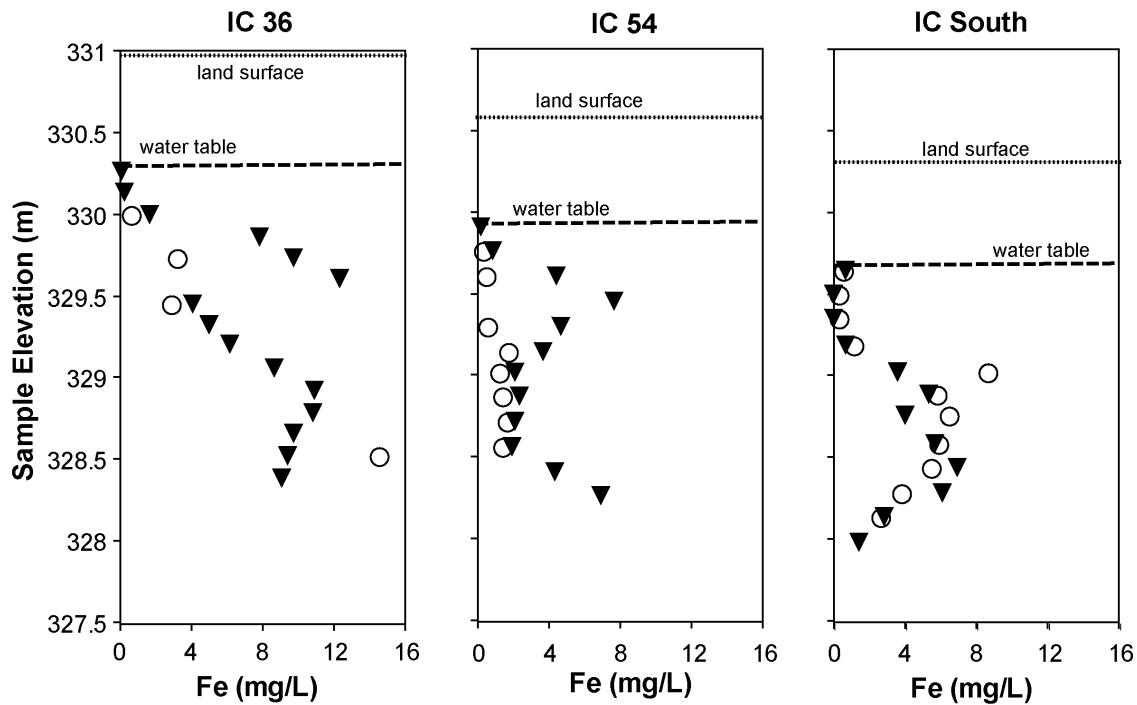


Figure 9. Depth variation in iron concentration of ground water from the wells at IC 36, IC 54, and IC South in October 2004 (○) and April 2005 (▼).

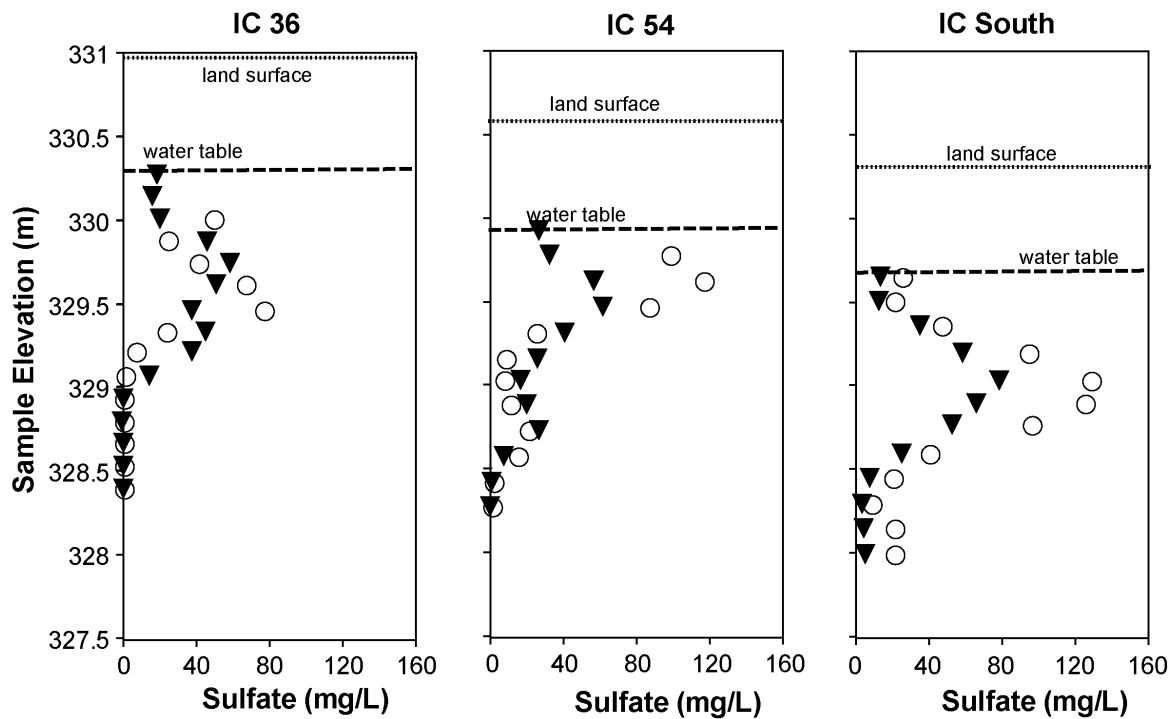


Figure 10. Depth variation of sulfate concentration of ground water sampled from the wells at IC 36, IC 54, and IC South in October 2004 (○) and April 2005 (▼). Elevation of water table during the April 2005 sample collection is shown by the dashed line.

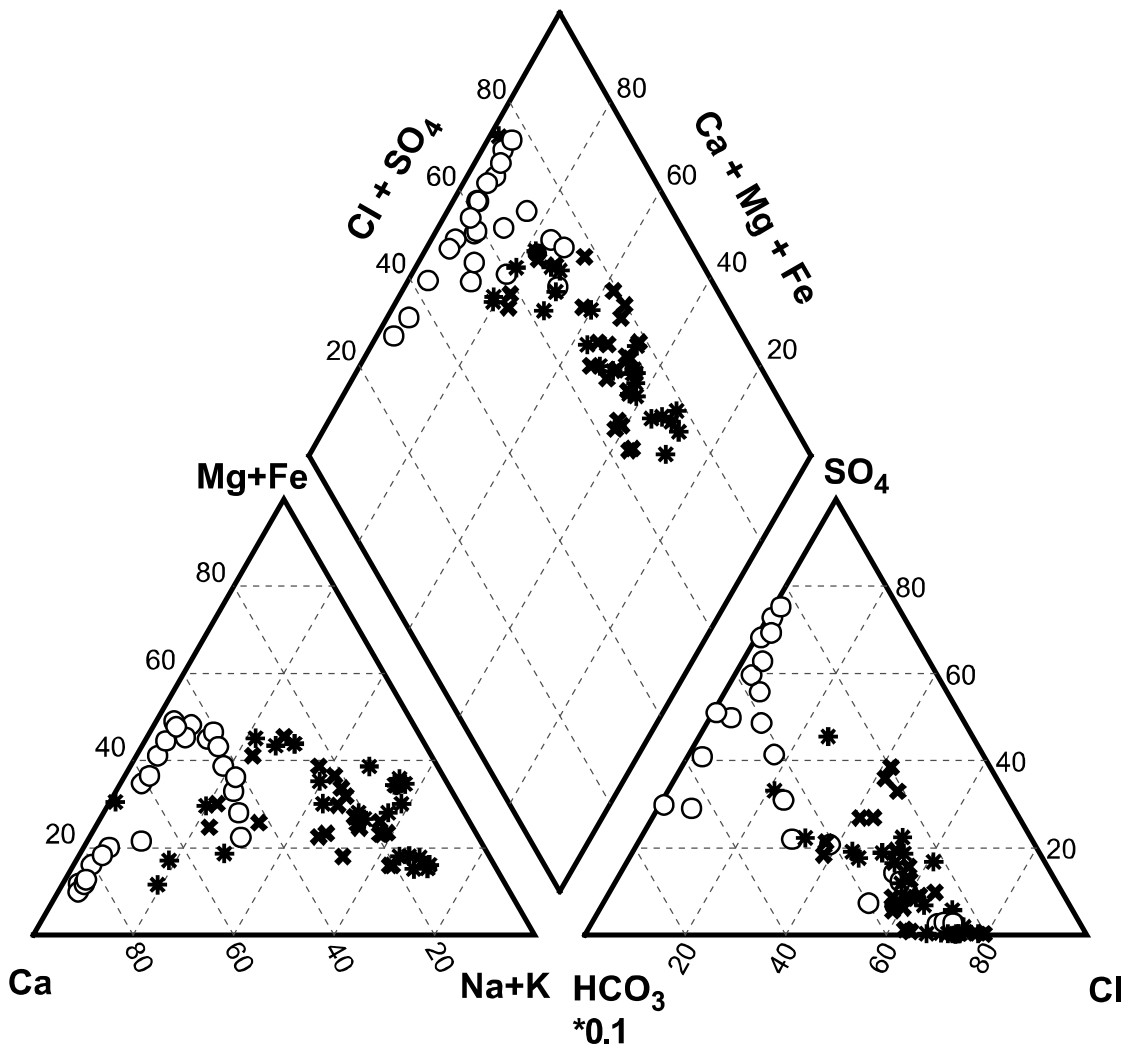


Figure 11. Piper diagram of major ion composition water samples collected from the wells at IC 36 (*), IC 54 (x), and IC South (o). Note that the bicarbonate concentration is multiplied by 0.1.

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Appendix 1. Depth intervals of the wells sampled at IC 36, IC 54 and IC South. Elevations are in meters and are relative to the North American Vertical Datum of 1988.

Well Site	Well Number	Top Elevation	Bottom Elevation
IC 36	3	330.33	330.19
IC 36	4	330.20	330.06
IC 36	5	330.06	329.93
IC 36	6	330.94	329.80
IC 36	7	329.80	329.66
IC 36	8	329.67	329.54
IC 36	9	329.52	329.38
IC 36	10	329.39	329.25
IC 36	11	329.27	329.13
IC 36	12	329.13	328.99
IC 36	13	328.99	328.85
IC 36	14	328.85	328.72
IC 36	15	328.72	328.59
IC 36	16	328.59	328.45
IC 36	17	328.45	328.32
IC 54	3	329.99	329.84
IC 54	4	329.84	329.69
IC 54	5	329.69	329.54
IC 54	6	329.54	329.38
IC 54	7	329.38	329.23
IC 54	8	329.23	329.07
IC 54	9	329.10	328.95
IC 54	10	328.95	328.80
IC 54	11	328.80	328.64
IC 54	12	328.64	328.49
IC 54	13	328.49	328.34
IC 54	14	328.34	328.19
IC South	3	329.73	329.58
IC South	4	329.58	329.43
IC South	5	329.43	329.28
IC South	6	329.27	329.12
IC South	7	329.11	328.96
IC South	8	328.97	328.82
IC South	9	328.84	328.69
IC South	10	328.67	328.52
IC South	11	328.52	328.37
IC South	12	328.37	328.22
IC South	13	328.22	328.07
IC South	14	328.07	327.92

Appendix 2. Elevation and description of cored sediment samples. Elevations are reported relative to North American Vertical Datum of 1988.

Sample Number	Sampled Interval		Sediment Description
	Upper Elevation (m)	Lower Elevation (m)	
IC 36-04-A	330.80	330.69	red-brown, laminated, clayey, fine sand; overlying sand is mixed with landfill trash fragments
IC 36-04-B	330.69	330.55	red, laminated, fine sand
IC 36-04-C	330.55	330.41	red-brown, laminated, clayey, fine-medium sand
IC 36-04-D	330.41	330.28	red-brown, laminated, medium sand
IC 36-04-E	330.28	330.14	brown, laminated, clayey, medium sand
IC 36-04-F	330.14	329.97	gray-brown, massive, medium sand
IC 36-04-G	329.56	329.33	gray-brown, massive, medium sand
IC 36-04-H	329.33	329.11	gray-brown, massive, medium sand
IC 36-04-I	329.11	328.93	gray-brown, massive, medium sand
IC 36-04-J	328.93	328.66	red-gray-brown, massive, medium sand
IC 36-05-A	330.4	330.32	red-brown, fine sand
IC 36-05-B	330.32	330.19	red brown, bedded, medium sand
IC 36-05-C	330.19	330.04	red-brown, bedded, fine-medium sand
IC 36-05-D	330.04	329.94	red-brown, massive, medium to coarse sand
IC 36-05-E	329.94	329.82	gray-brown, medium sand with sparse clay partings
IC 36-05-F	329.82	329.68	gray-brown, medium sand with laminae of dark-brown clay and fragments of carbonized plant fragments
IC 36-05-G	329.68	329.53	gray-brown, massive, medium sand
IC 36-05-H	329.53	329.41	gray-brown, massive, medium sand
IC 54-04-A	330.56	330.38	Brown, fine sand overlying brown silt
IC 54-04-B	330.38	330.16	red-brown, fine sand with interbedded brown clayey silt
IC 54-04-C	330.16	329.96	red-brown, laminated, silt and fine sand
IC 54-04-D	329.96	329.75	red, laminated, medium sand
IC 54-04-E	329.16	328.97	gray-brown, massive, medium sand
IC 54-04-F	328.97	328.82	gray-brown, medium sand
IC 54-04-G	328.82	328.66	brown, massive, medium sand
IC 54-04-H	328.66	328.51	gray-brown, medium sand
IC 54-04-I	328.40	328.30	intraclast conglomerate of red-brown, clayey, silt clasts, and pale red-brown coarse-medium sand with carbonized wood fragments
IC 54-05-A	330.12	330.01	red-brown, fine sand
IC 54-05-B	330.01	329.89	red-brown, silt

Appendix 2. Elevation and description of cored sediment samples —Continued.

Sample Number	Sampled Interval		Sediment Description
	Upper Elevation (m)	Lower Elevation (m)	
IC 54-05-C	329.89	329.77	red-brown, laminated, fine sand
IC 54-05-D	329.77	329.69	red, medium sand
IC 54-05-E	329.69	329.54	red, faintly bedded, medium sand
IC 54-05-F	329.54	329.40	light red-brown, medium sand
IC 54-05-G	329.40	329.23	gray-brown, medium-fine sand with distorted bedding
IC 54-05-H	329.23	329.05	gray-brown, massive, medium sand with a few granules
IC South-04-A	330.01	329.71	red, laminated, fine sand
IC South-04-B	329.71	329.47	red, laminated, fine-medium sand interbedded with red, silty-sand
IC South-04-C	329.47	329.31	red-brown, laminated, clayey, fine sand with dark red-brown clay granules and small carbonized wood fragments
IC South-04-D	329.31	329.15	red-brown, clayey, fine sand
IC South-04-E	329.15	329.00	red-brown, fine sand with silty laminae
IC South-04-F	328.61	328.40	red, massive, fine sand
IC South-04-G	328.40	328.26	red, massive, fine sand
IC South-04-H	328.26	328.18	red-brown, fine-medium sand
IC South-04-I	328.18	327.92	upper 8 cm is red-brown, medium sand; lower 9 cm is gray-brown, upper medium sand with minor mud-clast fragments
IC South-04-J	327.92	327.78	red-brown, upper medium sand, with deformed clayey silt laminae
IC South-04-K	327.78	327.63	red-brown, upper medium sand with mud clasts and snail shells
IC South-04-L	327.63	327.51	red-brown, upper medium sand with mud clast
IC South-05-A	329.69	329.58	red-brown, laminated, fine sand with carbonized wood fragments
IC South-05-B	329.58	329.43	red, faintly laminated, fine sand
IC South-05-C	329.43	329.33	red-brown, laminated, fine sand
IC South-05-D	329.33	329.12	red, with faintly visible distorted bedding, fine sand
IC South-05-E	329.12	328.93	brown, medium-fine sand

Appendix 3. Results of the grain-size analysis of sediment samples.

Sample Number	>500 μm (wt.%)	500 to 250 μm (wt.%)	250 to 100 μm (wt.%)	100 to 62 μm (wt.%)	<62 μm (wt.%)
IC 36-04-A	<1	1	10	79	10
IC 36-04-B	<1	6	18	64	12
IC 36-04-C	<1	13	65	19	2
IC 36-04-D	1	19	50	26	4
IC 36-04-E	1	33	57	8	<1
IC 36-04-F	<1	20	50	27	2
IC 36-04-G	1	15	54	28	2
IC 36-04-H	4	19	45	29	3
IC 36-04-I	<1	32	55	11	1
IC 36-04-J	<1	44	43	11	2
IC 36-05-A	<1	13	46	34	7
IC 36-05-B	<1	14	56	27	3
IC 36-05-C	<1	11	54	33	3
IC 36-05-D	1	41	44	12	1
IC 36-05-E	1	23	58	17	1
IC 36-05-F	<1	7	54	36	3
IC 36-05-G	2	34	55	7	2
IC 36-05-H	1	38	48	12	2
IC 54-04-A	12	4	24	29	31
IC 54-04-B	<1	<1	33	27	40
IC 54-04-C	<1	<1	9	42	49
IC 54-04-D	<1	20	50	27	2
IC 54-04-E	1	23	73	2	1
IC 54-04-F	<1	26	71	3	1
IC 54-04-G	<1	19	74	6	1
IC 54-04-H	<1	21	66	13	1
IC 54-04-I	4	35	47	7	8

Appendix 3. Results of the grain-size analysis of sediment samples—Continued.

Sample Number	>500 μm (wt.%)	500 to 250 μm (wt.%)	250 to 100 μm (wt.%)	100 to 62 μm (wt.%)	<62 μm (wt.%)
IC 54-05-A	<1	1	21	27	51
IC 54-05-B	<1	<1	26	29	44
IC 54-05-C	<1	2	37	55	6
IC 54-05-D	<1	32	65	3	1
IC 54-05-E	<1	46	46	7	<1
IC 54-05-F	5	60	32	4	<1
IC 54-05-G	<1	24	61	13	1
IC 54-05-H	1	37	57	4	1
IC South-04-A	<1	4	26	64	6
IC South-04-B	1	7	45	39	8
IC South-04-C	<1	1	34	60	4
IC South-04-D	<1	<1	21	79	1
IC South-04-E	<1	<1	15	79	6
IC South-04-F	<1	<1	21	71	8
IC South-04-G	<1	<1	34	62	4
IC South-04-H	<1	2	55	41	2
IC South-04-I	3	21	46	28	2
IC South-04-J	10	59	26	4	1
IC South-04-K	7	60	28	3	1
IC South-04-L	10	66	21	2	1
IC South-05-A	<1	<1	27	69	4
IC South-05-B	<1	<1	23	72	4
IC South-05-C	<1	<1	14	80	5
IC South-05-D	<1	<1	23	70	7
IC South-05-E	<1	1	35	59	5

Appendix 4. Concentration of iron sequentially extracted from the sediment samples using 0.5 N HCl and Ti³⁺-EDTA. Concentrations were calculated on an air-dried sediment basis.

[ppm, parts per million; Fe²⁺, ferrous iron; Fe_{total}, sum of ferrous and ferric iron]

Sample Number	Fe ²⁺ 0.5 N HCl (μmole g ⁻¹)	Fe _{total} 0.5N HCl (μmole g ⁻¹)	Fe ²⁺ /Fe _{total} 0.5N HCl	Fe Ti ³⁺ -EDTA (μmole g ⁻¹)
IC 36-04-A	0.3	6.3	0.05	21.1
IC 36-04-B	0.5	5.4	0.10	27.4
IC 36-04-C	0.5	6.1	0.09	33.7
IC 36-04-D	0.6	2.3	0.26	21.8
IC 36-04-E	0.4	1.2	0.32	12.2
IC 36-04-F	0.5	1.3	0.42	7.9
IC 36-04-G	4.7	4.7	1.00	15.2
IC 36-04-H	4.8	4.8	1.00	14.7
IC 36-04-I	4.8	4.8	1.00	8.2
IC 36-04-J	3.9	3.9	1.00	11.1
IC 36-05-A	0.7	3.4	0.20	24.5
IC 36-05-B	0.5	3.4	0.14	19.9
IC 36-05-C	0.6	2.3	0.28	15.9
IC 36-05-D	0.8	2.0	0.42	7.5
IC 36-05-E	2.1	2.7	0.80	11.8
IC 36-05-F	3.0	3.0	1.00	17.4
IC 36-05-G	6.1	6.1	1.00	8.1
IC 36-05-H	4.5	4.5	1.00	10.9
IC 54-04-A	1.6	7.9	0.20	37.6
IC 54-04-B	6.8	12.5	0.54	43.3
IC 54-04-C	0.5	7.0	0.07	49.6
IC 54-04-D	0.6	1.4	0.42	26.0
IC 54-04-E	1.8	2.1	0.83	8.4
IC 54-04-F	2.1	2.3	0.92	8.4
IC 54-04-G	2.3	2.5	0.93	8.4
IC 54-04-H	3.0	3.2	0.94	11.5
IC 54-04-I	19.0	18.8	1.01	20.6

Appendix 4. Concentration of iron sequentially extracted from the sediment samples—Continued.

Sample Number	Fe²⁺ 0.5 N HCl ($\mu\text{mole g}^{-1}$)	Fe_{total} 0.5N HCl ($\mu\text{mole g}^{-1}$)	Fe²⁺/Fe_{total} 0.5N HCl	Fe Ti³⁺-EDTA ($\mu\text{mole g}^{-1}$)
IC 54-05-A	1.2	6.1	0.20	34.0
IC 54-05-B	9.8	15.0	0.66	45.7
IC 54-05-C	1.3	4.5	0.28	33.3
IC 54-05-D	0.7	2.7	0.28	6.8
IC 54-05-E	0.5	1.7	0.28	6.4
IC 54-05-F	1.0	2.1	0.45	6.1
IC 54-05-G	2.9	3.0	0.94	5.7
IC 54-05-H	3.0	3.0	1.00	10.2
IC South-04-A	0.6	3.4	0.19	29.2
IC South-04-B	0.4	7.2	0.06	27.4
IC South-04-C	6.4	9.8	0.65	29.2
IC South-04-D	1.0	3.6	0.29	22.9
IC South-04-E	4.5	4.7	0.96	22.7
IC South-04-F	3.8	3.9	0.95	21.1
IC South-04-G	2.9	3.2	0.89	20.2
IC South-04-H	3.0	3.4	0.89	17.5
IC South-04-I	3.0	3.2	0.94	12.5
IC South-04-J	3.0	3.0	1.00	9.0
IC South-04-K	2.3	2.3	1.00	7.9
IC South-04-L	7.7	7.7	1.00	22.6
IC South-05-A	2.0	4.5	0.44	20.8
IC South-05-B	1.6	3.4	0.47	24.7
IC South-05-C	4.7	5.0	0.93	23.3
IC South-05-D	4.7	4.8	0.96	23.3
IC South-05-E	4.8	5.2	0.93	19.3

Appendix 5. Concentrations of water-soluble sulfate sulfur, acid-soluble sulfate sulfur, acid-volatile sulfide and disulfide sulfur extracted from the sediment samples and their sulfur isotope composition ($\delta^{34}\text{S}$). Recovered water-soluble sulfate was insufficient for isotope analyses. Acid-soluble sulfate extracted with hot 6 N HCl. Concentrations were calculated on an air-dried sediment basis.

[nd, not determined; sulfate-HCl, acid-soluble sulfate; AVS, acid-volatile sulfide; DI, disulfide sulfur]

Sample Number	Sulfate-Sulfur H_2O ($\mu\text{mole g}^{-1}$)	Sulfate-Sulfur 6N HCl ($\mu\text{mole g}^{-1}$)	AVS ($\mu\text{mole g}^{-1}$)	DI ($\mu\text{mole g}^{-1}$)	$\delta^{34}\text{S}_{\text{sulfate-HCl}}$ (per mil)	$\delta^{34}\text{S}_{\text{AVS}}$ (per mil)	$\delta^{34}\text{S}_{\text{DI}}$ (per mil)
IC 36-04-A	0.2	1.0	0.1	<0.1	10	nd	nd
IC 36-04-B	<0.2	0.5	0.1	<0.1	9.6	nd	nd
IC 36-04-C	<0.2	0.8	<0.1	<0.1	10.2	nd	nd
IC 36-04-D	<0.2	0.7	<0.1	<0.1	11.4	nd	nd
IC 36-04-E	0.2	0.8	<0.1	<0.1	11.6	nd	nd
IC 36-04-F	<0.2	0.7	<0.1	0.1	12.7	nd	nd
IC 36-04-G	<0.2	0.7	1.7	2.2	13.3	-16.8	-24.0
IC 36-04-H	<0.2	0.6	2.8	1.2	15	-8.8	-19.3
IC 36-04-I	<0.2	0.7	1.7	0.6	13.7	-3.9	-19.1
IC 36-04-J	<0.2	0.6	0.6	0.5	11	-13.6	-17.7
IC 36-05-A	nd	0.2	<0.1	<0.1	7.6	nd	nd
IC 36-05-B	nd	0.5	<0.1	<0.1	10.6	nd	nd
IC 36-05-C	nd	0.4	<0.1	<0.1	12.1	nd	nd
IC 36-05-D	nd	0.8	<0.1	0.2	15.2	nd	nd
IC 36-05-E	nd	0.7	0.7	1.2	14.9	-16.3	-16
IC 36-05-F	nd	1.0	2.3	5.5	14.7	-18.5	-25.1
IC 36-05-G	nd	0.6	2.8	1.1	15.1	-7.6	-21.5
IC 36-05-H	nd	0.2	1.3	0.7	nd	-4.9	-13.3
IC 54-04-A	0.2	1.6	0.2	1.2	4	nd	-9.1
IC 54-04-B	70.2	2.4	0.2	<0.1	7.3	nd	nd
IC 54-04-C	70.2	1.7	0.1	<0.1	6.5	nd	nd
IC 54-04-D	0.3	0.7	<0.1	0.1	14.8	nd	nd
IC 54-04-E	<0.2	0.4	0.6	0.2	14.1	-15.0	nd
IC 54-04-F	<0.2	0.4	0.8	0.2	15.8	-13.1	-18.8
IC 54-04-G	<0.2	0.7	0.4	0.3	20.1	-12.0	nd

Appendix 5. Concentrations and isotopic composition of sulfur species—Continued.

Sample Number	Sulfate-Sulfur H₂O ($\mu\text{mole g}^{-1}$)	Sulfate-Sulfur 6N HCl ($\mu\text{mole g}^{-1}$)	AVS ($\mu\text{mole g}^{-1}$)	DI ($\mu\text{mole g}^{-1}$)	$\delta^{34}\text{S}_{\text{sulfate-HCl}}$ (per mil)	$\delta^{34}\text{S}_{\text{AVS}}$ (per mil)	$\delta^{34}\text{S}_{\text{DI}}$ (per mil)
IC 54-04-H	<0.2	0.7	0.9	0.4	19.0	-11.2	-15.9
IC 54-04-I	0.5	1.9	3.8	15.0	10.8	4.3	3.6
IC 54-05-A	nd	1.4	0.1	0.1	nd	nd	nd
IC 54-05-B	nd	0.5	<0.1	0.1	nd	nd	nd
IC 54-05-C	nd	0.3	<0.1	0.1	nd	nd	nd
IC 54-05-D	nd	0.4	0.1	<0.1	12.8	nd	nd
IC 54-05-E	nd	0.5	<0.1	<0.1	13.7	nd	nd
IC 54-05-F	nd	0.8	0.1	<0.1	13.6	nd	nd
IC 54-05-G	nd	0.5	0.5	0.3	18.1	-12.4	-16.4
IC 54-05-H	nd	0.6	1.1	1.1	11.3	-2	2.7
IC South-04-A	<0.2	0.8	<0.1	0.2	7.0	nd	-16
IC South-04-B	<0.2	1.1	<0.1	0.1	6.2	nd	nd
IC South-04-C	0.2	1.3	0.4	0.5	5.5	-18.5	-19.7
IC South-04-D	<0.2	0.8	0.1	0.1	6.8	nd	nd
IC South-04-E	0.2	0.8	0.5	0.3	5.7	-14.4	-18.8
IC South-04-F	0.2	0.9	0.4	0.6	5.5	-15.4	-22.4
IC South-04-G	<0.2	0.6	0.6	0.4	7.3	-17.8	-17.4
IC South-04-H	<0.2	0.7	0.6	0.4	9.7	-21.5	-15.4
IC South-04-I	<0.2	0.8	0.7	0.5	10.2	-17.8	-20.3
IC South-04-J	<0.2	1.4	2.4	1.4	9.5	-7.1	-16
IC South-04-K	<0.2	0.9	1.5	0.3	12.4	-7.8	-12
IC South-04-L	<0.2	1.2	2.1	2.0	11.4	-7.1	-13.3
IC South-05-A	nd	0.8	0.1	0.2	7.3	nd	-17
IC South-05-B	nd	0.7	0.2	0.3	7.2	nd	-19.7
IC South-05-C	nd	0.8	0.8	0.6	7.1	-13.3	-22.1
IC South-05-D	nd	0.8	0.5	0.3	6.6	-18.4	-23.5
IC South-05-E	nd	0.6	0.6	0.5	6.8	-17.4	-19.9

Appendix 6. Results of chemical and isotopic analyses of ground water samples from wells at IC 36, IC 54, and IC South.

[nd, not determined; S.Cond., specific conductance; $\mu\text{S}/\text{cm}$, microSiemens per centimeter; DOC, dissolved organic carbon; mg/L, milligrams per liter]

Well	Sample Date	Elevation of Well Screen Center	pH	S. Cond. ($\mu\text{S}/\text{cm}$)	Ca (mg/L)	Mg (mg/L)	K (mg/L)	Na (mg/L)	Fe (mg/L)	NH_4^+ (mg/L)
IC 36-5	13-Oct-04	329.995	7.40	760	91.8	12	47	29	0.61	4.8
IC 36-6	13-Oct-04	329.870	7.27	1040	67.2	18	0.6	0.60	nd	52
IC 36-7	13-Oct-04	329.730	7.33	1410	81.8	30	87	92	3.1	64
IC 36-8	13-Oct-04	329.605	7.34	2330	238	89	108	245	12.3	31
IC 36-9	13-Oct-04	329.450	7.22	2330	134	66	151	292	2.9	17
IC 36-10	13-Oct-04	329.320	7.20	3350	90.2	48	188	303	nd	50
IC 36-11	13-Oct-04	329.200	7.01	3350	84.2	54	213	354	nd	46
IC 36-12	13-Oct-04	329.060	7.02	4340	87.8	63	228	395	nd	294
IC 36-13	13-Oct-04	328.920	7.06	4660	90.0	66	189	371	nd	138
IC 36-14	13-Oct-04	328.785	6.98	4800	130	86	208	473	nd	169
IC 36-15	13-Oct-04	328.655	7.02	4653	133	78	153	437	nd	335
IC 36-16	13-Oct-04	328.520	7.03	4630	164	95	172	531	14.5	361
IC 36-17	13-Oct-04	328.385	7.02	4570	nd	nd	nd	nd	nd	289
IC 36-3	19-Apr-05	330.260	7.28	584	86.6	7.4	13	19	0.09	<0.7
IC 36-4	19-Apr-05	330.130	7.28	584	106	14.1	21	22	0.22	2.10
IC 36-5	19-Apr-05	329.995	7.07	928	125	23	28	38	1.7	2.6
IC 36-6	19-Apr-05	329.995	7.07	928	176	48	51	99	7.8	2.0
IC 36-7	19-Apr-05	329.730	7.07	928	219	70	82	170	9.8	17
IC 36-8	19-Apr-05	329.605	7.07	3104	238	89	108	245	12.3	63
IC 36-9	19-Apr-05	329.450	7.08	2826	202	83	142	273	4.1	88
IC 36-10	19-Apr-05	329.320	6.78	3691	171	73	182	352	5.0	148
IC 36-11	19-Apr-05	329.200	7.23	4068	120	54	200	379	6.2	206
IC 36-12	19-Apr-05	329.060	7.49	4424	108	59	229	466	8.6	186
IC 36-13	19-Apr-05	328.920	7.10	4313	82.0	57	198	472	10.9	192

Appendix 6. Results of chemical and isotopic analyses of ground water samples from wells—Continued.

Well Sampled	DOC (mg/L)	HCO₃⁻ mg/L	Cl⁻ (mg/L)	SO₄²⁻ (mg/L)	δ¹⁸O_{SO4} (per mil)	δ³⁴S_{SO4} (per mil)	δ¹⁸O_{H2O} (per mil)	δ²H_{H2O} (per mil)
IC 36-5	nd	nd	21	50	6.6	10.7	-5.66	-34.70
IC 36-6	9.4	nd	43	25	13.6	36.0	-5.83	-34.00
IC 36-7	16	nd	90	42	15.0	43.0	-5.94	-33.20
IC 36-8	29	nd	130	67	16.6	38.9	-6.04	-31.30
IC 36-9	49	nd	210	78	16.5	36.5	-5.98	-27.90
IC 36-10	57	nd	230	24	17.6	41.3	-5.89	-26.70
IC 36-11	69	nd	270	7.9	nd	nd	-5.84	-23.00
IC 36-12	80	nd	340	1.4	nd	nd	-5.71	-20.80
IC 36-13	86	nd	360	0.5	nd	nd	-5.74	-19.20
IC 36-14	92	nd	390	0.6	nd	nd	-5.62	-17.00
IC 36-15	88	nd	400	0.6	nd	nd	-5.66	-16.80
IC 36-16	85	nd	400	0.8	nd	nd	-5.63	-18.50
IC 36-17	82	nd	400	0.5	nd	nd	-5.57	-17.80
IC 36-3	6.0	336.9	8.9	18.9	nd	14.7	-6.96	-41.30
IC 36-4	16	442.9	18.4	16.9	nd	20.6	-6.68	-39.19
IC 36-5	20	533.7	35.8	21	nd	27.4	-6.84	-39.02
IC 36-6	11	1008	91.2	47	nd	29.8	-7.12	-41.30
IC 36-7	19	1417	143	60	15.2	30.1	-6.96	-36.03
IC 36-8	18	1768	185	51	20.0	46.2	-6.82	-33.66
IC 36-9	30	1897	203	38	18.5	50.4	nd	nd
IC 36-10	50	2071	244	46	17.6	49.3	-6.31	-26.83
IC 36-11	55	2180	274	39	16.9	48.0	-6.06	-25.11
IC 36-12	70	2332	329	15.2	nd	nd	-5.81	-21.18
IC 36-13	80	2224	348	1.0	nd	nd	-5.48	-20.64

Appendix 6. Results of chemical and isotopic analyses of ground water samples from wells—Continued.

Well	Sample Date	Elevation of Well Screen Center	pH	S. Cond. ($\mu\text{S}/\text{cm}$)	Ca (mg/L)	Mg (mg/L)	K (mg/L)	Na (mg/L)	Fe (mg/L)	NH_4^+ (mg/L)
IC 36-14	19-Apr-05	328.785	6.95	4030	82.7	56	166	447	10.8	226
IC 36-15	19-Apr-05	328.655	7.10	3694	84.8	53	146	440	9.8	159
IC 36-16	19-Apr-05	328.520	7.02	3639	87.4	51	135	425	9.4	141
IC 36-17	19-Apr-05	328.385	6.97	3592	86.4	50	119	414	9.1	119
IC 54-4	14-Oct-04	329.765	7.10	1580	150	51	7.5	125	0.31	0.9
IC 54-5	14-Oct-04	329.615	7.15	1580	118	44	7.6	187	0.47	0.8
IC 54-6	14-Oct-04	329.460	7.24	1470	96	35	7.8	191	nd	0.7
IC 54-7	14-Oct-04	329.305	7.28	1160	76	28	7.2	130	0.59	2.0
IC 54-8	14-Oct-04	329.150	7.42	nd	67	28	7.2	138	1.7	4.3
IC 54-9	14-Oct-04	329.025	7.42	1120	54	24	6.7	181	1.2	4.3
IC 54-10	14-Oct-04	328.875	7.70	1160	50	22	5.8	183	1.4	5.8
IC 54-11	14-Oct-04	328.720	7.70	1250	52	24	5.1	180	1.7	5.6
IC 54-12	14-Oct-04	328.565	7.50	nd	65	29	5.9	214	1.4	6.2
IC 54-13	14-Oct-04	328.415	7.60	1290	58	26	4.8	194	nd	6.5
IC 54-14	14-Oct-04	328.265	7.50	1360	61	28	4.5	211	nd	6.6
IC 54-3	19-Apr-05	329.915	7.07	1580	168	44	9.1	77	0.27	1.1
IC 54-4	19-Apr-05	329.765	6.75	1322	159	48	5.8	77	0.93	<0.7
IC 54-5	19-Apr-05	329.615	6.84	1471	159	55	6.4	113	4.5	2.1
IC 54-6	19-Apr-05	329.460	6.84	1539	146	51	8.0	160	7.8	0.6
IC 54-7	20-Apr-05	329.305	6.83	1426	96	35	6.3	168	4.8	0.6
IC 54-8	20-Apr-05	329.150	7.10	1410	87	34	6.8	197	3.8	0.7
IC 54-9	20-Apr-05	329.025	7.10	1442	83	33	6.7	207	2.2	0.7
IC 54-10	20-Apr-05	328.875	7.30	1586	91	37	6.3	238	2.4	2.8
IC 54-11	20-Apr-05	328.720	7.25	1696	103	42	6.0	262	2.2	4.0

Appendix 6. Results of chemical and isotopic analyses of ground water samples from wells—Continued.

Well Sampled	DOC (mg/L)	HCO₃⁻ mg/L	Cl⁻ (mg/L)	SO₄²⁻ (mg/L)	δ¹⁸O_{SO4} (per mil)	δ³⁴S_{SO4} (per mil)	δ¹⁸O_{H2O} (per mil)	δ²H_{H2O} (per mil)
IC 36-14	90	2037	328	0.2	nd	nd	-5.31	-20.94
IC 36-15	88	1886	314	0.5	nd	nd	-5.17	-20.88
IC 36-16	82	1851	300	0.5	nd	nd	-5.02	-19.72
IC 36-17	81	1840	306	0.5	nd	nd	-5.09	-20.90
IC 54-4	6.6	nd	84	98	13.9	15.6	-3.80	-22.80
IC 54-5	9.8	nd	94	117	nd	nd	-3.43	-21.20
IC 54-6	15	nd	89	87	16.0	26.4	-3.24	-21.80
IC 54-7	19	nd	68	25	nd	nd	-3.05	-19.60
IC 54-8	32	nd	64	8.6	18.6	42.9	-2.78	-18.50
IC 54-9	21	nd	63	7.7	nd	nd	-4.04	-18.21
IC 54-10	22	nd	64	10.6	21.4	61.5	-3.02	-19.20
IC 54-11	25	nd	73	21	nd	nd	-2.56	-16.50
IC 54-12	25	nd	75	15.1	22.0	65.7	-2.49	-17.50
IC 54-13	25	nd	73	1.6	nd	nd	-2.62	-18.20
IC 54-14	24	nd	82	1.0	nd	nd	-2.89	-19.20
IC 54-3	2.1	813	41	27	nd	49.4	-5.36	-32.88
IC 54-4	5.4	820	43	33	nd	14.2	-5.21	-31.79
IC 54-5	20	870	65	57	nd	18.4	-4.62	-28.57
IC 54-6	13	858	74	61	nd	24.8	-3.91	-24.13
IC 54-7	16	766	81	40	nd	nd	-3.26	-21.17
IC 54-8	27	765	86	26	nd	nd	-2.90	-19.96
IC 54-9	14	812	87	16.5	nd	nd	-2.44	-17.32
IC 54-10	17	883	110	21	nd	74.1	-1.70	-14.29
IC 54-11	17	807	130	27	nd	nd	-1.27	-9.2

Appendix 6. Results of chemical and isotopic analyses of ground water samples from wells—Continued.

Well	Sample Date	Elevation of Well Screen Center	pH	S. Cond. ($\mu\text{S}/\text{cm}$)	Ca (mg/L)	Mg (mg/L)	K (mg/L)	Na (mg/L)	Fe (mg/L)	NH_4^+ (mg/L)
IC 54-12	20-Apr-05	328.565	7.23	1822	105	43	5.8	272	2.0	6.1
IC 54-13	20-Apr-05	328.415	7.28	2032	118	49	5.3	280	4.4	8.6
IC 54-14	20-Apr-05	328.265	6.87	2368	144	59	5.6	309	7.0	8.6
IC South-3	14-Oct-04	329.655	7.80	797	144	11	2.1	5.6	0.51	0.7
IC South-4	14-Oct-04	329.505	7.20	928	200	13	2.4	6.4	0.28	1.5
IC South-5	14-Oct-04	329.355	7.20	907	182	13	3.1	8.1	0.29	<0.7
IC South-6	14-Oct-04	329.195	7.20	913	183	16	4.2	11	1.1	1.3
IC South-7	14-Oct-04	329.035	7.10	979	201	20	5.1	13	8.6	0.9
IC South-8	14-Oct-04	328.895	7.21	1010	170	19	4.9	12	5.7	3.3
IC South-9	14-Oct-04	328.765	7.20	993	161	19	4.8	14	6.4	3.6
IC South-10	14-Oct-04	328.595	7.20	930	141	20	4.9	23	5.9	3.6
IC South-11	14-Oct-04	328.445	7.30	972	141	23	5.2	42	5.4	2.6
IC South-12	14-Oct-04	328.295	7.35	1140	142	32	5.2	66	3.7	5.4
IC South-13	14-Oct-04	328.145	7.40	1210	128	35	4.8	74	2.6	5.0
IC South-14	14-Oct-04	327.995	7.40	1320	142	40	4.9	99	nd	4.6
IC South-3	20-Apr-05	329.655	7.50	800	104	12	2.1	17.2	0.64	<0.7
IC South-4	20-Apr-05	329.655	6.63	628	123	8.1	1.3	5.4	0.02	<0.7
IC South-5	20-Apr-05	329.355	6.63	676	138	11	1.7	6.7	0.03	<0.7
IC South-6	20-Apr-05	329.195	6.71	781	154	14	2.7	8.4	0.64	2.5
IC South-7	20-Apr-05	329.035	6.58	888	178	17	3.7	11	3.6	<0.7
IC South-8	20-Apr-05	328.895	6.75	922	185	19	4.0	13	5.3	3.2
IC South-9	20-Apr-05	328.765	6.78	918	181	19	4.1	14	4.0	1.0
IC South-10	20-Apr-05	328.595	6.81	934	153	19	4.0	25	5.7	1.2
IC South-11	20-Apr-05	328.445	6.61	1082	158	24	4.5	52	6.9	<0.7

Appendix 6. Results of chemical and isotopic analyses of ground water samples from wells—Continued.

Well Sampled	DOC (mg/L)	HCO₃⁻ (mg/L)	Cl⁻ (mg/L)	SO₄²⁻ (mg/L)	δ¹⁸O_{SO4} (per mil)	δ³⁴S_{SO4} (per mil)	δ¹⁸O_{H2O} (per mil)	δ²H_{H2O} (per mil)
IC 54-12	26	969	160	7.9	20.2	62.1	-1.09	-8.28
IC 54-13	20	1030	210	1.1	nd	nd	-1.60	-8.75
IC 54-14	24	1160	260	0.2	nd	nd	-2.14	-10.55
IC South-3	7.3	nd	1.3	27	nd	nd	-5.19	-33.00
IC South-4	7.2	nd	0.3	22	12.0	-6.88	-5.18	-30.60
IC South-5	10	nd	0.3	48	nd	nd	-5.30	-31.10
IC South-6	nd	nd	0.7	96	14.0	-5.9	-5.36	-31.60
IC South-7	7.3	nd	0.9	130	nd	nd	-5.41	-30.70
IC South-8	5.9	nd	1.4	127	13.5	-4.8	-5.38	-29.80
IC South-9	6.5	nd	2.3	99	nd	nd	-5.20	-28.70
IC South-10	6.8	nd	12.6	42	18.0	11.1	-5.07	-25.70
IC South-11	7.0	nd	22	22	nd	nd	-4.88	-26.80
IC South-12	4.6	nd	53	9.7	18.1	31.3	-4.55	-25.00
IC South-13	11	nd	65	23	nd	nd	-4.28	-24.30
IC South-14	9.2	nd	78	23	20.5	40.1	-4.01	-23.00
IC South-3	4.9	278	8.5	14.8	nd	14.7	-6.09	-39.50
IC South-4	2.8	414	2.4	14.5	nd	nd	-5.59	-34.24
IC South-5	7.6	432	2.1	36	nd	nd	-5.28	-32.58
IC South-6	3.4	482	2.3	60	nd	nd	-5.37	-32.85
IC South-7	1.3	550	3.5	80	14.5	-1.3	-5.41	-33.05
IC South-8	3.7	586	6.0	68	15.3	31.8	-5.42	-31.75
IC South-9	1.1	581	8.7	54	nd	nd	-5.33	-29.99
IC South-10	1.6	575	31	23	nd	nd	-5.12	-28.28
IC South-11	4.9	638	60	9.3	nd	nd	-4.54	-25.48

Appendix 6. Results of chemical and isotopic analyses of ground water samples from wells—Continued.

Well	Sample Date	Elevation of Well Screen Center	pH	S. Cond. (μS/cm)	Ca (mg/L)	Mg (mg/L)	K (mg/L)	Na (mg/L)	Fe (mg/L)	NH₄⁺ (mg/L)
IC South-12	20-Apr-05	328.295	6.07	1343	169	31	4.6	67	6.1	1.3
IC South-13	20-Apr-05	328.145	6.62	1412	173	44	4.5	102	2.8	<0.7
IC South-14	20-Apr-05	327.995	6.97	1507	167	45	4.1	110	1.4	1.9

Appendix 6. Results of chemical and isotopic analyses of ground water samples from wells—Continued.

Well Sampled	DOC (mg/L)	HCO₃⁻ mg/L	Cl⁻ (mg/L)	SO₄²⁻ (mg/L)	δ¹⁸O_{SO4} (per mil)	δ³⁴S_{SO4} (per mil)	δ¹⁸O_{H2O} (per mil)	δ²H_{H2O} (per mil)
IC South-12	5.5	756	104	4.7	nd	nd	-4.65	-28.40
IC South-13	3.8	786	116	6.0	21.5	69.5	-3.96	-21.53
IC South-14	5.8	829	134	6.5	nd	69.5	-3.68	-19.93