

Plant Nitrogen Dynamics in Shortgrass Steppe under Elevated Atmospheric Carbon Dioxide

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ABSTRACT

The direct and indirect effects of increasing levels of atmospheric carbon dioxide (CO₂) on plant nitrogen (N) content were studied in a shortgrass steppe ecosystem in northeastern Colorado, USA. Beginning in 1997 nine experimental plots were established: three open-top chambers with ambient CO₂ levels (approximately 365 μ mol mol⁻¹), three open-top chambers with twice-ambient CO2 levels (approximately 720 μ mol mol⁻¹), and three unchambered control plots. After 3 years of growingseason CO2 treatment, the aboveground N concentration of plants grown under elevated atmospheric CO₂ decreased, and the carbon–nitrogen (C:N) ratio increased. the same time, aboveground biomass production under elevated atmospheric CO2 conditions increased the net transfer of N out of the soil of elevated-CO₂ plots. Aboveground biomass production after simulated herbivory was also greater under elevated CO₂ compared to ambient CO₂. Surprisingly, no significant changes in belowground plant tissue N content were detected in response to elevated CO₂. Measurements of individual species at peak standing phytomass showed significant effects of CO₂ treatment on aboveground plant tissue N concentration and significant differences between species in N concentration, suggesting that changes in species composition under elevated CO₂ will contribute to overall changes in nutrient cycling. Changes in plant N concentration, could have important consequences for biogeochemical cycling rates and the long-term productivity of the shortgrass steppe as atmospheric CO₂ concentrations increase.

Key words: Carbon dioxide; rangelands; plant nitrogen; *Bouteloua gracilis*; *Pascopyrum smithii*; *Stipa comata*; C₃ grasses; C₄ grasses; nitrogen yield.

The effects of increases in the atmospheric concentration of carbon dioxide (CO₂) on nitrogen (N) cycling will shape the way that ecosystems respond to elevated CO₂ because plant productivity in most ecosystems is limited by N availability (Hungate 1999). Specifically, ecosystem dynamics will be altered as a result of changes such as increased plant

nutrient-use efficiency, altered plant species composition, altered plant resource allocation, and increased plant water-use efficiency (Drake and others 1997; Stitt and Krapp 1999; Polley and others 2000). Productivity, herbivory, nutrient cycling, and decomposition rates will be affected as a result. Previous studies also suggest that the physiological responses of C₃ and C₄ plant species to elevated CO₂ will differ (Bazzaz 1990; Morgan and others 1994; Wand and others 1999; Kimball and others 2002). Increased carbon–nitrogen (C:N) ratios in the

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aboveground plant biomass grown under elevated CO₂ conditions (Chu and others 1996; Ball and Drake 1997; Rogers and others 1999) will likely have a negative feedback on net primary production (NPP) because increased immobilization of N in the soil will limit NPP (Rice and others 1994; O'Neill and Norby 1996; Williams and others 2001; Gill and others 2002). However, the results of Hu and others (2001) suggest that reduced availability of soil N will slow microbial decomposition and increase plant N utilization as plants and microbes compete for N (Kaye and Hart 1997). Similarly, Hungate and others (1997) found that elevated CO2 increases plant N uptake. Although many studies have shown changes in the C:N ratios of green plant biomass and the C:N ratios of litter under elevated CO₂, only a few studies have found evidence that decomposition rates are directly affected by elevated CO₂ (Kemp and others 1994; O'Neill and Norby 1996; Franck and others 1997; Cotrufo and others 1998; Norby and others 2001), and other mechanisms by which elevated CO2 may affect decomposition continue to be explored (Dukes and Hungate 2002). Thus, the effects of rising atmospheric CO2 levels on N cycling and on N distribution in ecosystems are not well understood. Our ability to predict changes in N cycling will require an understanding of the mechanisms by which elevated CO₂ can change N cycling (Hungate 1999; Zak and others 2000).

The shortgrass steppe represents an important grassland ecosystem of the Great Plains of North America. This ecosystem is comparable to regions that account for approximately 11% of global land area (Bailey 1979). The shortgrass steppe occupies about 280,000 km² in the central Great Plains and extends from western Texas to the Colorado-Wyoming border (Lauenroth and Milchunas 1991). Approximately 40% of the shortgrass steppe region remains in native grassland and is economically valuable as grazing land for livestock. Changes in plant tissue quality (for example, N content) can affect both domestic and native herbivores, as well as overall ecosystem productivity. Reduced forage quality can result in lower animal weight gains and lower reproductive success as digestibility decreases (Owensby and others 1996).

We investigated the effects of elevated atmospheric CO₂ on changes in plant N content in short-grass steppe after 3 years of experimental treatment (1997–99). This is the first *in situ* study of the effects of elevated atmospheric CO₂ on plant N concentrations in shortgrass steppe. Based on results of studies in other ecosystems (Campbell and others 2000), we hypothesized that increased aboveground bio-

mass production under conditions of elevated atmospheric CO₂ would decrease N concentration in shortgrass steppe vegetation. In addition, we hypothesized that changes in plant N concentration would be influenced by changes in plant species composition and relative differences among species in changes in plant N concentration due to growth under elevated atmospheric CO₂. Our study addressed the following questions: (a) How does growth under elevated CO₂ affect plant N concentrations at the species level and at the plant community level? (b) Are there interactions between elevated atmospheric CO₂ and simulated herbivory in determining aboveground plant productivity and plant tissue N concentration?

METHODS

Study Site

This study was conducted at the US Department of Agriculture- Agricultural Research Service (USDA-Central Plains Experimental (40°50'31"N, 104°42'50"W) located approximately 60 km northeast of Fort Collins, Colorado, USA, at an elevation of 1650 m a.s.l. The dominant plant species at the site are Bouteloua gracilis (H.B.K.) Lag. (blue grama), a C4 grass; and Pascopyrum smithii (Rydb.) A. Love (western wheatgrass) and Stipa comata Trin and Rupr. (needle-and-thread grass), two species of C₃ grasses. The soil is a Remmit fine sandy loam (Ustollic Camborthids). Meteorological data are recorded by the USDA-ARS station, which is located approximately 200 m from the study site. Mean annual precipitation is 32 cm, of which approximately 70% falls between the months of April and September. Mean seasonal air temperatures range between 0.6°C in winter and 15.6°C in summer.

Experimental Design

Nine experimental plots were established in 1996 to study the effects of elevated atmospheric CO₂ on shortgrass steppe. Baseline measurements were made in 1996. Open-top chambers were placed on plots, and CO₂ treatments were started in 1997. Open-top chambers (4.5-m diameter, 3-m height) constructed of a galvanized steel frame and enclosed with clear polycarbonate sheet were placed on six of the plots. Three chambered plots were maintained at ambient atmospheric CO₂ levels by circulating outside air through the chambers. The other three chambered plots were maintained at elevated CO₂ levels (approximately 720 µmol mol⁻¹) by addition of pure CO₂ to ambient air

passing through the chambers. The chambers were placed on the plots in March and removed in October of every year. The three unchambered plots served as controls for monitoring the effects of the chambers themselves. Further details on the experimental design have been described by Morgan and others (2001).

Sample Collection

The south half of the ground area of each experimental plot was designated for aboveground biomass harvests. A metal wire grid was designed to exactly relocate sampling sites at each harvest. The grid consisted of 56 quadrats, each 40.5×15.3 cm (total area, 3.46 m²). To simulate herbivory, the grid was placed over the south half of each plot in late July, and aboveground vegetation in every other quadrat was clipped to crown level, separated by species, dried at 60°C, and weighed. Species representing a small fraction (less than 5%) of the biomass in the sample area were grouped in general categories by plant growth form (C₃ grass, C₄ grass, forb). This defoliation procedure removed approximately one-half of the green vegetation in one growing season and was equivalent to a moderate to heavy level of grazing. In late October, after the chambers were removed, the grid was placed over the same area in each plot. By October, all plants were past maturity and had senesced. The quadrats that were harvested in July were harvested for regrowth growth after simulated herbivory, and the remaining 28 quadrats were harvested to assess end-of-season phytomass. This pattern was reversed each year, such that quadrats that were not clipped the previous July were clipped in July at peak standing phytomass as well as at the end of the growing season, whereas the alternate quadrats were harvested only at the end of the season in October.

To collect soil cores for belowground biomass measurements, steel cylinders (20.3-cm diameter) were inserted to 60-cm depth in the north half of the ground area of each experimental plot. To minimize disturbance caused by inserting the cylinders, all cylinders for the study were installed before the CO₂ treatments began. Two cylinders containing soil cores were removed from each experimental plot in October of each year to coincide with aboveground biomass sampling. Cores were brought back to the laboratory, where roots and crowns (bases of stems) were separated from the soil, dried at 60°C, and weighed.

Dry plant biomass collected aboveground and belowground was ground through a 0.02-mm sieve (Retsch, Haan, Germany), and representative subsamples were analyzed for total carbon (C) and N using an automated C/N combustion analyzer (PDZ Europa, Cheshire, England). Ash content was determined by combustion at 600°C. Standing crop of N (also reported in the literature as N uptake) was calculated by multiplying N concentration by biomass.

Data Analysis

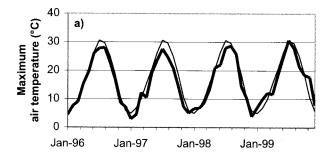
The study was a randomized block design. Data were analyzed using the SAS PROC MIXED procedure (SAS Institute, Cary, NC, USA). We performed two-way repeated-measures analyses of variance (ANOVA), with main effects of CO₂ treatment (unchambered, ambient CO2, and elevated CO2) and year (1997, 1998, and 1999), consistent with Morgan and others (2001). Aboveground biomass was separated by species only in July; therefore, species was also included as a main effect in the analysis of July measurements. Belowground biomass was not separated by species in this study. The main effects were evaluated using Tukey-Kramer post hoc means comparison tests to determine significant differences among means when the main effects were significant ($P \le 0.05$). Analysis of covariance was performed to examine the effect of precipitation, calculated for annual period (January–December) or growing season (April–October) or spring season (March-June), as a covariate.

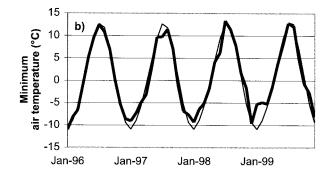
RESULTS

Comparison of monthly averages with the 80-year mean based on measurements since 1912 shows above average precipitation in all years of this study (Figure 1). These measurements also show slightly cooler than normal maximum daily air temperatures during our study, as well as slightly warmer than normal minimum daily temperatures, especially during the winter months.

Aboveground Plant Biomass

Aboveground plant biomass and plant N measurements were made in 1996 before CO_2 treatments began. Data collected in 1996 are presented here as a reference to background conditions. In 1996, total aboveground biomass production was not different between plots designated for ambient CO_2 and elevated- CO_2 chambers. The CO_2 treatments were begun in March 1997 and were applied from March to October of each year. There were highly significant treatment (unchambered, ambient CO_2 , and elevated CO_2) and year effects on July, October, and regrowth biomass (Table 1, Figure 2). In July (Fig-





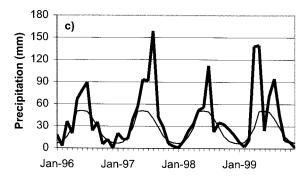


Figure 1. Meteorological data were recorded daily at (a) station located approximately 200 m from the study site. Monthly values are presented for a average daily maximum air temperature, (b) average daily minimum air temperature, and (c) precipitation. The thick solid line represents measurements; the lighter line represents an 80-year average for the site, taken as normal.

ure 2a), during peak standing phytomass, aboveground biomass production was significantly higher (35%) in elevated-CO₂ chambers than ambient-CO₂ chambers, as reported by Morgan and others (2001). The same pattern of increased biomass production under elevated CO₂ was observed in the October harvests (Figure 2b). Aboveground biomass regrowth after simulated herbivory was significantly higher in the elevated-CO₂ plots than in ambient-CO₂ and unchambered plots in all 3 years of CO₂ treatment (Figure 2c). Biomass production varied from year to year, and total aboveground biomass was highest in 1998.

Aboveground Plant Nitrogen Content

There were no differences in N concentration in October and regrowth biomass in 1996, before the CO₂ treatments started. Nitrogen concentration in biomass collected in July and regrowth harvests varied significantly among treatments, and there was a significant effect of year on N concentration of all three plant biomass harvests (Table 1). Nitrogen concentration of aboveground biomass in July was significantly lower (by 21%) in elevated-CO₂ plots compared to ambient-CO₂ plots (Table 2). The percentage decreases in July in aboveground plant tissue N concentration under elevated CO2 compared to ambient CO₂ were 22%, 17%, and 25% in 1997, 1998, and 1999, respectively. In October, N concentration of aboveground biomass did not differ significantly among CO₂ treatments (Table 2), but there was a slight trend in the N concentration consistent with July and regrowth biomass N concentration. The October results were as we expected for senesced plant biomass. Nitrogen concentrations in regrowth biomass in elevated-CO₂ plots were on average 11% lower than in ambient-CO₂ plots (Table 2). Across all CO₂ treatments, overall N concentrations in aboveground plant biomass were lowest in 1999, and N concentration of July and October aboveground biomass decreased in each year of the study.

The total standing crop of N in aboveground biomass in October and regrowth harvests tended to be greater (P = 0.12 and 0.09, respectively; Tukey-Kramer means comparisons) in elevated-CO₂ plots compared to ambient-CO2 plots (Table 2). There was no difference between ambient and elevated CO₂ treatments in total standing crop of N in July (P = 0.9). Aboveground biomass production was higher in elevated-CO2 plots compared to ambient-CO₂ and unchambered plots and more than compensated for lower N concentrations. Overall, the total standing crop of N was highest in July, when plants were green (1.2 \pm 0.1 g N m⁻²), whereas biomass collected in October had senesced and contained less N (0.8 \pm 0.1 g N m⁻²). The total standing crop of N in regrowth biomass was low (0.4 \pm 0.1 g N m^{-2}), reflecting the low availability of N in the latter half of the growing season. There was also a significant effect of year on total N in aboveground biomass (Tables 1 and 2). Total standing crop of N in regrowth biomass decreased over the course of the experiment. These changes in N concentration and standing crop of N in aboveground plant biomass resulted in a steady overall increase in the C:N ratio of plant biomass (average \pm SE) from 38.4 \pm 3.4 in 1997 to 46.0 ± 3.4 in 1998 and 54.4 ± 6.9 in 1999.

Table 1. Results of Repeated Measures Analysis of Variance (ANOVA) Showing Main Effects and Interactions for Biomass, Tissue Nitrogen Concentration, and Standing Crop of Nitrogen for 3 Years of Experimental Treatment (1997–99)

	Biomass (g m ⁻²)		Nitrogen Concentration (%N)			Standing Crop of Nitrogen (g N m ⁻²)			
	July	October	Regrowth ^e	July	October	Regrowth ^e	July	October	Regrowthe
CO ₂ treatment ^d	b	b	с	С	NS	ь	а	ь	b
Year	b	NS	а	С	С	С	b	а	С
CO_2 treatment \times yr	NS	NS	NS	NS	b	NS	NS	NS	NS

 CO_2 , carbon dioxide; $^aP \le 0.05$

From the covariate analyses, there were no statistically significant relationships between precipitation (annual, growing season, or spring season precipitation amounts) and aboveground plant biomass, N concentration, or standing crop of N.

Species Responses

Biomass harvests were separated by species in the July harvest only. We analyzed biomass and N content of the three dominant grass species in our study site, B. gracilis, P. smithii, and S. comata. These three species collectively represent approximately 85% of the total aboveground biomass at the site. The measurements of biomass of each of these individual species are presented in Figure 3.

We observed a significant CO₂ treatment effect on July biomass, as described above. Our data show that species had an important effect on July biomass (P = 0.02) (Table 3). Species biomass varied significantly between years, and differences between years depended on species (Table 3). Biomass production was higher overall in 1998 compared to 1997 and 1999. This difference was driven by higher productivity of both P. smithii and S. comata in 1998. Overall, the productivity of *B. gracilis* decreased slightly over the 3 years of CO₂ treatment. The biomass of *S. comata* was greater under elevated CO₂ compared to ambient CO₂ and was also highly variable between plots.

In general, N concentration of aboveground biomass of individual species decreased with time in all treatments (Table 3 and Figure 4). Differences in N concentration among years were larger than differences among CO2 treatments (unchambered, ambient CO₂, and elevated CO₂). The N concentrations of all three species were lower during the experiment than in 1996, before the CO₂ treatments began. In B. gracilis, the N concentration was 9% lower in the elevated-CO₂ plots compared to ambient-CO₂ plots. The N concentrations of P. smithii were on average 20% lower in elevated-CO2 plots compared to ambient-CO₂ plots; in S. comata, the N concentration was 25% lower in elevated-CO₂ plots compared to ambient- CO_2 plots (P = 0.06).

The effects of CO₂ treatment on total standing crop of N separated by individual species were not statistically significant (P = 0.08) (Table 3 and Figure 5). Trends in the data suggest that the N content of aboveground biomass of *S. comata* increased with elevated CO₂, whereas the N content in B. gracilis decreased. Total standing crop of N was strongly affected by year. The total N in B. gracilis biomass steadily decreased through time and was approximately 50% lower in 1999 compared to 1997. Total N in P. smithii standing biomass was variable between years. Total N in S. comata standing biomass stayed relatively constant through time due to a decrease in N concentration and an increase in biomass production through time.

Belowground Plant Biomass and Nitrogen Content

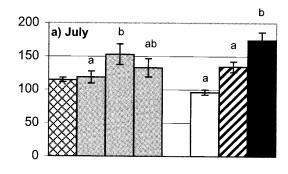
Roots and crowns were sampled from soil cores collected in October of each year, after the plants had senesced. Measurements of root and crown biomass did not show statistically significant effects for treatment (P = 0.3 and 0.1, respectively) or year (P = 0.1 for both roots and crowns) (Figure 6). There was a trend toward higher root and especially crown biomass in elevated-CO2 compared to ambient-CO₂ chambers. There were no differences in standing crop of N in roots or crowns among treat-

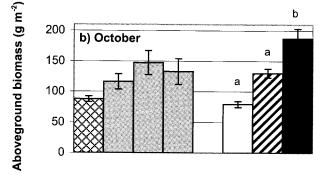
 $^{^{}b}P \leq 0.01$

 $^{^{}c}P \leq 0.001$

^dCO₂ treatment includes unchambered, ambient-CO₂, and elevated-CO₂ plots.

^eRegrowth after simulated herbivory





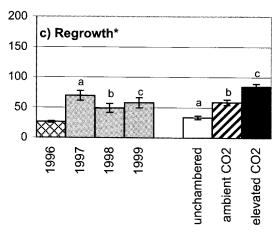


Figure 2. Aboveground biomass shown for main effects of year and carbon dioxide (CO₂) treatment (a) in July, at peak standing crop; (b) in October, at the end of the growing season; and (c) regrowth in October (*after simulated herbivory in July). Measurements in 1996 were made before the experimental treatments began. They are provided here for reference but were not included in the statistical analyses. Error bars represent SE (n = 3). When the main effect was significant, treatment differences were determined at $P \le 0.05$. Bars within each main effect marked with the same letter are not different.

ments (P = 0.4 and 0.8, respectively) or years (P = 0.1 and 0.2, respectively) (Table 4). As expected for senesced plant material, standing crop of N in roots and crowns was higher than in aboveground biomass overall.

DISCUSSION

Aboveground Plant Biomass

Aboveground biomass production was significantly higher under elevated CO2 compared to ambient CO₂ in the first 2 years of CO₂ treatment. Phytomass in elevated-CO₂ plots was 30% higher than in ambient-CO₂ plots in 1997 and 47% higher in 1998 (Morgan and others 2001). In the analysis of variance, year explained a significant amount of the variance in July and regrowth biomass without an interaction with CO2 treatment. Although precipitation did vary from year to year, annual precipitation, growing-season precipitation and spring precipitation were not significant covariates for July, October, or regrowth biomass. Although we expected precipitation to be a primary driver of biomass production in this semiarid environment (Milchunas and others 1994), annual precipitation amounts were above average in all years from 1996 to 1999. Therefore, there may be other environmental factors varying on time scales shorter than a year or growing season that account for the interannual differences in biomass production. In a California grassland elevated-CO2 study, Hungate and others (1997) found that changes in soil moisture under elevated CO2 best explained increased plant N uptake and plant productivity. Simulated herbivory in our study may have interacted with precipitation to determine aboveground biomass production by increasing precipitation-use efficiency, as described by Varnamkhasti and others (1995).

Our results confirm previous laboratory growth chamber studies that showed enhanced productivity of the dominant grass species of shortgrass steppe under elevated-CO2 conditions (Hunt and others 1996; Morgan and others 1998). Other studies have reported similar, but generally smaller, increases in aboveground biomass following exposure to elevated CO₂ (Owensby and others 1993b; Leadley and Stöcklin 1996; Lüscher and Nösberger 1997; Dijkstra and others 1999; Kimball and others 2002). In an open-top chamber study of tall grass prairie in Kansas, Owensby and others (1999) observed differences in aboveground biomass production only in years of below average precipitation. Their results suggest that plants exposed to elevated CO₂ have greater water-use efficiency and improved soil water status, which allows the plants to continue to photosynthesize longer during periods of water stress. Thus, plants exposed to elevated CO₂ are more productive during dry years than plants exposed to ambient CO₂. In our study, measurements of soil moisture indicate that soil water

0.32 (0.04) c

	Nitrogen Conc	entration (%N)		Standing Crop of Nitrogen (g N m ⁻²)			
CO ₂ Treatment or Year	July	October	Regrowth ^e	July	October	$Regrowth^e$	
Unchambered	0.98 (0.04) a	0.68 (0.05) a	0.82 (0.07) a	0.94 (0.05) a	0.54 (0.06) a	0.28 (0.04) a	
Ambient CO ₂	0.94 (0.05) a	0.65 (0.05) a	0.79 (0.05) a	1.24 (0.06) ab	0.85 (0.07) ab	0.47 (0.05) b	
Elevated CO ₂	0.74 (0.04) b	0.60 (0.03) a	0.70 (0.05) b	1.28 (0.09) b	1.12 (0.09) b	0.59 (0.05) b	
1996	1.21 (0.02)	0.76 (0.02)	0.92 (0.02)	1.39 (0.05)	0.68 (0.05)	0.24 (0.01)	
1997	1.04 (0.04) a	0.75 (0.02) a	0.88 (0.02) a	1.21 (0.05) a	0.86 (0.08) ab	0.60 (0.06) a	
1998	0.86 (0.03) b	0.69 (0.03) b	0.88 (0.04) a	1.28 (0.09) a	0.98 (0.11) a	0.41 (0.05) b	

0.55 (0.01) b

0.98 (0.07) b

Table 2. Aboveground Plant Nitrogen in July, October, and Regrowth Biomass (SE in parentheses)

Values in each group followed by different letters indicate significant difference ($P \le 0.05$).

0.49 (0.01) c

0.76 (0.04) c

1999

content increased under elevated CO₂ (Morgan and others 2001), probably as a result of increased water-use efficiency in plants under elevated CO₂. Growth chamber studies of *B. gracilis* suggest that enhanced growth is also a result of a positive photosynthetic response to elevated CO₂ (Hunt and others 1996). In other growth chamber studies, *P. smithii* also showed a positive photosynthetic response to elevated CO₂ (Morgan and others 1998), but *in situ* measurements of photosynthesis in this study showed no direct benefit for either species (LeCain and others 2003).

Aboveground Plant Nitrogen Content

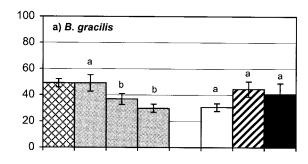
Aboveground plant tissue N concentrations decreased under elevated CO2, although the differences evident in July disappeared in senesced phytomass by October. Decreases in aboveground tissue N concentration under elevated CO₂ in this study are similar to values reported for forest ecosystems (reviewed by McGuire and others 1995) and grasslands (Owensby and others 1993a; Zanetti and others 1997; Wand and others 1999). The results from our open-top chamber study are in agreement with elevated-CO₂ studies that used the Free Air CO₂ Enrichment (FACE) approach (Kimball and others 2002). Insignificant treatment differences in N concentration in senesced biomass (collected in October) compared to green biomass (collected in July) indicate that these perennial plants efficiently reallocated N resources belowground at the end of the growing season. Nitrogen leaching from herbage during plant senescence is not likely to be significant. Precipitation is a primary determinant of N loss through leaching in this ecosystem (Hook and Burke 2000), and precipitation amounts in September and October are generally half of precipitation amounts in May through August. Similarly, the N concentration of senesced plant tissue in a Swiss grassland FACE experiment was not significantly changed under elevated CO₂ (Blum and others 1997). Significant decreases in green aboveground plant tissue N concentration under elevated CO₂ may result from a reduction in the amount of carboxylating enzymes required by the plants (Conroy 1992; Read and Morgan 1996). Growth chamber studies of *B. gracilis*, and *P. smithii* also showed decreases in plant N concentration under elevated CO₂, which were described as a result of greater plant N limitation under elevated CO₂ (Hunt and others 1996; Read and others 1997).

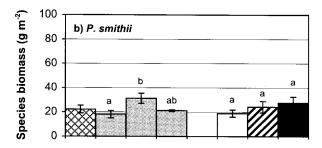
0.67 (0.12) b

Higher biomass production more than compensated for lower N concentration in elevated-CO2 plots, so that total N in aboveground biomass was greater on average in elevated-CO₂ plots than in unchambered and ambient-CO2 plots. The difference in total aboveground plant N may be linked either to increased root biomass production, and therefore greater N acquisition, under elevated CO₂ or to increased N use or uptake efficiency. Increased N mobilization and N mineralization rates, as suggested by Carnol and others (2002), may account for the higher N content in plant biomass under elevated CO2. Increased soil organic matter decomposition under elevated CO2 may also have enhanced N mineralization. A laboratory growth chamber study of B. gracilis showed decreases in overall plant N content but also a significant increase in total root N content under elevated CO₂ (Morgan and others 1994). In an open-top chamber study in Kansas tallgrass prairie, Owensby and others (1993a) observed lower N concentrations in

Data from 1996 included for reference. a, b, c, dCO₂, carbon dioxide

^eRegrowth following simulated herbivory





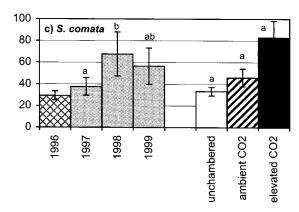


Figure 3. Species biomass of the three dominant grasses, (a) *B. gracilis*, (b) *P. smithii*, and (c) *S. comata*, collected in July of each year. Measurements in 1996 were made before the experimental treatments began. They are provided here for reference but were not included in the statistical analyses. Error bars represent SE (n = 3). When the main effect was significant, treatment differences were determined at $P \le 0.05$. Bars within each main effect marked with the same letter are not different.

plants grown under elevated CO₂ in some years of their study, but total standing crop of N was also higher under elevated CO₂. This result was attributed to increased N acquisition and increased N retention by the plants.

We observed significant differences in aboveground plant N concentration and standing crop of N by year. These differences could not be explained by annual or growing season or spring season precipitation as a covariate. The differences are proba-

Table 3. Results of Repeated Measures Analysis of Variance (ANOVA) showing Main Effects and Interactions for Species Biomass, Tissue Nitrogen Concentration, and Standing Crop of Nitrogen for Three Years of Experimental Treatment (1997–1999).

	Biomass (g m ⁻²)	Nitrogen Concentration (%N)	Standing Crop of Nitrogen (g N m ⁻²)
CO ₂ treatment ^d	b	b	NS
Year	С	С	С
Species	а	С	а
CO ₂ treatment			
× year	NS	NS	а
CO ₂ treatment			
× species	NS	NS	NS
Year × species	С	С	b
CO ₂ treatment			
× year			
× species	NS	NS	NS

CO₂, carbon dioxide

bly driven by a more complex combination of precipitation and other environmental factors. Soilavailable N in the experimental plots did not change detectably over the course of this study (A. R. Mosier unpublished). In addition, N cycling rates, as reflected by N trace gas fluxes, did not change significantly under CO₂ treatment (Mosier and others 2002). However, the amount of N in plant litter is small (1.2 g N m $^{-2}$) compared to the total soil N pool (230 g N m $^{-2}$ in 0–20-cm depth), and it is possible that small changes in soil N that would account for differences in plant N could not be detected. Biomass clipped from the experimental plots in all harvests was not returned to the plots in this study. Although plant biomass had senesced at the time of October harvests, removal of all aboveground biomass represents relatively severe grazing or disturbance. The harvest technique could explain some of the year effects observed in the analysis of variance. The net removal of biomass (and therefore N) from the system may have contributed to the decreases in total N in plant biomass over the years.

Defoliation Effects

Recovery from simulated herbivory (defoliation) was significantly higher under elevated CO₂ com-

 $^{^{}a}P \leq 0.05$

 $[^]bP \le 0.01$

 $^{^{}c}P \leq 0.001$

 $^{^{1}}$ = 0.001 d CO₂ treatment includes unchambered, ambient CO₂, and elevated CO₂ plots

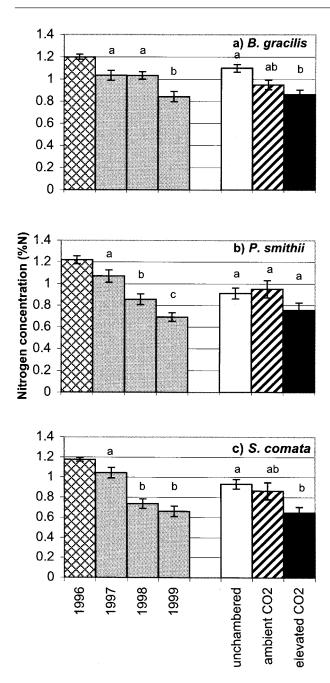


Figure 4. Aboveground plant biomass nitrogen concentration of three dominant grasses, (a) *B. gracilis*, (b) *P. smithii*, and (c) *S. comata*, collected in July of each year. Data are presented as in Figure 3.

pared to ambient CO_2 . Aboveground biomass production after simulated herbivory was higher in elevated- CO_2 plots compared to ambient- CO_2 plots by 26%, 47%, and 65% in 1997, 1998, and 1999, respectively. At the same time, the N concentration of aboveground regrowth after simulated herbivory was only slightly lower in elevated- CO_2 plots compared to ambient- CO_2 or unchambered plots, sug-

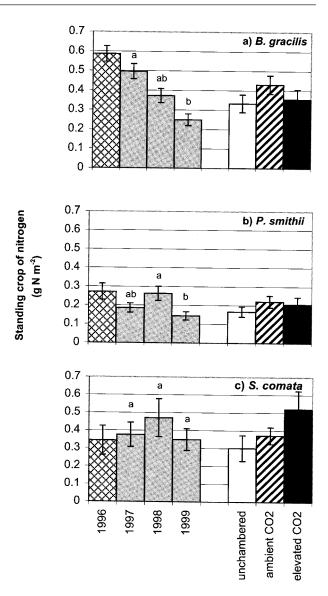


Figure 5. Total standing crop of nitrogen (N) in aboveground biomass of three dominant grasses, (a) *B. gracilis*, (b) *P. smithii*, and (c) *S. comata*, collected in July of each year. Data are presented as in Figure 3.

gesting that N mineralization or remobilization from belowground organs may have been greater in elevated-CO2 plots. Past studies of long-term grazing of shortgrass steppe found that increased soil water availability had negative effects on grass N concentration, but the effects depended on grazing intensity (Milchunas and others 1995). Aboveground productivity in historically heavily grazed systems was not significantly affected by simulated herbivory or water supplementation treatments (Varnamkhasti and others 1995), but plant N concentrations tended to be lower under these treatments (Milchunas and others 1995).

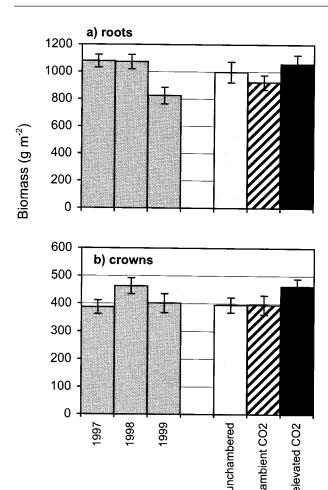


Figure 6. Biomass of **(a)** roots and **(b)** crowns collected from soil cores taken from 0-60-cm depth. Error bars represent SE (n = 3).

Table 4. Belowground Standing Crop of Nitrogen in October

SE in parens

	Standing Crop of Nitrogen (g N m ⁻²)				
CO ₂ Treatment or Year	Roots	Crowns			
Unchambered	11.02 (0.90)	4.65 (0.46)			
Ambient CO ₂	9.77 (0.67)	4.54 (0.45)			
Elevated CO ₂	11.12 (0.70)	4.96 (0.21)			
1997	11.63 (0.64)	4.44 (0.40)			
1998	11.14 (0.83)	5.40 (0.39)			
1999	9.15 (0.60)	4.32 (0.27)			

These results suggest that plants subjected to herbivory may respond positively to the direct effects of elevated CO_2 rather than the indirect effects of

improved soil water status or other CO₂-induced factors. Greenhouse studies have shown that regrowth after simulated herbivory is increased under elevated CO₂ for *P. smithii* but decreased for *B. gracilis* (Skinner and others 1999). In that study, remobilization of total nonstructural carbohydrates and soluble N from root and crown tissues after simulated herbivory was lower under elevated CO₂ in *B. gracilis* but higher under elevated CO₂ in *P. smithii*. The differences in the contribution of remobilized N for shoot growth after simulated herbivory between *B. gracilis* and *P. smithii* may account for some of the decrease in biomass of *B. gracilis* relative to *P. smithii*.

Plant Nitrogen Allocation and Plant Tissue Quality

Changes in aboveground plant N concentration may be driven by changes in the production of different structural tissues (lignin, cellulose). Laboratory analyses of B. gracilis and P. smithii indicate higher total nonstructural carbohydrate concentrations in response to elevated CO2 (Read and Morgan 1996; Morgan and others 1998). Increased production of nonstructural carbohydrates would tend to decrease plant N concentration (Conroy 1992; LeCain and Morgan 1998; Stitt and Krapp 1999). Studies of CO₂ effects on forage chemical composition have documented changes in lignin content (Fritschi and others 1999). Because lignin has a relatively high C:N, increased lignin could account for some of the differences in C:N between CO₂ treatments. The increase in tissue lignin concentrations that we measured in October in elevated-CO₂ plots (D. G. Milchunas and others unpublished) is similar to results reported in some studies (Cotrufo and others 1994); however, in other studies, such changes or decreases in lignin concentration with elevated CO₂ were observed (Chu and others 1996; Ball and Drake 1997). Reduced tissue N concentrations and tissue quality could have significant impacts on microbial decomposition rates, as well as herbivore nutrition. Although aboveground biomass production may increase under elevated-CO₂ conditions, grazing ruminants in this system need to increase intake rates to meet their N demands.

Species Responses

We observed a significant effect of year on species biomass and a significant year-by-species interaction. The data show an overall increase in *S. comata* and a decrease in *B. gracilis* over time. Biomass production of the three dominant grasses was significant production of the three dominant grasses was significant effect of year on species biomass and a significant year-by-species interaction.

nificantly affected by CO_2 treatments (unchambered, ambient CO₂, and elevated CO₂). Although differences between ambient and elevated CO2 could not be detected within individual species, the trend in the data suggests increasing productivity of S. comata under elevated CO₂. The three dominant grasses differed in N concentration at peak growing season, perhaps reflecting differences in N acquisition or N-use efficiency. Variation in plant structural composition, such as lower ratios of leaves to stems, which are generally lower in N concentration, in S. comata, may also account for differences in N concentration. Divergent species responses to elevated CO₂ have important implications for the shortgrass steppe. Changes in the relative abundance of the three dominant grass species in this ecosystem, as shown by data after 5 years of elevated-CO₂ treatment (Morgan and others in press), suggest that species composition may change under long-term elevated CO₂. A shift from a community dominated by the C₄ shortgrass B. gracilis to a community dominated by the C₃ midgrass S. comata would after ecosystem N-cycling dynamics.

The data on total standing crop of N by species reflect such a shift in species composition. Both N concentration and biomass production of *B. gracilis* decreased between 1997 and 1999, which is reflected in the downward trend in total N content in *B. gracilis*. On the other hand, the N concentration of *S. comata* also decreased significantly between 1997 and 1999; however, increased biomass production of *S. comata* (Figure 3) compensated for the N concentration decrease, so that the total standing crop of N in *S. comata* remained unchanged. In this way, *S. comata* outpaced *B. gracilis* in the acquisition of ecosystem N resources, regardless of CO₂ treatment.

Our results are similar to those from a Swiss grassland study in which significant interspecific differences in response to elevated CO2 were observed, whereas to intraspecific differences in response to elevated CO2 were detected (Lüscher and Nösberger 1997). Leadley and Stöcklin (1996) also reported important species-specific responses of calcareous grassland species to elevated CO2 that would be expected to cause changes in community structure under increasing atmospheric CO2 concentrations. Important species effects have also been observed in many other studies (Garbutt and others 1990; Diaz and others 1993; Owensby and others 1993a, 1993b; Chu and others 1996; Hungate and others 1996; Grünzweig and Körner 2001; Joel and others 2001; Reich and others 2001). Berntson and others (1998) suggested that plants' abilities to take up N may drive changes in community composition under elevated CO₂. The variation in responses of individual species to elevated CO₂ in terms of multiple characteristics—such as leaf area, leaf biomass, plant phenology, biomass of reproductive parts, and seed N content—complicates estimates of ecosystem responses to elevated CO₂. Predicting ecosystem responses to elevated CO₂ will require a better understanding of many of these responses at the species level.

Belowground Plant Biomass and Nitrogen Content

Although aboveground plant biomass production increased significantly under elevated CO2, similar responses were not observed in belowground biomass. Many studies have reported increases in root growth and root-to-shoot ratios under elevated CO₂ (Rogers and others 1999), but relatively few of those studies focused on natural ecosystems or grasslands in particular. One study in annual grasslands showed a slight trend of lower root-to-shoot ratios with increasing atmospheric CO2 (Berntson and others 1998). We did not calculate root-toshoot ratios because the aboveground and belowground biomass data reported here were collected from separate areas. However, our measurements of overall increases in aboveground biomass and no change in belowground biomass suggest lowered root-to-shoot ratios under elevated CO₂.

The wide range of root-to-shoot ratio responses to elevated CO₂ in the literature suggests that analysis of root-to-shoot ratio alone may be an inadequate means of describing the effects of elevated CO₂ on plants (Norby 1994). Changes in root-toshoot ratios under elevated CO2 could depend strongly on other conditions, including nutrient and water availability (for example, see Sindhφj and others 2000). Results from the tallgrass prairie study conducted in Kansas suggest that changes in root biomass were related to changes in species composition (Owensby and others 1993b). In our study, shifts in plant species composition, combined with long average root turnover times of 5-7 years (Milchunas and Lauenroth 1992, 2001), might explain why changes in root biomass are too small to be detected. In addition, Kimball and others (2002) indicate that open-top chambers may have a negative effect on root growth when compared to the methods used in FACE studies. Similar to root biomass, the biomass of plant crowns did not change significantly under elevated CO₂ compared to ambient CO2. These data suggest that greater aboveground biomass production under elevated CO₂ resulted primarily from the increased leaf area

of individual plants rather than an increase in the number of plants.

Changes in N concentration in aboveground biomass may be the result of changes in plant allocation of N between aboveground and belowground tissues (Gorissen and Cotrufo 1999); however, root and crown N contents were not significantly altered under elevated CO₂ compared to ambient CO₂. At the end of the growing season, we expect that senesced plants have reallocated N to their roots and crowns. The increase in aboveground productivity under elevated CO₂ may mask any impacts on root and crown standing crop of N despite changes in aboveground plant N concentration (Table 2). Shifts in species composition may also make it difficult to detect changes in root and crown N in response to elevated CO₂. Important species differences in response to elevated CO2 have been reported for root biomass and N content. Curtis and others (1990) found that whereas a C3 species showed increased root biomass and decreased N concentration in root biomass, a C4 species in a similar environment showed no root growth or N content response to elevated CO_2 .

CONCLUSIONS

Nitrogen concentrations of aboveground plant biomass decreased under elevated CO2, but increased aboveground biomass production under elevated CO₂ resulted in higher total standing crop of N in elevated-CO₂ plots. Aboveground biomass production after simulated herbivory was also greater under elevated-CO₂ compared to ambient-CO₂ conditions. At the same time, elevated CO₂ did not result in significant changes in belowground biomass production or belowground plant N content after 3 years of CO2 treatment. Despite obvious aboveground plant responses to elevated CO₂, our results indicate that overall changes in ecosystem N cycling in response to elevated CO2 will occur slowly and will be difficult to detect in grasslands, where the majority of plant biomass is located belowground and turns over on 5- to 7-year time scales. Differences in aboveground tissue N concentration between species and changes in the proportion of total biomass represented by each species under elevated CO₂ suggest that ecosystem N cycling has been altered as a result of community composition changes under elevated CO2. These indirect effects of elevated CO2 on ecosystem N cycling through longterm changes in community composition may be more important than the direct effects of elevated CO2 on biomass production and plant nutrient content.

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