#### UNITED STATES DEPARTMENT OF AGRICULTURE

#### FOOD SAFETY AND INSPECTION SERVICE

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# ADVANCES IN POST-HARVEST INTERVENTIONS TO REDUCE SALMONELLA IN POULTRY

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February 23, 2006 9:00 a.m.

The Loudermilk Center Atlanta, Georgia

FACILITATOR: DR. DANIEL ENGELJOHN

Deputy Assistant Administrator, Office of Policy, Program and Employee Development, Food Safety and Inspection Service

#### PARTICIPANTS:

- DR. SEAN ALTEKRUSE
- DR. PATRICIA BENNETT
- MR. DANE BERNARD
- DR. STAN BAILEY
- DR. MARK BERRANG
- DR. JEFF BUHR
- DR. KEN BYRD
- DR. JOHN CASON
- DR. PATRICIA CURTIS
- DR. MARTY EWING
- DR. RANDY HUFFMAN
- DR. LAURA HULSEY
- MR. LOREN LANGE
- DR. BARBARA MASTERS
- MR. DAVID McNEAL
- DR. JULIE NORTHCUTT

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## PARTICIPANTS: (CONT.)

- DR. ROBERT O'CONNOR
- DR. KEN PETERSEN
- DR. RICHARD RAYMOND
- Dr. John Rice
- DR. RICHARD ROOP
- DR. SCOTT RUSSELL
- MR. MICHAEL RYBOTT
- DR. BRUCE STEWART-BROWN
- DR. ROBERT W. WILLS

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## I-N-D-E-X

AGENDA ITEM	PAGE
Opening Remarks:	7
Daniel Engeljohn, PhD, MS, Deputy Assistant Administrator, Office of Policy, Program and Employee Development, Food Safety and Inspection Service, USDA	ı
Richard A. Raymond, MD, Under Secretary, Office of Food Safety, USDA	10
Barbara J. Masters, DVM, Administrator, Food Safety and Inspection Service, USDA	28
FSIS Overview of the CY05 Broiler and Ground Poultry Salmonella Data:	38
Loren Lange, MS, Deputy Assistant Administrator, Office of Public Health Science, Food Safety and Inspection Service, USDA	i
Proposed FSIS Salmonella Verification Categories for Broilers:	48
Sean Altekruse, DVM, MPH, PhD, DACVPM, Deputy Executive Associate, Office of Policy, Program a Employee Development, Food Safety and Inspection Service, USDA	
Application of Systematic Review Methodology to Salmonella Intervention Strategies in Broiler Production and Processing:  Robert W. Wills, DVM, PhD, DACVPM,	60
Mississippi State University	
Panel Q&A Session:	76

# **NEAL R. GROSS**

Controls from FSIS Perspective:	85
Laura Hulsey, DVM, Technical Assistance & Correla Staff, Technical Service Center, Food Safety and Inspection Service, USDA	
Common Findings During Comprehensive Food Safety Assessments in Poultry Establishments:	96
Kenneth Petersen, DVM, MPH, DACVPM, Assistant Administrator, Office of Field Operations, Food Safety and Inspection Service, USDA	
PANEL Q&A Session:	113
Overview of Antemortem Controls and Establishment Sanitation:  Salmonella and Campylobacter in Broiler	_
Transport Cages:	136
Mark Berrang, PhD, MS, Research Microbiolog Agricultural Research Service, USDA	ist,
Effect of Bacterial Load on Chickens Entering the Processing Plant on Final Carcass	ng
Contamination:	151
Stan Bailey, PhD, MS, Microbiologist, Agricultural Research Service, USDA	
Overview of Sanitizers Currently in Use with Emphasis on Pre-operational Sanitation to	<u>h</u>
Ensure That Pathogens Are Not Surviving Cleaning and Sanitizing:	164
Scott Russell, PhD, MS, University of George	าล
· · · · · · · · · · · · · · · · · · ·	

## **NEAL R. GROSS**

Assurance Manager, Sanderson Farms

	•
Processing and Sanitation Issues Unique to Very Small Establishments:	191
Patricia Curtis, PhD, MS, Auburn University	
Salmonella Interventions Unique to Turkey Processing Establishments:	204
Michael Rybolt, Doctoral Candidate, Manager, Scientific and Technical Affairs, National Turkey Federation	
PANEL Q&A SESSION:	211
Overview of Slaughter Dressing Processing Control	<u>.s</u> :
Limits on the Effectiveness of Antimicrobial Treatments:	<u>-</u> 222
John Cason, PhD, Animal Physiologist Scienti Agricultural Research Service, USDA	.st,
Scalding, Defeathering and Rehang as Primary Sources for Redistributing Salmonella Typica With No Antimicrobial Intervention and the Featherless Broilers:	_
Jeff Buhr, PhD, Animal Physiologist Scientis Agricultural Research Service, USDA	st,
<pre>Mechanics of Poultry Processing (First Processing):</pre>	253
David McNeal, MS, Product Manager, Meyn Amer	rica
Reprocessing of Fecal Contaminated Carcasses And the Use of Antimicrobials:	<u>5</u> 267

# Agricultural Research Service, USDA

Stan Bailey, PhD, MS, Microbiologist,

Impact	of	Chilling	on	Poultry	Carcass	
Microbi	iolo	ogy:				285

Julie Northcutt, PhD, MS, Food Technologist, Agricultural Research Service, USDA

Managing pH for the Maximum Antimicrobial

Effectiveness of Chlorine in Processing
Water:

299

Ken Byrd, DVM, Director of Regulatory Affairs
and Plant Applications, Mionix Corporation

Interventions at Further Carcass Processing
(Parts) to Control Salmonella, Including
Grinding, and How the Choice of Packaging
Material Impacts the Prevalence of Salmonella
and Cross-Contamination:

311

Scott Russell, PhD, MS, University of Georgia

Panel Q&A Session: 328

Closing Remarks/Adjournment:

Daniel Engeljohn

#### P-R-O-C-E-E-D-I-N-G-S

(9:00 a.m.)

DR. ENGELJOHN: Good morning. This is the second in a series of meetings that we're having on the control of *Salmonella* in raw products, with this particular focus on poultry, and broilers specifically today.

Just a few housekeeping issues before we get started. We are net-casting the presentation today so that individuals who are not able to attend can at least hear the presentations and see the PowerPoint presentations. And then our intention is to make available copies by CD and other formats to anyone who would like a copy. We're going to make copies available to all the establishments that we regulate, but, certainly, we'll make the information available to anyone else who requests it.

But as we get started today -- we do have 28 speakers over the course of the day-and-a-half that you're here, and that's intentional. We have a lot of information we want to ensure that you have access to, and so they're rather short presentations, and we're

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going to hold the speakers to their time limits.

But for other issues related to those of you here in the room, the restrooms are outside the door to the left. There is no refreshment here in this building. So if you go out the building to the right to the United Way building just directly opposite of this building, there are food and beverage facilities there. And then if you hang a left out of this building and go back up Auburn, there are a number of food court choices up the street there.

With that, I think we'll get started.

Today we will have opportunity for questions and answers from the audience.

We are transcribing the meeting so that all the information from this meeting is available to the public. And so I do ask that if you have a question, you queue up in front of this microphone in the center of the room at the appropriate time when we have questions and answers available and that you give your name and the association that you represent so that we can get that into the official record. And then we'll make every attempt to answer your questions

today. And if not, we will find another format to be able to answer questions that are raised.

(Pause.)

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DR. ENGELJOHN: They're not able to hear us on the phone lines. So we're hoping that we can get that corrected and you'll let us know if you hear differently. And if you can't hear us, be sure to let us know, as well.

I do want to get us started. Dr. Richard Raymond, the Under Secretary of Agriculture for Food Safety, will be our first presenter.

Dr. Raymond was appointed as the under secretary on July 18, 2005, and he's responsible for overseeing the policies and programs of the Food Safety and Inspection Service. And he chairs the U. Code Codex Steering Committee, which provides quidance to U. S. delegations to the Codex Alimentarius Commission. He has extensive experience in developing and implementing policies and programs designed to improve public health.

Prior to joining USDA, Dr. Raymond served as the director of the Nebraska Department of Health

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1	and Human Services Regulation and Licensure Division,
2	where he oversaw regulatory programs involving
3	healthcare environmental issues. He also developed
4	several anti-bioterrorism initiatives and a statewide
5	healthcare alert system. Dr. Raymond also played a
6	major role in the development of local health
7	districts that serve Nebraska's 93 counties.
8	Please welcome Dr. Raymond.
9	(Applause.)
10	DR. RAYMOND: Thanks, Dan.
11	Before I start, let me just speak into
12	this microphone and see if the telephones pick it up,
13	just in case that solves our problem.
14	(Pause.)
15	DR. RAYMOND: We're doing a mic check for
16	the telephones.
17	(Pause.)
18	DR. RAYMOND: Maybe they all left.
19	(Pause.)
20	DR. RAYMOND: We can't hear them? Okay.
21	They can't hear us, and we can't hear them.
22	Well, good morning, everybody. And thank

I	
1	you for coming to this very important meeting to
2	discuss advances in post-harvest reduction of
3	Salmonella
4	(Pause.)
5	DR. RAYMOND: They're hearing this now?
6	Okay. Great.
7	in poultry. You don't know how good it
8	feels to come to a group which for the most part has
9	had nothing to do with shipping hotel rack veal to
10	Japan.
11	(Laughter.)
12	DR. RAYMOND: It's nice to have a
13	different venue and talk about Salmonella for a couple
14	of days. And then we'll go back and talk about veal,
15	I'm sure.
16	(Laughter.)
17	DR. RAYMOND: As most of you know, I hail
18	from Nebraska, as do several other people that have
19	followed the Secretary to the USDA for the second term
20	of the Bush administration. And some of us were back
21	home a few weeks ago and gave a little talk.
22	We were on a panel the three of us.

And when we opened up questions and answers, the reporter said, What would you like people to say as they walk by your casket about your time in public service? And the first individual said, I would want them to say I worked long, hard hours and was totally committed and dedicated. The second person would say, I would want them to say, "He was an honest man."

And it was my turn, and I said, I feel I must apologize a little bit; if I had a more time to think about this than the Secretary and the other gentleman, well, what I would want them to say is, I think he's still breathing.

(Laughter.)

So after the last month of DR. RAYMOND: dealing with hotel racks, we are still breathing, and it's time to move on to another subject. So today and tomorrow, we're going to dedicate our time t.o discussing new research and new insights and then learn from practical experience on how we can reduce this prevalence of Salmonella post-harvest. And I also hope you do some networking and exchange best practices, which I know the industry is looking at

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very seriously, in helping each other out to control this problem.

Everybody needs to understand that there's going to be changes in how the Office of Food Safety and the Food Safety and Inspection Service approach this important issue of reducing Salmonella.

You probably heard in 1996, when the rules were published, that it was the goal of FSIS to lower Salmonella rates. I don't think that ever happened. You're hearing it again, but it's a new world, we've got a new Administrator, a new Deputy Administrator, a new Under Secretary and a new Secretary, and we believe strongly that this is going to happen this time. And we're going to tell you how, and we're going to tell you why.

So just accept that change is coming. And be prepared to work with us, not against us, and we can all make this happen together. Your participation particular today and tomorrow but, also, in the future and in the past is very critical to us at FSIS to develop these best practices to combat Salmonella.

We, the Office of Food Safety and FSIS,

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have been at the forefront of the battle to reduce the prevalence of foodborne pathogens in meat, poultry and products, and we've that through egg done communication, cooperation and collaboration with the industry, with consumers and with scientists. trying to be open and transparent as we develop these new policies, and Salmonella will be no exception to trend that we have set under Dr. Masters' leadership the last year-and-a-half or so.

As you have heard me say before -- those of you who have heard me talk -- this is not new news. My first day on the job, Secretary Johanns told me this should be one of my top priorities: To get our arms around Salmonella and lower those rates and protect the public. And believe me, it is one of my top priorities, and we will get this done.

We want to explain these slides to you in just a little bit of detail. My job today is to explain to you kind of the groundwork of where we're going.

We have been surveying and doing stats in over 100 large plants since 1998, seven years' of

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experience of doing Salmonella stats. These slides -the 20 percent line there is the performance standards
that have been established for carcasses. The middle
line is one-half of the performance standard, twice as
good as you have to be to pass the performance
standard.

What this slide shows you is in Category 1, we have 25 percent of our plants that have always had their Salmonella stats come in below one-half of the performance standard, six or fewer positives. We know this can be done. Those plants show us that it can be done.

The second category, the majority, 45 percent of the plants, have never exceeded the performance standards in those seven years. At times they've been below the 10 percent, at times, they've been above the 10 percent, but they've never exceeded the performance standards.

And then we have 30 percent of our plants, who at times are above the performance standard and at times are below the performance standard. What this graph doesn't really truly represent is sometimes

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they're way below the performance standard; sometimes they're at 5 percent. So the squiggles should be bigger, but that -- I apologize for that.

The point is almost two-thirds to three-fourths of our plants performed better than the performance standard in the last seven years. So to the ones that say, "We can't do this; we can't get the Salmonella rates down," I ask you to meet with those plants on those bottom two squiggly curves. I think they'll give you perhaps a different viewpoint.

I know that if I go to a restaurant tonight and eat a chicken breast, I don't know which plant it came from, and I don't know what the performance sets were. Now, the consumers want to They want to know who's up there in that know that. top 30 percent, and they want to know who's down there in that bottom 25 percent. And we haven't done that. But it's one of those little carrots and sticks that you're going to hear about that we may entertain if we can't get some movement within the industry to get the 30 percent coming on down so they more closely mirror the ones with the good performance sets.

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Now, this is different than the plants. This is the actual sets. I mentioned some of those plants that are in the more variable categories will occasionally have a set that gets the 5 percent or lower.

And in actuality, in those same seven years, in all of the sets that we performed, the majority, over 50 percent, were at lower than one-half the performance standard. More than half the time, it was down to below 10 percent. We need to find out what happened the other 49 percent of the time, when they fell into Category 2 or Category 3, and it's not consistent amongst the plants.

We had one plant that had a 30 percent rate on the performance set. We did a food safety analysis, and we worked with that plant to point out areas that they should and could improve. In the next set we went in, they had a 2 percent. Now, that is dramatic. What we need to do is move the trends that direction in all of our plants that are in the poultry business. That's our goal. I firmly believe that we can get that done.

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I know that there are going to be some nay-sayers out there. I've heard them. They say it just can't be done. But you know what? We've got consumers out there that say, You're not doing enough, and you're not doing it fast enough. So this isn't a plan that's going to please everybody. We have listened to the consumers and we have listened to the industry; we've listened to your representatives, we've been in the small plants, and we've listened to the scientists. And what we have tried to do is come up with a plan that has a little bit in it for everybody to get everybody on board.

The worst thing that could happen is to have someone lay down on the tracks and say, We're not going to go with this; we're going to stop it on the Hill; we're going to stop it in the Secretary's office, and you know what; we're going to keep doing business like we're doing. And that just isn't good enough.

So we have tried to accommodate. We've tried to compromise. We've tried to bring enough people to the table that we can sell this and we can

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make it work. And again, we'll give you the details later today on how it is going to work.

I had one plant owner -- honest to God -- who told me in a public meeting that it was the consumers' fault because they didn't cook their chicken and they didn't cook their turkey to the right temperature and it was our fault because we didn't educate the consumers. That's a plant owner.

Now, I recognize that is the minority, not the majority, but there's still some outreach that needs to be done, because those people can get very vocal. But I think most or the people we have talked to have the same goal, and that is to reduce the Salmonella loads. And I think, together, we can get this done.

Back in 1996 to 1998, I'm sure the beef industry said the same thing that some people in the poultry industry may be saying: It can't be done; we can't get *E. Coli* rates down to less than one per one-hundred thousand; it is impossible; it's inherent; it's the nature of the beast; it's a part of the plant. But they had two things happen to them that

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made them roll up their sleeves and work together and work with us, and they got it done because of the Jack-in-the-Box and because *E. coli* 0157 was declared an adulterant.

Now, Salmonella, as you know, has not been declared an adulterant. And, thank God, we haven't had a Jack-in-the-Box scenario -- or Chick-Fil-A -- or something like that. We don't want that to happen. But if it does, believe me, the pressure will be intense on the industry. We would rather do it now because it's the right thing to do.

We don't have a Jack-in-the-Box scenario for Salmonella that I'm aware of, but we do have 14.5 people out of every hundred-thousand Americans get sick with culture-proven Salmonella every That's 42,000 people a year. The CDC estimates it's actually 1.3 million people a year that get sick with Salmonella; they just don't sick enough to get a stool culture. And 400 people die with Salmonella. It's just that we're used to it because those numbers have been there a long time. We don't have a Jack-in-the-Box.

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I talked to a reporter yesterday who said,

I don't think this will work; I'm not hearing

anything -- I'm not hearing bells and whistles, and

nothing's happening. And I said, It's because we

haven't had a Jack-in-the-Box, but the industry is

going to make this happen; stay with us and watch our

progress. And I hope she -- hopefully, she'll write a

nice article about us.

These are positive regulatory samples for *E. coli* over the last five years. The same -- this is human illness in *E. coli* over the last seven or eight years. This last one right here is 0.9. Healthy people in 2010, the goal for the year 2010, is 1.0 persons per hundred-thousand. We reached that goal in *E. coli* in 2004, six years ahead of schedule.

When it was up here at 2.5 and 2.4, that's when the beef industry was saying, It just can't be done; we can't get down to that; we've set an unrealistic goal for us. But they did it, and I hope we can all take notice of that. And I hope we can all be motivated by what they were able to do.

Listeria? The same example, without the

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Jack-in-the-Box. But their sample products have gone down, and the human infection rates have gone down -with Listeria. Now, we've got a few risk-based initiatives we took on a couple of years ago when that number popped up there that are going to keep the Listeria numbers going down. And that's all good news the people that consume these ready-to-eat We need to work on the chicken and turkey products. products just a little bit, however.

We do believe that those experiences, again, lend proof that this can be done. When the industry works together with the scientists, with the Agency and with consumers and shares best practices, we know we can get this done. Now, we have a long way to go, though.

The healthy people in 2010 goal is 6.8 infections per 100,000 people. We're at 14.5 people. E. coli was at 2; their goal was 1, and they made it. You're at 14.5, and the goal is 6.8. You can make it. You can make it. It's going to take awhile, but we've got six years to get to the healthy people for 2010.

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What you're going to hear speaker after speaker after speaker is the ways we have learned, things that we have learned that are working and the research that has been done. And you're going to hear about our new rollout that we put on the web two days ago that will be posted in the <u>Federal Register</u>, I believe, on the 26th. You're going to see our new stats for 2005.

2005 stats coming out in February? That's unheard of. That's how this Agency looks at things now days. We aren't going to wait a year to release data; we're releasing them right now because they'll show if we've got a problem.

And you'll hear how we're going to release set data in a different fashion to not wait a year to address issues. We've got the issues to address now. I'll probably be gone in three years or less. I want to see change while I'm here; I don't want to go back home and say, I tried, but we just didn't get anything done. So we want you to work with us.

In closing, I do want to say we do have a strong system in place. I'm not saying the sky is

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falling. We have a great system, but we can always make it better. Any system that doesn't move forward is a system moving backwards. You can't tread water and try to maintain it. It just doesn't happen. The bugs get smarter, and the bugs get more resistant. And we need to work for that.

That's something that -- public health is always changing. If we don't change as the bugs change, we lose ground. A hundred years ago, the life expectancy was 45 years when you were born in America. This year, it's 75 years. That's 30 years we've gained in life expectancy, and that's not because of medical science; for the most part, it's public health.

1906, one out of five coffins was filled with the body of a child that never reached his birthday; they died of her fifth infectious In the '40s, we invented penicillin -- I diseases. shouldn't invented it -discovered say we we penicillin. Bacteria could now be attacked. Pneumonia didn't kill little babies, and meningitis didn't kill young children.

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In 1955, the polio vaccine was licensed. Kids no longer died from polio or were crippled for the rest of their lives. They were not neurologically impaired. Their parents could let them go swimming in the swimming pools without worrying about polio. None of us in this room, I don't believe, had children before penicillin was discovered; there may be one or two that had a child born before the polio vaccine was discovered, and I won't ask you to raise your hand.

But, you know, we don't know what it was like for parents a hundred years ago to have children, realizing that children in your community died on a regular basis; it was just an expected event. They died of diarrhea, dysentery, enteritis, diphtheria, smallpox and polio, things that we can now prevent.

Nobody would have thought 100 years ago that we would have vaccines that would wipe out childhood diseases. No one would have thought we would have bullets that would kill bacteria. But we've done those things. And so when you say, "I don't think we can do better with Salmonella," suck it up. We're going to do better with Salmonella.

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We don't know what that magic bullet is going to be, but one of the reasons kids now live past their fifth birthdays and the life expectancy has gone up is because -- we have those scientific inventions, but we also have safer food, we have safer water, and we have sewage treatment and disposal.

But now days, when a child dies of foodborne illness, it's а disaster. It's а catastrophe. It was expected 100 years ago that it would happen. Right now, it's a problem with product or the handling of the product in the home or the some place along the road. restaurant or and we must do better, because it is truly something that is totally preventable.

We all have the same goal. And if we just remember who we're working for here and if we just remember who we're trying to protect, I'm pretty sure we can get there together.

So once again, I thank you for attending this conference; I know you're dedicated and committed or you wouldn't be here. We look forward to a healthy exchange over the next two days. Thank you very much.

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DR. ENGELJOHN: Now, I just want you to know -- just so you don't think that I'm really lax in my job -- I'm giving an exemption to Dr. Raymond and Dr. Masters for how long their speeches take. So I'm not holding them to their time lines, but, all you other speakers, you other 26 speakers, I'm going to hold you to it.

DR. RAYMOND: You didn't let me start until ten after, Dan.

(Laughter.)

DR. ENGELJOHN: Well, we're going to move on. And thank you very much, Dr. Raymond, for those remarks.

Our next speaker is Dr. Barbara Masters. Dr. Masters was named the Administrator of the Food Safety and Inspection Service in August 2005. In this position, she is responsible for leading FSIS in its mission of protecting public health through food safety and [inaudible due to failure of in-house PA system].

Dr. Masters began her FSIS career as a

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veterinary medical officer in 1989 near Hot Springs, Arkansas, and has since held a variety of posts field throughout the Agency, both in the and headquarters. Since March of 2004, Dr. Masters served as the Acting Administrator. And during that time, she raised the scientific training investment in the 10,000-employee work force to a record \$20 million, as well as enhanced communications with both internal and external audiences.

Please welcome Dr. Masters.

(Applause.)

DR. MASTERS: Thank you, Dan.

Good morning, all. I'm certainly pleased to be here today to participate in this important meeting.

I certainly want to thank our FSIS Office of Policy, Program and Employee Development for hosting this meeting. It's no small challenge to put on a meeting of this magnitude, and it adds to the challenge when they have to put it together in netcast. So I also want to thank those folks that are working to do the net-cast portion of this meeting.

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I think it adds to the value when we're able to not only bring those of you that are interested in being here but we're able to reach many more of our stake holders that have an interest in this topic when we're able to do the net-cast. And we're also able to save it.

I talked to many of you after the preharvest portion that said, There was just a few of
those presentations I would have loved to have shared
with many of my folks back home in the plant. So
we're hoping that we're able to add value to this
meeting by having the net-cast portion and also being
able to put it on the CDs for you to take back with
you so that you can show portions of this to your
folks back at home.

So we do believe we're reaching many more of our stake holders by having this format. So thanks to those of you that worked very hard to get this meeting put together.

As you're aware, as an Agency, we've been working on our farm-to-table approach to food safety.

While most of our regulatory authority lies in the

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plants, we realize that food safety begins long before the product reaches the processing or slaughter establishments.

We've had so much information to share on the topic of Salmonella. That's why we realized last year we needed to approach the Salmonella topic with two meetings. That's why we got together last year in August and had our pre-harvest meeting in Athens.

And we had such a long and fruitful discussion on Salmonella and the trends and the research relative to Salmonella and the discussion that we had on the pre-harvest topic. We discussed controlling Salmonella to the maximum extent practical and the impact that that control has at pre-harvest and on the levels of Salmonella coming into the plant.

We're hopeful that at this meeting, we can talk about the latest data and have discussion at the in-plant level for combating Salmonella, because we recognize most of you will use a combination of that information for the solutions for combating Salmonella. We want you to know that we have not forgotten all the information that was shared at the

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pre-harvest meeting and we're very soon going to be issuing some compliance guidelines that will come out of that pre-harvest meeting.

discussed in Athens, there we are things that can be done prior to that product reaching the plant, but we also believe there are many things that can be done at the in-plant level. Again, we recognize most of you will do a combination of things pre-harvest and at the in-plant level, but the bottom is we need to make sure that we're doing the Salmonella everything practical to control It's up to you as an industry holistically to take on that challenge.

As Dr. Raymond mentioned, we are aware that individual plants have been and can control Salmonella and have been meeting the performance standards. We know this because we have data that represents that plants have been consistently meeting the performance standards.

In my opening remarks in Athens, I talked about the fact that we would be using the model that we used for  $E.\ coli\ 0157:H7$  in the beef industry. Dr.

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Raymond talked about it at a high level in his presentation, and I want to get a little bit more into detail, when we say we're using that model, what we really mean by that.

We're using the E. coli model in the sense that what we did for the beef industry is that -- we conducted a risk assessment for E. coli Based on that risk assessment, we had all of these beef establishments re-assess their own HACCP plan. After they had re-assessed their HACCP plans, we went out as an Agency and conducted food safety assessments of those re-assessed HACCP plans. The results that we found have been reductions in the positives in our regulatory samplings, as well as reductions in foodborne illness.

I think the significant point we need to understand is that it was -- the industry taking on the challenge to re-assess their food safety programs is what we believe has really made the difference in looking at the reductions in our positive regulatory samples, as well as the reductions in foodborne illness.

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When we went out and verified for our food safety assessments the changes in the programs, the industry had taken on that challenge of re-assessing their programs, and we saw significant differences in those plants. That was the crux. The industry-wide initiative of re-assessing their programs is when we saw drastic changes industry wide. We believe that the poultry industry can see similar changes if they apply a comparable model.

The challenge that I have to you as the poultry industry is to use the data that you're going to gain at this meeting as well as the industry information that we're going to share with you in the form of compliance guidelines from the pre-harvest meeting to significantly start decreasing the prevalence of Salmonella in your plants. You do not need to wait on us as an Agency to require you to reassess your HACCP program; you can re-assess your HACCP program any time you desire.

You're going to have general information coming to you at this meeting relative to best practices, relative to lessons learned from ongoing

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food safety assessments that we've done; you're going to have information coming to you from the literature reviews that we've done. You're going to have specific plant information that we're going to be providing to you.

You do not need to wait on this Agency to conduct a food safety assessment. You can take on the challenge to look at your own food safety systems and make the necessary changes in the design of your food safety programs. And we believe we're going to be providing you the type of information to make the necessary changes, based on the information at the pre-harvest meeting and again at this meeting, to make the changes in your program to control Salmonella in your establishment.

Please don't misunderstand me. We do understand that there are different ways to control Salmonella. We understand that plants are using a variety of ways to control Salmonella and that there are a lot of different ideas and different approaches to controlling Salmonella.

Dan mentioned there's well over 20

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speakers at this meeting; that's because we recognize there's no one-size-fits-all. There's not one magic way to say, Ta-da, we're going to control Salmonella in the plant this way. We want you to look at what works best in your plant environment and apply that in your plant environment; the challenge to you is to listen to the speakers, look at your own plant environment, re-assess your program and figure out what works best in your plant environment.

Don't wait on us to come into your facility and say, Why didn't you take advantage of the information. You don't need to wait on a food safety assessment for that to occur. Again, you have options, and we encourage you to take advantage of those.

We're going to be very transparent with data. You're going to hear from our speakers. Again, we're going to share lessons learned from previous food safety assessments. Learn from those lessons learned.

You're going to hear consistently from Dr. Petersen about plants that failed to control their own

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processes. We're going to share with you, again, compliance guidelines from the pre-harvest meeting. We've done an extensive literature review, and we're going to share that with you.

My favorite I -- so many people have heard me talk about an AVMA meeting where we -- there was a great literature review: "Ain't Nothing Good Ever Happens at Picking." Yet, when I walk into many of your poultry operations, I see picking fingers that haven't been changed in months. Take advantage of this information and apply it.

We're going to be sharing with you your own Salmonella data on an individual -- result by result. You're going to hear that from us. If you're getting those results back and not using them to your own advantage, then you're not going to be making the necessary changes in your own operation. And again, that's the kind of information we're expecting you to use and apply, and you can determine what the best controls are in your own operation.

Again, the CDs will be available from this meeting. Transcripts will be available from this

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meeting. And so we're just imploring you to take the advantage and re-assess your own programs to make the necessary changes. And you need not wait for a food safety assessment for that to occur.

Improvements are expected. As Dr. Raymond indicated, changes will occur. We do expect to see significant improvements in Salmonella. We do believe the poultry industry is up for it. We've started to see some changes; unfortunately, we're not seeing Salmonella numbers go down. You'll see that when the Salmonella results are posted. That's not what we want to see. We want to see the prevalence go down.

And we do believe you're up for that challenge. You're here, and that's a good sign. You've been meeting with us as an Agency, and we do recognize that you're starting to work together as groups. And so we do believe you're up for the challenge. We want to work with you. We want to make information available to you. So we know that you're up for the challenge, but it is up to you to make that difference.

So again, thank you for being here. And

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1 we look forward to watching those numbers come down over the next year. Thank you very much. 2 (Applause.) 3 DR. ENGELJOHN: Thank you, Dr. Masters. 4 Our next speaker is Mr. Loren Lange; he's 5 the Deputy Assistant Administrator for the Office of 6 7 Public Health Science. Loren came with the Agency back in 1979 8 and has held several leadership positions with us, and 9 10 he had also worked at FDA prior to that. He has his degrees in mathematics from Iowa State University and 11 a master's degree in applied mathematics from Johns 12 13 Hopkins University. Loren's going to talk to you about the 14 On our web page, you have access to all of 15 2005 data. 16 data up through 2004, including the serotype Loren's going to give us information 17 information. about broilers and ground products for 2005. 18 19 Loren? 20 MR. LANGE: Thank you, Dan. Good morning. It's a pleasure to be here 21 to speak to you about -- oh. 22

1	I've got to clip it on here? Okay.
2	(Pause.)
3	MR. LANGE: Right here? Does that work?
4	THE REPORTER: Yes.
5	MR. LANGE: Our speakers have mentioned
6	that we've got a new Administrator and new Deputy
7	Administrator. And Barbara knows that
8	Barbara, you were a little over a year
9	ago, I got a new boss.
10	And you've got the Under Secretary and the
11	Secretary and the Deputy Secretary.
12	And I was out for dinner one night, and
13	someone said, Well, how are things going at work. And
14	I said, Well, there's two constants: President Bush
15	and me. So at least there's some constant at work.
16	(Laughter.)
17	MR. LANGE: So I'm glad to be here this
18	morning. I'm going to present a brief summary of our
19	2005 results from testing poultry products for the
20	three products you know, broilers, raw ground
21	chicken and raw ground turkey that are covered by
22	existing performance standards.

The results of all seven product categories that we test for are being posted on our web site today; they should be up there by noon, by my understanding. And if you haven't heard or seen it, I'm just going to mention that last Friday, we put up or serotype data for all the seven years, 1998 through 2004, and we'll soon be updating that to include our 2005 data.

Now, before I present the results, I do want to just point out a little bit about the nature of the data that we post on our web site. either positive or negative results from verification testing; the results from are not statistically designed baseline studies. Thus, estimates of results not national product are prevalence. However, we do consider that the data do give us an indication of the trends.

The data I will present are summaries of individual samples from what FSIS calls A sets. These are the sets that are routinely scheduled to verify compliance in establishments that are either new or past their previous verification step. The data do

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not include the follow-up steps to verify corrective actions. Thus, I will be presenting the data that are exactly the same that we've been posting on the web site since this program was initiated in 1998.

This presentation will cover two topics.

I'll first summarize what we found in 2005, and then I have a few slides showing that the results from 2005 didn't follow what we have seen as some historical patterns.

First, the summary of the 2005 poultry This slide shows seven years -- no. results. I'm results This slide shows the 2005 for sorry. In the A sets, the percentage was 16.3 broilers. This is the third year that we have seen the percent. percentage of positive samples go up in broilers.

This next slide shows the broilers by year of the percentage of passing sets. And you see -- one sort of observance here is that as the first couple of years that the percentage of positive samples was going up, there really wasn't a huge change in the percentage of the sets that were passing.

In fact, we see that actually from 2003 to

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2004, the percentage of sets that passed actually went up a little bit. But when we got into last year, when the percentage got as high as 16.3 percent over the year, we obviously had a lot more higher percentage of sets that were failing. And it dropped 9 percent, from 90.3 down to 81.3 percent, of sets that were passing.

This next slide shows our ground chicken results per year. The percentage was up a little bit in 2005 from 2004 -- well, actually, it was up 32 percent. But notice the numbers of samples are very small here. So we really don't consider the ground chicken data to be merely as good an indicator of trends certainly as the carcass data are, but we do put it up on the web, and I am presenting it here for completeness.

The next slide shows ground turkey by year. Again, ground turkey went up from 2004 to 2005, up from 19.9 to 23.2 percent. Now, with ground turkey, we do get, you know, approximately a thousand samples every year. And as you can see, generally, we've been averaging around a thousand samples. So we

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consider this a little better indicator of trends.

We sampled ground turkey in, I would say, roughly half of the major turkey slaughter operations.

So we're getting -- our information indicates at least half produced this ground product, and we have been sampling in about half the plants.

Now, the next slide's where I'm going to point out what I said my second topic is. We have results showing how 2005 did not exactly follow historical patterns by month and by quarter.

This slide shows the broiler data over seven years. Just to make sure everybody's clear, what it's showing is that like -- April/May/June, it shows that broilers have averaged 10 percent. Well, that's the April/May/June data from 1998, 1999 and all the way up to 2004 summed up, the percentage of positive samples over seven years.

We see this trend that the low has been April/May/June over those seven years, 10.2 percent, and the high has been the fourth quarter, October/November/December, 14.6. Now, relative to the absolute levels, 10 percent and 14 percent, that

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difference of 4.4 percent, you know, we would say is considerable. So there certainly has been this historical difference between the months.

We went back and looked at just the last three years, and we have seen this is, you know, about -- I'm sorry -- the three years, 2002 through 2004. And there we saw that, again, April/May/June was 10.4 percent; October/November/December, 15.3 percent. So we saw this same pattern: Low in the second quarter; high in the fourth quarter.

This is the 2005 data by quarter. It certainly looks different.

Here the second quarter was actually 19.7 percent, the highest, where it had always been the And the fourth quarter wasn't the lowest. lowest. The third quarter was a little low, but the fourth quarter was down to 14.5 percent, clearly a different Something changed, and we hope this -- it's pattern. in certainly, you know, the last change two quarters -- is a change in the right direction. And we hope this is the beginning of a new trend.

Next I have a couple slides showing the

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same thing for the broilers by month. And we can see
that the lows were always April this is the seven-
year data. Excuse me. The lows of 9.8 percent were
April/May. And we had three months,
September/October/November, that over seven years were
always above, you know, 15.3 percent. You know,
there's tons of data. I mean every time I see this, I
think, You don't find real-world data that follows
such a nice S curve. I just you know, for someone
that has spent his whole life looking at data and
trying to analyze data, I'm always amazed by you
don't see curves like this in real-life data. Very
nice.
But here's 2005. Again what that's not
a nice curve oh.
I've got two minutes? Okay.
Nice curve, not a nice curve. But guess
what. Those low months where the highest in fact,
May was 21.7 percent. And we were down November
was the lowest at 13 percent.
I've got two minutes.

Ground turkey? This goes back to the

1	quarters we saw the second quarter. Just in
2	contrast to broilers, ground turkey was highest in the
3	second quarter, where broilers had been lowest. But
4	this is 2002 to 2004, three years of data, and it
5	changed. Something had happened. The last three
6	quarters were all about the same. And this is what
7	happened last year.
8	We saw it increasing by quarter. Now,
9	there's not a lot of samples in each quarter, so we
10	don't know if this is really trend. But we did see
11	the fourth quarter higher than it had been, you know,
12	over the 2002 to 2004. There may be some concern
13	there.
14	Thank you. That's the end of my
15	presentation. I made my time limit.
16	Dr.RAYMOND: Just one more thing.
17	[inaudible due to failure of in-house PA system]?
18	MR. LANGE: No, not here. Dan gave me the
19	two minutes.
20	DR. RAYMOND: For those who don't know
21	Loren well I owe him one, because he got me once.
22	But this is the first time I've ever heard Loren walk

1	away without saying, Oh, just one more thing.
2	MR. LANGE: I do have one more thing while
3	I'm walking back
4	DR. ENGELJOHN: That's enough, Loren.
5	Thank you, though, very much.
6	(Laughter and applause.)
7	DR. ENGELJOHN: And I do want to move on.
8	All of this information is available to you. So just
9	so you know, it will be available.
10	Our next speaker is Dr. Sean Altekruse.
11	Sean is our Deputy Executive Associate in the Office
12	of Policy, and he's responsible primarily for
13	coordinating the statistical and technical support for
14	our policy development. He's also a captain in the U.
15	S. Public Health Service and has his veterinary degree
16	from the University of Georgia, a master's in public
17	health from the University of South Carolina and a PhD
18	from the Virginia/Maryland Regional College of
19	Veterinary Medicine.
20	I do want to preface Capt. Altekruse's
21	presentation with a note that, as Dr. Raymond
22	mentioned earlier, on Tuesday, we did post a <u>Federal</u>

Register document that was submitted to the office of the Federal Register for publication. And it will publish in the Federal Register on Monday, the 27th, but it is available to you. It may have some slight formatting changes from what the published is, but it is available to you -- which will clearly articulate what Dr. Altekruse is actually going to present to you now.

### So, Dr. Altekruse?

DR. ALTEKRUSE: Good morning. So what I'd like to talk about today is the Salmonella categories that are described in the document that has been posted to the web, which -- our Under Secretary, Dr. Raymond, described the three categories -- and also, the type of data that we're going to be sharing, which our Administrator, Dr. Masters, mentioned, specifically, the serotype information.

So specifically, the Salmonella categories are -- the current casts the sets into two groups.

One is those that are less than 50 percent of the standard, and the other is that they're above 50 percent of the standard without failing the standard.

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And then the third category is the existing category of exceeding the standard. And I'd like to talk about why we think these three categories make a lot of sense.

So just briefly, we have looked at data for a variety of product classes. And today, I'm going to be presenting information specific to large broiler establishments, but the same patterns are seen for small broiler establishments and for other product classes, as well. And also, this presentation is specific to data through 2004, although we have looked at data through 2005 and the same patterns continue to occur.

So establishments are tested about once a year. One broiler rinse is collected per day, and there are 51 rinses per set. This should be fairly familiar. And then those rinses have results of:

Less than 50 percent of the standard, which would be six or fewer positive tests per set; greater than 50 percent, which would be seven to 12 positive tests per set, and; exceeding the standard as having 13 or more Salmonella-positive tests in a set.

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So these these categories really make some sense. Category I shows consistent *Salmonella* control is possible. Category II suggests to us that these plants are doing the right things but, with a little bit of perseverance, can do even better. And Category III is what it always has been, which is failing to meet the standard.

And why have we selected these categories?

The reasons are really pretty straight forward.

First of all, Category I is the normal scenario.

There are -- and Category II accounts for about 25 percent of sets. And Category III is really -- it's an outlier in terms of what we're seeing over the historical time. Less than 10 percent of sets are in Category III.

And furthermore, if we look at Categories ΙI and III, that's where the majority of the Salmonella-positive tests are. And, even important, it's also where the serotypes that are most commonly associated with human illness are most likely to occur.

So it's really a testimony to the hard

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work of the great FSIS work force and what they've accomplished. Over the last seven years, they have completed Salmonella A tests in about 135 large broiler establishments, so we have -- about once a year. So 762 completed sets. And from that ongoing prospective sampling and verification program, we have serotype information, phage type information and pulsed field data on isolates.

So this brings up some questions. What does this historical information tell us? First of all, the vast majority of establishments are in Category -- of sets are in Category I.

This schematic shows the distribution. So a huge number of sets are in Category I. And then Category II is about a quarter of all sets. And Category III is way out there. And in a few sets, we've seen as many 30 isolates per set.

Now, this slide is intended to provide a little definition about common serotypes. The Centers for Disease Control publishes the list of the 20 most common Salmonella serotypes in people each year. And when we look at that list, we see some of the

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serotypes which also occur in poultry rinses that we collect, particularly Heidelberg, Typhimurium, and Enteritidis, those top three.

Now, each one of these serotypes has its own little reservoir. So with Typhimurium, we see it in a lot of product classes; it's not unique to poultry by any means. Heidelberg is -- it does occur in some other classes, but primarily in poultry. And Enteritidis is -- it's really the exception to find it in other product classes, although never say, Never. So -- but really, these serotypes do have their own unique reservoirs.

Now, what can we -- what knowledge can we gain from our A sets about these common serotypes? First of all, this first question, "Is the percent of the common serotypes the same in each category," is an important question because if the driver for being in Categories II and III is a serotype that usually doesn't commonly cause human illness, we should know about that.

Secondly, are these common human illness serotypes more likely to be found in one of the

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categories or another? Third, is any category resident for the Lion's share of the total serotypes?

And the last question is, How many isolates can you normally expect to see in an A set of these serotypes?

So let's go through those questions systematically.

Do these serotypes account for the same percent of Salmonella? Well, you'd be very surprised if the percentage was absolutely identical across all classes. However, it's very close. It's within 5 percent. And when you look at the confidence intervals around the point estimate, they all capture the overall average, which is 48 percent, and they all capture 50 percent, as well.

So in other words, just shy of 50 percent of all Salmonella that we get from broiler rinses are in these common human serotypes, and there's no statistical difference between the two groups.

Now, the second question was, Are any of these categories more likely to be positive, to have sets that are positive, for *Salmonella*? And this would apply -- the table shows, "Was there a human serotype in the set," with the answer yes or no. And

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you can see for Category I about a third of all sets tested negative for human serotypes. In contrast, in Categories II and III, it was about 5 percent of all sets that tested negative for human serotypes.

And so if you look at the odds ratios for those associations, what you see is that there's a very robust statistical association between Categories II and III and the likelihood of testing positive for a human serotype.

The third question was, Is there category that accounts for the lion's share of these And the answer to that is also serotypes? Categories II and III, although -- remember it was 24and 8 percent respectively of all sets that fell into those two categories. So they -together they account for 32 percent of sets. They account for 63 percent of common human serotypes.

And I think an important additional point is that Category II, the middle category, which is sort of the new category that we're describing here, has 737 positives for the common human serotypes. So that category contains the most common human

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serotypes.

And then the last question on that introductory slide was, How many isolates of common human serotypes should you expect within a given A set? And the answer is very few. Twenty-five -- this slide shows the percent of all sets and then the isolates of common human serotypes with -- per set. And you can see that 25 percent of A sets had no human common serotypes in them.

And then if you go to, "1," of the common human serotypes, the next increment on the X axis, that's 49 percent of all sets that had one or fewer human serotypes. So I had to say that there were two or fewer -- most sets had two or fewer, because you have to add in two or fewer to get to 64 percent.

And then going on out the scale, "3," is 75 percent. "8," common human serotypes within an A set is something that you would not expect to see in 95 percent of sets. "12," is -- you would not expect to see that in 99 percent of sets. And you have to go to 30 common human serotypes per set to reach the maximum.

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So in summary, most sets have no or very few common serotypes of human illness in them. And Categories II and III, although they account for only one-third of all A sets, accounted for two-thirds of the common human serotypes. So we really think that those two categories, II and III, are an important place to focus.

Now, I've talked about sets, but I'd like to talk about something that Dr. Raymond mentioned, as well, which is the performance of individual establishments over time. We have data on more than 100 large plants over seven years, and a quarter of those establishments could demonstrate consistent control of Salmonella throughout the entire period of follow-up.

Most sets had no or -- wait. So this is looking at the large establishments with five or more sets through 2004. We see that a quarter of the establishments could control *Salmonella*, as defined by consistently being in Category I. And the others were in Category II or III. And most often they were in Category II or III at least twice, but there was a

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gradient of performance. Some just barely -- some only were in Category II one time out of the entire follow-up period. And some failed as many as three sets.

So this graphic sort of depicts that distribution over time, and the biggest category is the middle category. The way I would interpret this is that this is a group of plants that is sort of on the line and, with perseverance, they can move in the direction of the establishments depicted in green, which have lower than 50 percent. But if just allowed to drift, there's also the potential that they could begin to have sets that they fail. And we want to encourage the first and discourage the second.

I'd like to also focus specifically on the plants that never exceeded half the standard, because I think this is an important group of plants. These plants, after five, six or seven tests, have never failed a plant. And I think that this isn't accident. This occurs because of leadership within the establishments: The executives, the quality employees, control management, the plant the

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testing and the validation and infrastructure they're using that control Salmonella. And so I would suggest that these plants are the leaders and that they are an important industry to helping resource for the meet challenge.

So in summary, what we see in looking at the data over time is that there is a tendency for patterns to emerge, that plants can demonstrate the very best control or an intermediate level of control variability in their or have some control of And I showed this slide earlier, but I'll Salmonella. show it again, because maybe it means a little more after my presentation.

Category I indicates consistent Salmonella control. Category II suggests that they're moving in the right direction, but some improvement is possible.

And category III continues to be failing to meet the standard.

Really, this data is the product of seven years of hard work by the FSIS work force, and I'd like to acknowledge just a few of the people who have

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1 helped to make this presentation possible. Thank you. 2 (Applause.) Thank you, Dr. Altekruse. DR. ENGELJOHN: 3 4 Well, I hope that gave you a bit of perspective as to why we have a need to talk today and 5 to continue the dialogue. But to pull all this 6 7 together, we have Dr. Robert Wills from Mississippi State University, who has done some extraordinary work 8 on pulling together the literature to define, 9 10 interventions are available, and how effective are 11 they. We think this information is absolutely 12 13 critical for you to hear. We're delighted that this through one of 14 work was being done our sister 15 agencies' grant programs; the CSREES within USDA 16 helped fund part of this work, and we think it's just And we know that there's other work 17 outstanding. related to Campylobacter that's going to be underway 18 19 soon. Dr. Wills since 2001 has been an associate 20 professor of veterinary epidemiology in the Department 21

of Pathobiology and Population Medicine at Mississippi

1 State University. He previously was an assistant 2 professor in the department of veterinary biomedical sciences at the University of Nebraska at 3 4 Lincoln, in Lincoln, Nebraska. He received his doctor of philosophy from 5 Iowa State University, a college of veterinary 6 7 medicine degree in Ames, Iowa, and a doctor of of veterinary medicine from the University 8 Missouri/Columbia College of Veterinary Medicine in 9 10 Columbia, Missouri. Dr. Wills, we're delighted to have you 11 here today. 12 13 DR. WILLS: Well, thank you. I was a little worried. 14 I thought maybe we'd catch up pretty quick here when we couldn't find 15 16 my presentation, but we did find it. I want to talk about a method that was new 17 to me; about a year-and-a-half ago, I guess, I've 18 19 known about it. I think it has great potential to 20 help us to fully utilize the literature that's available -- scientific literature. And I think it's 21

a good tool that we can apply to figuring out what the

best intervention strategies are controlling Salmonella.

I want to take the moment to acknowledge the people that I'm working with on this project: Hart Bailey, who's here; Kris Clements, who's with Mississippi State, as well, and that's helping us with this. I also want to acknowledge Jan Sargeant at McMaster University in Canada, who's really a leader in developing this methodology for use in food safety.

I think the premise that most people agree with is that the production of safe food involves intervention strategies to be implemented at all stages of production, from farm to fork. We've heard about that already today. In order to do that, decision makers need information: What strategies, and where to apply them.

I think the scientific literature is a good resource for this information, but there are some problems I see with using the literature -- some stumbling blocks. One is sometimes a lack of literature: It's difficult to find information on a particular subject; it may be out thee, but it's not

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readily accessible.

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And the other is kind of the flip-side of that, that you have a large body of information and, in fact, so much information that you have an overwhelming quantity of it. And it's difficult to synthesize that and summarize it, and, even when you do, you find conflicting conclusions.

So systematic reviews are a way to help us deal with this scientific literature and come up with some answers from it. It's a method of identifying effective treatments or processes based the available evidence from a variety of sources. Ιt differs from traditional narrative or critical reviews of literature. And I'll go into a little more detail about that in a moment. But it gives us a transparent and replicable scientific methodology to collect, assess and synthesize all available information on a subject.

And one of the key components of this is that it could be reproducible in that in the final paper or document you come up with with the systematic review -- within that, the methods should be outlined

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sufficiently that a person could take that through the same systematic review and, hopefully, come up with the same conclusion.

Now, the reason I became interested in systematic reviews was as an outgrowth of the Food Safety Research and Response Network, which is USDA funded, a sub-project of the Food Safety CAP. And we have a sub-project in that looking at Salmonella and preventions, and Jan Sargeant, as I mentioned, is leading this effort. Annette O'Conner and Jim McKean at Iowa State University are conducting the systematic review of Salmonella interventions in pork or -- in pigs. And then Hart and I are working on one in -- looking at Salmonella interventions in broilers.

I wanted to put up this slide, and you may not be able to read it. But it's "A Guide Conducting Systematic Reviews in Agri-Food Public Health", and it's kind of а how-to manual conducting systematic reviews on food safety topics, and it was produced by Jan Sargeant. And I wanted to put it up here because it's a good resource and I took a lot of the information for this presentation from

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it. I want to acknowledge that.

So in looking more at our rationale and significance of systematic reviews, how they've been used and why, they're very commonly used in human medicine fields and used in evidence-based medicine.

And they're used primarily to identify effective interventions to reduce disease burden.

But they may also be used to identify knowledge gaps that target additional research, and that's one reason I think there's great promise in systematic reviews. Even if you don't come up with the final answer, one of the byproducts of it is that you find out the gaps that you need to know in order to find those answers. So I think that's useful.

It can also identify methodological strengths and weaknesses in the available research. We can find out why we don't have the answers. Perhaps it's because the research wasn't conducted appropriately. And that will help us identify, once again, where we need to go with more research. And it can also encourage best study practices development for intervention research. We can find out how to do

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the research to find the answers.

So Jan Sargeant put this slide together, and it shows that there's a lot of systematic reviews used in health science, lots of those. If you narrow it down and look at systematic reviews for public health, there are still quite a few, but it's a reduced number. And then when you get to looking at systematic reviews in microbial food safety, there are really very few of those available.

Now, this isn't an exhaustive list here, but I wanted to make a point of it because it's a recently published systematic review for development or -- for control of Campylobacter in broilers. It focuses primarily on contributing factors and sources of Campylobacter in Great Britain, but I thought it might be of interest. And it demonstrates that these methodologies are being used in the field of food safety.

I mentioned I make a comparison of systematic versus traditional reviews. And generally, systematic reviews have a more focused study question; it's defined, and it is a foundation of the review.

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Systematic reviews also have a more explicit and comprehensive search strategy; the articles are selected according to uniformly applied and specified criteria.

quality of articles is formally, rigorously and consistently assessed. And а quantitative summary is made if possible; it's not a requirement for a systematic review, although the true goal, if you could, would be to do a Met analysis of multiple studies for your final product And if that's -- if the studies systematic review. aren't there that allow that, the systematic review still be successfully done and done can productivity.

I wanted to give you a schematic here of the systematic review process, just to get a general idea of it. And then I'll go through it in a little more detail. We start out with the study question, and I've put this in the center of the schematic because I think the whole systematic review really revolves around the question that's being asked. And it has to be a well defined question.

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Then the next step is to do a literature search and try all the information that can be found, all the primary research that can be found, for that study question. I have a step process of relevance screening so that you find the articles that are relevant to that study question. Once they pass that, they go through a process of quality assessment. If they make it through that, the data is extracted from those articles, that information is synthesized or summarized, and then the final product is a written report.

So start off here with the development of a focused study question, and the question needs to be clearly defined *a priori*. And that's, I think, a critical part of this.

You don't do the review and then decide what you wanted to find out from it; you start with the question in mind at the very beginning. And that question will include components on the population that's being studied, what intervention is being looked at, what outcome is being assessed, and it specifies the system

level or sector of agriculture to be reviewed.

And this -- the development of this study question is accomplished by forming a panel of content experts. And they decide how to write this question so that the subject of interest is captured within that question.

The next step is a literature search. want to generate a complete list of all primary research that is relevant to the question. And the search terms, once again, are based those components that make up the study question: Population, intervention, outcome agriculture and sector.

And we're looking at published literature. To do this, we use an exhaustive list of search terms in multiple electronic databases. You'd also need to hand-search journals that -- if you can identify relevant journals that we feel may have the papers on this topic, but they are not listed in any of the electronic journals, we'd still want to go through those and hand-search them.

Also -- in addition to the published

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literature, gray literature is also searched. Now, gray literature is completed-but-unpublished research. This can be found through scanning the internet, electronic and hand searching of conference proceedings and contacting researchers, national and international experts in the field directly.

Well, once we have this huge amount of information -- and this -- we may end up with several thousand -- it's quite likely we'll end up with several thousand abstracts after our search -- we then go through a process of screening these for relevance. We want to determine if an article has potential to answer the study question through this screening process.

A priori criteria for subject relevance and inclusion of material into the review is established, and these are a series of questions that have to be answered based on information in the abstract to determine if the abstract is relevant to answering the study question.

And another point here that's unique, I think, with systematic reviews as compared to

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traditional reviews is that at least two reviewers independently assess each article or abstract to determine whether or not it's relevant. If they have conflicts, if one excludes it and the other one includes it, then they, those two reviewers, have to meet and resolve those conflicts to decide which way the abstract should go.

I've put up here a shot of the screen on the software program we're using in our systematic review. I just wanted to point out that we have a series of questions on the right-hand side that we can check off. And the information we have available to make these decisions are the authors, the title, and then an abstract if available. Sometimes abstracts don't come through, and we have to do a little more searching to track those down.

In our particular one systematic review, we're categorizing the information, thinking that we will probably come up with several systematic reviews on intervention strategies in broilers, but our first screening process is to divide it into broilers and layers and then different segments of the poultry

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continuum and then into different intervention strategies.

But once we've had an article successfully pass the screening process and at least two reviewers have deemed it relevant, then we find the full article and do a second, stricter level of screening using full copies of the articles. There's a standardized procedure developed for each type of study design, and that's used to evaluate each article. And once again, two reviewers use that standardized protocol to review that particular article.

And just a list here of the items that are used in this assessment: A look at the study objectives, the population that's being looked at, intervention strategy -- make sure it's appropriate -- a look at the outcome assessment, how withdrawals or loss of data was handled, and, also, how the data was analyzed, and then the conclusions, if they were appropriately made or not.

Different types of study designs provide different levels of evidence, and I have a list here.

I think I'll skip over it other than that within a

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systematic review, different types of designs have different values or levels of evidence, and you can restrict it to a systematic review to a particular level if you have sufficient articles; if you don't, you may have to look at everything, and that will make it more difficult to compare the data.

So once we've had an abstract to go through the screening process and we get the full article and we've screened it or assessed its quality, the next step is data extraction, where that paper that has made it all the way through is read and we take the different components of it and put those in the data repository so that the can be summarized and analyzed as a whole.

And once again, we'll have a form that we use to fill out. It includes information: Descriptive data of the article in context, study characteristics -- what design was used, how many animals, and those sorts of things -- and then study results.

Then this information is synthesized or summarized to take the results from multiple primary

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studies that have met the quality assessments and put them together and analyze them using either qualitative methods or quantitative methods when possible.

The data is presented in this step so that similarities and differences between the studies and the level of evidence can be visualized, and the results are presented to show if intervention consistent and effective. And that's -- the final bottom line that we'd like to achieve is to make a determination of how well that intervention works and, possible, how well it worked in different situations and different conditions.

Now, one of the challenges that we have in doing this is just managing all of the data and the information. You start out with a very large volume of abstracts -- several thousand. We started out with one pass that had over 13,000 and dropped it to around 2,500 as our final starting point. But that's a lot of abstracts to keep track of. And if you have to have paper copies of all those, it really becomes a challenge.

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And then once you move those through the multiple stages of valuation, it becomes even more difficult. And if you have reviewers -- multiple reviewers potentially from different places, then it becomes even more of a challenge.

There's some software available -- the one we're using is a web-based program -- that can handle Data's loaded into -- the literature's loaded It contains the standardized into the program. protocols, so you can have check-lists through it. It keeps track of what reviewers have it automatically tracks the articles, it keeps track of what the reviewers have said about those articles and identifies discrepancies between them and keeps track of all the information all the way through data extraction.

I have a few things here. I've mentioned the systematic reviews that have been done in pork and the one we're working in broilers. And there's also one being done on *E. coli* 0157 in cattle at McMaster and also one looking at the association between Johnes and Crohn's disease in Canada.

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Just a few slides here on the strategy or -- on the systematic reviews we're using in intervention strategies in broilers. I mentioned we're taking an approach where we're looking at pre-harvest/post-harvest, looking at broad field -- a lot of different interventions, categorizing those so that we can either look at them at that level or focus in on a fewer number of interventions and do a complete review of those.

And these are just the screening process slides that we have here, where we're dividing up into broilers, layers -- we do keep track if a turkey article -- if it refers to turkeys as well as broilers or layers. Then we divide it into the thicker segment of a production continuum. Then we have a list of intervention strategies that we're keeping track of.

And then the final question here is, Is this primary research? This is sometimes used as the very first question. If it's a review, you may use that as a source of more articles, but it's not included in the systematic review, only primary research.

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I'd like to thank Dr. Jay Levine from FSRRN -- he's director of that program -- and Dr. Mary Torrence, who's program manager for the funding for that project. Thanks for your attention.

(Applause.)

DR. ENGELJOHN: Well, thank you very much for all the information this morning.

And as I said, I think we are particularly interested in the work that Dr. Wills is working on with broilers. I think he is nearing completion of what he is doing for broilers, and then there'll be some activities associated with that and then a process for making that information available, because we as the Agency have a special interest in making sure you as the industry and regulated customer have access to that important information.

That completes our first segment of today.

We have 20 minutes set aside for a break. I'll call

you back in when we're ready to go. I know there's a

water fountain outside to the left, as are the

restrooms. And then there are beverages down the

street to the right.

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(Whereupon, a short recess was taken.)

DR. ENGELJOHN: Okay. Some of you have some questions you'd like to ask. And so I thought I'd just take this point while we're still uploading a presentation. We have time for maybe one or two questions -- at least one question.

So I've received a couple I'm going to answer. It -- but if somebody has a question that you'd like to ask the panelists, would you please come up to the microphone? And get ready to say who you are and which association you're with, and then we'll get started.

There have been questions about whether or not we're making the CD available and how quickly we're making compliance guide information available. Just so everyone knows, as soon as this meeting is over, we will download the information from the internet. The transcripts will not be immediately available, but all the PowerPoint presentations, speakers' notes and so forth will be.

And as quickly as we can get that done, we'll make those available. So you should expect that

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1	within the next couple of weeks, that information
2	would be available.
3	I think we have somebody ready to ask a
4	question.
5	If you would, give your name and
6	association.
7	(Pause.)
8	DR. ENGELJOHN: And the question is for
9	Dr. Altekruse. So is he here?
10	(Pause.)
11	DR. ENGELJOHN: It looks like he stepped
12	out. So we'll try to answer it.
13	MS. JOHNSON: I'm Patricia Marsh Johnson
14	with V. E. T. Solutions. My question was for Dr.
15	Altekruse actually.
16	So I was confused. In one point in his
17	slides when he was talking about the isolates that are
18	commonly found in human illness, he was stating that
19	about 50 percent of the Set A isolates are serotypes
20	that are common to human illness, but then in a
21	subsequent slide, he said that the vast majority of
22	sets have less than two isolates per set.

1 So I'm not understanding how that jives, I guess, because less than two isolates per set is far 2 less than 50 percent of the isolates being part of 3 4 those that are commonly found in human illness. DR. ENGELJOHN: I think we'll wait for 5 Sean to get back to answer that one --6 7 (Laughter.) DR. ENGELJOHN: -- rather than me give 8 9 you an answer, which I would probably attempt to do, 10 but I think I will not. So we'll keep that one on the record, and we'll try another one. 11 Does somebody else have a question? 12 13 MS. NESTOR: I'm Felicia Nestor with Food 14 and Water Watch. And I'm just wondering, after seeing the seasonality in the Salmonella data, what will you 15 16 do -- it sounds like from reading the new Federal 17 Register notice that you're going to be testing on a more consistent basis. 18 19 Now, how will you deal with the fact that 20 some plants that are going to be tested in certain months are going to be getting a higher percentage 21

just because of the seasonality effect?

22

If I were in

1	the industry, I would definitely be arguing me if you
2	were testing me in the worst months.
3	MR. LANGE: When we if you've read the
4	Federal Register notice
5	DR. ENGELJOHN: Who are you?
6	MR. LANGE: What?
7	DR. ENGELJOHN: Who are you, for the
8	transcript?
9	MR. LANGE: Oh. This is Loren Lange,
10	Office of Public Health Science.
11	As we speak Dan could answer this,
12	too we have a team of people that we're assembling
13	that beginning at the implementation of the change
14	will review each set result as it's complete. And
15	they'll look at the number of positive sets, and
16	they'll look at the number of serotypes of human
17	health concern in that set.
18	And this team is going to recommend based
19	on certain criteria a pattern for rescheduling. So
20	some establishments could get, you know, re-sampled in
21	a very quick period of time. Others that have either
22	booked low levels or low levels of human isolates will

change. We -- certainly, you know, to the seasonality, I mean we expect reduction all across the year. And at the end of the year, we'll sort of be evaluating.

Now, it will be -- the one thing I didn't mention is we posted the 2005 results. That is the last year of having any data in that format, because as we're shifting our resources to focus more on establishments where there are higher levels of public health concern due to the serotypes, we are going to change the nature of our data. So we are losing that, but we think it's a good decision to make.

DR. RAYMOND: I want to add a couple other things. This is Dr. Raymond, for the record.

Felicia, when I go eat chicken, I don't want to worry about which plant it came from or what month the chicken was slaughtered; I want to know that chicken is safe. And so our goal is not to have that S curve that Loren likes so much as a statistician. We want a flat line.

And I think you've already seen -- when we were looking at Loren's bar graphs, we looked at the

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last two quarters. Maybe perhaps we've seen -- it's too early to say this, but, hopefully, we've seen a change. Hopefully, the industry has created some changes that will drop those numbers down and we won't see that seasonality.

The second thing. Not to steal thunder from tomorrow, but when we're doing these sets, every time we have a positive culture, we will be letting Rather than wait until after the 53 the plant know. culture samples have been taken and then scrub those for a month and then tell them, they're going to know right off the bat that if they have positive, you first four out of five, they're know, tests the probably looking at another set to be done very So they'll start making changes right then, quickly. hopefully, rather than three months down the road.

So I think your question is a very good one. I mean I agree with you. I would not want to get tested in October if I had a choice of May or October if I was a plant -- with the current statistics. But we want to change those.

DR. ENGELJOHN: We do have our

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presentations ready. So I'll hold off on more questions. But I --

Dr. Altekruse, a question was asked. And if we can, we'll get the person who asked the question to just re-ask the question.

(Pause.)

MS. JOHNSON: Dr. Altekruse, you had in your slides -- on one slide that 50 percent of isolates were those that were common human serotypes in a set. And then in a subsequent slide, you mentioned that there were usually less than two isolates of those, which is far less than 50 percent. I obviously am not understanding the difference in the way the slides --

DR. ALTEKRUSE: Oh. The 50 percent is the ratios of all Salmonella isolates that are those human serotypes. Now, a lot of A sets have zero Salmonella in them. That's not an uncommon finding. So the average number of human serotypes is two, and that would suggest that, you know, the average is somewhere in the four to six range. There's some variability around it. But there are some plants that have many

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1	more you know, they're skewed out to the right with
2	30 Salmonella-positive tests. And one of those
3	every single one of those <i>Salmonella-</i> positive tests
4	was a human health serotype.
5	But the what that slide was trying to
6	portray was, How many human serotypes can you
7	typically expect to see in a set? And the answer to
8	that is very few. Typically, you know, 50 percent of
9	sets have two or fewer in them. So and but the
10	averages you know, they don't express the ranges.
11	So that average the overall average
12	would include all the sets that have zero and the few
13	sets that have 30. And so the total average for all
14	Salmonella would be, you know, in the range of about
15	six isolates.
16	MS. JOHNSON: Thank you.
17	DR. ENGELJOHN: Okay. I just heard a cell
18	phone. So I want to make sure that everyone has
19	turned off their cell phones. I haven't heard many
20	yet.
21	And so that we can keep our conference

line, we're going to start out now with our second

1	part of this morning's presentation with Dr. Laura
2	Hulsey. She is a veterinary staff officer at our
3	technical service center in Omaha, Nebraska. She has
4	her expertise in poultry slaughter, and she received
5	her degree in veterinary medicine from Oklahoma State
6	University and had a small animal practice in
7	Washington state, as well as working a private
8	practice in Jackson, Wyoming. Please welcome Dr.
9	Hulsey.
10	(Applause.)
11	DR. HULSEY: Thank you. My presentation
12	today is going to be an overview of poultry slaughter,
13	and this is a step-by-step
14	You can't hear me?
15	(Pause.)
16	DR. HULSEY: Okay. So this, my
17	presentation, will be an overview of poultry
18	slaughter.
19	(Laughter.)
20	DR. HULSEY: Again and again? Okay.
21	(Pause.)
22	DR. HULSEY: And this will take us step by

1	step
2	(Laughter.)
3	(Pause.)
4	DR. HULSEY: okay to set us up for
5	the presentations that follow today.
6	The information for the concerns and the
7	controls came from a literature review that we did at
8	the tech center over the last year, and the focus will
9	be on Salmonella. Each step in the process will be
LO	covered by about three slides, an in-plant picture
L1	that's typical of that process and a slide
L2	highlighting concerns at that step for Salmonella and,
L3	also, a slide that lists possible controls for
L4	Salmonella.
L5	We'll try to point out at each step where
L6	Salmonella can be introduced, amplified, decreased and
L7	where it has been found at the highest levels and some
L8	of the factors that influence the levels at these

This slide review that we have done is

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steps. And we'll do that briefly, because each of the

presentations today delve more deeply into those

subject areas.

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less formal in structure than Dr. Wills has planned, but it has provided a foundation for identifying decision criteria for risk-based inspection systems, and it will pull together information from the papers that describe the interventions and best practices. Those interventions and best practices we are trying to capture in compliance guidelines for use by the industry, and especially for small and very small plants.

Okay. So as I said, step by step: Live receiving and hanging, stunning and bleeding, scalding, feather removal, evisceration and chilling.

Okay. This is a slide that Dr. Fisher at the technical service center put together. And the purpose of the slide is to show the general trend of Salmonella levels through the slaughter process, and it's pretty much what you would expect.

He took the averages and found a mean prevalence for *Salmonella* at each process step. This was taken from 23 different research papers. So high at receiving, lowest numbers at scalding, highest at feather removal, moving on down through evisceration

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and chilling. So we would hope for the lowest levels at chilling. In this particular group of papers, we didn't see that.

Okay. So this is a typical picture of the live receiving where we stage the trucks or -- where you stage the trucks and unload. Some of the concerns at live receiving are, of course, the high level of Salmonella coming into the plant. The incoming loads can overwhelm in-plant interventions, and they are carried forward to the subsequent steps.

In one study that we looked at, there were feather samples recorded at 6.7 logs per gram, and skin samples at 5.9. And as many of you know, the crop has a large concentration of the *Salmonella*, along with the cecum, colon and the cloaca.

Controls that can be implemented at live receiving are, of course, feed withdrawal times and coop sanitation and cleaning programs, also unloading and holding area sanitation programs. The holding times, of course, affect the number or the amount of litter ingestion, as do employee traffic patterns in and out of the facility.

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And I'm sure that you've all had the experience of the live hang employee coming in through the back door of the plant while a white coat comes out of the other door. So traffic patterns can get to be a problem, bringing contamination into the plant.

Another factor is the air flow that moves into the plant from live hang. You want a positive flow from inside to outside.

The next step is stunning and bleeding, a typical picture of that electrical stunning. Concerns at this step are that the immobilization causes voiding of feces and further contamination of the carcass.

We had one study that we looked at that made a comparison of the withdrawal times. And they had a table that showed that the volume of excreta increased as the feed withdrawal time increased. And therefore, shorter withdrawal times may lead to a of the broilers producing lower percentage an excretion. That contamination at stunning, of course, is carried through forward to the scalder and the picker.

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This is a shot of a scalder without a counter-current flow. And a scalder that doesn't have a good flow to it, of course creates a bath of cross-contamination. The best idea is to have a continuous flow of water that moves the contamination and the feces away from the carcass instead of along with it.

Some more of the concerns that we have are that it washes much of the dirt and feces off, more microorganisms are removed during scalding than in any other step, but they affect the quality of the scald water.

Salmonella and Campylobacter are the most common organisms identified. We looked at a study that 75 to 100 percent -- of the samples of the prescalding identified 75 to 100 percent Salmonella prevalence. And of course, they accumulate over the shift.

Okay. Scalding controls. Brush systems are one that we saw throughout the literature review that had a good impact -- also, rinses before the scalder, counter-current flow and multi-tank systems. In one three-stage system in a multi-tank system,

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there was a report of 3.4, 2.0 and 1.2 logs CFU per mil. coliform reduction.

Temperature is important -- and pH. The literature papers reported on both high pH and low pH that decreased the number of microorganisms. In one report, they decreased the pH to 4.3 with 1.1 percent acetic acid and increased the death rate of Salmonella Newport and Typhimurium by 91 percent in the scald water.

This is a bird scrubber that I just got a picture of before we came here -- at a plant, that they installed. And you can see quite a significant difference from before and after. I don't have any numbers on this, but I hope to have them before we do the compliance guidelines.

The next step is typically the highest amount of cross-contamination and the highest level of Salmonella -- at feather removal. So we've seen up to 100-percent incidence of Salmonella at the feather removal step. The picking fingers and the feather follicle are implicated most often at this step and may drive Salmonella into the follicle. Also, the

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cloaca is emptied by the rubbing action of the picking fingers, which increases the contamination.

The controls this step, from the at literature, included the National Chicken Council's preventing the build-up **GMPs** that recommend feathers, continuous rinses for the equipment and the carcasses and, also, equipment adjustments to minimize cross-contamination.

One study talked about if contaminated water from the scald step's driven into the follicles by the picking fingers and then it moves forward into the chiller. It's the chiller cold water, and the follicles contract and hold that contamination in there.

Other interventions at this step include post-feather-removal rinses at 160 degrees Fahrenheit, chlorine rinses, acetic acid rinses, hydrogen peroxide rinses and other adjuvants; these produced mixed results.

Okay. We're going to move into the evisceration step. The crop removal seemed to be the area of highest concern. The greatest percentage of

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Salmonella was recovered from the crop, also from the ceca, but the highest incidence of carcass contamination occurring at pre- and post-crop, due to rupture and spillage at that step.

Okay. One study by Byrd and Hargis in '02 recovered marker organisms from broiler crops prior to live hang. They recovered 92 percent pre-crop and 94 percent post-crop. So that's a pretty big recovery rate. Also, GI leakage from the equipment and manipulation of that equipment and from the GI tract contaminates the carcasses and the equipment.

Controls at this step include, again, the National Chicken Council GMPs, which are -- focus on the most ideal feed and water withdrawal prior to slaughter, the maintenance and the adjustment of your equipment and continuously rinsing and sanitizing evis. equipment. Also, employee hygiene plays a role in this, as do whole-carcass water rinses.

Carcass rinses in the studies decreased Campylobacter, coliforms and E. coli. And you can see a 36-1/2 percent decrease in Salmonella when rinses are used, compared to 20.5 percent without; they are

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not effective against attached pathogens, however.

So we have a presentation later today on on-line re-processing in Salmonella that will delve a little more deeply into it, but these are some of the on-line reprocessing methods that are reviewed in the papers. At least 23 parts per million Free Available chlorine decreased the incidence from 5- to 2 percent in one study. 10 percent TSP decreased 1.3 logs. And then we went through several others here that we'll touch on later today: The 2 percent lactic acid, and so on.

The last step for an intervention to be applied is the immersion chiller. Concerns that were covered in our paper are that lipids are 84 to 98 percent of the filterable solids in the chiller; they consume the available chlorine, and they can protect the microorganisms.

It's also the major site of cross-contamination between positive and negative flocks, and it can increase the incidence by 20.7 percent on average. Salmonella-negative broilers will remain negative if you don't put a flock of Salmonella-

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positives through there first. So that says a lot for tracking pre-harvest before the birds get to the plant.

Okay. Other controls. The National Chicken Council, again, recommends proper water replacement, quality and temperature. The chiller health also depends on the pH generally of 6.5 to 7.5, Free Available Chlorine at one to five parts per million, and trying to minimize organic solids with a high flow rate, counter-current direction cleanliness.

bottom line the So the over through all these steps is that the appropriate interventions applied effectively can decrease Salmonella in the slaughter process.

I don't know if I mentioned just as a final note at the introduction as things were trying to get going here that we are working on the compliance guidelines that should be out shortly and will capture the information that we have in this lit review. Thank you.

(Applause.)

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DR. ENGELJOHN: Thank you, Dr. Hulsey, for that overview of what we're going to spend the rest of today and tomorrow morning talking about with regard to the production process and slaughter and [inaudible due to failure of in-house PA system] processing.

Our next presenter is going to give us some lessons learned, I think, from the perspective of what we've found when we as an Agency conduct our food safety assessments. Dr. Petersen is now the assistant administrator for the Office of Field Operations; he took that role in December of 2005. He recently had been the deputy administrator for the Office of Field Operations.

He served here in the headquarters. And then, prior to that, he worked as a senior staff officer in our Office of Public Health Science. He was in private practice for [inaudible due to failure of in-house PA system] time, as well, and has both a doctor's of veterinary medicine and a master's in public health, as well.

Please welcome Dr. Petersen.

DR. PETERSEN: Okay. Thanks, Dan.

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1	Actually, before I get started, I want to
2	circle back I can't believe I'm the only one who
3	missed it to Dr. Wills' bio. He works at
4	Mississippi State. And I have an administrator who's
5	a big-time Mississippi State grad, and I thought he
6	did some training in Nebraska. And I have an under
7	secretary that if anybody thinks he's not a major-
8	league Corn-husker, they need to spend about five
9	seconds with him.
10	So, Dr. Wills, where ever you're sitting,
11	I think you kind of hit all of the numbers. And so
12	very good.
13	VOICE: And Iowa State.
14	DR. PETERSEN: And Iowa State. Boy, he's
15	all over the place.
16	(Laughter.)
17	DR. PETERSEN: Okay. What I want to talk
18	about is things we're finding in our comprehensive
19	food safety assessments that are done by the folks in
20	the Office of Field Operations. And food safety
21	assessments of course, we have a directive out on

that now. The directives are information that apply

to our folks. That's at Directive 5100.1. That outlines the food safety assessment methodology.

The food safety assessments are basically a comprehensive look by specially trained people. They look at -- let me hold this.

(Pause.)

DR. PETERSEN: Okay. The food safety assessments are conducted by specially trained people, as most of you know, and they look at the structure and design of the program and, fundamentally, What is the theoretical basis for your program, what is the rationale for it, and are you delivering that in your food safety system, and are you making necessary —adjustments when necessary when things go awry in your process.

I want to start with a few definitions to make sure we're all on the same page. A food safety hazard: Any biological, chemical or physical property that may cause a food to be unsafe for human consumption. Your HACCP analysis, which is conducted to determine the food safety hazard [inaudible due to failure of in-house PA system] occur in the production

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process and identify the preventive measures that can be applied to control those hazards.

And then "reasonably likely to occur" are for those hazards where the establishment would establish controls either because they've historically occurred or they're reasonably likely to occur in the absence of those controls. Okay? Those are all regulatory references to be confused with not statutory definitions, statutory definitions for adulteration.

And since we're here to talk about poultry, we're working within the Poultry Products Inspection Act. There are multiple definitions of adulteration, but I really just pulled out two that are the most germane for us here today.

What we call a G-1, that's a particular section of the Act, and it basically has two parts to it: "Either bears or contains any poisonous or deleterious substance which may render it injurious to health," or, a little more commonly, in case it's not an added substance, "The article," meaning the poultry product, "shall not be adulterated under this clause

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if the quantity of such substance in or around the article does not ordinarily render it injurious to health."

So we've got a couple things in the second part there. It's not an added substance -- the quantity -- and not ordinarily injurious to health. For your friends on the beef side, *E. coli* 0157 -- this is where our authority drives for that. No one would consider an *E. coli* infection anything other than injurious to health.

So we don't quite have that with poultry. Some use the reference, "Naturally occurring." But I think, theoretically, at some point -- not today -- we could have a Salmonella that has such hospitalization rates, high attack rates, high infectivity rates, high case fatality rates, and then I may wonder if that Salmonella is not ordinarily injurious to health.

So not today, but that's a possibility if some of the virulence factors continue to evolve.

Really more likely what we're dealing with and what we'll talk about as we get into these food safety assessment discussions is the G-4 reference,

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"Insanitary Conditions": "Where it's prepared, packed or held under insanitary conditions whereby it may become contaminated with filth or where it may have been rendered injurious to health."

Insanitary practices such that I find the product adulterated, meaning I can't put the marks of inspection on that product. Adulteration not to be confused with food safety hazards. Two different references.

Okay. So what did we find? Food safety assessments -- these are the assessments. We've done about 31 of them or -- 31 that we've looked at that we have initiated since October 1, 2005. Briefly, the results. Ten of which led to a notice of intended enforcement, basically, where I had questions about the adulteration status of that product.

Interestingly, having heard Dr. Raymond's first slide earlier, where he -- one of his early slides showed about 30 percent of plants were in that Category III. These aren't the same plants, but, interestingly, about 33 percent of these assessments in plants led to an NOIE.

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Eleven non-compliance records were issued, meaning there was some miscellaneous non-compliance, but not sufficient for me to pursue an enforcement action. Nine had no action, so were within compliance regulations. 30-day rethe And one had а assessment letter issued. Yes, we do still do 30-day re-assessment letters. Those are not regulatory They basically mean I have a question about issues. some supporting documentation that you're unable to provide or explain, and we give you a period of time to describe that.

What I want to do with these next slides is -- of course, we've got some folks in the room who are quite familiar with our regulatory frame work, and we have others who are less so. And so I've characterized these findings kind of in between those two levels of knowledge.

And general observations. And we'll work through these observations, not necessarily synonymous with the regulatory requirements, but I think you'll see some common themes as we go forward.

Many establishments do not sufficiently

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identify Salmonella as a food safety hazard reasonably likely to occur, and, yet, when you look at their supporting documentation, all of the supporting documentation says it is likely to occur. But they do identify microbial growth, enterobacteriaceae, pathogenic microorganisms, as the hazards.

Inadequate consideration of incoming Salmonella loads. I see references to on-farm practices, on-farm vaccination protocols, but more listed just for the fact of listing them rather than using that information or integrating that information into their thought process.

Process control steps. Inadequate consideration of incoming Salmonella loads and process control steps that affect Salmonella levels. We just heard some of that a minute ago on the last slide. Multiple steps in the process can have multiple impacts, but not considered there at all. And intervention's not validated to address Salmonella.

Inconsistencies between the hazard analysis and the selection of your critical control points and critical limits. Hazards identified in the

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hazard analysis. And they're simply identified. No indication on whether they're reasonably likely to occur. No supporting documentation for decisions that the hazard is reasonably likely to occur. It's just there.

And for those who are using prerequisite some level, insufficient records programs at describe what it is they do and how they affect your decision making. And of course, we look to -supporting documentation to demonstrate the scientific regulatory basis for your program informs what it is you're trying to implement, what it is you're trying to execute.

Salmonella. Where it is identified and is reasonably likely to occur at some process steps -did that's your hazard analysis -but not subsequently indicate where in fact it would reduced. prevented, eliminated or So we have Salmonella that's floating out there. At some point, if it's reasonably likely to occur, where are you dealing with it?

Some plants put controls in place and take

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actions consistent with the hazard, but not indicated as reasonably likely to occur. So addressing it in fact, but not addressing it in the thought process and the design of what it is you're trying to do.

And interestingly, CCP has established a control of food safety hazard for a hazard that's not even deemed reasonably likely to occur. That's a difficult concept.

Key steps without identified hazards. A couple examples: Biological hazards not identified at processing steps -- that doesn't mean they have to be, but some processing steps, we think, may have some biological hazards; Red-water chilling and some of the chilling steps. And if they're not identified, there's no rationale, no subsequent documentation, to justify how those decisions were reached. You can justify the decisions, but you need to have a basis for it.

Off-line steps, whether it be wash-out, fecal air sac, IP cut-up or whatever, where we have incoming birds with a hazard reasonably likely to occur, but, somehow, when we get to the more risky

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parts of the process, the hazard -- there's no hazard identified or none is specifically referenced. And so what is the thinking and what is the theme cutting through your program? And what is the thought process at each step of the way?

Water re-use. You are expected to have measures sufficient to prevent or reduce physical, chemical or biological contamination to the extent necessary to prevent contamination or adulteration of product. So have you considered that? Have you considered the impacts of water re-use on your HACCP plans -- subsequent HACCP plans? That, of course, is a regulatory expectation.

No supporting decisions for selection of CCPs and critical limits. You've selected them. You have a critical limit. And here largely we're talking about various temperature critical limits, and, of course, there are some common ones: 36 degrees, 50 degrees and 55 degrees. Well, why is that? What is -- what are you targeting? What are you trying to accomplish?

No supporting documentation for the

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monitoring and verification frequency selected for the CCPs. Why are you doing your checks at a certain frequency? What is the basis for it, and what do they tell you? How does it tell you the status of your process on an ongoing basis?

That's true where the common controls are involved -- whether it be typical fecal contamination, of course, is a common control -- or temperature controls. What's the frequency for your temperature controls, monitoring and verification, and why do you do it at that frequency?

CCP validation. Particularly, of course, involving an antimicrobial intervention, is it effective to reduce the identified hazard at the entry point? And your supporting documentation in your hazard analysis of food safety hazard reasonably likely to occur. Where is it dealt with, and how do you know it?

Chlorine identified in the hazard analysis at steps as a control measure to prevent the food safety hazard. However, when you go to those steps, chlorine's not even applied. We need some thematic

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thinking, some systematic thinking, to what we're trying to design and implement. And then, of course, zero tolerance and temperature are not uncommon CCPs.

Corrective actions following a critical limit deviation are not implemented, particularly for temperature controls. Say if your temperature is 50 degrees and you exceed it, 51 degrees. Well, that's your critical limit. What are you doing? What should you be doing? Are you doing it on an ongoing basis? And if you don't, you need to look at your system.

Corrective actions not documented, not implemented, or the preventive measures were ineffective. And here we'll work mainly through some SSOP issues.

Repetitive corrective actions: Clean, sanitize, inspect. Pre-op, for example. On a -- at some level at some point when that happens with some frequency on particularly some product contact zones, what does that tell you about either the effectiveness or the design of your SSOP program?

Repetitive documentation of temperature

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deviations. Well, it's only one degree, so maybe it's not so bad. Well, it is a critical limit. And if they're exceeded, there are some expectations on how we deal with it.

Recurrence of a deviation should inform you of something. And what do you do with that information, and how do you improve your program accordingly?

And so all of which leads to not conducting appropriate re-evaluation or modification of the design of, say, your SSOP, or what does it tell you about the execution of your SSOP? You can improve the execution if you think the design, of course, is fundamentally satisfactory. But with repetition, one of those should result.

So the next couple of slides kind of pull this all together in a Salmonella context. Sanitation performance standards. And these are some common themes particularly but not exclusively in the plants that led to enforcement actions.

Just fundamental employee hygiene issues.

And we heard that with the last slide. Restroom

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issues, product handling, hand washing and that type When supplemented with some pest control know, unaddressed you pest control concerns -concerns -- cockroaches and that type of thing -superimposed with employee hygiene concerns. And ventilation, of course, refers to condensation controlled in a manner sufficient to prevent creation of insanitary conditions product or adulteration.

Control and equipment and facilities. the equipment contaminating product? Is it leading to fecal contamination of product at some ongoing frequent basis, and why is that acceptable? That's superimposed with facility concerns: Pipings with the, you know, insulation displaced over product zones, superimposed with the wall sanitary issues. And so these things tend to multiply when we look at what is being executed in that facility.

Water re-use considerations. Of course, an SBS issue. We touched on that.

So taking that, the findings in those plants, it doesn't -- I'm not suggesting it needs to

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be all, but when you start to add in facility questions with ventilation questions with employee hygiene questions.

And then development and implementation of your SSOPs. What did you want to do, and is it working? And if not, are you making adjustments? That's maintenance and effectiveness, taking appropriate corrective actions. And what do those corrective actions tell you about what's occurring? And then, of course, appropriate documentation.

And so those two things, SPS and SSOP, with what we walked through on some fundamental inconsistencies on what it -- what your program says or does. Hazards without controls. Hazards that are reasonably likely to occur with process steps that are not validated for that hazard.

The monitoring frequency. How do you know it works for you? And then validation verification and re-assessment. I thought I heard Dr. Masters say something using the words, You don't have to wait for us. And you don't. These are the issues we're finding, and my strong preference is that you find it.

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You don't need me to find it. And so you have that 1 2 opportunity. And then the record keeping. That goes 3 4 with everything else, to document what your system is delivering. 5 heard earlier that Salmonella-6 So 7 positives have some reflection on the status process control, Categories I, II and III, and then, 8 of course, exceeding the performance standard. 9 That 10 was the previous bench mark. And a facility with some perhaps serotypes 11 of human health concern now superimposed perhaps in an 12 13 establishment with of these repetitive some SPS issues, these SSOP issues, these HACCP issues. 14 And so 15 what is your program? How is it constructed? 16 make sense? Is it consistent? And this is kind of the -- I won't call it 17 a hierarchy, but this is kind of the sequence of 18 19 events that can be an entry point for us to look at 20 your food safety system. And when we've done that -what I outlined is what we typically find. 21

just, I think -- I know you can

And I

1	design and implement a HACCP system. And so we need
2	to look at the thought basis behind that and look at
3	the scientific basis and get it done.
4	And that's it.
5	(Applause.)
6	DR. ENGELJOHN: Okay. Can you hear me on
7	this microphone?
8	(Pause.)
9	DR. ENGELJOHN: We're going to have to
10	find a microphone that we can put down there that we
11	are sure that you can ask questions from.
12	But if I can have the members from this
13	morning's presentations come up to the table and
14	we'll take some questions from the audience. And then
15	I will also ask whether or not there are any questions
16	from our telephone callers. So there were
17	Dean, I saw you up here.
18	Somebody needs to come up to the
19	microphone.
20	(Pause.)
21	DR. ENGELJOHN: Please state who you are
22	and your association, and ask your question.

1	MR. BENSON: Okay. Mike Benson with
2	Jennie-O Turkey Store. I'm asking this question as a
3	consumer, though, of poultry products. My question is
4	for Dr. Masters.
5	In the presentation Laura had earlier, it
6	was showing the use of a lot of chemicals in the
7	processes. And it would appear from the agenda and
8	from the discussion that the FSIS is recommending or
9	implying the recommendation that we should be applying
10	more and stronger acidifiers, alkalizers, oxidizers,
11	sanitizers, quaternizers and more of these chemicals
12	to our food products.
13	My question is, Dr. Masters, do you think
14	that these are safe? And is that the direction that
15	FSIS is looking for processing companies to go forward
16	with?
17	DR. MASTERS: I think that in my
18	presentation, I indicated that there is
19	This is Barb Masters. There is a variety
20	of options that you can use to control <i>Salmonella</i> in
21	your establishment. I'm aware of some establishments
22	that use no chemicals in controlling <i>Salmonella</i> . I

think that Dr. Hulsey was presenting to you some of the interventions that are available. I think Dr. Wills presented to you that they are doing a systematic approach to some of the interventions that are available; many of those will be non-chemical interventions, hopefully.

And so I'm certainly not suggesting to you that the only approach that you can use is to continuously apply more and more chemical interventions in your process. I think you need to start at pre-harvest and look at what you can do pre-harvest.

You need to look at control in your process. And I think that's -- what Dr. Petersen was trying to say to you is you need to look at every part of your process, starting with your employee hygiene, looking at your SSOPs, looking at your sanitation process control and looking at your control of fecal contamination. And you need to look at all of those things in combination.

What Dr. Hulsey was providing to you was some of the areas that you might look at some

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1	interventions, and that would be your option to look
2	at which of those might be useful in your
3	establishment. But certainly, we're not suggesting
4	that you need to only control Salmonella through the
5	use of increased chemical controls in your
6	establishment.
7	DR. ENGELJOHN: Yes. Next question.
8	MR. DANILSON: Thank you, Dan.
9	Probably to Loren, but I don't know. It
10	might be
11	DR. ENGELJOHN: Who are you?
12	MR. DANILSON: Pardon?
13	DR. ENGELJOHN: Who are you?
14	MR. DANILSON: Oh. Thank you, Dan. Dean
15	Danilson with Tyson.
16	I found it very interesting the data
17	that you showed in your red zone plants, the low
18	the disparity of the number of A sample sets that have
19	been taken on those red zone plants relative to middle
20	zone and green zone. As your efforts in 2006 and 2007
21	increase in sampling activity on those red zone plants

relative to the data trend that we're looking at, we

are very likely to see a continued increase in Salmonella incidents as reported in your trend analysis, because of the bias that will push back up there, which will not look good to whoever is looking at that.

And I -- we need to be aware of what that next two years is going to show us on that trend. And I hope that increasing trend doesn't take away from our interest and desires for promoting and making incentives for the green zone plants and take away from those efforts. So, you know, I just ask that that be kept in mind.

MR. LANGE: I have a response to that.

I -- one thing we will do is have the staff at FSIS looking at all different ways in which we can display the data.

And just thinking off the cuff, it may not be something we can do, but we also do have -- through animal disposition reporting system, our have we numbers of carcasses slaughtered. We could generate a weighting statistic like the last set new by production weight for each plant.

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I'm just throwing out an idea, but we'll consider a lot of different ways of presenting the data because, as I said earlier, this ends that era of 1998 to 2005 in terms of at least having some consistency in what we've been posting on the web.

DR. RAYMOND: And I'm going to add to that Loren.

In that we've talked about this and we recognize very clearly that the trends may get skewed, we also recognize from some of the people who advise us from the outside that we shouldn't even be using these as trends because it's not a statistically significant sample. As Loren said right up front at the very start of his presentation, this is not intended to be a statistically significant sampling of the product; it's to sample perhaps verification and to judge individual plants, but it doesn't say what the Salmonella load really truly is from the product.

That said, we've also discussed changing how we sample and that it would also skew the statistics even more significantly. Instead of always sampling on the first shift, I strongly believe we

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need to look at the sampling of other shifts. And that will change our data, also, but we won't know how much it'll change until that we can have а statistically significant number of first shift samples and second shift samples in the same plants to find out what the risk of a carcass is going through the second shift versus the first shift.

I mean those are issues that have been raised to me, and I agree that they're issues that need to be addressed. So I don't think we'll be able to say anything from trends in the future, you know, looking backwards. We'll have a new trend data, and we'll use that.

But very most importantly is not how many sets are positive or how many carcasses test positive; it's how many people test positive. And so we still have that baseline data going: The number of human beings who have culture-proven Salmonella infections per year. That's really the bottom line for me. I mean it's important -- what's going on in the plants, but the bottom line is what's going on when people eat the product.

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1	DR. ENGELJOHN: And I'll just point out,
2	too, just because we haven't mentioned it yet but we
3	will later, that we are in fact going to start another
4	national baseline study, which is statistically
5	designed, that's going to start this year, as well.
6	So it will serve as a basis to give us a more accurate
7	picture of true prevalence.
8	Could we have the next question?
9	DR. BYRD: Ken Byrd, with Mionix.
10	Actually, I have a question for Dr. Hulsey.
11	You had mentioned something about a study
12	where in the scald the pH was I believe you said
13	4.1 and you got a certain reduction in Salmonella. I
14	didn't get that number. Could you repeat that,
15	please?
16	DR. HULSEY: That's a good question, and
17	I'll have to look. And I'll get back to you on that.
18	DR. BYRD: Okay. Thank you.
19	DR. HULSEY: Okay.
20	DR. ENGELJOHN: I think could I ask the
21	operator? Are there any questions being queued by the
22	telephone callers?

(Pause.)

DR. ENGELJOHN: While we're waiting for that, go ahead and ask your question.

DR. O'CONNOR: Bob O'Connor from Foster Farms, and I had a question for Mr. Lange.

The seven-year data that you looked at for what we term seasonality -- basically, you broke it into quarters. How many companies within those seven years were tested during the same quarter over those seven years? So if I was Company A and it was Year One and I was tested in Quarter 3, did I get tested in that same quarter the next year? If I was tested five times, how many times did I fall into the same quarter?

MR. LANGE: There is a pattern of when -that sort of grows from when we started the program in
1998, where it initiated. We scheduled everybody that
was a large establishment early in the year. And then
by only scheduling one set a year, we did get in the
pattern that if you were large, in the early years,
you tended to get sampled in that March/April/May/June
period.

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In the last couple of years, we've been trying to sort of, as you can see, diversify that a little bit. And you can see we actually have the largest number of samples last year in the fourth quarter. So it did happen.

I can't, you know, tell you -- but, you know, there's a lot of plants that probably found themselves getting sampled at the same time of the year each year. But that will change. Now, so I mean -- but that won't happen in the future, because rescheduling will be based on, you know, public health considerations.

I'd like to add one other comment to what Dr. Raymond said. When this issue of a second set came up, I went back and looked at the report of the original 1994-to-1995 baseline that established the 20 percent. All those samples were collected on first shift samples. All those samples were Monday through Thursday.

So in trying to do studies, I -- we have both microbiologists and statisticians. And they don't always come from the same perspective as to when

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1	samples get collected and when samples get shipped,
2	because the microbiologists always want consistency
3	and the statisticians would like it spread across the
4	board. And it's you have to find that common
5	ground between the two.
6	DR. O'CONNOR: Okay. I think the reason
7	I'm asking the question is because we kind of deemed
8	that seasonality. And I think from a biological
9	standpoint, if we deem it seasonality, we might try to
10	look for a biological reason for that. But if it's
11	not truly seasonal, if it's more a scheduling incident
12	or coincidence, then I probably wouldn't concentrate
13	on the seasons of the year.
14	MR. LANGE: Yes. There
15	DR. O'CONNOR: For interventions. That's
16	what I'm saying.
17	MR. LANGE: You would have to do a very
18	sophisticated analysis of trying to account for all
19	the potential variables. Was it the plants that were
20	getting scheduled in that April/May/June period that
21	just always kept that down, or was it a real change?

But I -- you know, there's probably both happening,

1	like anything in the real world.
2	DR. ENGELJOHN: Any other questions?
3	MR. POTTER: Bill Potter with George's.
4	My question is about upcoming baseline studies and
5	maybe future Salmonella standards and how they're
6	developed.
7	My question is, In the upcoming studies,
8	has there been any consideration of the enumeration of
9	the Salmonella colonies on the carcass? Most of us in
10	research are really interested in the enumeration.
11	In other words: Is the carcass containing
12	one DNA strand of Salmonella, which would make it
13	positive under the current testing, or does it
14	contain I don't know a million? To us many
15	of us, that is of significance, and it would be good
16	to know. Is that being considered in future studies?
17	MR. LANGE: Yes. As Dan just mentioned,
18	we are in the final process of designing a new broiler
19	baseline. We will quantitate Salmonella and
20	Campylobacter on all samples. And we have over the
21	last year, we have had the National Advisory Committee

on Microbiological Criteria for Foods recommend a new

1	Campylobacter method that we will use in our baseline
2	studies, and it is currently undergoing validation at
3	the contract lab that will do our baseline study.
4	So we hope to start a shakedown period
5	we hope by the end of March. But if not, we should
6	start in April of this year for young chickens. And
7	probably then the next thing I'll have the staff work
8	on is a turkey carcass baseline.
9	MR. POTTER: Okay. Is excuse me. If I
10	may, is the enumeration just for Campylobacter?
11	MR. LANGE: And Salmonella.
12	MR. POTTER: And they'll okay. Thank
13	you.
14	MR. LANGE: They will enumerate for
15	Salmonella, too.
16	DR. ENGELJOHN: This is Engeljohn. I'll
17	just add a little more to that.
18	We will also be looking at some [inaudible
19	due to failure of in-house PA system] organisms. And
20	I would also point out that we've changed the
21	structure of this baseline. We traditionally had done
22	baselines in which we looked at the point most

immediate to sale or to the consumer, meaning the post-chill sample. You should expect this time we will do a sample at re-hang before the carcasses are chilled and we are going to do it at post-chill in the same facility.

And as we get better and have more ongoing baseline studies, which we do intend to do for all the classes of products, we will add more points in the production process by which we will take those samples so that we can see what's happening in the entire production process. But this baseline study is designed to have two points in the production process by which we pull samples.

MS. NESTOR: Felicia Nestor, Food and Water Watch. Loren, did I understand you to say that the original baseline was done first shift, that most of the samples were done first shift Monday through Thursday?

MR. LANGE: Yes. If you go on our web site and read -- what is it -- June '94 to '95, the actual baseline is posted on there. And there's a sentence that says -- it says, All samples were first

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1	shift and Monday through Thursday. So it was back
2	then, it all tied in to Fed-Ex delivery schedules.
3	And we were shipping carcasses back then, and the
4	microbiologists all wanted the carcasses shipped the
5	same day they were sampled.
6	MS. NESTOR: And what months did that
7	MR. LANGE: That would have been across
8	that would have been designed to so it weighted
9	production by month. So as a baseline study, it is
10	based on production volume to get your national
11	product estimate. So if production was actually
12	changed, you would expect more samples in the month
13	where production was higher. And I don't know if
14	you'll see that fluctuation in production. But it
15	would be evenly across all months, yes.
16	MS. NESTOR: So it's a one-year baseline?
17	MR. LANGE: Yes. It was one year, yes.
18	MS. NESTOR: Okay. And were all sizes of
19	plants tested? Because I know in ground beef, the
20	very small plants weren't tested.
21	MR. LANGE: In all except the very
22	smallest. The sampling frame, I think, covered 99.9

1	percent of production. So they were all subject to
2	sampling, but then the number of times the way the
3	baselines in the '90s were done, the number of times a
4	plant would be sampled was entirely proportional to
5	its production volume or, at least statistically,
6	that's how the number of samples was derived.
7	MS. NESTOR: Thank you.
8	DR. ENGELJOHN: While we have the
9	gentleman coming up to the phone, could I ask the
10	operator again?
11	Has anyone identified that they want to
12	ask a question?
13	FEMALE VOICE: Yes. [inaudible due to
14	failure of in-house PA system].
15	DR. ENGELJOHN: Loren, were you able to
16	capture that?
17	MR. LANGE: I captured
18	DR. ENGELJOHN: Could you repeat the
19	question? And we'll just
20	MR. LANGE: Yes. The second part of it
21	was, Is I tried to show both the seven-year data,
22	the 2002 to 2004, I've looked at 2001 to 2003. That

1	pattern of broilers being low in April, May and June
2	and higher in October, November and December appears
3	to have been consistent across all seven years through
4	2004.
5	And the point I was trying to make is it
6	really changed in 2005. And it changed because the
7	third and the fourth quarters actually ended up being
8	the lowest.
9	Now, that answers part of it. I didn't
10	catch the whole thing.
11	DR. ENGELJOHN: The first part of the
12	question that I think I heard was also that there
13	appeared to be the 2.8 percent increase from last year
14	to this year, I think, was the question and that it
15	appeared to be rather significant. Is that true?
16	MR. LANGE: We don't have the ability to
17	sort of test to say, "Was the increase from 13.5 to
18	16.3 a statistically significant change," because the
19	data isn't set up so that you can run that type of
20	test. But we certainly, it was a larger increase
21	than we've seen in previous years.

DR. RAYMOND: This is Raymond. And I

would say, Caroline, what bothers me the most is that it wasn't a one-year increase on the carcasses on the broilers. It has been four years in a row or three years in a row that it has gone up. Starting four years ago, each year has shown an increase. And I don't think you can deny, if you look at a four-year trend, that there's not a problem.

DR. MASTERS: And this is Barb Masters. And I guess that was the real reason we started these meetings last August, and that's why we're here. And that's -- what we're hoping to do with these meetings and the sharing of information and with the policies that we put out that will be in the <a href="#Federal Register">Federal Register</a> is to turn that trend the other direction. And that's -- what we're hopeful we saw in the last two quarters is the trend going the right direction and that we're, you know, challenging the industry. And we're optimistic that we can start seeing the numbers go the other direction.

We do believe that it's a trend in the wrong direction. Whether it's significant or not significant, it is in the wrong direction, and we

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1	believe the data needs to go the other direction. And
2	we're optimistic that with the policy changes that
3	we have put that we're recommending and that we're
4	putting in place are the kinds of things that can make
5	that trend go the other direction.
6	DR. ENGELJOHN: Do I have other questions
7	here in the audience?
8	MR. YANCY: Al Yancy, Gold Kist. This
9	question's for Dr. Petersen.
10	I think I heard you say that there had
11	been 31 FSAs since October 1 of '05. The maybe the
12	only question is, Are those is that statistic
13	does that speak only to targeted FSAs for failed A
14	sets, or is that routine FSAs, or is that some mix
15	thereof?
16	DR. PETERSEN: We gave the district
17	managers just some general guidance. They didn't
18	really target for A sets [inaudible due to failure of
19	in-house PA system] performance. [inaudible due to
20	failure of in-house PA system] district managers were
21	aware of some plants where [inaudible due to failure

system] they

had

some

of in-house

PA

22

lasting

Salmonella control over time. And at the starting point, that's where we want [inaudible due to failure of in-house PA system].

MR. YANCY: Okay.

DR. PETERSEN: And so they had [inaudible due to failure of in-house PA system] the ones that [inaudible due to failure of in-house PA system] their minds. And once we better characterize the food safety assessment methods in a, you know, Salmonella-[inaudible due to failure of in-house PA system] commodity in all establishments, that's really [inaudible due to failure of in-house PA system].

MR. YANCY: The reason I'm asking, as a follow-up is I wondered if there was any direct correlation between the ten NOIEs that were issued and Salmonella incidence in those plants. In other words, did the ten plants that got NOIEs -- did they -- if we were to have the Categories I, II and III at present, would they have all fallen into a Category II or III, or do we know?

DR. PETERSEN: Well, I can look at that. But that wasn't -- if you look at the basis for any of

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these actions, it's the exception [inaudible due to failure of in-house PA system] issue. It's that [inaudible due to failure of in-house PA system] of issues that leads you up to failure [inaudible due to failure of in-house PA system] if there was a failure [inaudible due to failure of in-house PA system] these things come together over time.

MR. YANCY: Okay. Thank you.

DR. ENGELJOHN: If I could -- Dr. Hulsey, if you could, respond to the question that was answered earlier. And repeat the question if you could.

I believe your question was, DR. HULSEY: What was the decrease in pH in the scald water for the Salmonella killed in water -- if that was correct. That was from a paper by [inaudible due to failure of in-house PAsystem]. And he determined t.hat. [inaudible due to failure of in-house PA system] water with a pH of 4.3 [inaudible due to failure of in-house PA system] 4.1 [inaudible due to failure of in-house PA system].

DR. ENGELJOHN: And I would also just

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1	reiterate on the point that the information that Dr.
2	Hulsey is sharing with you will all be fully
3	articulated in the compliance guideline that we expect
4	to have ready to go within a matter of days, not
5	weeks, as I typically try to characterize things.
6	Any other questions in the audience here?
7	(Pause.)
8	DR. ENGELJOHN: If I could ask the
9	operator?
10	Is there any other question on the phone
11	line?
12	(Pause.)
13	DR. ENGELJOHN: Okay. We have no further
14	questions. It's about lunch time. We will be back
15	here at one o'clock for the afternoon session with
16	some research information. Thank you very much.
17	(Whereupon, at 12:00 p.m., this conference
18	was recessed, to reconvene at 1:00 p.m. this same day,
19	Thursday, February 23, 2006.)
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22	

we're

### A-F-T-E-R-N-O-O-N S-E-S-S-I-O-N

This

ENGELJOHN:

DR.

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(1:05 p.m.)

afternoon,

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going to actually concentrate on actual research and real-life experience that can be shared with you. Our first speaker this afternoon is Dr. Mark Berrang. He graduated from Virginia Tech with a bachelor's in '86 and then from the University of Georgia with a master's in '88.

He began working with the USDA's agriculture research service in Dr. Nelson Cox's lab in '89. He completed a PhD in food microbiology from the University of Georgia and took a senior scientist position with the Agricultural Research Service. And his specialty has been in the area of poultry food safety microbiology.

During the course of his career, he has conducted research at all stages of production and processing, ranging from the hatching egg to further-processed, ready-to-eat poultry meat, and these efforts have resulted in the authorship on 87 peer-reviewed articles published in scientific journals and

1	90 scientific abstracts.
2	Please welcome Dr. Mark Berrang.
3	(Applause.)
4	DR. BERRANG: Thank you, Dan.
5	Can you all hear me okay?
6	VOICES: Yes.
7	DR. BERRANG: As Dan said, I'm going to be
8	talking about Salmonella and Campylobacter in broiler
9	transport coops. And most of my own work in transport
10	works has actually been done specifically with
11	Campylobacter, but I will discuss some information
12	about Salmonella today where I can and where it's
13	appropriate.
14	We all know the kind of coops we're
15	talking about: The dump coops that carry broilers
16	from the farm to the plants. And thanks to our
17	interaction with local commercial cooperators, we've
18	been able to get coops to work with so we can use
19	real-life surfaces to sample.
20	We have several different ways that we
21	sample these coops in our experiments and our studies.

And the most important surface in my opinion in a

transport coop is going to always be the floor, because that's where most of the fecal matter ends up and that's where the birds tend to settle onto and contact during transport.

So pathogens in transport coops. It's no big surprise that Salmonella and Campylobacter have been detected in transport coops. And Stan Bailey published a paper a few years ago, where he found that 5 percent of the coops that he sampled were positive for Salmonella prior use; 10 percent were positive after use.

And in another, Belgian, study, 56 out of 128, or close to 44 percent, of their transport crates were positive for Salmonella prior to use. And these authors showed that the prevalence of Salmonella-positive broilers was increasing during transport, and they suggested the transport crates that they used in Europe were an important source. And of course, similar findings and suggestions have been made relative to Campylobacter.

So what is the source of Salmonella and Campylobacter in these coops? Obviously, Salmonella

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and Campylobacter are present in the gut and feces of broilers from a positive flock. And feed withdrawal tends to change the microflora, and transport tends to increase excretion. And that's how they get into the coops.

Coop flooring and carcass microbiology.

Jeff Buhr and some other folks at the Russell Research

Center did a study a few years ago where they compared

broilers that were transported on solid flooring,

traditional fiberglass flooring, to those that were

transported on an elevated wire floor. The wire then

allowed the fecal matter to drop through, and there

was less contact between the bird and the feces.

What they found is that feathered carcasses that were transported on the solid flooring had noticeably more fecal contamination right here on the breast than the ones that were on the wire floor and higher numbers of E. coli, also. But what they also found is that after those same carcasses had been scalded and defeathered, all the differences in the carcass microbiology had disappeared. So we started Well, to wonder, how really important is this

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contamination in the transport coop?

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I did a study with the help of some other folks at Russell Research Center where we looked at cross-contamination in transport coops. And what we examine the possibility wanted do was contaminated feces left in these dump coops can cause transfer of Campylobacter to birds that were previously free of Campylobacter.

did what we is -we got some transport coops -- new transport coops that had never been used before, loaded them with Campylobacterpositive broilers, let them stay in those coops for pulled Campylobacter-positive eight hours, the broilers out and replaced them with birds from a Campylobacter-negative flock. So we were able isolate the fecal contamination in the dump coop as the only source of Campylobacter to these previously Campylobacter-free carcasses.

And what we found was that after just two hours in that dump coop, more than half of the defeathered carcasses from the test flock, the Campylobacter-negative flock, had now become positive

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for Campylobacter and -- albeit it was at lower numbers. It was still a substantial number on the rinses from these birds that -- carcasses that were from a Campylobacter-negative flock.

And the important point is that we sampled them after defeathering. So this contamination was transferred in the dump coop and was maintained in the carcass through scalding and picking.

So Campylobacter can in fact be spread to previously negative broiler by contact with contaminated feces remaining in the dump coop, and this contamination can remain on the carcass through scalding and picking.

Now, keep in mind that Salmonella is a much more hardy organism than Campylobacter. And I feel confident in saying that if Campylobacter can be transferred by this route, the Salmonella can be, too. In fact there's similar findings that have been recorded in the literature for Salmonella.

"So should we be washing and sanitizing cages," is the question to pose. And is that -- is washing and sanitizing dump coops the answer?

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Right now, according to Julie Northcutt and Dena Jones, about 28 percent of U. S. broiler plants have a coop wash or sanitizing procedure in use. There's a much higher percentage of processors in Europe who are washing and sanitizing crates, and they have been for quite awhile.

Nevertheless, even though they've been washing and sanitizing crates in Europe, it's not hard to find reports coming out of Europe where they have detected Salmonella or Campylobacter or both on their transport crates -- even after washing and sanitizing them. And the authors of these studies usually point out human error and the mixing of the chemicals or equipment breakdown or not getting all of the fecal matter off the crates as reasons why they still have to detected these organisms after washing.

Having said that, though, there are literature that reports in the show that some experimental washing and sanitizing procedures But when you look at this research, before you run out and adopt a washing and sanitizing procedure based on experimental results, you need to evaluate

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these results carefully.

For instance, were the chemicals inactivated properly before the samples were taken? There's studies that are published where the target bacteria might actually be being inadvertently being killed after sampling during culture, because the chemical has not been properly inactivated.

Another thing to consider is, How realistic or commercially viable is the procedure, and how expensive or unwieldy is the equipment? If it involves big immersion tanks or high temperature application of chemicals, it might be more money than is really reasonable.

Julie Northcutt and I did a study at a commercial processing plant that does have a coop washing procedure in place. And what we did is -- we measured the numbers of total aerobic bacteria and E. coli on the floor surface of dump coops, and we also measured the presence of Salmonella and Campylobacter.

And what we found is that by washing these coops with water, just by spraying them out with water, they were able to reduce the numbers of aerobic

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bacteria and  $E.\ coli.$  And then later they applied a sanitizer, which lowered the numbers even more.

And the prevalence of Campylobacter and the prevalence of Salmonella were also lowered, but, even so, after the washing and sanitizing procedure, we were still able to detect Campylobacter on two out of 27 coop floor samples, or about 7 percent. I feel like if we had sampled a higher number of coop surfaces after washing and sanitizing, we probably would have found Salmonella in some of them, as well.

So we decided to look at some novel approaches and try to see if we could find other ways to possibly decontaminate transport coops. And one thing we tested was allowing the coop to dry out and be stored between use for extended periods of time. And our thinking there was that by allowing the fecal matter left in the cage to dry out and be exposed to atmospheric oxygen, we would lower the numbers of Campylobacter that we were able to detect in that feces.

So what we did is -- we got dump coops, and we loaded them with *Campylobacter*-positive

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broilers, left those broilers in there for eight hours, removed them and then put the dump coops in a pole shed, where we just left them for 48 hours, and periodically scraped out all of the feces from some of the openings and cultured that for *Campylobacter*.

And this graph shows some of the results from that study. The Y axis is marking the number of Campylobacter that we recovered per opening. Across the X axis is time.

What we found is that we started out with nine 10 to eight to between the and 10 the Campylobacter per opening and that stayed steady for the first eight hours of sampling, but, by 24 hours, we saw a two-log decrease, down to about 10 to the six per opening, a two-log being about a 99-percent By 48 hours, we were down to our limit of decrease. detection or even where we were unable to detect any Campylobacter from those -- the feces that was left in these dump coops.

The Campylobacter counts closely mirrored the percent moisture in the feces that was left behind. And this graph shows how the percent moisture

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in the fecal matter tended to go down as the coop was left out to dry.

So by allowing these transport coops to dry out and not be used for 24 to 48 hours, we were able to lower the numbers of *Campylobacter* in that feces, but we were not able to reliably eliminate the *Campylobacter* altogether.

And it's important to note that the expense required to maintain enough coops for this would be really impossible to justify. I'm not suggesting that every company needs to go out there and buy three to four times the number of coops and build a huge building and store them. That's not at all what I'm trying to say.

Julie and I also did a study looking at washing and sanitizing of coop flooring. And this was designed to test the efficacy of spray washing with tap water followed by an immersion I chemical sanitizers to eliminate *Campylobacter* on the coop flooring material.

And this was similar to the field study that we did, but it's a much more controlled

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environment, because we were doing it in a lab so we had a repeatable way to contaminate the floor surface and a much more repeatable way to wash it and sanitize it.

What we were doing in this study is -- we used little pieces of coop flooring that we cut from a large sheet that we got from a commercial cooperator, and we intentionally contaminated these little squares with gut contents from Campylobacter-positive broilers with known amounts of Campylobacter in them. allowed the gut contents to dry, and then we'd wash it off with tap water at a known water pressure. dipped the little squares then we with whatever remaining fecal matter was still on them into a sanitizer to see what would happen.

This graph shows what we found when we did the water spray wash followed by immersion in chemical sanitizers for 15 seconds. And the control bar shows the number of *Campylobacter* on a square that was not washed and was not dipped with sanitizer.

Where we had about ten to the seven Campylobacter on that little square, just washing it

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with water lowered those counts by two logs, down to ten to the five. But immersing those little squares of flooring into a quat or a chlorine for 15 seconds did not improve the reduction of the *Campylobacter* at all.

And we found the same kind of thing when we immersed them into the sanitizers for 60 seconds. In fact, when we immersed them in the sanitizers for five minutes, the numbers we found from the little squares that were immersed actually went up. And I think they were just getting so wet that the remaining fecal matter was washing off -- coming off of the sample much easier.

So spraying the floor surface with tap water lowered the numbers of *Campylobacter*, but adding an immersion of 200 parts per million quad or chlorine did not help.

So we know that water spray can lower the counts, and we know that drying can lower the counts. We decided to look at those two together and measure the effectiveness of the water spray followed by an extended dry time to eliminate Campylobacter, and this

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graph shows the results we found in that study.

Here in this study, we counted Campylobacter, which is shown on the blue bars, and we also counted E. coli, shown on the red bars. And, now, Salmonella is closely related to E. coli. So I would expect Salmonella to respond in a similar way to E. coli to this kind of treatment.

With our control -- again, this was done with little squares of flooring. With the control squares, we found about 10 to the 7 Campylobacter and close to 10 to the 6 E. coli. Just applying the spray -- low-pressure water spray, we were able to lower those counts significantly, down to 10 to the 4 Campby and about 10 to the 3 E. coli.

The 24-hour dry was -- that's just drying and not spraying. That lowered the counts even better than a spray by itself, down to about 10 to the 1 Campylobacter and half-a-log E. coli. But when we used the spray followed by the dry, we had our best reduction; we were not able to detect any Campylobacter or E. coli.

Notice the one red bar there with the

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little asterisk on it. That's to point out that when the little -- when the fecal matter had dried for 24 hours and it was re-wet -- we sprayed it after 24 hours to simulate that coop going out on a wet day -- the numbers of *E. coli* do rebound up to about Log 2, not the size they were originally -- nowhere near the size they were originally, but it does rebound somewhat. *Campylobacter* did not rebound. But I would expect *Salmonella* to behave like *E. coli* in this kind of situation.

So the low-pressure tap water spray lowered the numbers of E. coli, but simply allowing the gut contents to dry out was more effective, and using them together in concert was very effective. But the re-moistening -- again, if that dried-out fecal matter becomes re-moistened, coli E . and probably Salmonella can rebound.

So is floor surface drying a reasonable sanitation treatment? It can certainly lower the numbers of *Campylobacter* that we recover from these floor surfaces, but it would really require, obviously, a major change in thinking relative to coop

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1	design or management, and we can't do it the way
2	things are right now.
3	We cannot store cages for 48 hours. We
4	would have to either have removable floors that could
5	be stored separate or have a fast way to dry out the
6	surface.
7	So overall, coop washing and sanitizing is
8	an expensive proposition. You have to worry about
9	water costs, your personnel and the time involved.
10	You have to worry about what are you going to do
11	with your runoff water? Are you causing cross-
12	contamination in that area by spraying that material
13	around?
14	The efficacy is questionable. Even in
15	Europe, where they've been doing this a long time,
16	they're still finding pathogens on the surfaces after
17	sanitizing. And we're looking at new ways to sanitize
18	these coops now, and we'll see what we come up with.
19	And I think I'm about out of time.
20	(Applause.)
21	DR. ENGELJOHN: Thank you, Mark, very much
22	for that helpful information on getting the birds to

the facility.

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We're going to move now into the facility. And we have Dr. Stan Bailey, a microbiologist also with the Agricultural Research Center, with USDA. is the lead scientist and Bailey research microbiologist for USDA Agricultural Research Service, where he has directed research toward monitoring, controlling, reducing and ultimately eliminating contamination of live poultry by human enteric pathogens.

During his 21-year career, he has authored or co-authored about 500 publications on food microbiology. And today, we have Dr. Stan Bailey to talk about the processing plant and final carcass contamination. So thank you very much.

(Applause.)

DR. BAILEY: Thank you, Dan. But that's 31 years, not 21. Don't take them away; I need them for retirement.

(Laughter.)

DR. BAILEY: Thank you for the invitation.

And the first thing they asked me to talk about today

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was, What does the effect of what's coming off the farm, the load of bacteria on your chickens, have on what you can do in the processing plant? So I went through some of the work that we've done and some of the literature.

But I think if we're going to talk about what's on the chicken, then we need to have just a little basic understanding of what's going on. And the first thing I wanted to talk to you about was, What are the factors that affect Salmonella colonization of poultry?

And you'll notice I might say, Colonization. And that's because, for the most part, the vast majority of strains of Salmonella don't infect the chicken in the classic sense; it's not causing a disease. I mean there are a few strains that will. But for the most part, we're talking about it just setting up shop and growing in the intestinal tract.

And there are a number of factors that affect that. Probably the first and foremost is the age of the chick or the chicken. A new-hatched chick

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is highly susceptible to becoming colonized with Salmonella. If you take -- each strain may be slightly different.

But if we take the same strain and give it to a day-of-hatch chick, probably 50 to 75 percent of those chicks will become colonized with Salmonella with maybe ten to 20 cells. By the time that bird's three or four days old, it'll take maybe 10,000 to 100,000. By the time it's a week or more old, it might take a million cells of the same strain of Salmonella. Of course, all of that can be affected by stress and bird health, the third factor I show there.

Then there's -- how is the Salmonella getting there? There's a couple of primary ways that we're always concerned about. I think first and foremost is what's coming out of the breeder stock.

If you have your breeder stock contaminated and if they take it into the hatchery and spread it around and, therefore, you're putting birds on the ground that are *Salmonella* colonized, then you're almost beat before you start, because anything you do from that point on -- for the most part, you're

going to have a hard time preventing or getting rid of that Salmonella.

And then, of course, you can get it onto the farm free of *Salmonella*, and then you also have to worry about feed and environmental exposure. And each of those can play a separate role. And I could talk for the 15 minutes I have about all of this, but let me move on.

theoretically So what seeing are we happening on farms? For the most part, if you don't have extra stress and other factors affecting you, this graphic would probably indicate what's happening most of the time. You'll have some low level of Salmonella coming out of the breeder's farm or else in the environment or the feed when they first get there, and that will explode up in the first two weeks or so.

In around two to three weeks, you'll see your maximum level of *Salmonella*. And if you don't do anything and if they don't have any extra event that causes significant stress or anything, then you'll see that intestinal colonization of *Salmonella* starting to

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drop off over time to the point they're six to seven weeks old. Again, without any extra stress pressures, it'll drop down pretty low, sometimes 1 or 2 percent, 5 percent or 10 percent, just depending on the situation.

But it's not just what's in the bird that we're concerned about. Remember, we talked about there at Week 2 or 3, that might be your maximum level. Well, what happens is, unlike campy, which Mark was just talking about, Salmonella is not very fragile. It sticks around for a long time. So if you ever get it in your flocks, if you get it on the skin or on the feathers, it's likely to stay there and take it into the farm or into the processing plant.

I did two studies several years ago where we looked at that a little bit. In the first one, we challenged the chicks the day of hatch with Salmonella, and we followed them for five weeks or six weeks, to the time they would go to the processing plant. And at that time, they had been reduced down to that 5 percent level we were talking about intestinal colonization. But of those same birds, 53

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percent of them were carrying Salmonella on the outside of the bird.

Another set of studies we did similarly had 15.5 percent of the birds intestinally colonized percent 50 with Salmonella compared to the feathers. And as we're talking about load coming into the plant, I think that's particularly pertinent because -- most of you are aware -- that know the literature and know what we've seen through time -that probably 90 to 95 percent of the birds in a processing plant that are Salmonella-positive are carrying very few cells of Salmonella probably less than a hundred and often times less than 50 cells of Salmonella, on the whole carcass. So it result of probably is а some kind of crosscontamination at transport maybe.

So those figures I was just talking about then were what was going on with the birds when they left the farm. There has been several studies done -- and I didn't put them all here, for the sake of time. But to summarize, we see in general an increase in the Salmonella carriage, either internally or

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externally, of between 20 and 40 percent for those birds after transport compared to before transport.

So being in that coop, being exposed to the fecal material, just the aerosoling, the sitting in the materials, or whatever, will increase the carriage of *Salmonella* from 20 to 40 percent during transport.

Another area that we can talk about where I can show you a little bit of the issues that have to do with the carriage had to do with a study we did several years ago, and I thought I'd highlight just a few points of that. And the objective of this study was to characterize on a multi-state basis the prevalence of Salmonella from numerous sources in chicken production and processing.

We looked at plants in four states each season of the year, and we artificially defined high and low production. We let the companies do that. And those would be production parameters: Feed conversion, just generally good operating plants versus those that weren't perceived as being so good. And actually, in this study we found a very low level

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of Salmonella overall.

And I just wanted to show this to you to kind of see the kind of variability we would see. But you can see within plants, there is a big variability by season and by plant. But I threw this slide in for a particular reason. If you want to monitor what's going on in your farm level and you want some really simple way to do it, we've found that used fly strips can catch your flies. They're a natural filter, and they monitor. And we found the best sensitivity of knowing what was going on on our farms just by looking at what was going on with the flies.

To the point that Mark was just talking about, transportation coops, look at the difference that we see here. In three of the four times that we looked at this, we had significantly higher levels of Salmonella detected in the coop swabs after transport than before. In that particular study, to some of the things that Sean Altekruse was talking bout this morning, we identified 36 different serotypes. Those frequently were Senftenberg, Thompson and Montevideo.

And we found one strain that we -- had never been

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isolated from poultry before.

Now I want to spend just a couple of minutes with you talking about a study that we've been doing -- and just completed a couple of months ago -- for the last year with FSIS which again is illustrative of the type of load that we're bringing into the plant.

With this particular study, we asked some basic questions. One was, Can a reliable measure of process control be determined from one or more postchill samples, or do samples have to be paired from within a given flock post-pick compared to post-chill?

And, if you want to answer that question, how many -- if it requires either one at the end or a pair, how many samples would you have to look at to have an indication of what's truly going on to take out statistical variability?

And then finally, is there a relationship between the reduction in *E. coli* counts during processing and the reduction in *Salmonella* and *Campylobacter* during processing?

We looked at 20 randomly selected plants,

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again, across all four seasons. FSIS collected the samples and sent them to us in Athens in a blinded manner. I have no idea what plants they were and where they came from. And in this particular study, we looked at ten carcases post-pick and ten carcases post-chill.

In the study, we looked at quantitative E. coli and coliform and Campylobacter levels, qualitatively Salmonella. We used Petrifilm for the E. coli and coliform and direct plating on Campy Cefex for the Campy, and we used the FSIS procedure of a BAX PCR. And if we got a positive screen, then we used cultural procedures to get our Salmonella isolate. When we finished the study, we had done a total of 6,400 analyses, 1,600 for each of the four organisms.

Just to give you a little bit of an idea of the kind of load, as I'm supposed to be talking about incoming load, when we looked at the rehang immediately after the picker, we saw about 72 percent positive for Salmonella across all of the samples and all of the seasons, but you can see that, obviously, the processing plants were doing something right,

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because we had about 19.6 percent of the birds positive by the time they got out of the chill tank.

So we did have a pretty significant load coming in, but the processing plant seemed to be doing a reasonably good job. And you can see some variability in the seasons, too, particularly more on what's coming out of the plant than what was going in.

I did want to finish with just a couple of slides as we're talking about the importance of what's on the birds as they come in. And I'm going to talk later about some chemical disinfection things, and you'll have other talks this afternoon.

But if we go back to the point of, "What's the load coming in," being particularly important -- I did a study with Tonya Roberts a couple years ago where we looked at Salmonella control in Scandinavia, particularly Sweden and Denmark. And I'm not going to get into that whole study, but we obviously don't have the same type of industry. We can't do the same control program, but I think we can learn from some of the principles that guide that program and maybe apply some of those to help us here on the farm.

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And the basic principles of the Swedish program is: If broilers are never exposed to Salmonella, then they can't become colonized. In their system, they started off by eradicating all the positive breeder flocks and continue to do so over time, and a number of other things. But we can't do that. That wouldn't be practical. But what we can do is learn from that and know that we have to be able to control what's coming out of our breeder flocks, and there's ways and things we'll look at.

Then the other point that comes from that is: Their entire control program -- they have zero Salmonella -- or pretty close to zero. Their entire control program is on the farm. They use no chemicals in their processing plant. They use the same basic equipment we do, but they just use water.

So what potential U. S. are some intervention strategies that don't involve eradication? Well, on the farm or -- on the breeder farm to start with, we can control Salmonella, believe, to a great extent -- and I know this to be true because I've seen it done -- by controlling

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what's going into the feed, with biosecurity, with an effective vaccination program, an effective competitive exclusion program, and, particularly, we have to pay attention to moisture control.

In a broiler production, again, we can look at the feed, biosecurity, competitive exclusion and moisture control. And we won't get into all the issues we talked about in the fall about getting some of these products approved.

So in conclusion, most chicken flocks in S., unfortunately, carry some load of Salmonella. It's to our -- what we have to work on is getting that load as low as possible. And again, transportation appears to increase both the internal and external carriage of Salmonella. We have seen and will see more today that chemical treatments in the plant can reduce Salmonella on processed chickens, but it's important to keep the level of Salmonella low in and on the chickens as much as possible, and that will eventually require going back to the farm.

So that's all I have. And thank you very much.

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DR. ENGELJOHN: Thank you, Dr. Bailey.

We're going now to on to move а presentation on sanitation and the sanitizers being We have Dr. Scott Russell, who is since 1994 a used. professor of poultry processing microbiology in the department of poultry science at the University of Georgia; he also has had real-life experience as a production manager and microbiologist at a processing facility in Gainesville, Georgia. And with him -- he has done a lot of research that will be helpful to you and I'm sure would be good as research --

So Dr. Russell?

DR. RUSSELL: Thank you. I'd like to thank the Organizing Committee for having me. It's a real pleasure to be here with you today. And in the [inaudible due to failure of in-house PA system] I have, I'd like to go ahead into the subject.

And as [inaudible due to failure of inhouse PA system], numerous microbes can adhere to processing equipment surfaces, as most of you know, and they may concentrate and grow in crevices or

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joints of rubber gaskets that remain moist. And wear on these surfaces can also impact the ability of the organisms to adhere, and any remaining food material that may be left on those surfaces can then provide a growth medium for the bacteria.

So part of the whole idea of cleaning and sanitation involves removing a lot of that material to avoid any sort of growth material being there for those organisms.

So just by way of a general overview for about cleaning sanitizing, how to go and important for the large pieces of trash and things to be picked up and electrical connections to be covered. And then they want to go through a pre-rinse with warm or hot water and then apply usually an alkaline cleanser, as applied through a central system using pretty warm water, and then five to 20 minutes of exposure, and then usually the ceilings and things like that -- floors, walls, equipment -- rinsed with a cleaner.

And then it's inspected, and touched up as necessary. And only after all of those things have

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been conducted, that's when the sanitizer's applied.

And I want to go into kind of more in depth as to why

it's so essential that we get rid of all of that

stuff.

Now, most of the commonly used detergents -- and I won't go into this in detail -- are alkaline detergents by far. They're especially useful in the poultry industry because of the types of soils that are deposited there. And there are a number of categories of those types of products.

Acid detergents are generally used to remove strongly encrusted surface matter, and they're good for cleaning the scale on a fairly regular basis, as well. Some synthetic detergents that are out there are used occasionally -- such as quaternary ammonium -- and then there are soaps that are used -- a variety of different things that can be used to do cleaning.

Now, with regard to sanitizing, there are a number of different products that have been used in the industry, but I want to point out to you and, as we go through this, I want to make it real clear that

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I don't -- the sanitizer isn't nearly as important as removing the biofilms in the cleaning process. It is important to use a good sanitizer, and all of these things give you an opportunity to see what's out there, for example quaternary ammonia, a very popular sanitizer used throughout the industry.

Some companies use industrial-strength some will use a chlorine-dioxide mixture. bleach; Some use iodine compounds especially in hand-dips and things like that. Some have gone to encapsulated lysozyme, and I'll talk a little bit about how that works. Ozone systems, steam systems, vapor, peroxide peroxyacetic and cetylpyridinium chloride are all being used in these contexts throughout the industry at different -- in different areas.

This -- I don't want you to read this. I didn't put this here for you to read it or memorize it. There won't be a test afterwards. Okay?

The idea here is for you to see what types of characteristics are needed for a sanitizer to be considered effective. Really, you -- the companies

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that develop these things have to jump over numerous hurdles to make this thing worthwhile to the industry.

I mean look at all of these different things. The kill rate, the toxicity, stability, speed, penetration, film-forming, and all of these kinds of characteristics -- proclivity -- have to be -- it has to pass all of those tests in order for it to be effective. And then, of course, we have to look at what area that the product's going to be used in.

There are concerns such as, What kind of equipment is in that area? Is it aluminum? Is it stainless steel? Is it wood? Is it plastic? And in all of those areas -- concrete floors, these kinds of things -- it's important to determine, What's the best sanitizer or best cleaning method for those types of equipment? Of course, we don't have a lot of time to go into that.

There are other concerns, too. What type of water is coming into the plant? Is it high in iron? Is it hard water? All of these kinds of things are really important. And of course, the cost of the

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sanitizer is very important, as well, as most of you know -- and then the type of material you're applying it to -- walls, tile walls, rubber belts, and so forth.

Now, why is it that the sanitizer -- I mentioned earlier that I don't think sanitizer is nearly as important as removing biofilms removing the bacterial colonies that are already there on the equipment. I want to show you some things as we go along.

And this is one of the big problems that we see throughout the food processing industry. Bacteria can be transferred from floor drains, from walls, ceilings and these kinds of places, where they will accumulate and grow -- bacteria like Salmonella and Listeria -- and they will -- they can easily be transferred by aerosolization.

These -- a lot of the employees, as you know, like to use high-pressure hoses. And they'll spray the floor, and these bacteria will aerosolize, and they'll land on a piece of equipment. It could be stainless steel, it could be rubber belting or what

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These bacteria will then begin to communicate with one another through a processing called quorum sensing, and they send out a signal. Usually, this is an N-acelated-homoserine lactone, AHL for short.

And these chemicals basically are signaling to the other bacteria, Hey, I'm not real happy here; there's not many nutrients here; we have got to get together and build a house to protect ourselves. That's in a nutshell what they're doing. Now, I'm not sure if they say it in those words, but that's, you know, my best estimate, anyway.

of bacteria But number do this. Salmonella, E. coli, Pseudomonas -- a lot of them do that. And they form these biofilms sort of -- if you envision in your mind -it's like an ant. it's raining outside, Ants -and the ants together, some signal goes out, and they begin to build an ant mound. It's a very similar type of process.

Now, as I mentioned to you, they do this

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to protect themselves, and they do this to prepare an organized communal structure. And it is similar to an ant mound. And I have some pictures here for you to see.

early stages, the biofilm the is composed of a cell layer attached to a surface, and the cells grow and divide, and they form a dense mat layers thick. The bacteria use numerous quorum sensing to signal each other to reorganize, therefore forming a very complex array of pillars and irregular surface structures. And these structures are connected by convoluted channels that deliver food and remove waste, and we're going to see pictures of this.

Now, why is this significant? Because if you go along and spray a sanitizer on an already formed biofilm, you may only kill 10 percent to 50 percent. And, believe it or not, 50 percent in the bacterial world doesn't mean anything. Okay?

We've talked about a significant reduction in bacteria being at least a log; that's 90 percent. If you go and only kill 10 percent or 50 percent, you're not even removing by a log. And that doesn't

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do you much good. So it's essential that these biofilms be removed, because they will protect bacteria, and they will keep them from being killed by sanitizers.

This is a picture of how biofilm forms and how short a time it requires. In the first window up here at the three hour stage, you see single bacteria there. But by as early as eight hours, you start to see the yellow biofilm formation around the colony and, by ten hours, the bacteria almost wholly encased in that biofilm. So you can see, in a very short period of time even during an individual processing day, these bacteria can easily become encased in a biofilm and can become protected from sanitizers.

Look at how complex this structure is.

There can be everything from yeast, bacteria -- many different strains or species of bacteria can get in this structure. And they're all communicating in there, and they're forming these organized channels.

What happens then is -- this is sort of a good diagram. You can see that the aerobic bacteria live on the top of the biofilm, and oxygen can

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penetrate down to them. But down below them, anaerobic bacteria are living, as well. Food and moisture are able to go through those channels that you could see in the previous slide down to the bacteria and allow them to survive just fine. The waste materials are excreted out the bottom of the biofilm.

It's just like an ant mount; it's a very organized communal structure. It makes it very, very difficult to penetrate that with chemical sanitizers, and that's really our challenge. That's one of the problems that we see throughout the industry.

This is an example of how the biofilm -the bacteria that form these things can be sprayed and
they can aerosolize because of high-pressure hoses and
so forth. They fly through the air, they land on a
piece of equipment, and they go through these stages.
And once they reach like Stage 5 here, as you see on
the slide, then they rise to the top.

The thing can break off or be rubbed off by incidental surface contact or sprayed off with a hose. Next thing you know, these bacteria are flying

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to other areas within the cooler or within the processing plant and forming other bacterial biofilms, which, again, makes it very, very difficult to remove these things.

So as I mentioned before, you can clean and sanitize. And if these things are in biofilms and you don't remove the biofilms, you can still have only 10 to 60 percent elimination, which isn't much, in those cases. And most cleaning programs involve four days with alkaline cleaner and one day with an acid cleaner. And authors Jessen and Lammert found that the effective detergents on biofilms was negligible whereas the most efficient disinfectants were able to eliminate L. monocytogenes. Very high strength and long reaction times were required under these conditions. Acid disinfectants composed of hydrogen peroxide and peracetic acid were more chlorine-type efficient than the compounds or oxidants.

Similar results are reported by other authors, while others find the efficacy of the two disinfectants or even the opposite efficacy. So

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there's conflicting studies as to which is better, chlorine or oxidant-type compounds or like peracetic acid-type oxidant compounds combined with acids.

recommendations for removing Now biofilms. This kind of has gone through some evolution over time, but Parker and others found that chemicals that attack the polysaccharide matrix of the biofilm -- that is that glycocelic structure -- were particularly effective in killing and removing cells in a biofilm. Treatment of this biofilm on a clean -of either of the biofilms on a clean, stainless steel surface with lysozyme, which is an enzyme produced by a number of different types of cells, killed biofilm cells and prevented the attachment of any bacteria to the surface.

Now, lysozyme can easily be isolated from egg shell membranes, in case you're wondering where we might get something like that in large volume. The authors suggested that lysozyme may have potential as an alternative control method for biofilms of bacteria. Gibson and others reported that cleaning produced a .91 log reduction and high spray -- and

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again, that's not that much. That's not even a whole log reduction, where -- and they were saying that high-pressure sprays and mechanical methods -- actual scrubbing methods -- were the best.

But it required quite a high-pressure spray; 17.2 bars of spray were required just to start to remove biofilms of *Pseudomonas* and *Staphylococcus*. Increasing spray times didn't seem to have any effect on the biofilms, and acidic or alkaline or neutral detergents didn't increase the removal of biofilms. However, the acidic and alkaline cleaners or products affected the viability of the organisms and then minimized the spread of contamination later on.

So now, we did some studies where we were able to dramatically reduce fully formed *Listeria* biofilms on stainless steel products. And this product was applied by electrostatic spraying. It was created in an all natural way; in fact, the product could be consumed by itself in a fully concentrated form without any problems.

Log 10 reductions in our study ranged from 3.3 to 7.2 logs when compared to the controls, and

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caused the *Listeria* biofilm literally to float off of the stainless steel coupon. It was really interesting to see that.

Here are the data if you'd like to see them, but we saw reductions anywhere, again, from 3.3 logs all the way up to 7.2 logs on fully formed biofilms of *Listeria* on stainless steel coupons. So we had very good results there.

Testing for biofilms. Sampling is very difficult because they adhere to equipment so tightly, and scraping or high-pressure swabs are recommended. Traditional methods for microbiological testing are very slow; most of them are 48 hours, and they just provide a retrospective assessment of cleanliness. Rapid real-time methods are much more appropriate.

You can see here how difficult it is to do a swab technique, and it takes 48 hours. This is a Petrifilm contact method, to give you an idea, another 24 hours. The traditional contact RODAC plate takes another 24 hours to get results. A Hycheck's another way to do that.

These are new carbohydrate and protein

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1	test strips that are out there. They're real time.
2	And these other carbohydrate test strips are real
3	time, as well. A lot of people have switched to ATP
4	bioluminometers, because you get real-time assessments
5	of these things. And I don't have time to go into
6	here.
7	Conclusion? Proper cleaning and
8	sanitizing is absolutely essential in preventing the
9	cross-contamination of pathogenic bacteria from
10	equipment to food products. Choosing the proper
11	chemical for the food matrix is important in terms of
12	removing that food from the surface.
13	And biofilm formation is a serious
14	problem, because they're difficult to eliminate once
15	they've formed fully formed biofilms. And as I
16	mentioned before, it doesn't take much time to do
17	that. Microbiological monitoring programs should be
18	real time and should not be retrospective. Thank you.
19	(Applause.)
20	DR. ENGELJOHN: Thank you very much.
21	We're now going to hear from Dr. Marty
22	Ewing, who is a graduate from the University of

Georgia College of Veterinary Medicine in '87 and practiced in the private clinical medicine before joining USDA FSIS. She then moved to Florida, where she began working for the state department of agriculture and managed the national poultry improvement plan.

We're glad to have her here today. She's going to talk to us about a natural disaster and things that she was able to find out from that.

So thank you.

DR. EWING: Well, I appreciate it, everyone. Thank you to the Organizing Committee again for inviting me to speak here today.

I think you might find this presentation a little (different) from some of the others. This is more going to be about the experiences and concerns that we had following what was probably one of the most catastrophic natural disasters we've had.

But first, for those of you that aren't familiar with Sanderson Farms, while we now have facilities in Texas and Georgia, we are primarily a Mississippi-based company. In south Mississippi, we

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1	have four processing plants. In one just north of New
2	Orleans in Louisiana, there are four hatcheries, three
3	feed mills and over 600 contract poultry houses. And
4	in that region we process approximately 4 million a
5	week.
6	Okay. It's a little ironic, but, almost
7	six months to the day ago, on August 26, we were
8	traveling home from what was then the pre-harvest
9	meeting on Salmonella interventions. And at that
10	time, a small Category 1 hurricane was skirting south
11	Florida, and we went home and did the standard
12	hurricane preparations.
13	We filled all the fuel tanks and discussed
14	with our growers, checked generators
15	(Pause.)
16	DR. EWING: Okay. I'll try to talk
17	louder.
18	We checked all the generators in the
19	hatcheries, plants and poultry houses. We try to
20	deliver as much feed to the growers as we can in
21	advance and, also, pick up any hatching eggs. We

communicate with our growers on feed, water, fuel and,

again, drainage, to prevent flooding of the houses.

And generally, within 24 hours of where the path -- we can distinguish where the storm is going to hit, we'll notify the district office of any plant closures. Unfortunately, on Sunday, we woke up to a Category 5 behemoth, and her name was Katrina.

This slide depicts that path of Katrina as she roared across the Mississippi coast line. The star is Laurel, Mississippi. That's our corporate headquarters. And all of our facilities in Mississippi were either affected by hurricane or tropical force winds.

After Katrina, there was no power south of I-20. I-20 goes through Jackson, and all of our facilities are south of there. There was no phone service and no communications except for one radio station that we could get intermittently out of Jackson. There was no water in most areas. There was no gas or diesel fuel. And if there were, we couldn't pump it, because we didn't have any electricity.

Curfews were ordered in most localities.

And the extent of the devastation, although we didn't

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know it at the time, was over 90,000 miles.

Now, the Gulf Coast? It was incredible. If you go west of the Gulfport/Biloxi area, houses that are left standing are uninhabitable. And if they're not uninhabitable, they're just gone. West of Gulfport past Christian, there is nothing but front steps and foundations. And it's still that way. Fortunately, where were -- we were about 90 miles inland -- we were not hit, although we did have tremendous wind damage.

This is the road to my son's high school, outside of our house, completely blocked. And blocked roads were everywhere. This is a church down the street from that house. If you'll notice, the windows are very well taped. The joke around our town now is, Gee, they really should have taped the steeple down.

Now, the damage in Laurel was particular devastating from the high winds, and many trees were knocked down. This actually belongs to one of our employees in Laurel; this is her home. Fortunately, she and her family are fine, but her home was devastated. We had about five or six others in our

offices that had experienced the same thing.

And of course, once we got the trees cleared, we had downed power lines everywhere. And something I want to share with everybody because -- I didn't realize this. But even though there's not any power for hundreds of miles, if somebody has a generator hooked up and it's hooked up wrong, these lines can still be hot. So always respect downed power lines.

So, anyway, the good news? There was no loss of life to any of our Sanderson employees, contract growers or contractors. And there was also no major facility damages to any of our feed mills, hatcheries or plants, although we did have some live haul sheds that were damaged.

The bad news? Seventy-one poultry houses were just totally gone. 90 more houses had substantial damage in that they would take three to six months to repair. And virtually (every) farm in Mississippi sustained some damage. Now, in the houses that were partially damaged but the chickens were still contained, we were able to pick those up and

move them either to other houses on that farm or maybe even to another farm. Injured and unconfined birds were euthanized.

Here's one of our houses and yet another and another. And you can see the chickens.

Bird losses. We lost 3 million birds outright and another 5 million due to reduced placements and egg sets.

So housing. In the houses that were left standing, management was a challenge. Most of our houses are tunnel ventilated. We in many places were not able to do that because of curtains, roofs or wall damage. We did experience some heat loss in our older birds because they weren't acclimated to the heat. And unfortunately, September was the hottest September on record in Mississippi.

Because of the feed constraints, we had to feed a single ration, and we also couldn't pelletize it. The birds -- instead of just pre-feed, we had to feed them only twice a day. But we were slowly about to get them back on to a full feed by mid- or the end of September.

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Now, obviously, we had birds that needed to get desperately out of the field. And in many cases, the processing plant was the best way. But there were concerns about re-opening the plants. First we had to make sure that there was no flood or water damage, which, fortunately, we did not experience.

There were power outages. So there was potential product abuse. So we had to dispose of 4.7 million pounds of product in our coolers and cold storage. Now, a large majority of that was stuff that was stored down in the New Orleans cold storage facilities.

Now, prior to reopening the plants, we also had to assure water potability. So it took us -Hammond and McComb were able to come back online that next Saturday, September 3, Collins and Hazlehurst on that Sunday. And at Laurel, because of water potability, even though, ironically, Laurel had power two days after the storm hit, we couldn't start running and processing birds there until the next Wednesday, a week later.

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Of course, the poor communications and blocked roads, some people were just completely displaced, and employee attendance was low. To help employees when we were able to communicate to them that the plants had reopened, meals were provided. We gave them free ice.

Our human resources people were trained to file to help our employees file for FEMA and Red Cross assistance. We also provided fuel to not only our contract growers to run their generators but also to employees just to get to work. And it continues to be a problem in our region. The industries -- not only ours -- continue to be plagued by labor shortages.

Obviously, the birds' condition -- this was a challenge. We had -- we were processing birds aging from the range of the low 40s to mid-70s. We normally process birds that are either 50 days of age or 60.

Obviously, we had an increase in variability. And this was particularly apparent when you had multiple-age flocks, because we had to move one age of birds into houses with other ages of birds.

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There was a moderate -- not as bad as I thought it would be -- increase in condemnation and trim. Wе did slight increase in see а IP, Inflammatory Process, particularly in those flocks that we had to pick up and move. And there was a slight increase in Zero Tolerance deviations, but, fortunately, that was minimal.

And of course, as Dr. Bailey was talking about, there was stressed birds. There was concern about maintaining our Salmonella performance So USDA started 51-day windows in all standards. plants in the regions. And I'm pleased to say that all of the plants in our regions -- not just ours -met the performance standards. And there was also a special FSIS team sent to survey the plants, and I believe those results are to be published. But basically, what they did is -- they took pre-evis. samples at the rehang table and post-chill samples. The pre-evis samples they tested of *E. coli*. The post-chill samples they tested for  $E\,.$ coli and Salmonella.

The paired samples that we got from --

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that we shared with the USDA or they shared with us. We had no Salmonella-positives post-chill. The average reduction on the E. coli amounts in those samples we had -- in each of the plants, you can see fairly, you know -- well, relatively high CFUs for the incoming pre-evis and -- but really low levels, you know, less than 200, for -- well, less than 50 for colony forming units post-chill.

Salmonella isolations. We routinely monitor pre-evis and post-chill just to see where we're at. This depicts the dates just prior to Katrina hitting, and you can see where she hit. That's where that dip on the pre-chill is, where -- the pre-evis numbers.

You see that dip and where it goes down to zero. That's because no samples were being submitted.

And what you can see is, you know, we have a fairly -- it fluctuates the incoming, what we're seeing, but fairly -- we were able to maintain a fairly low level post-chill.

This is another one of our plants. And, again, you can see the dip when Katrina hit. And this

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is another plant in which -- they did quite well.

Now, as a company, we don't like to just depend on the positive/negative aspects of looking just at incidence levels. We also like -- we also monitor MPNs. And we don't do this for every sample, because this is quite arduous.

We use an AOAC-approved method. It's a nine-tube method. So each one of these samples represents nine Salmonella isolations, which means over a thousand. But we use this to try to assess what kinds of numbers we're having coming out of the chiller.

Now, I know there are a lot more people in here that know a lot more about MPNs. So if I say something incorrect, please correct me.

But basically, my microbiologist -- she's not going to report anything as sterile without autoclaving or irradiating. So on this first column, the orange one, the less than two basically states that they couldn't find anything. The next column is colony of a most probable number of one to ten, the next one greater than ten to 100. The next one, the

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3.7, is greater than 100 to -- it's from 100 to 1,000.

And we had zero MPNs greater than 1,000.

So with that, in conclusion, we feel that we were able to maintain process control and produce a quality product for our customers.

(Applause.)

DR. ENGELJOHN: Thank you very much, Marty. And that was very helpful information. I know we -- as an Agency, we're particularly interested in what happened during the hurricane. And it's something we all have to be prepared for in the future.

Our next presenter is going to talk about processing and sanitation issues unique to very small establishments. We have with us Dr. Patricia Curtis; she's a professor and Director of Poultry Product Safety and Quality at Auburn University. She has worked with the International HACCP Alliance in terms of presenting a course, and she has a special interest in distance learning.

With that, she has received her PhD and master's of science degrees from Texas A&M University.

We are pleased to have with us Dr. Patricia Curtis.

DR. CURTIS: I'm going to talk to you very briefly about the differences in working with small plants or very small plants and the large plants and some of the challenges that they face in trying to meet Salmonella standards. If you look at the very small plants, most of those plants -- when we're talking about poultry, there is a very, very small number of plants that actually do the slaughter.

Most of those raw poultry plants are purchasing product from the larger plants. But I want to tell you they have no clue as to -- they think, you know, We're getting it from a HACCP-approved plant, so this product is fine. They don't have a clue if you're in the low level, Category I to Category III, of the contamination level, and they don't have a lot of clue in many cases about how to control that when it comes into their operation, because they don't have the scientific background.

And what they -- when they got into business, they got into business more as an art than a science; their family may have been in business, and

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that's the way that they've always done things. So it creates a little bit of a concern when you're trying to teach some of the concepts. They're relying very heavily on those inspectors that were in the plant to tell them what they needed to do and how they needed to do it.

Yes, there's exceptions to all of these.

And my view may be a little biased, and I'll state this up front. I work primarily with large plants, and I only get called in to the very small plants when there's a problem. Okay? So I'll say that up front.

And if any of you are from very small plants and this doesn't apply to you, I apologize, but that's the reason I wanted to say up front that I deal mostly with the plants that are having the problems.

So what I see is that they have that real lack of the scientific background, which causes issues, because, as Dr. Russell was saying, when you're trying to do cleaning and sanitizing -- I've been in many of the plants who are going, Well, I want a cleaner and sanitizer combined; why can't I do that and save a step. And it's really hard to make them

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understand.

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And if they start cleaning, they will take -- and they'll get -- Oh, well, if I can clean at a certain temperature, I can double that temperature it'll even be better Ι double and or can the concentration and it'll be better. And obviously, we know that that's not true. Ιf you raise temperature too high even during pre-cleaning, you'll start denaturing the protein on there protecting the bacteria, and you've got other problems.

So they depend a lot on the sales people that come through, because they get very, very frustrated now that the FSIS is not providing them with some of the information on how they need to do things.

Often times, they create wider а product. They will purchase products in, and then be simply portion-control they may sizinq handling that product in some manner. Or they may be creating ten different products or 15 different products from that raw product that's coming in.

Most of their operations are manual, so

there's more opportunity for that cross-contamination.

And I've seen many, many of the processors,

particularly the very small, where it's a mom-and-pop

operation or there's just one. They want to know,

Well, why can't I go down to Wal-Mart and buy my

supplies?

You know, they want to go down and get Clorox for their sanitizers; they don't understand the difference between household cleaners and some of the commercial cleaners that are available. They don't know which cleaners work best in certain situations. And other than just seeing the scale forming from the hard water, many times, they don't understand the impact of what that hard water will have on anything from the sanitizers to even how their product may react — that they're making.

They don't have a clue about the pHs and the effectiveness of the pH so that your sanitizers are effective. And they've many times [inaudible due to failure of in-house PA system] the biofilms.

Since they have fewer employees, it's very difficult for this group of people to be reached for

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training programs. And even when they are able to come in and do training programs -- I know that when I was in North Carolina, we spent one summer -- there was a whole group of us extension people that went out and did training for the very small plants.

And they couldn't be away from their facility for three days in a row because there was nobody back home to keep the business going. So we would have to go in and do one day a week for three weeks, or something like that, for HACCP training.

So the training assistance has to be more creative for the small plants, but the thing that's even more important is being able to follow up with that person after they've attended the training, because they often get back home and they go, Well, that situation really didn't work for me, because I have something a little bit different. And then they have to find someone who can answer those questions for them.

And even more recently, the very small plants that I've seen that are trying to start up an operation -- they don't even have a clue of where to

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turn. They don't have an idea of where they need to find -- Well, you said that I just can't put back that these are germs, you know. And I'm going, No; you have to be more specific. "Well, I don't know what's on there; how do I find out what's on there.

So the question of just being able to identify the hazards becomes a very, very big issue. And the smallest of the plants are usually not members of associations. So they don't have that backing of the associations that many of the larger plants to do provide the information that will help them as they're trying to create a HACCP plan. And they certainly don't have the expertise.

A lot of them will have a consultant come in and develop their HACCP plan, which is fine until they have the first deviation or the first problem that's associated with their HACCP plan. And then they really don't know how to fix that problem.

So -- and as FSIS comes back each year and looks a little bit more critically at their HACCP plan, they're not prepared for that. They don't really have a clue unless their inspector in their

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plant is really, really helpful to them and provides them some scientific expertise. And we're seeing a movement away from that supplying of information. So those people are having a much more difficult time.

And you might say, Well, look at all the information on the internet; there's all kinds of things out there. But let me tell you. I teach HACCP classes, and I have a mixture of large-plant and small-plant people in my classes. And recently, I spent a lot of time in introductory HACCP.

all of you that have done the HACCP classes know you have to write a HACCP plan. And you spend a lot of time writing all that stuff down and then putting it on charts to share with the class. So I thought, Okay, this is great; I'll just buy some tablet notebooks, and I'll pass them out, and the groups can use these; and, you know, we can go in and we can look at some of the FSIS web pages and some other resource web pages.

Well, I found that the people from the larger plants are very comfortable with that. They're used to using technology. And the people -- and I

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can't say this is 100-percent true, but I've found that many more of those very small-plant people will just kind of scoot that tablet under the table and get out their notebooks and keep writing, because they're uncomfortable even with that technology.

So we can't assume that just because we post it on the internet, that's the way that we're going to reach everybody. I mean that's a good way to share information, but it's not particularly the best way for all the people to obtain the information that they need.

The -- also, the people who are from these very small plants -- they're usually making very unique products in many cases. And they may be making chicken jerky or something. I mean, you know, it's amazing, some of the products that they come up with.

Well, where do you go even when they call you or find somebody that has some experience in some micro-background or some processing background and their product is so unique that you're going, Well, I don't know anybody that has ever done any research in that area. And they're at a loss, because they still

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are expected to come up with validation information.

Hopefully, there's a safe harbor that you can point
them to, but that's not even always the case.

So this validation information that's required by small plants is a very, very tough task for many of those. So they had -- and they have no idea how to run validation studies in their own plant and, if they did, they didn't have the equipment to do it.

I was amazed a few years ago in North Carolina. As I said, when I was doing this training. We were talking about calibrating different kinds of thermometers. And can you believe there were some very small plants that didn't even own a thermometer at that time, which is kind of scary?

But you have to -- they have certain ways, and I think that we're past that and many of the plants are moving in that direction. But just the expertise of how to handle certain equipment, how you plan out validation experiments that would really work for those operations, is just beyond their expectations. So they need lots of help, but I'm not

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sure how we get that help to them.

The other thing that you run into often times in very small plant operations is that you have only one or two people in the plant. So you have the same people monitoring HACCP that are doing the reviews.

And if any of you have ever written things and you've gone back to check your own writing, it's very difficult to find your own mistakes. It's a lot harder to go back and proofread something that you did than to proofread something somebody else did, because you keep reading things in there that -- not intentionally that weren't there -- but it's just harder to catch.

So it makes it more difficult for small plants when they go back and are trying to do preshipment sign-offs and things to make sure that everything gets done.

So they definitely are much more dependent on the in-plant inspectors for -- to provide updates on regulatory information and to provide some of that scientific knowledge that they may need in the

processing of their products.

I don't know exactly how's the best way to go about reaching small plants to try to teach them how to do validation studies, but I think as we look for more and more validation requirements onto the small plants, there's going to have to be some material that is provided to them to help them figure out, What is a validation study.

I know that even working with large plants, you wind up with people going, Well, what exactly are the inspectors looking for when they're talking about validation? Well, imagine that even if you're able to tell them, then when you go to the very small plants, they have no clue what you're talking about. And they have no way of getting the scientific magazines to provide some of it or even finding the people with the right expertise to help them design some way in order to do their validation.

So basically, I guess the thing that I want to summarize here from this presentation is that there's a lot of very small plants out there and they're producing a lot of very good product. But

they have a lack of scientific knowledge to tell you why that product is safe. That's not to say their products are not safe. I don't want to imply that at all, because many of them -- what they're doing is a very good process, but they can't explain to you why that's a good process.

And in a lot of the presentations that we heard earlier today, we were seeing that there's being a request for more documentation on why we're doing the things that we're doing. And so I think that that is a key area that we need to pay particular attention to for the very small plants.

the sanitation And from area, those sanitation decisions, hopefully, the sales men that are producing and selling the sanitizers and the providing detergents good credible are some information to these small plants and visit them more and help the small plants understand why they need to use more commercial products than trying to rely on some of those household products.

And then from the processing expertise, I think it would help a lot if some of the inspectors

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would emphasize to these small plant processors the importance of continuing education, because I think they truly don't understand what they would gain. They see it as time that is being taken away from their production, from their profits and from their livelihood, but they don't understand that if they don't participate in some of these trainings, it is going to cause them more problems in the future.

With that, basically, I'd like to say -conclude with that the small plants are very different
than the large plants in the help that they need. I
think that they can produce some of the product that
is equally as good and safe as large plants', but they
need a lot more assistance from the federal government
than do the large plants.

(Applause.)

DR. ENGELJOHN: Thank you very much, Dr. Curtis. And we'll certainly take that as a challenge to the Agency to make sure that we focus on getting the proper type of information to the audience that we have to serve. And we do think that the large plants have a lot of that expertise that they could share, as

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1	well. And so we certainly will be looking to see what
2	partnerships we can form there to get the right kind
3	of information out.
4	We're now going to hear something about
5	the turkey industry. We have with us Michael Rybolt,
6	who is the Manager of Scientific and Technical Affairs
7	at the National Turkey Federation. Michael has his
8	background with his work at Mississippi State
9	University.
10	So we're glad to have you here, Michael.
11	MR. RYBOLT: Thank you, Dr. Engeljohn.
12	Yes, you do have another Bulldog in the
13	house.
14	What they asked me to come and talk to you
15	about today are some of the food safety best
16	practices.
17	(Pause.)
18	MR. RYBOLT: Oh, I'm sorry. You can't
19	hear me? I thought everybody could hear me. I'll
20	talk about the food safety best practices that the
21	turkey industry has.
22	A little bit about the National Turkey

Federation. Everybody knows we're a national trade association based in D. C. We are the only trade association representing the turkey industry and its allied industries exclusively.

Food safety is considered a high priority for the turkey industry and for the National Turkey Federation. To prove that, several years ago, the turkey industry developed the "Food Safety Best Management Practices for the Production of Turkeys." And this is more of your own farm live production BMPs. It's in its second edition now, and it was updated recently, in 2000.

The process is actually pretty intensive.

It starts out at your foundation and multiplier breeders, and it goes all the way through to the feed manufacturing and delivery. It covers your live haul, and it also covers your meat bird production and growout section.

Here's a snapshot of the whole program itself, and it has a flow diagram of each one of the processes and has a control point that you should evaluate in each one of those steps. And it also has

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a monitoring and feedback component.

So it follows the HACCP-like program and is fairly intensive, and it covers everything from disease diagnostics, disease prophylaxis, whole quality, [inaudible due to failure of in-house PA system] servicing, biosecurity, which is big, vector control, drinking water and sanitation. So it covers a lot of the different live production components that you would -- that are important for food safety.

We also have developed our ground turkey good manufacturing practices; this was developed by the Tech & Reg Committee. The ground turkey GMPs cover everything from receiving, storage, tempering, grinding, packaging, finished product storage and distribution. It doesn't necessarily have the HACCP flow chart that the live production or the production BMPs have, but it does cover all the different components for producing ground turkey.

We also have developed -- we contracted with two researchers, Dr. David Caldwell and Audrey McElroy. David is at Texas A&M, and Audrey is a Virginia Tech Hokie now. We contracted with them to

do a chiller study, and it's just a snapshot to see what was going on in five different establishments to see what sort of best practices they use within their chiller systems and [inaudible due to failure of inhouse PA system] for the Salmonella and Campylobacter levels to see what works best.

And I'm not going to go through the whole study, but I just want to talk about -- the objective the emergent chilling and best was to assess management practices and their effects microbial quality of carcasses, whole turkeys, coming out of the chiller, and then to develop the best recommendations for practices or the industry follow.

Some of the conclusions that the group found were that you need to maintain your total chlorine at 15 to 25 ppm. That there is a discrepancy [inaudible due to failure of in-house PA system] some of the other literature that's published, but this is what was going on in these particular plants at that time.

There were different types of chlorination

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being used, and the type of chlorination in this particular study didn't matter; what really mattered was that you maintained your pH appropriately so that you have the effective form of chlorine available.

And what -- the main part of the BMPs was that the chiller can serve as an effective part of your overall pathogen control program as long as you're managing it appropriately.

The next step for us is to do a process control mapping. I think you'll hear some other talks later on about that. But the NTF Micro group decided that we needed to do this within the turkey industry, and we decided that we would go to different processing points and measure for different organisms to see what was going on at that particular process.

I tried to get them to do every single process, but, of course, money comes into play and you can't do that. But we did settle on some of the key points or key processes within the slaughter section of the plants: Before scald, after scald, after the pickers, at rehang, pre-crop, post-/pre-chill rinse or antimicrobial application and then, of course, post-

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chill.

The plants are going to take five wing samples at each one of those locations, and the samples are going to be pooled. And we're going to test for Salmonella-positive or -negative at this point in time. We're also going to quantify generic E. coli and do APCs, as well. And the pilot is going on right now.

We have a handful of plants that are going through this protocol at this point, and we're planning on doing this for about 30 processing days so that we can get enough data and look at the data to determine, Where do we need to concentrate? Do we need to concentrate only at, you know, rehang? Do we need to concentrate only at, you know, pre-chill? And then we'll go back and launch this industry wide to other plants and let them start sending in data.

Again, once we're done with the pilots, we'll take the data and analyze it and present it to the entire committee and industry and try to get everybody to participate with shared data to NTF so that we can collect all this data, analyze it and

discuss it and talk about what practices the companies are using, what works best and what doesn't work best to try to develop some BMPs there and to build on what we already have in place at this point in time.

We have, you know, our live side. And we have our ground BMPs. And now we're going to build onto those with the process mapping.

I do have just a snapshot of data. I don't have enough to really show this, but I did -- I thought it was interesting to show what we do have at this point in time. Like I said, we're just in the very beginning of this; we only have a few days of results in.

But here you have the APC and generic *E.*coli results from some of the plants. And this is -
the red is the APC, and the yellow is the *E. coli*.

And you can see a reduction in the process from prescald all the way down to post-chill.

So we are showing process reduction. I don't have any Salmonella data. I know this is a Salmonella meeting, but I don't have anything to show you at this point, and I do apologize for that. But I

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1 guess this is -- Dr. Engeljohn hasn't shown me a sign 2 yet. So we're doing good. just to emphasize, The -- again, this 3 4 mapping study is in its infancy. We're just getting started, and we're only a few days into this. 5 But the goal of all this is to build onto 6 7 the already -- the programs that we already have in place within the industry and to further develop our 8 9 best management practices for the production 10 turkeys. And, you know, this is going to help in the hurdle approach. You know, there's no one silver 11 bullet, unfortunately. So it's going to build on the 12 13 programs that we already have in place and, hopefully, help decrease the Salmonella. 14 15 (Applause.) 16 DR. ENGELJOHN: Thank you, Michael, for that presentation on turkeys. And the fact that you 17 don't have information on Salmonella -- did you know 18 19 we are going to start [inaudible due to failure of in-20 house PA system] turkeys? MR. RYBOLT: I didn't know that. 21

Well, we are.

DR. ENGELJOHN:

# (Laughter.)

DR. ENGELJOHN: Well, we now are [inaudible due to failure of in-house PA system] check with the people on the phone. But if you would, as we did this morning, if you have any questions of any of the panelists from this afternoon's session, please come up to the microphone and identify your name and association and then ask your question.

MR. WALTHER: Hi. I'm Jeff Walther from Mount Air Farms, and I have a question, I think, for Stan Bailey and maybe a little bit for Mike.

And that's, Would you think I was crazy to think that we could -- we're trying to save our birds during the live-haul process by a technique I learned, you know, a million years ago. And we go out there, and we drench the birds coming out of a house on a 90-degree day, and the birds come out of the house white and they arrive at the plant tan.

And, you know, our *Salmonella* -- we do a fairly expanded sampling for *Salmonella*. It's just presence or absence. But I expected to see some tremendous numbers in the summer time when we did

1	that, and I haven't. And is it because I just I
2	should be looking at colony forming units, or, you
3	know, am I just missing it, or have you got any idea?
4	Is that just a is that a bad practice?
5	DR. BAILEY: Can you hear me?
6	(Pause.)
7	DR. BAILEY: Can you hear me now?
8	VOICES: Yes.
9	DR. BAILEY: Maybe I could get a job on
10	the Verizon commercial.
11	(Laughter.)
12	DR. BAILEY: It's a mixed bag. The good
13	thing is you're doing that for a reason, and that
14	keeps the birds from getting stressed. And anything
15	that keeps the birds from getting stressed is going to
16	keep them healthier and keep their intestinal tract
17	more intact. And so that's a good thing.
18	Any time you're adding moisture to
19	anything to do with Salmonella, it makes it worse.
20	The fact that you're not seeing elevated levels of
21	Salmonella because of the process probably indicates

that you've got a very low level coming in.

22

That

1	would be my biggest guess.
2	MR. WALTHER: We do not have a low level
3	coming in. I'm sorry.
4	DR. BAILEY: But well, then if you've
5	got a very high level, then it doesn't have that much
6	room to go up. So I don't
7	(Laughter.)
8	MR. WALTHER: Okay. That's great.
9	(Laughter.)
10	DR. BAILEY: Other questions?
11	(Pause.)
12	DR. ENGELJOHN: If we could ask the
13	operator if there's anyone on the line that may have a
14	question?
15	(Pause.)
16	DR. ENGELJOHN: Does anyone else in the
17	audience have something?
18	MR. SANCHEZ: Marcos Sanchez from Texas
19	A&M. I just want to follow up on the same question he
20	just had on the humidity levels and the increase in
21	Salmonella.
22	I know that I mean when I did some work,

too -- I mean the drier air we had at -- we had less numbers, too, but we still were able to recover some of it. And I know that there were some publications recently or some findings recently that -- actually, when you have humid environments, you have more competition, too. That competition keeps those numbers down -- of Salmonella. And so I was just wanting to know your perspective on that, too.

DR. BAILEY: Well, it's true that if you have more moisture, everything's going to grow better. But everything that I've personally worked with and all of the literature that I've seen in the past, from Ed Mallenson's [phonetic] work at Maryland to some of the industry data that has been looked at internally by some of the companies, would suggest that your biggest problem with Salmonella is always moisture.

Anything you can do to keep an environment dry is probably the greatest thing you can do to reduce the prevalence of *Salmonella* in your chicken houses. Surely, any -- you know, if you increase the good bacteria or your competitive flora, then you'll be making the situation better, but I don't think it's

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1	worth the trade-off of worrying about doing that via
2	moisture management.
3	If you want to increase the good bacteria,
4	there are some other ways. You can look at some
5	prebotics or, if you can find it, an approved
6	competitive exclusion-type product. But I don't think
7	I would be attempting to do that with moisture
8	management.
9	DR. ENGELJOHN: While the gentleman is
10	coming up to ask a question, if the panelists have
11	anything you want to add or you to anyone's
12	presentation or anything else, also, raise your hand.
13	MR. BENSON: Hi. Mike Benson from Jennie-
14	0.
15	Dr. Bailey, I was intrigued by your
16	measurement of Salmonella in the houses with the fly
17	strips. In Minnesota, we don't have many flies in the
18	winter time. I was wondering if you have any
19	suggestions for other measurements on what the load is
20	coming into our processing plants other ways of
21	measuring what the live load is.

DR. BAILEY: Well, there's a number of

ways people go about that. As you -- and you can do fecal or cecal grab samples or litter samples. I think probably the most commonly used sampling methods are either drag swabs or using, like they do in Scandinavia, kind of a foot-sock which is basically an oversized wristband that you put over the bottom of your feet and walk around the house with. That has proved to be a particularly effective measure.

There -- each of these methods have some advantages and disadvantages. I would suggest that, whatever method you use, you use it consistently, it will -- you will able be to historical perspective of what you're doing whatever the positives or negatives of the method you're using will cancel itself out over time and you'll be able to see relatively what you're doing.

I personally -- some of the companies I know also do antimicrobial testing on the birds a week or so before they go to the slaughter plant. And they kill, say, three birds in the plant and look at the fat pads for antibiotics. And they take those birds and look at the cecal content of those birds. So

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1 that's another potential alternative. As I say, there's a number of different 2 ways you can do it, and I would just be consistent. 3 4 Decide what you're going to do, and use the same thing all the time. 5 DR. RUSSELL: Yes. I'd like to add to 6 7 that. I saw a paper I edited one time where they compared the use of drag swabs to just using those 8 9 surgeons' booties that you put over your feet and 10 walking around the house. And Ι think they recovered -- 2 to 3 percent of the samples 11 positive for Salmonella using the drag swabs and maybe 12 13 11 percent with the surgeons' booties. And they concluded that that was a much more effective way to 14 15 do it. So just sort of to add to that. 16 MR. COUGHLIN: I'm Michael Coughlin from 17 Johnson Diversity. And this question is for Dr. Bailey. 18 19 the prevalence Can you speak to of Salmonella within the feather follicle pre- and post-20 picking, if in fact that microbiology has been done? 21

DR. BAILEY:

22

I know that in all the 30

years -- 32 years I've been working, people say that the Salmonella gets driven into the follicles. But as I'm sitting here really thinking about it, I can't honestly say that I personally know of a research paper that has demonstrated that. But maybe somebody else in the audience -- I think it probably is true, but I don't know.

Mark?

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I was involved in some work DR. BERRANG: where we were -- a graduate student developed a method visualize Campylobacter to live and dead --Campylobacter, now -- live and dead cells at different heights down into a feather follicle. And what she found was that she was able to find some viable Campylobacter deep down in the feather follicle, but the numbers were much, much higher on the upper surface of the skin.

And most of what I see in the literature -- and Dr. Buhr might have something to say about this, but most of it seems to be anecdotal, you know: "The feather follicles open up, and then they close, and, boy, I'll bet you that's where they are."

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1	But what I see with the <i>Campylobacter</i> data suggests
2	that the upper surface of the skin is really where we
3	find more numbers of viable <i>Campylobacter</i> , anyway.
4	MR. COUGHLIN: Right. And this is what
5	I've noticed, too: That there's very little
6	information in the literature. I was wondering if my
7	search was inadequate.
8	But I'm also wondering if the types of
9	populations in the feather follicle would be dependent
10	upon the oxygen content, like a biofilm, maybe
11	anaerobic at the bottom and aerobic at the top.
12	DR. BERRANG: That was our thinking with
13	Campylobacter. That you know, since it's a
14	microaerophilic organism, it might really be happier
15	down in there. But, again, it didn't seem to be borne
16	out by the data in that particular study.
17	DR. BAILEY: I would think that would also
18	be very dependent on when you chose to look, because
19	that feather follicle until the feather is pulled out
20	is going to be full.
21	So if you went in, you know, very soon
22	after that, the atmosphere or the lack of oxygen or

1	whatever the micro-environment is there would not have
2	had a whole long time to select for a given
3	population. But if it was, you know, a week or two
4	later after processing, you might see a different
5	situation then than you would immediately after
6	processing.
7	DR. ENGELJOHN: How about on the phone?
8	Did we hear anything?
9	(Pause.)
10	DR. ENGELJOHN: No one from the audience
11	has anything they want to follow up on?
12	(Pause.)
13	DR. ENGELJOHN: Well, with that, then
14	let's take a break. We'll come back in at 2:45.
15	(Whereupon, a short recess was taken.)
16	DR. ENGELJOHN: We'll go ahead with the
17	afternoon session. We get to leave early if we get
18	all the presentations done. So
19	(Pause.)
20	DR. ENGELJOHN: Well, welcome back to the
21	afternoon session. I notice that there are a few
22	empty seats not that you're not coming back to the

presentation and that people might be standing outside.

But I do want to remind everyone that if in fact you're not going to be back tomorrow or you're going to leave early today -- we do have evaluation forms out on the table, and we would like you to fill them out to give us some input as to how we can enhance the future public meetings that we have. So please stop by, pick one up and fill it out.

Our next presenter is Dr. John Cason, who is an animal physiologist scientist with the Agricultural Research Service at USDA. He has worked for ARS for 16 years, with most of his research focusing on poultry processing. He has his degrees from the University of Georgia.

We're pleased to have John Cason with us. Thank you.

(Applause.)

DR. CASON: Thank you. Good afternoon to everyone. I'm glad to be here, and I want to thank FSIS for the invitation to make a presentation. Basically, I'm going to be talking about factors that

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limit the effectiveness of antimicrobial treatments.

And in case you can't read this in the back of the room, this is a "Dilbert" cartoon, and he's in his cubicle making a PowerPoint presentation on his computer. And he says to himself, "I no longer feel the need to change the real world as long as I can change these bullet points." And I'm sure everyone here has done a PowerPoint presentation and had that feeling before. To avoid that in my case, I want to start with a photograph of a chicken to keep me connected with reality.

(Laughter.)

DR. CASON: Okay. So now we're all set.

So the question I want to ask is, Why can't we remove or kill all of the bacteria on poultry carcasses? And of course, outside of cooking and irradiation, there really is no way to either wash off the bacteria or kill the ones that are on the carcass or to do that to all of them, anyway.

And this is an anecdotal sort of graph here from a paper by Huda Lillard from about 25 years ago -- well, my math might be off a little bit -- 20

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years. And this is -- the three lines here show carcasses that were hand-picked before being scalded on the top line. And the middle line is carcasses after scalding and picking, and the bottom line is a carcass that has been eviscerated and is about ready to go into the chiller.

And these are numbers of aerobic bacteria recovered in consecutive carcass rinses of the same carcass. And you can see that the higher levels -- when you have a dirty carcass, there is a bit of a curve to the line, but as you get down through the plant and the carcasses are cleaner, the lines are relatively flat. So that -- each consecutive rinse washes off about the same number of bacteria.

And this is the results for the same carcasses, showing the number of enterobacteriaceae removed from carcasses. And of course, this family of bacteria has several pathogens that are of interest to us.

And I want to direct your attention to the bottom line here, which is the carcasses just about ready to go into the chiller. And if you look at the

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level there -- I don't know if we have a pointer up here, and, if we did, I wouldn't know which screen to point at. If you look at the tenth rinse -- there's maybe 4.7 logs of enterobacteriaceae that were removed from that carcass in the rinse.

And I did a little calculation down to the 40th -- I'll just point at my screen. I did a little calculation down to the 40th rinse. And from ten to 40, about a million enterobacteriaceae were removed from that carcass. But a carcass rinse down there at Number 40 was unable to detect that difference. And so whenever we do a sample, we have to keep in mind that our interpretation has to be based on how that sample is taken.

Besides that study that I just showed you, there have been five studies that I could find where they did consecutive rinses on enough carcasses to do some statistical analysis. And the table here shows the kind of bacteria that they were checking for and the number of times the carcasses were rinsed, and that's a total of 32 comparisons between two consecutive rinses.

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And of those 32 comparisons, there was only one report of a significant difference, and that was in aerobic bacteria. Now, I suspect that if you did enough carcasses, there would be many more significant differences found, so I think the problem is not that the number doesn't decline slightly with each successive rinse, but that the level of variation is so great that it's just about impossible to find a significant difference.

So why do bacteria persist on carcasses so that we can rinse carcasses 40 times and still get off the same number of enterobacteriaceae? And I'm going to talk about some reasons that have been given in the scientific literature. And I was really pleased to hear some of the earlier discussion.

Number One: Bacteria in feather follicles. It was suggested as far as I know first by a British microbiologist in the 1960s.

And there was a similar suggestion in a study of turkey carcasses by Avens and Miller where they couldn't understand why they were able to keep on getting bacteria off the carcasses, and they said that

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maybe some of the bacteria are subcutaneous. The feather follicle idea has been repeated dozens of times; it's in some pretty high-powered publications by national committees and investigatory bodies that should know all about things.

And this is an electron micrograph that I found in a paper by Thomas and McMeekin from 1984. And this is chicken breast skin before water immersion. And that little bar down in the lower left is 30 microns. The usual size range for *Salmonella* is about 1-by-3 microns. It's a little rod.

And so end to end, you could probably put about ten Salmonella bacteria along that black bar. And so if you have the idea now of the size of a Salmonella, you can see lots of places down in those little crevices where they could hide.

And the next photograph shows the chicken breast skin after it has been soaked in water for 30 minutes. And you see the skin takes up water and the crevices become much deeper. And now there are many more places for the bacteria to hide. And I'm not suggesting that they hide consciously, but it's an

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artifact of -- well, maybe they do, since they can talk to each other, as somebody else said.

(Laughter.)

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DR. CASON: And SO Ι did some for calculations. standard formula There's а calculating the surface area of a carcass. And so I did a 2-kilogram carcass. That's а 4.4-pound eviscerated carcass going in the chiller. It would take about 2-1/2 million of these photographs to show you the entire surface of the skin of that carcass.

And on the way over here this morning, I came with some really smart people. So I was asking them how many feathers a chicken has. And the estimates that we have seen in the literature -- this thing is turning on and off up here.

The estimates in the literature range from 5,000 to 9,000, so I used 10,000 as a convenient number. And that's really a high estimate. I would have to show you on average 250 of these slides for us to see one follicle. Now, granted, some of the follicles are really thick -- the flight feathers and the tail. It would take about four of these high and

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four wide to show one entire follicle on some of those larger ones.

And so even though I could argue that just in terms of numbers follicles should not really be very important, it's more important to do some experiments and ask the chickens.

And for the last four or five years at the Russell Center, Jeff Buhr has been maintaining a flock of featherless chickens. Now, this was a spontaneous mutation that arose in a flock somewhere, and Jeff got someone to send him some eggs.

And this is a single recessive gene. And if both parents are heterozygous, 25 percent of the chicks are hatched without feathers and -- they're not entirely without feathers. I think you can see there are a few little tufts of feathers here and there. But we've been using this as an experimental model to see what differences it makes if a chicken has feathers or it does not. And of course, if they don't have feathers, they don't have follicles.

So we've done at least three experiments, and there are probably several more in the pipeline.

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And basically, our research shows that follicles don't make any difference.

There may be a short in this mic -- or something.

We tested de-feathered and chilled and then carcasses that had been stored for a week for lots of different kinds of bacteria. And basically, the chickens without feather follicles are not any different.

In agriculture, I don't know if we have urban legends; we probably have rural legends.

(Laughter.)

DR. CASON: And so I think that feather follicles don't make any difference at all. Of the studies Ι that know about where they published photographs of Salmonella and Campylobacter bacteria in feather follicles, in both cases, the follicles soaked for several were hours concentrated suspension with millions of bacteria. And so I think that neither of those studies with the photographs really reflects the real world.

Physical attachment of the bacteria to the

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carcasses has been suggested. Biofilms have been mentioned, but a traditional biofilm is a mat of bacteria on a physical surface growing out into a liquid. And so I don't think that biofilms really match up with poultry.

Physical attachment to the surface to me doesn't seem to be terribly convincing even though I know it happens. And the main reason is that the surface changes so much while the carcass is going through processing. The cuticle is moved in most plants, and part of the epidermis below that, but, yet, the bacteria still persist on the carcass.

Some people also suggest there might be a clumping effect and that the clumps are there and every time you rinse, one or two cells break off the clump, and so the numbers don't change. That may be possible, although, I think, in a lot of photographs, there seem to be relatively small numbers of bacteria in groups on the surface, not just big clumps.

Surface chemistry has been suggested. The presence of fats and oils can protect the bacteria. The pillae and flagella -- the bacteria -- have

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sections that recognize chemical receptors on the surface of the skin; however, when rinses have been done with surfactants and detergents and enzymes such as lypasis and proteinasis, the increase of numbers of bacteria that come off the carcasses is really very modest.

So I don't think that surface chemistry -even though this may have some effect, I don't think
it really explains why we can get so many bacteria off
of carcasses.

I think surface physics is one of the most convincing explanations. People talk about surface tension in the water, the formation of water layers around the carcass, the lack of ability to get shear forces right up on the skin of the carcass to remove bacteria. And there's a possibility that you might get into an exchange-in-equilibrium situation where as many bacteria are coming back onto the carcass as are coming off as you're doing your rinse.

And I have a cartoon here, and, luckily, it's showing up well on the big screens if not on the little screen. I made this up; this is sort of a

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"Dilbert" sort of thing.

And what I have here is -- you can see the yellow chicken skin. And there are some crevices with some bacteria in them, and there's a water layer once the carcass starts through processing. It has been dipped in water and the scalder, and it has been sprayed several times, and there's a water layer that can't be removed; it can be evaporated, but you can take the towel, and you can't get that off.

Okay. Now, in this one, this has more blue up above it; you've added the water to do a carcass rinse. And so you start to shake, and the arrows show that there's a good velocity of water out in the rinse, but as you get closer to the surface, there's less and less velocity. And in fact, by the time you get down to those bacteria in those little crevices, they don't even know that somebody's doing a carcass rinse out there.

Now, several physical things have been done to carcasses to try to increase their removal or the elimination of the bacteria, including bubbling. Bubbling does increase the removal of bacteria from

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carcasses; however, it has a side-effect. In a lot of applications, it increases water take-up by the carcasses. So there might be some problems there.

It does kill bacteria. However, ultrasound treatments are used by the people who study biofilms to break up the biofilms so that they can count individual cells, and so ultrasound might be a way where you can kill lots of bacteria and then get a worse result when you do your carcass rinse.

Brushing has been tried. I think it recontaminates the carcass about as fast as it removes the bacteria. And I want to show you some preliminary results of some scraping experiments that we've done at Russell just recently.

Here's a cartoon again showing that water layer that can't be removed. And we scraped with about a 2-1/2-inch piece of stainless steel. And you can see a lot of material -- perhaps fat and liquid and bits of the cuticle -- can be removed. And there were large numbers of bacteria in this material on the little blades, but when we tested the carcasses by a

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whole-carcass rinse after chilling, there were no differences.

And this is a little cartoon that's supposed to represent the stainless steel scraper blade going over the surface. And the pressure flattens out everything on the surface and closes up the crevices so that bacteria down in the crevices are completely protected. It may remove some from the surface, but, again, it does not reach all of the bacteria on the surface.

And so I went over some of the things here that have been suggested as the reasons for why you can't get all the bacteria off of the carcasses. I think a lot of research has been done on looking for a silver bullet, an antimicrobial chemical.

But the trouble is not that we haven't found the right silver bullet; it's that we can't get the antimicrobial chemical into contact with the bacteria that are in those crevices on the carcasses.

So I think that new approaches are needed to improve the efficacy of antimicrobial treatments.

And I just got the two-minute warning. So

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we're actually running ahead of time, I think. Thank you.

(Applause.)

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DR. ENGELJOHN: Thank you very much for that presentation.

Our next speaker is Dr. Jeff Buhr; he's an animal physiologist scientist with the USDA's Agricultural Research Service. He has his degrees in avian sciences and a PhD in veterinary anatomy from the University of California, Davis. His research for the past nine years has focused on broiler processing, specifically feed withdrawal, de-feathering, removal and bacterial decontamination, and currently is looking at environmental lighting programs and their effect on colonization by pathogens.

DR. BUHR: Well, thank you very much. Thanks for the invitation.

I've been asked to talk specifically about scalding, de-feathering and rehang. And my topic will mainly stick with <code>Salmonella</code>, but we also have to talk about <code>Campylobacter</code>, because I don't have data on everything. And I'm not going to talk about any

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silver bullets or antimicrobials. And at the end, we'll have a little bit with the featherless chickens.

If you look at the relative levels -- and this is something to remember -- when we're talking counts or colony-forming units, how many bacteria, this is easy to do on a whole-carcass rinse on a feathered bird or when it's in a dump coop shoot. However, when we're talking incidence, if we didn't enrich samples post-chill, we're not going to find very much Salmonella. So we need to keep -- in "incidence" or "prevalence," we're talking about percentage here.

If we look through processing and the presentations we've had this morning, in general, scalding is going to decrease numbers, and it may or may not decrease incidence. And I'll show you some data to support that.

Everybody agrees de-feathering is going to increase numbers, increase incidence, specifically related to the decrease we just had with scalding. Rehang is pretty neutral. Most of it's automatic. It's not going to really increase numbers that you

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can't wash off. It may increase incidence if you don't wash off.

Here are some data from Mark Berrang, looking at Campylobacter. Now, this one's different from the Salmonella we had this morning. This is individual flocks through a processing plant. And if you look at the pink one on the top, you'll see that it tends to be the highest one as we go from pre-scald to post-chill. But the important thing to remember, also, is as we go into post-chill, everybody ends up down there in the same bottom part about one log, whole-carcass rinse, except for the pink one.

So it's important when you're sampling a processing plant to make sure you're sampling the same flock as it goes through the plant. Otherwise, you may get increases or decreases that are flock related, not necessarily plant related.

This one talks about scalding: Tripletank, counter-flow, S-shaped. What does scalding do? Scalding, as we've heard, loosens feathers. Ιt removes feces, litter from the process in house, and, bacteria. also, Ιt also enables us to loosen

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1 feathers, and it's also going to take off most of the epidermis. 2 What are the positive aspects? The good 3 4 thing -- when I say, "Positive," I mean good, not positive meaning a positive sample. 5 There's some physical equipment advantages with processing. 6 7 Multiple tanks. We've seen triple-tank can have three scalders; with three tanks, you 8 9 different temperatures. We're not convinced that 10 temperature is all that important as opposed to -three tanks appear to do just as good a job. 11 Counter-flow. The water's coming in when 12 13 the chickens are going out. So clean water is getting on the cleanest chickens. 14 Each tank, the birds are 15 Triple-pass. 16 exiting the tank the opposite from where they entered. The negative aspects of scalding. 17 It's a Any time you have a common bath or 18 common bath. 19 common surface, there's a possibility for cross-20 contamination. It's immersion. There are very few spray scalders or steam scalders out there that stay 21

in the plants for any length of time. We have the

potential for internal contamination.

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Another disadvantage of making this presentation: Very seldom do people report data or prepare something with scalding without the combination of picking.

This is a reference to what Mark said we -- if you'll look at the chicken on the left this is the dirty chicken from here -the conventional processing fiberglass floor. The one on the right was on the elevated wire, a lot cleaner. And we've seen birds a lot dirtier than this. And yes, these birds have a lot more bacteria -- on the left.

So to aid with this, which has also been mentioned, people have added brush machines, either pre-scalding or post-scalding. They have a preventer now that squeezes the bird's empty cloaca. Electrical stimulation is supposed to induce defecation. All claim to have reduced fecal matter going into the scalder so you have a cleaner bird to start with.

Here are some samples from John Cason where we're looking at a triple-tank scalder. The

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birds enter from the left and go to the right. We've shaken these samples up. Visibly cleaner water.

Now, if we look at the micro on this, we have the triple-tanks left to right in the columns, we have a single-temperature tank up on the top row, triple-tanks on the bottom. You can see, as you go from left to right, the recovery of Salmonella in the water decreases. So we're not recovering very much Salmonella.

Now, if we look at the carcasses, all the way over to the right, three quarters of the carcasses still positive. Half the carcasses scalding. It isn't doing that much for Salmonella crosscontamination. And we're actually reducing it with this triple-tank.

What about the respiratory tract? We talked about immersion scalding. It was mentioned this morning about stunning, electrical immobilization, where they're aspirating contents. What happens in a scalder where it's immersion?

This is a study we did again with Mark Berrang. We looked at *Campylobacter*, coliforms, *E*.

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coli and total aerobes before and after scalding and
picking -- before and after scalding. We didn't pick
these birds. I'm sorry.

If we look at Campylobacter, it's very low. If we did a whole-carcass rinse on these birds, 30 out of 30 were positive for Campylobacter. In the respiratory tracts, only 11 out of 30 were positive.

7 is our minimum level of detection. That was one colony on two plates -- I mean a one is one colony on one plate.

So Campylobacter isn't really a problem in scalding. Other bacteria do increase during scalding. Can we prevent this increase? We thought, "Well, can we prevent this increase. What if we put a black cable-tie around the neck during bleed-out? Will that stop it, or is this some other factor breaking up clumps and increasing the number of bacteria in the respiratory tract?"

When we looked post-pick -- I'm sorry -- post-scald, we saw the blue bars on the right. We completely eliminated that increase. Well, the increase due to immersion scalding is a passive aspect

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during going to water. The question was raised:

Well, what does this have to do with respiration?

Could these birds be struggling? What if they were

deader? Would it have an effect?

And we said, Well, for a bird to

ventilate, it has to have neural reflexes from the brain stem; so let's decapitate the birds. We decapitated the birds. No longer ventilation. We'll see if this is purely mechanical or it's a biological effect.

If we look at the light blue and the dark blue columns, these are the birds that we decapitated. And we go from pre-scald. The light green are the ones that weren't decapitated -- stunned and bled. Light blue, there's no difference. If we look at coliforms, you know, 4.3 and 4.2, decapitation.

So this is purely passive. We're putting the bird under water, increasing pressure and then releasing that pressure. It has physics -- it has nothing to do with the bird being -- ventilation.

Let's talk about picking. The positive aspects? It removes the feathers; it also removes the

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epidermis. But it also removes the bacteria on the feathers and epidermis that were put on there in the house and also in the transportation coops.

Negative aspects? As we've heard today, it expresses cloacal contents into the picker and then spreads it around.

Here's a study we did with Mark Berrang where we wanted to demonstrate the increase post-scald during the picker of Salmonella and Campylobacter.

The left side is Salmonella, and the right side is Campylobacter.

We put ten-to-the-seventh Salmonella in a gel capsule into the cloaca of the bird before scalding. That's the left column. We did a breast swipe post-picking, all negative. The birds that we -- I'm sorry. The left side is post-scald prior to picking. The dark yellow one is after we picked them. We have 84 percent positive for Salmonella, 57 percent positive for Campylobacter, indicating that contents are squeezed out of the cloaca.

But this is Salmonella and Campylobacter in peptone, not in fecal matter. So we did another

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study. We added ten-to-seven Salmonella to one milliliter of cecal contents and spread it on the breasts. We did this every other bird. If you'll notice, the second bird and the third bird have a dark streak on them. We had a leader bird and a tailer bird. So we didn't have an effect of bouncing around the picker.

And we also did this with the featherless birds. You see the first featherless bird and then the third featherless bird have a fecal sample on them.

We took breast squares off the birds postpick. You can look at the results.

If we look at the two left bars --100 percent of the birds that we put Salmonella on we recovered it from. That's pretty good. In 100 percent of the birds that we didn't put Salmonella on we didn't recover any Salmonella.

After we picked them, the left column, 81 percent were the birds we put Salmonella on; 88 percent of the birds next to that we didn't put Salmonella on. It didn't matter what position, didn't

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matter if they were featherless or feathered; the picker is spreading Salmonella around.

If we look at the histology section on the left here, we have skin prior to scalding and picking with Salmonella on top. You see the epithelial layer, the base membrane right here -- the basal cells. After scalding and picking, that is literally ripped off the chicken. So in addition to redistributing the cuticle, it's redistributing the Salmonella on the carcass.

We came up with some possible ways for, Well, what can we do to prevent this. And we came up with cloacal plugging. Mike Musgrove did this in '97.

And it would decrease Campylobacter. We can show you the results here.

This is, again, pre-pick and post-pick plugged. In the ones pre-picked, we didn't get any Campylobacter. In the third column, we didn't plug them; 100 percent of these birds were positive for Campylobacter after picking. In the right column, they were plugged; only 11 percent. So we decreased it, but we didn't get 100 percent. Why didn't we get

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100 percent? Well, sometimes the plugs leaked. And sometimes we have to go a little bit harder. If we look Campylobacter data from at that some transportation study, we had the dirty birds, which would be in the green, the clean-wire birds in the We saw Campylobacter dramatically decrease white. after scalding and picking.

We plugged these birds and removed the heads and the feet. So scalders are cleaning it up, but we're not getting it down to zero.

If we look at featherless birds here -this is where the featherless birds come in -- the left column would be normal birds that weren't plugged. In the right column, they were plugged and sutured closed. We sutured them closed. And you can see we virtually eliminated Campylobacter from a postpick carcass if we sutured the vent closed. And you can see how we can do that.

In a normal bird, on the left, sewing the vent closed, you get some feathers in the way. It's not easy to do. On the right, it's easy to sew the vent closed with a dead, featherless bird.

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This shows them after picking. This slide also on the left side shows how big this feather follicles are. Okay. Everybody can see those. I didn't see anybody say they could see Salmonella. So Salmonella in and out of those feather can qo follicles if it wants to, but it's not really a big factor.

What happens with Salmonella while in scalding and picking if you eliminate vent leakage? Nothing. The scalder knocks down Campylobacter. These are the same birds pre-scald and post-scald. It didn't do anything at all.

Well, we thought, What if we pick the birds longer? If we're redistributing the *E. Coli* and the *Salmonella*, what if we did it twice as long? We'd get rid of more of it.

The left two bars, we picked them for 30 seconds; right for 60 the two bars seconds. and E. coli, first Campylobacter the bars are feathered and featherless. Ιt didn't make any difference. So picking them longer or more pickers isn't going to make a difference.

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The third topic I'm supposed to talk about is rehang, where we're going from the kill line to the evisceration line. This is mainly machine transfer now, physically transferring from your de-feathering line to your evisceration line. It's going to minimize external surface cross-contamination if they make a correct transfer.

Positive aspects? Physically different lines, plus we're removing the heads and feet from the evisceration line -- sources of contamination.

Negative aspects? We can get leakage from the vent, and we can also get leakage from the esophagus and the crop. Time on the rehang table can be a problem especially if you have a slowed-down evisceration line.

If we look at the digestive tract of the bird. on the left side, we see the end of the realize esophagus and the crop, but that the [inaudible due to failure of in-house PA system] And the bird is hanging upside-down, so it can leak contents.

Similarly, on the right side, we have the

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vent and the ceca contents. I'll show you some data that -- feed withdrawal doesn't do much to their content.

Let's start with the ceca and some work we did with Arthur Hinton. We did feed withdrawal from zero hours, on the left, to 24 hours. The green bars show the weight of the ceca. We see feed withdrawal had no effect. We've seen that ten minutes postmortem the digestive tract is still active.

Now, on the other hand, we're looking at Salmonella, in the pink bars. Salmonella's still there. So if we're going to leak contents any time during processing, we're going to leak Salmonella.

Now, we've all looked at crops that are empty and full in birds. Similar data with crops. However, if we look at the left, after about 12 hours have of feed withdrawal, we empty an crop. Unfortunately, during time that period, the concentration of the counts of Salmonella are going We've removed the lactic acid bacteria feed up. source, pH has gone up and Salmonella is going to bloom.

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Now, at the end, I was also supposed to talk about the featherless birds. John mentioned them. It is an autosomal recessive that came in a Leghorn background. We had to make three out-crosses to commercial broiler breeders. It's described in <a href="Poultry Science">Poultry Science</a> if you want some more detail.

This is the second generation, a breeder male on the left, a female on the right. They do have a few down feathers; we haven't gotten rid of 100 percent of the feathers. We treat them just like normal broilers. You can see they have leg bands.

And as John says, when we did these studies, we paired equal body weight of feathered and featherless birds. The only thing they need is a heat lamp in this type of area in this type in the winter time.

Let's look at post-chill data, feathered birds on the left, featherless on the right. Pick your bacteria. There's no difference.

Let's look at the Salmonella and Campylobacter we're supposed to talk about. These are counts -- okay -- direct plates, 22 to 28 percent

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1	positive. This was one out of 8 percent positive
2	for Salmonella with direct plating, Campylobacter up
3	there at 97 and 94 percent. If we enrich the samples,
4	it jumps up there.
5	So <i>Salmonella</i> is now at 83 and 77 percent;
6	it doesn't matter if they're feathered or not
7	feathered. <i>Campylobacter</i> was 100 percent. Now, we
8	made these birds positive by challenging them the week
9	before.
10	So the absence of feathers and empty
11	feather follicles does not result in lower carcass
12	bacteria recovery from post-chill carcasses. It's
13	when we're giving the carcass to the consumer.
14	And that's the end of my presentation.
15	Thank you.
16	(Applause.)
17	DR. ENGELJOHN: Thank you, Jeff. That was
18	very interesting.
19	Now we're going to talk about the
20	mechanics of poultry processing, and we have with us
21	David McNeal from Meyn America. David is from here in
22	Georgia. He has been with Meyn since 2004, and he has

1 a food technology background. So welcome. 2 MR. McNEAL: Thank you for the invitation. 3 [inaudible due to failure of in-house PA 4 system] we manufacture poultry processing equipment. 5 And Ι asked to speak today about reducing 6 was 7 digestive tract contamination on carcasses processing from an equipment standpoint. 8 I've listed several factors which affect 9 10 broiler carcass contamination. The microbial load of the live bird which arrives at the processing plant. 11 This includes the bacteria on the skin and inside the 12 13 digestive tract, which is out of the processor's control. 14 15 conditions and content of t.he The 16 This includes how much time the birds have tract. 17 been off feed and water and what they have been eating. 18 19 Processing equipment which is not set up This includes parts and equipment of the 20 properly. wrong size and equipment not adjusted properly. 21

Maintenance of equipment.

22

such

Parts

as -- that come into contact with the broilers -- with the carcasses that can be bent or scratched up or moving parts that can be worn out.

All of these factors are related to minimizing broiler carcass contamination. Processing plants will always be challenged with differing grades of these factors.

Here's a picture of the digestive tract of a chicken. I've listed the most critical parts which pertain to contamination.

Once feed enters the esophagus, it will first travel to the crop. If there's food already in the gizzard, then the feed will stay in the crop until the gizzard is empty. We know if broilers are without feed for too long, then the crop conditions will change, and the amount of Salmonella can greatly multiply.

Once food exits the gizzard, it will pass into the duodenum. This portion of the intestine is important to the process because of its location in the carcass. The duodenal loop lies just below the abdominal surface, and this is the portion of the

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intestines that is most often cut during the opening process.

We also know the ceca and colon are locations in which high numbers of *Campylobacter* and *Salmonella* can be found. The colon or bing area is the area most often damaged with the bing cutting machine.

There are differences between breeds and strains which affect size and shape of broilers. Size, live average weights and shapes of birds greatly influence equipment setup and adjustment and ultimate performance. For example, a breed raw 708 is bred with a long breast and has longer legs as compared to a Taw, which is thicker with shorter legs.

Other factors which affect conformity of a flock are differences in feed conversion between breeds and gender differences. All of these affect the efficiency of the process.

Machines are designed specifically to operate within a given weight range, usually within three to four pounds. And generally, a machine setup is only needed if the average live weight changes of

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the flock more than one pound. If there's too much variation in a flock, then equipment will have to be set to a happy medium; this results in a reduction in equipment performance and an increase in process contamination.

Now, these two photos I have here are of -- I've got -- the photo on the left is of vent cutting blades. I've got five different sizes, and it may be hard to tell the difference. But the smaller diameter blades are used for lighter weight broilers, and larger ones are used for heavier birds.

The picture on the right is of two different lifting units for an eviscerator. The one on the left has a different angle, and more material is used to position smaller birds. The one on the right is designed to allow for more space when lifting the bird for positioning and machine function. If the processor's live weight changes dramatically, then changes greater than the machine's adjustment may be needed. Parts may need to be changed.

The weight variations in this slide I got from our U. S. install list. As you can see, there's

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a large range of weights which fit into the young broiler class of inspection. We're often asked to deliver systems which can process live weights from 3-1/2 to 8-1/2 pounds. The 3.6 to 5.4 weight range represents markets targeted for cut-up, and weights above 5-1/2 pounds are mainly used for deboned product.

Traditional inspection systems, which include Streamline Inspections Systems, SIS, and New Evisceration Line Speed, NELS, were established around the same time the introduction of the 180-degree machines or round machines, as we call them, and were used to improve the performance and quality of the evisceration process.

last advances in In the ten years, Stork Nuova technology such as Meyn Maestro and evisceration systems have established a process which completely removes the viscera pack from the bird and it separately from the for presents carcass inspection. By separating the viscera pack from the carcass rather than draping it over the back of the it was microbiologically proven to reduce carcass,

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internal and external pathogens.

Here I have a picture of two different evisceration shackles. The shackle on the left is made of stainless and is what we call a rigid or one-piece shackle. When a carcass is hung in this shackle, the shackle hangs straight, and the carcass, therefore, is tilted out. This can impede the operations of some machines because the machine cannot lift the carcass with complete manipulation.

The shackle on the right is plastic. It is hinged from a building. And at the bottom, there's a 90-degree break. This is used to allow the carcass to hang straight for more manipulation and placement by the machine. The more carefully and consistently birds are aligned, the greater the performance of that machine.

I want to focus on three pieces of equipment in the evisceration department: The venting machine, the opening machine and the eviscerator. These three all deal with eviscerating the viscera pack and digestive tract in the bird, from which a carcass can be contaminated.

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The venting machine is the first machine. It's job is to remove the vent and bursa Fabricius, often called rosebud, and to position it over the back of the carcass. If this is not executed correctly, then the opening cut cannot be achieved and there will have to be manual evisceration; this can cause increased number of carcasses be an reprocessed.

Some possible reasons for a vent not being removed. The bird may not have been fed into the units of the machine. This could be a timing issue with the machine or a misaligned in-feed guide bar.

The rosebud could still be vent or attached to the bird. Possible solutions for this are sharpening of vent cutter blades, change of positioning of the bird or lowering the cam to be -the lower cam could be too low.

Cutting the intestines and/or back, kidney or hip damage could be causing the machine being adjusted to the wrong height or wrong size parts for average live weight.

I've got several pictures here of

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carcasses just coming out of the vent machine. In the first one, you can see the carcass is hanging straight and it's in a two-piece shackle.

The second one is of a carcass with the vent and rosebud hanging over the back of the carcass.

And as it has been vented properly, if there's any fecal leakage, it will not drip onto the surface of the carcass. The next picture is just a top view, and this opening created by the vent machine is what the opening machine needs to open the abdominal skin.

Opening machine. The function of this machine is to open the abdominal cavity to prepare the carcass for evisceration. It's output demands are no cut guts, no damage to keel or breast meat and to leave sufficient breast meat coverage of the keel area.

There are many different kinds of opening machines. There are those which use a blade very similar to a box-cutting blade, which enters at the opening of the vent and springs toward the keel in a swinging motion.

There's a scissor-type opener which cuts downward

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through the skin and membrane in a clamping action.

And there's a cross-cut style opener, used mostly in

Central and South America.

The picture at the bottom is an example of what can happen with the box-cutter-style opener if it's not adjusted properly. As you can see, the skin is exposed, downgrading the product and exposing the breast meat.

Here I've listed performance failures for the opening machine. A bird can miss the units and not feed into the machine properly. The same solutions as before: Check the timing of the machine and in-feed guide bars.

Cut guts from the opener could be caused from improper timing of the cam with an overhead line or a proper height of machine. A long or short opening or cut keel can be caused from improper positioning of the bird. For example, if the bird is too big to fit into the machine or too small to be positioned, then the cut will not be uniform.

All of these pictures here are of a scissor-type opening machine. As you can see, the

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photo on the left is the unit in an open position. The blade is open. In the middle photo, you can see that -- there's a picture of it closed. And you can also see that the bottom guide bar of the blade prevents the blade from cutting the guts. And the third picture is just of the opening machine in operation.

Draw machine. This type of evisceration machine is often called a draw machine because it draws the viscera pack and digestive tract out of the cavity of the bird. It uses a spoon to pull the guts out of the bird and drape them over the back of the carcass. It works off of a central cam.

The working principle of this machine is that the spoon enters the abdominal cavity near the keel area, it travels past the liver and positions itself just below the gizzard. As the machine turns, tension is created with the use of the central cam, and the spoon scrapes the viscera pack and digestive tract out of the bird. Each viscera must then be manipulated into a presentation for inspection personnel.

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The original patented technology complete viscera removal was developed by Meyn. With the eviscerator, the viscera pack is transferred to a pack take-over device, which is then distributed to different colored pans. The different colored pans then presented inspection personnel are to in a predefined position along the conveyor line with the appropriate carcass.

Key actions of the Maestro eviscerator include accurate positioning of the carcass and the spoon, capturing the trachea and esophagus, and complete removal of the viscera pack. Key performance indicators of the Maestro are: than 99 Greater percent removed viscera packs, properly presented packs with viscera and carcasses to inspectors for disposition, and a less than 10 percent liver damage.

The working principle of the Maestro spoon is that it enters the abdominal cavity near the keel area, travels past the liver and stops between the crop and the gizzard. The viscera pack is then pushed through the spoon with pressure from the central cam. The spoon rotates to close tightly, clamping the

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esophagus before the spoon lifts out the viscera pack.

The photo on the left just shows the Maestro in operation. You can see the spoon is inside the cavity and it's in an open position. The picture on the far right shows the pack take-over device as it deposits the viscera pack onto the trays. And the photo in the middle shows the carcass and viscera pack in line for inspection.

I've discussed the operational functions and performance criteria for the machines which deal with gutting the bird, the ones most critical for controlling contamination. But I would like to briefly discuss the methods of prevention, with method of kill, removal of fecal matter and reduction of contamination with an inside/outside bird washing.

Here I've pictured a cross-section at the base of a head. Here you can see the location of the jugular veins and carotid arteries in the neck, of which all or some must be severed at the death of the animal. You can also see the locations of the esophagus and trachea.

A shallow cut will result in less than an

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optimal bleed-out and will also leave the crop intact until the head is removed by the head and trachea remover just after picking. Removing the head in this manner will often tear the crop, and its contents will know from spill upon the carcass. We previous that depending on the length of feed research withdrawal, the conditions of the crop could change to favor the growth of Salmonella.

If the animal's killed by using a deep cut, a faster bleed and death can be realized. The connection of the crop to the head will be severed, and the crop can be removed by the Maestro eviscerator without spilling the crop contents. The cropping machine at the back of the line actually becomes more of a trachea machine.

Processors that do whole-carcass deboning already utilize this method of kill. The head must be completely removed because the carcass shell and neck will be mechanically deboned. No skeletal fragments may remain.

Fecal removal machine. This machine can be used as a preventive measure. The machine is

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located in the slaughter line after bleed-out and prior to scalding. This machine expresses fecal matter and rinses it off of the carcass.

We know carcasses will deposit fecal contents in the skull and/or in the pickers, thereby spreading and cross-contaminating each carcass which passes through them. The machine is designed to help reduce the organic load at the earliest stage in the process.

Ever since the implementation of the Mega-Reg in '96, plants have been required to meet a zero tolerance regulation for physical fecal contamination.

Uses of inside/outside bird washers have greatly increased; most plants have two per line. This machine uses many nozzles and various water pressures and consumptions to clean the inside and outside of the carcass.

The use of this machine is designed to reduce contamination; it is not designed for prevention. It's only performance criteria is 100-percent cleanly rinsed birds.

In conclusion, broiler carcass

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contamination is influenced by the condition of the bird when it arrives at the slaughter plant. Processors can use preventative methods when possible and maintain equipment properly -- which is used for evisceration to reduce carcass contamination. Thank you.

(Applause.)

DR. ENGELJOHN: Well, thank you.

That leads us to our next presentation, dealing with reprocessing of fecal contaminated carcasses and the use of antimicrobials. We have Dr. Stan Bailey back with us. He is a microbiologist with the Agricultural Research Service at USDA.

Stan?

DR. BAILEY: Thank you, Dan.

(Pause.)

DR. BAILEY: Thank you. The second area they asked me to talk about this afternoon was reprocessing. And I guess maybe the reason they asked me to do that is because we did in our laboratory -- I wasn't involved directly in the very first project I'll talk about, but the others after that. We did

the very first work in this area back in the '70s.

A little bit of background before I get into the actual reprocessing. In the early '90s, '92 and '93, after the Jack-in-the-Box *E. coli* 0157:H7 issue with beef and with recurring issues with Salmonella and other issues with poultry and other animal species, there was a concerted effort made by USDA to begin to try to improve this situation.

What came out of that led to a lot of meetings and other things and the Mega-Reg, the HACCP document that we still refer to today. But one thing that was a central tenet of those discussions at that time was fecal contamination, the assumption being that if carcasses were fecally contaminated, then they would have a greater propensity for having Salmonella and other pathogens.

At about the same time -- well, I'll get into that in a second. With fecal contamination, there's a couple of issues. Obviously, there's the aesthetics of the situation. It's not particularly palatable to think that the meat product that you're buying or want to eat would have feces on it. And

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then there's microbiological concerns, the assumption being made that if you have fecal contamination, then you have a higher likelihood of *Salmonella*.

And one of the central tenets that was put forth by USDA at that time was that all species would be treated equally in terms of allowability of fecal contamination. That is not a bad theory and idea, but it's somewhat problematic when you think that species aren't exactly equal. Many types of products have the skin removed, and that's an entirely different situation than poultry which has skin on. So there's a lot of issues that we could discuss about that.

But to the idea of reprocessing, about 1972 or maybe even '73, Ken May, who at that time was with Holly Farms, came and talked to Roy Blankenship, who was the research leader in our unit at that time, and asked him if we could take a look at an issue that was really causing a lot of issues for the poultry industry.

And at that time, approximately 1 percent of the birds were having to be reprocessed because of visible fecal contamination. And the only thing you

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could do at that time was to trim the product if it had fecal contamination on the outside. Or if it had internal feces, you had to discard the product.

And so we -- that was just slightly before I started working with them. But they initiated a study in 1975 that was the first to show that inspection-passed and offline re-processed broilers were basically microbiologically indistinguishable.

And this is one of the data slides from that study. And you can see if you look at the inspection-passed or -condemned product on an external swab -- this is looking at enterobacteriaceae, which is the family that <code>Salmonella</code> is in -- there was no difference. And if you washed it with water again, there was no difference.

In internal swabs, the condemned were a little higher, and you got somewhat lower, but not microbiologically significant, after washing. And so you see what happened with the initial study.

So then a few years later, FSIS had taken that under consideration and rewrote the rules in the Code of Federal Regulation, which allowed that under

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the supervision of a USDA inspector, reprocessing treatments were allowed, including trimming, vacuuming, washing or a combination of these. If internal contamination's present or treatments other than trimming are used, the entire carcass must be washed with water containing 20 parts per million.

So what that means in simple terms is that after this regulation was passed, visually fecally-contaminated birds as identified by an inspector were allowed to be pulled off the line and washed and put back on the line. When that process went into place, there was a lot of concern by consumer groups and others that this was maybe not a good idea.

So we revisited that issue and published in 1993 another study showing the microbiological quality of conventionally processed and reprocessed broilers. And you can see here that they're basically microbiologically indistinguishable between the two. And again, I wanted to point out that this study was done with just water alone, no chemical treatments.

About this same time, Amy Waldroup and some of her co-workers also looked into this issue as

for commercial reprocessing of broiler chickens. And they used 20-parts-per-million chlorine in the carcass wash water and determined the effect also that reprocessing was having on *Campylobacter*.

So they were the first ones that looked at the Campylobacter issue in reprocessing, and they found that there was some plant variability, but they concluded that current reprocessing procedures were microbiologically justified and that on reprocessed carcasses, there were no significant differences in Salmonella prevalence or numbers and that either affected Campylobacter were not significantly lower than in commercially processed birds.

And then in 1997, Dan Fletcher at the University of Georgia published a study where he showed that on-line reprocessing reduced the need for off-line reprocessing by 73 to 84 percent.

He found that Salmonella and Campylobacter incidences were not affected by on-line, versus off-line, treatments, there was no significant differences in treatments between the effects on aerobic plate

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counts, Campylobacter or coliforms, and summarized that on-line processing of visually contaminated carcasses could greatly reduce the number of carcasses being subjected to off-line reprocessing without negative effects on bacteria and pathogen counts.

And then in 2003, our next speaker, Julie Northcutt, published a paper where she showed the effect of bird washers on carcass microbiological characteristics. And in this study, she was looking just at the effect of washing and the equipment itself with water, not the chemical effect, and found no differences were found in coliforms or *E. coli* counts due to washing in an inside/outside bird washer, that total aerobic plate counts were lower on carcasses from one plant, but not on carcasses from two others, and washing in water alone did not significantly change carcass bacteriological characteristics.

So that brings us to the second half of what they asked me to talk about, and that is the use of chemicals. So we have shown historically that -- pretty much almost every study that has ever been down has shown that you can reprocess carcasses and

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microbiologically it'll be indistinguishable from inspection-passed carcasses.

So that has led to the use of a lot of chemicals in the plant. Either it is reprocessing aids or, just in general, to try to get down to meet the Salmonella performance standards. And the chemicals -- and I'm sure I left somebody's off. And if you're in this room and I didn't put your chemical on here, I apologize.

But the ones that are fairly commonly seen is: Chlorine, probably the most widely used of all of the chemicals -- can be up to 50-parts-per-million product contact; a fairly new chemical on the market place, Cecure, cetylpyridium chloride; Inspexx, which is Peroxyacetic acid; Safe<sub>2</sub>O, which is an acidified calcium sulfate; Sanova, which is an acidified sodium chlorite, TomCO, which is a CO<sub>2</sub>/chlorine system, and then; TSP, or trisodium phosphate.

And each of these have been used or tested a great deal, and there's a good bit of publications on them. And I didn't have time to really go into the individual use of each of these, and so I won't try to

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do that. But I think there are a couple of considerations as we think about all these chemicals that we really have to consider and keep in mind.

One, what is the organic load that you're going up against? A lot of these chemicals will kill things in a test tube. The key is: Can you get the active ingredient in the chemical to the bacteria that you're wanting to kill? So the amount of organic load on a bird can be imported. And some people -- it has been variable results with the benefits of using brushes and scrubbers, but there are -- some people have found that using brushes to get that organic load minimized does help the process.

Then there's the issue of dipping. Can you -- and you see different permutations that people are using: Pre-chill dips, post-chill dips. The chiller itself, the immersion chiller, is really a big dip tank. I mean usually 30 to 45 minutes in a dip tank. So having that exposure to these chemicals in a dip tank can be effective.

And then there are sprays. There's continuous spraying of equipment. I was part of the

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team and led the studies that did the initial work back in the '70s that led to the regulation requiring continuous spraying of 20-parts-per-million chlorine to all common equipment surfaces that touch birds. And so we know that spraying of equipment can keep a build-up from happening over time.

There's external spray cabinets where you're just doing like the old-line final washes, where you're washing the outside of the birds. And then there's inside/outside spray cabinets, as you've just heard some talk about.

So all of these can play a role. And you see different companies using different permutations of these.

There's a lot of issues and concerns with chemicals, though, as we talk about it. Certainly, export markets are one. Depending on where in the world you might be exporting to, there's different rules and regulations. If you wanted to export to Canada or Europe, you certainly can't use the elevated levels of chlorine. And I don't know all the rules for all the countries, but I know that there are

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issues you have to deal with.

There's organoleptic quality or the potential issues with organoleptic quality for certain of these chemicals. Certain chemicals will do a good job of killing Salmonella, but maybe they might discolor or have some off-flavors or something that you'd be concerned about from an organoleptic point of view.

I think a really important factor that people have to consider is your water chemistry; all water is not created equal. If it's a municipal system, depending on what they're using -- are they using chlorine or chloramine or some other kind of chemical in the water already -- the hardness of the water, the amount of trace minerals in the water -- everything doesn't work the same as everywhere.

So if we go in as researchers or as regulatory agencies and we evaluate a plant and they seem to be having real good results, there's a lot of things that could be contributing to that. One, what is the load of pathogens coming in on those birds to start with? That might be a big factor.

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But another factor may be, Well, this chemical seems to work really well in this plant, but it doesn't work very well over there. Well, that might have to do with water quality or a lot of other issues.

There's also worker health issues when we talk about chemicals. Some of these chemicals are pretty toxic. If you go in a plant that's running maximum levels of chlorine, sometimes in those areas, it'll just about knock you over. And some of the other chemicals can have issues that we have to be concerned about for worker health.

And certainly, sampling technique is very important. We know from many of the early studies with some of the chemicals that looked particularly effective that what we were doing was not necessarily killing the <code>Salmonella</code> on the carcass.

What we were doing was -- that carcass carried some of that residual active ingredient of those chemicals with it. And when you did a rinse sample of the carcass to see if it had killed the Salmonella, you kept that residual chemical in your

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rinse water, and it would kill it there. It hadn't killed it on the carcass.

So when we're doing these studies, we have to be particularly cognizant of that issue and make sure that we neutralize the chemical during the sampling process.

I'm just going to pick a couple of -- I'm not trying to promote this particular chemical in what I'm doing here. I want to make that very clear. But I did want to show you a couple of selected studies where you can see that chemicals when used properly can be effective in helping you reduce your levels of Salmonella.

In a study that was published by Kemp and co-workers in 2001 -- and it was, I want to point out, a company-funded study, and I think that's always important to know, but -- looking at acidified sodium chlorite spray system, the microbiological quality of fecally contaminated carcasses found be was to significantly better than that of off-line reprocessed And all but two of 1,127 carcasses passed carcasses. the zero fecal tolerance test.

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So this allowed the -- this was taking the reprocessing step and putting it in a continuous online system and using a chemical to help you control the issue. And you were able to do that without having to take things off-line.

And this is some of the data from that study, and I won't go over all of it. But if we look at just the Salmonella line, after the eviscerator, the carcasses were about 37 percent positive. After the continuous on-line reprocessing, they were 10 percent positive as compared to the birds that were taken off-line for reprocessing, which were slightly lower than the post-evis, but still at about 32 percent.

And even if we take these birds that were continuously on-line reprocessing and going through the chiller, they were still only 12.5 percent positive. So as I say, I'm not trying to selectively pick out and advocate this chemical, but this is an example of some good results that we're seeing.

And then at Auburn University, they did also a study looking at the post-chill application of

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acidified sodium chlorite. And this is something I hadn't talked about earlier, but this is taking a dip after the chill tank.

So you've gone through the entire process, and you have a post-chill dip. And using this -- at Auburn University, using this acidified sodium chlorite, you can see the pretty good results that they were getting. The *Campy* levels were down and extremely low, and so were the *E. coli* levels.

I'll take about -- the last two minutes I have is almost exactly the amount of time I'll need. This very same study I talked about earlier I won't go over. And it was the 20 plants. We worked with FSIS plants in the post-pick and post-chill. And, again, the same things we looked at.

But the reason I wanted to go over this real quickly was to show you that for what -- we have survey data showing us what each of these plants were using, and they're all over the board. People are using different things, but I just wanted to show you what was happening in the plant.

If we look at the rehang station and we

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look at *Campylobacter* levels, we averaged across that study log 2.66. By the time we came out of the chill tank, we were at log .43; in *E. coli*, 3.28 to .92. If we -- you can look at the data by season and see the really pretty straight-line effects that you were getting, very consistent effects, with *E. coli*.

And if we look at Salmonella, which we're talking about today, you can see that at the rehang station, we were bringing in or after -- by the time they got through the picker, anyhow, about 72 percent of the birds were Salmonella-positive. But by the time we came out of the chill tank, over the course of the whole year that we did this study of 1,600 samples each, that -- we had about 19.6 percent positive.

So that's not great in terms of where we want to be, but I think it does show that the plants were doing something that was pretty good. I mean they had a very dramatic reduction.

The next and last slide. When you pair the samples as we talked about before, you can get an idea of how effective your process control is in your plant.

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But if you look at the slide here, you can't see all of the individual lines, I don't guess, but each one represents a plant. They represent what was happening at that plant and what the levels of *E. coli* were on those carcasses at the rehang station in the first side. And they represent what the rehang station was -- I mean after the chiller on the right-hand side.

And if you look at the slope of the line, you can tell how good a job those plants were doing in reducing the bacteria load of an indicator organism that's always there. The problem with doing this with Salmonella is it's sporadic; sometimes it's there, and sometimes it's not.

By taking -- as Jeff had referred to earlier, in taking paired samples within a flock -when you use within a flock, you eliminate a lot of those other variabilities you're concerned about. eliminate seasonality. You eliminate weather conditions, whether the carcasses got wet and all that. And you iust see what happens in that individual plant.

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1	And if you do this over time, you're going
2	to know what you would expect if everything's working
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3	properly. So that is a potential to measure process
4	control, we feel.
5	In conclusion, reprocessing and on-line
6	reprocessing can and do result in processed chickens
7	that are equal to or have improved microbiological
8	quality compared to inspection-passed chickens.
9	Chemical interventions in processing are resulting in
10	significant improvements in prevalence of Salmonella
11	and in reductions in Campylobacter levels in broilers.
12	But I do believe that we do need a
13	caution. And somebody asked the question earlier.
14	Continued use of large quantities of chemicals may
15	lead to increased concerns with export markets and
16	perceived public health issues by some people. So I
17	think that's something we need to keep in mind. And I
18	thank you very much.
19	(Applause.)
20	DR. ENGELJOHN: Thank you, Stan.
21	Realizing it's the afternoon and we've

been going for a little over an hour, now we are going

to take a ten-minute break. So come back at 4:05, and we'll finish out the afternoon and then have questions and answers.

(Whereupon, a short recess was taken.)

DR. ENGELJOHN: We have with us Dr. Julie Northcutt, who's going to talk about the impact of chilling on the poultry carcass microbiology. Dr. Northcutt is a research food technologist and lead scientist in the poultry processing research unit at the Russell Research Center, Agricultural Research Service with USDA. And Dr. Northcutt has her degrees in food science biochemistry from North Carolina State University and Clemson University.

Welcome.

DR. NORTHCUTT: Thank you, Dr. Engeljohn.

And I appreciate the opportunity to speak with you this afternoon. And I'd like to thank you all for staying toward the end here. Also, I'd like to thank the rest of the folks at the FSIS who have organized this, and specifically Dr. Patty Bennett and Dr. Bill Shaw.

As mentioned, I'm going to talk for the

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next few, 15, minutes about immersion chilling and air chilling and how they affect poultry microbiology. So as we know, poultry is chilled primarily to reduce microbial growth.

And the methods include, as I've already mentioned, traditional immersion chilling, which is the method that is most commonly used in the United States, although air chilling is becoming more popular. Air chilling can either be a dry air chill or what is called an evaporative or spray air chill. And in unique situations, there have been combinations of the two.

There have been a number of studies on immersion chilling and very few on dry air chilling or on evaporative air chilling and even fewer projects that have compared the two methods. Many of the methods, on a frustrating level, do not cite the complete information. So it's difficult to dig out through the literature and make a valid comparison because the conditions, the rates and the times are not always complete in some of these studies.

There are a few fairly good comprehensive

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review articles, and I'm sure there are a few others that I have not listed, but this slide shows a few of those. And if anyone is interested in getting a copy of some of these, I would be happy to provide those to you if you would contact me either after this or through e-mail.

So when we look at some of the previous studies, what we find is that in terms of Salmonella, most of the literature has focused on looking at prevalence of Salmonella -- that is: The number of positive carcasses -- and they have not looked at or reported the exact numbers or the counts.

Now, overall, Salmonella prevalence was reduced by immersion chilling and by air chilling, although for the air chilling, I would like to mention that was just one experiment that I was able to find that looked at Salmonella, because there are just so few data out there.

Campylobacter. We found that we get up to about a two-log reduction with immersion chilling, and that's when we use a whole-carcass rinse for recovery.

We get little change in Campylobacter with air

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chilling, but in that method -- that was a neck-skin maceration method from a processing plant that was in Kuwait.

And when we look at generic *E. coli* and coliforms, we get about a one-log reduction without chlorine; when we maintain our chlorine between 20 and 25 parts per million, we get between a two- and three-log reduction. That's with immersion chilling. We got no significant reduction with air chilling, but, again, that was the neck-skin maceration recovery method, which tends to recover higher levels than just a whole-carcass rinse.

I'd like to spend the next few minutes talking specifically about some research that we've done at the Agricultural Research Services and then end with another study, from the University of Bristol.

This was a study that I conducted with Mark Berrang, whom you've already heard from, Andrew Dickens, Nelson Fletcher and Nelson Cox in 2003. And what we were doing was looking at the effects of broiler feed withdrawal and transportation on levels

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of Campylobacter, Salmonella and E. coli on carcasses before and after immersion chilling.

And what we did was -- we went out and found a Campylobacter-positive commercial flock; at about 28 days of age, we tested the litter. And when we identified the flock, we then went back at 36 days of age and moved them to a university facility. We inoculated them one week before processing with a marker strain of Salmonella, and then we processed them at 42, 49 and 56 days of age.

We did a whole-carcass rinse after a manual final wash, and that was our pre-chill counts.

And then we did another whole-carcass rinse after chilling, and we maintained our chlorine level in the chillers at 20 parts per million.

This is just a picture of our little prototype tumble chillers.

And this slide shows the data with the log counts on the Y axis -- that's colony forming units per mil -- of the whole-carcass rinse. And the different categories of bacteria are on the X axis. The white bars that you see in the first column of

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each bacteria represent the pre-chill counts, and the black bars represent the post-chill counts.

And what we found is that with the chilling, we got a 1.2 log reduction in coliforms, we got a 1.3 log reduction in *E. coli*, we got a 1.3 log reduction in *Campylobacter*, and we only got a half-alog reduction in *Salmonella*.

If you'll notice, we started off with a very low level of Salmonella. And I think the reason is because we inoculated them one week before we have waited processed and we should and inoculated them about two days before we processed. So there were low levels to begin with even though that was a significant reduction and it was only half a log.

This next study is from an individual in our group that you've already heard from, John Cason, and co-workers. And in this study, John wanted to look at the effects of pre-chilled fecal contamination on the numbers of bacteria recovered from broiler carcasses before and after immersion chilling.

And what John did was -- John came up with

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this idea of dividing a carcass in half and using one half of the carcass as a control and the other half as a treatment. And this is an excellent way of doing a study because you statistically have the companion comparison, which is a far more superior statistical comparison than one carcass to another.

So he divided the carcasses half in two, and then he identified a 3-by-5-centimeter rectangle section on each half. And on one half of each pair, he put .1 grams of fecal material. Then he waited ten minutes, and then he washed it and chilled it in the same prototype chiller that I just showed you, then did a half-carcass rinse and then also recovered the section of skin that was macerated, that3-by-5-centimeter square of skin.

And I don't know if you can see this or not, but there are little dots designating the area on the skin here. And this is an example of a paired half, so he has got the 3-by-5 identified on the control carcass and then the fecal material on the other half.

So what John found -- and this is just the

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E. coli data, but he found the same thing for enterobacteriaceae and for coliforms. He did not look at Salmonella, unfortunately.

But for E. coli in the rinses on the control half that did not have the fecal material, he found 5.4 log; on the half that had fecal material, he found 5.5 log, not a difference. The skin halves or -- the skin sections that were macerated -- he got comparable counts on those, as well. And mentioned, the same results occur for enterobacteriaceae, which is the category of bacteria that includes Salmonella. And also, for coliforms -the same data. So no difference.

Another study I want to tell you a little bit about is from Doug Smith, John Cason and Mark Berrang, also with ARS. And in this case, they looked at the effects of fecal contamination in immersion chilling on *E. coli*, coliforms, *Campylobacter* and *Salmonella* counts on broiler carcasses.

This is a little bit of a complicated design. And I stole this slide from Doug, so I can't take credit for the handy drawing. But what Doug did

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was -- he got eight carcasses, and he divided them half in two.

If you'll look at the left side of the screen -- he took eight carcasses, divided them half in two. One half went into one chiller which he designated as the clean chiller. That companion half went into another chiller that he designated as the contaminated chiller.

Then he took another set of eight carcasses, and he divided those half in two. The one half that did not have fecal material on it went into the clean chiller; he put a tenth of a gram of feces on the companion half and then put that into the contaminated chiller.

And this is the results that Doug found from this study. The white bars represent the carcass that had the fecal material on it, the black bars halves represent the carcass that were the contaminated chiller, and then the green bars represent the halves that were in the clean chiller. And he found no difference in coliforms and E. coli among any of the treatments. He did find a slight

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difference with Campylobacter.

With Campylobacter, the control carcasses were about six-tenths lower in log Campylobacter than the direct contamination. They were half a log lower than the cross-contamination. So that's a minor difference, statistically significant, but, biologically, we always question whether or not a half a log is of practical significance.

In terms of Salmonella, he did not find any detectable levels of Salmonella in the control carcasses, and less than one log on either the direct or the cross-contamination.

I would like to point out that no chlorine was used in the chiller. And when I started trying to figure out how much of a volume of water he used, it worked out to be about .9 gallons per pound. So we decided to do another study where we wanted to look at volumes of water that were used in the chiller.

And in this first study, we wanted to use extremes that were not commercial volumes. So we picked a very low volume of water and then something that was eight times that to see if we were going to

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find a difference in bacterial counts.

So what we did was -- we used John Cason's method again of dividing a carcass half in two. And on one half of each pair, we put in .25 gallons per pound, which is typically about half the level that we would see commercially, and then we put the companion half in eight times that, which would be two gallons per pound. This, again, was non-chlorinated water.

And what we did was -- we put each half in an autoclave bag with zip-ties. And then this was submersed into a secondary tank of chilled water that had an air agitation in the bottom. And after 45 minutes, we then pulled those halves out. We let them drip for five minutes, and then we did a half-carcass rinse and looked at what we could recover.

So again, we've got the log counts on the Y axis and the different bacteria on the X axis. The white bars are the pre-chill counts, the low volume of water is in the black bar, and the high volume of water is in the green bar. And from pre-chill, using a low volume of water, we got a 1.5 log reduction. We gained another half-log reduction -- and that's total

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aerobic bacteria -- when we increased the volume of water.

For *E. coli*, we saw a two-log reduction when we used a low volume of water; this was then increased to 2.8 log with a high volume of water. For enterobacteriaceae, we saw a 1.2 log reduction with a low volume of water and a 2.2 log reduction with a high volume of water.

Campylobacter had our largest reduction.

We saw a 2.7 log reduction with a low volume of water and then a 3 log reduction when we used a high volume of water.

Interestingly enough, we also collected the water that was in the chill bags, and we looked at how much bacteria was in that. And we did it on a per-mil basis. And oddly enough, we found that on a per-mil basis, each mil of chill water had the same total aerobic bacteria count, it had the same *E. coli* count, it had the same enterobacteriaceae count and the same *Campylobacter* count.

So we are actually going to go back and do some additional volumes that are in between these two,

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but we've found this to be extremely interesting.

Maybe there is a possibility that a certain number of
bacteria get in each mil and that's going to be the
standard.

I wanted to end with a study from the University of Bristol by a fellow named Jeff Mead and his colleagues. And this was conducted in 2000. And he looked at the microbial cross-contamination during air chilling.

And what he did was -- he used a marker strain of *E. coli* that was a non-pathogenic strain. And he put this on one carcass, and then he ran it through a commercial system. And the commercial system was set up for an evaporative chill with 50 parts-per-million chlorine. Then they turned that system off. And they ran it through and did a dry chill.

And he evaluated the contamination in a plus-or-minus ten carcass direction, and he also evaluated -- I'm not going to show that today, but he evaluated it on a companion line to see if it would go in all four directions. And in fact, it did. But

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what he did was -- he put 10 mls of 10° colony forming units per ml, and then he sampled the different carcasses.

This slide shows the set of data for the evaporative chill. The zero position on the X axis is the carcass that was inoculated, and he went upstream and downstream. And then you will notice that he found at least on log even ten positions away. So we did get cross-contamination there.

The same thing for the dry air chill. Although the counts were lower, he was able to recover bacteria as far away as ten positions.

what know from this is So we that immersion chilling causes at least a one log reduction in carcass pathogenic bacteria; post-chill, when we fecally contaminated carcasses, have they are microbiologically equivalent to non-contaminated carcasses.

And the potential does exist for crosscontamination during immersion and for air chilling, particularly if antimicrobials missing or not used. And I wanted to mention that because I frequently hear

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that a lot of folks are saying, "Okay, we're not going 1 to get cross-contamination with air chilling," and I 2 think that's a miscommunication. So we will as long 3 4 as we are chilling chickens. So thank you for that. And I will 5 show you the place where we all work. 6 7 (Applause.) DR. ENGELJOHN: Thank you. 8 9 Our next speaker is Dr. Ken Byrd, 10 veterinarian with Mionix, who will talk to us about experience with managing pH and its effectiveness in 11 processing water. He comes to us with experience from 12 13 the field; he has worked with industry and is a former FSIS employee, as well. 14 15 So welcome. 16 DR. BYRD: Thank you very much. Ι appreciate the opportunity to be here. 17 Let me begin 18 simply by offering a disclaimer. FSIS does not 19 endorse any particular products. 20 Can you all hear me okay in the back? Ι 21 was -- can you hear me?

(Pause.)

DR. BYRD: Okay. I didn't know whether there was an issue with the sound or what. I was walking around during the break back there awhile ago and talking, and I heard one of the sound system folks say that they thought there was a loose screw in one of the speakers. So, you know, I don't know for sure how to take that.

The topic of my presentation does have to do with chlorine and some of the factors that are issues to make it work. Many if not most of the poultry slaughter operations use some form of chlorine in the plants, whether it's sodium hypochlorite bleach or whether it's calcium hypochlorite, some of the swimming pool-type tablets, or gas or whatever.

To make this work, I need to do just a quick review of the chemistry of chlorine. And I -- most of you all probably know all this, but there may be someone here that doesn't understand it all. And so I don't mean to insult anybody's intelligence, but I've kind of got to go back to zero and start over on this.

To really accomplish the disinfectant and

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oxidation from chlorine, you'd have to (have a) chemical reaction between the chlorine and the water to form hypochlorous acid. Now, the hypochlorous acid is the particular substance that does destroy the bacteria, and it usually does this by the process of oxidation or simply pulling electrons out of their cell membranes.

Now, the kicker to this is: When you produce the hypochlorous acid, it's a relatively weak and unstable substance, and it doesn't stay in that form very well; it will tend to dissociate into the chlorite ion. Now, what -- again, the hypochlorous acid is what is the antimicrobial. Key point: The chlorite ion is not relatively effective.

So why is pH control important in this whole scenario? PH is what drives the equilibrium back and forth between whether the hypochlorous acid stays in its chemically active form or whether it dissociates over into the relatively ineffective antimicrobial.

The higher the pH, the more the reaction's driven to the right, the less antimicrobial activity

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you have, because more of it is driven over to the chlorite. The lower the pH, the more it's driven back to the left to stay in the hypochlorous acid form.

If I had only one slide that I could present, this would probably be the slide because it pretty well puts into picture what is happening with the hypochlorous acid. For instance, look at if you've got a pH in your water of, let's say, eight. Only about 27 percent of your free available chlorine is actually going to be in the hypochlorous acid form. You're getting the beneficial effect out of about 28 percent of your chlorine.

Now, typically what do we do when we're using chlorine and we're not getting the effect that we want? Well, you know, call maintenance and tell Boudreaux to crank up the chlorine pump; let's add in some more chlorine, you know. When I first got in around the meat and poultry industry back in the midto latter '70s, you know, that was the thing to do: Let's just add more chlorine.

But what happens with this scenario so often -- and I've got a bet that there's somebody here

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in this room that has probably experienced this -- you don't get the results that you want. You add more chlorine. Then you start getting complaints from some of the workers or some of the inspectors: My eyes are burning; my throat's burning; you know, we've got too much chlorine. Okay. Go tell Boudreaux to turn it back down; you know, we're getting some gassing-off.

And so it becomes very, very frustrating.

What's the issue? You know, we add more chlorine
because we need it; now we can't use it because it's
gassing off. The key to it is not to add more
chlorine, but simply make what you've got work better.

And you do this simply by shifting the pH.

For instance, again, look at the pH of 8.

You have about 27 percent or so of your chlorine that's actually active. If you drop that pH to 6.5, you increase your hypochlorous acid from 27 percent to about 92 percent. What have you done? Have you added more chlorine or more chemical to that? No. You've simply just adjusted the pH and use what you now have already in the water. That's very, very important.

And you need to keep a good handle on

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this, you know. Used to be we used some of the swimming pool kits to measure pH and whatever; you'd pour some of this in and some of that in and you'd get a color reading, and you'd compare it to a chart or whatever. Today, with the new technology, the handheld pH ORP meters are very effective and very inexpensive. I bought one, oh, back in the summer, and I think it was -- well, it was less than \$150 for a handheld ORP pH meter that's quite accurate.

Again, the key to effective use of chlorine is to keep the chlorine in the hypochlorous acid form. As Dr. Bailey said awhile ago -- and I loved that comment -- not all water is created equal.

Well, that's very true.

Because how your acidifier reacts depends on your water quality, you need to take this into consideration. Does it come from a well, where it may have a lot of mineral content? Does it come from the surface? Where does it come from? What's the pH and the alkalinity? What's the hardness of it? This can help give you some ideas.

When I first got around the meat and

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poultry industry, our buzz words were, "How much chlorine have we got in there? Well, we've got, you know, 20 parts per million," or whatever. And we thought, "Well, okay, that's good." But this simply measures the total amount of chlorine, which is the bound chlorine, which can't react any more, and your free available chlorine.

A little bit later on, we got a little bit more sophisticated, so we started talking about, What is your free available chlorine? And we thought, Okay, now this is the cat's meow. You know, we -this is the buzz word. But there's still an issue that, because that with measures not only hypochlorous acid, but the hypochlorite ion. you know how much free available chlorine you've got, but you don't know whether it's active or not, because if your pH is 8.5 or so, you've probably got, you know, 10 to 15 percent that is actually active.

So the best way to actually monitor or to measure the sanitizing effect that's in your water is through oxidation reduction potential. It's not a direct measurement of the hypochlorous acid, but it is

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1	rather an indicator of the hypochlorous acid.
2	ORP is a conductive measurement; it's
3	measured in millivolts. To kill free-floating
4	pathogens in water, an ORP of 650 to 700 is usually
5	recommended. You know, we try to hit 700 or a little
6	bit more.
7	It's a range, not a fixed number. And
8	just as a side note, it has been reported that it's
9	not really a practical method for monitoring the
10	antimicrobial potential of water treated with hydrogen
11	peroxide or peroxyacetid acid. And I'm not chemist
12	enough to tell you why.
13	Also, there's it has been reported that
14	there's some caution against using citric acid as an
15	acidifier, because it some evidence indicates that
16	it may in some way interfere with the lethal action of
17	the HOCl.
18	Okay. I mashed the wrong button.
19	(Pause.)
20	DR. BYRD: Okay. An inorganic food grade
21	acid is what is recommended.
22	This is just an example of some of the

pathogens and the kill time with the different ORP readings. As -- for instance, if you'll -- look at Salmonella, for instance. At an ORP of -- I'm going to do like Dr. Cason; I'm going to point on my screen here, and you all follow me along.

For Salmonella, at an ORP of about 485, you can see it takes over 300 seconds for it to be killed. But if you increase your ORP over to 665, you get killing in less than 20 seconds.

Now, the acidifiers that are used to acidify the water come in different forms. There's benefits and there's disadvantages to all of them.

Dry powders? One of the advantages is it's less freight. Any time that you're shipping a dry powder, well, it's probably more cost efficient, but there are some draw-backs, you know. The dry powders must be mixed. Boudreaux may come in on Monday morning hung over pretty bad, and, you know, he may not be real good at mixing today.

There are -- any time you've got to mix something, there's increased labor, and there's higher risk of mixing errors and potentially inconsistent

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results. There may be some hazards to the workers.

And also, if you've got some undissolved solids, these
things can plug hoses and nozzles and that type of
thing.

There's also gas acidifiers such as  $CO^2$ . Some of the pros? No mixing is required. They're relatively inexpensive.

I am told that some of the things to be aware of in using it is the safety factor. You need to monitor the CO<sup>2</sup> in the air. It's not real easy to automate. And you need some sort of device or system in place so if you lose -- this is a free-flowing system, I'm told. And if you lose electrical power, this system continues to emit the gas. So you've got to have something there to shut it off.

Also, there are ready-to-use liquids. With the liquids, no mixing's required. So you don't have the extra labor or the potential of mixing errors and those types of things.

Now, any time you start to ship a liquid, you know, it's probably going to cost a little extra freight. One of the ways to address that is just to

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use bulk shipments. A very, very important point, though: Whatever you're doing, automate your system.

Automate your system.

When we first got started in acidifying chillers, we were doing it manually. And that was an extremely time-consuming thing, and there were spikes up and down, and whatever. We weren't getting real good, consistent results, and we quickly realized that this had to be automated.

And this is just one of our systems sitting on the side of a chiller, as you can see.

Again, I'll point on my screen, and you all follow.

But if you can make it out on the slide there -- there's a couple of little probes that you see sitting in a little box. One measures pH, and the other monitors ORP. These hook into a PLC which in turn goes to a couple of pumps to increase or decrease the acidifier, as well as the bleach, for chlorine source.

Where would you use pH-adjusted chlorinated water? Well, I think some of the speakers that have already been here today have pretty well

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covered that. Basically, anywhere that you're going to put chlorine in the water, why not use it and use it efficiently? Also, as Dr. Hulsey mentioned, just the scalder -- just the pH in the scalder showed some -- the pH reduction in the scalder showed some very encouraging results.

Avoid misuse. As with anything else, use as directed. Isn't that a catchy thing? You know, I have to remind myself of that every once in awhile. Use as directed. Use as written in the food safety documents. You need to educate the user and document the training.

Some of the material I've presented today has come from a publication by Dr. Trevor Suslow at the University of California. This is a real good reference on addressing ORP, how it works and how to measure it. And if you don't get this information written down here and you want a copy, well, holler at me, and I'll get this address for you.

So in conclusion, let me just simply say this. Chlorine is effective. It's readily available, it's relatively inexpensive, and it's very effective

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1	when properly managed. And, "When properly managed,"
2	is the key.
3	That's all I have. Thank you.
4	(Applause.)
5	DR. ENGELJOHN: And here we are at our
6	last presentation for the day. Dr. Scott Russell will
7	come back and give us some input on further
8	processing.
9	Dr. Russell?
10	DR. RUSSELL: Thank you, Dr. Engeljohn.
11	(Pause.)
12	DR. RUSSELL: Thank you. Stan says since
13	we had to give two talks, we get double the pay. So
14	I'm looking forward to that.
15	(Laughter.)
16	DR. RUSSELL: All right. These are some
17	of the topics that I'd like to cover. And again, I'd
18	like to say, like Julie did, thank you for staying so
19	late for the presentation. It's good to have a nice
20	audience at the end of the day.
21	But some of the things we want to cover in
22	this presentation are the regulations involved and how

the problem occurs in terms of post-process contamination on fully cooked products and some technologies used to apply sanitizers in these areas.

And we're going to look at clean rooms, employee hygiene, drain treatments that have been — that are very novel, biofilm abatement procedures and innovative surface materials and how some companies use in-process sanitation and novel packaging materials, as well. And then we'll talk a little bit about microbial testing.

As most of you know, the USDA has stated that official establishments that produce ready-to-eat meat products must prevent adulteration by pathogenic environmental *Listeria monocytogenes*. And the new directive also requires that the plants conduct verification procedures to make sure that the organism is being removed from these foods.

And there's a similar regulation with Salmonella, well, fully cooked regard to as on products, and it qoes sort of like this '381.150. I'll just read the part on lethality: 7-log reduction of Salmonella or an alternative

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lethality that achieves an equivalent probability that no viable *Salmonella* organisms remain in the finished product, as well as the reduction of other pathogens and their toxins...is necessary to prevent adulteration."

So the same sort of regulation there on both products. Now, how does this occur?

And the basic problem, again, very similar to what I mentioned previously, is that these organisms, either Listeria or, in some cases, Salmonella, equipment surfaces -can get on the particularly Listeria, because it lives well in the drains in the cooling areas. It can get on fans, and it can get on cooling units and on employees. can incidentally go into the air as an aerosol that can be blown by these fans in high areas where there's a lot of rapid air movement, and it can get on the equipment surfaces.

One of the new technologies that has been developed -- and I say it's new; it has been around for probably 20 or 30 years but not really been applied to the poultry industry this way. And in many

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of the food industries, it's just now starting to get real notice, and a lot of people are starting to see how this can work.

Essentially, this is the electrostatic spraying nozzle. And there's a high-pressure air stream pumped through the middle of the nozzle. The sanitizer is introduced into the air stream, where it's vortexed. And it goes through a round, very small aperture, and it's sheared off into about 30 micron particles.

After that, it goes through this silver ring that you see here on the diagram, and it's charged. And it works a little bit different than the normal electrostatic sort of painting-type procedures that you see where you charge the metal and charge the spray a different charge and they coat beautifully. This technology works on the basis that you're charging the spray and, as it approaches an object, the object takes on the opposite charge of the spray.

It's kind of an unusual thing, but here it is in action. And you can see this is pesticide being sprayed onto a leaf of a plant. And you can see that

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the spray is defying gravity, coating the underside of the leaf and the back side of the stem.

To see this a little bit more clearly, we have a demonstration sort of electrostatic sprayer.

Pretend that this is a chicken carcass or an egg or a piece of surface or equipment or food contact surface.

We spray the object with a powdery substance to mimic sort of what a sanitizer would look like for, let's say, eight seconds. This is what it looks like. It's sort of like if you sprayed the ball with a can of spray paint; essentially, only half the ball is covered.

Now, we can actually spray the same ball with, for only two seconds, much less material using the electrostatic sprayer with the charge on. And you see a much better coverage. Much better coverage. We've seen coatings in these areas in restaurants, food processing plants and further processing plants, where we can use 1/80th the amount of material. 1/80th.

Now, you might ask, Well, why in the world isn't everybody using this technology? Well, if you

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were a big company that produced chemicals and I came to you and said, "I can cut the amount of chemical that these companies have to use by 1/80th," what do you think you're going to do? Well, that's the look I get when I go talk to them about these kinds of things.

So you can see the difference here. We were in a room, with 400,000 eggs, about the size of this room. And we used one gallon of material to sanitize the eggs and the floors and the walls. It's pretty dramatic, the kind of results that you can get.

Now let's talk a little bit about clean rooms. This is a strategy that has been employed very well at a lot of further-processing plants. And what occurs here is that the raw area is truly separated from the cooked area. And when I say, "Truly separated," essentially, where the oven is is a very large wall structure.

And I don't have it on the diagram, but just imagine a wall between the oven and the IQF freezer. And the idea is that those chicken pieces or parts prior to cooking will have pathogenic bacteria

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on them, whether it's Salmonella, Listeria, or whatever, and as it goes through into that IQF freezer, it's a new world. Okay?

That IQF freezer is evaluated on a regular basis for biofilm formation. Extra effort is required to remove the biofilms in that freezer. Special antibiofilm agents are used in those freezers. So it's tested on a regular basis microbiologically to make sure that that freezer is free of pathogens.

So when it goes through that oven, it enters into a whole new space now, and it's in a clean room, essentially. The air that goes into that freezer and into the room after the freezer is handled differently.

it After comes out of the IOF, Individually Quick-Frozen, freezer, it goes into the true clean room. And in this room which is very it's almost like separate -going into laboratory -- there's no water on the floor. most of you know, in these plants, there's water all over the place in food and poultry processing plants, but not in this room. These floors are kept very dry.

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The employees from the raw areas are not allowed into those areas. The employees that come in there have to have clean, sanitized boots, smocks, clothes and so forth.

And they sanitize these rooms every two hours in some cases. They'll shut the process down, they'll remove all the food from the area, and they'll spray the walls, floors and ceilings with very high concentrations of quaternary ammonium or some other kind of chemical. And then they'll go in. And if it's too high to be legally used, they'll rinse it off. Okay?

Let's say they used 400 parts per million.

They have to go in there and rinse all that stuff
off. Then they begin processing again for two hours.

The air from the outside is filtered using hepafilters before it comes in there. It's evaluated on a regular basis for biofilm formation.

Again, extra efforts are made to control biofilms, and novel chemical systems are used in these rooms, but there it's almost like a sterile room.

It's not sterile, but it is controlled very, very

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carefully as a means of doing this. And some companies have had very, very good results in terms of Salmonella and Listeria control using these kinds of rooms.

Another area that's of concern is employee hygiene. They want mandatory handwash and sanitizing stations. The hand dips need to be changed on a very frequent basis. I've seen plants were the restrooms have access to the plant, and that's not a good idea. People just walk right out of the restroom right onto the plant floor, not a good idea.

A lot of plants you'll see, you go in, and the people that work in the plant also have cows on their farms. And they're out there managing the cows, and then they'll walk right into the plant with the same boots, same jeans and so forth. And they're wearing very short smocks. That's not a good idea.

I like to tell people to examine employees daily for illness. I used to walk through the plant when I managed the plant and -- further processing plant and make sure that they're not visibly sick or carriers.

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It's a good idea to question foreign employees that are coming back from other countries to see whether or not they may be -- may have become sick. I had to serve on a court case one time where 86 people went to the hospital and one lady died because of that. So it is important to monitor those kinds of things.

And, also, it's important to make sure your employees are familiar with American hygiene customs in the restroom. We have a problem with some folks from some countries that the toilets don't flush. And they'll use the toilet paper and stack it up next to the toilet. And that's, of course, not very hygienic.

So let's change subjects a little bit. A new drain treatment has just become -- has just been developed by Dr. Mike Doyle, who's a professor at the University of Georgia down at the Griffin Labs. And what they did was -- they selected two commonly used competitive exclusion bacteria, the two species in particular, Lactoccus lactis subspecies lactis and Enteroccus durans, and they treated floor drains in a

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poultry processing plant with these bacterial cultures.

The results over a five-week period showed a several log reduction in Listeria at temperatures of 4 to 37 degrees centigrade. And Ecolab now is working with UGA to license the technology. And of course, requests have been made from many major meat and poultry processors to use this to reduce those levels of Listeria in the drains, and, thereby, the incidental spraying of the hose into the drains and the incidental aerosolization of Listeria and Salmonella from those areas won't get the equipment.

So that holds some promise there. It's a new treatment.

As I mentioned in the previous talk, the big problem with biofilm is that you have to break up the matrix. And Lysozyme has been effectively used in this area.

We had some discussions after the last talk, and some of my colleagues mentioned that it may have difficulty with -- Lysozyme may not work so well

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on gram-negative bacteria such as Salmonella, whereas it may work better on some of the gram-positive bacteria. So that's a concern that I did not notice in the literature, but I appreciate that being brought to my attention.

And again, the same sort of thing here:

Mechanical methods have been shown to be very good.

High-pressure sprays have been shown to be very good.

But again, this requires a lot of labor. And it would be great if we could come up with better technologies, better chemistry and better cleaning methodologies for breaking down these biofilms, as opposed to having to use labor and hand-scrub these things off of the equipment.

Again -- I showed you this. Most of you were here earlier. And this was a very successful trial in the sense that we formed these *Listeria* biofilms on stainless steel coupons and we were able to get dramatic reductions on those biofilms over many, many reps. So that was very nice.

Now, as I mentioned before, this idea to me was fairly relatively new: That a company would

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shut down every two hours -- literally discontinue their processing, shut that whole plant down, have everybody removed from the processing floor, and go through a process of disinfection of all the equipment.

Imagine the labor involved in removing all of their product from the lines because, you know, it's suspect of being, well, contaminated with regard to the chemical.

And so they have to remove all of the stuff. The equipment, the wall and the floor is thoroughly rinsed or foamed with 400-parts-per-million quaternary ammonium, for example, and then they have to rinse everything down, bring everybody back and then begin production again. So that is a big bit of labor there, but it has been used to some good effect.

Now, there have been some innovative surface materials developed recently. One is called AlphaSan, also called Silveron. It's a product by Milliken and Westlake, a combination of the two companies. And what they do is incorporate this into plastic cutting boards.

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And we did some studies on this, and we could see -- in a very short period of time, we could see in only one hour a reduction in bacterial numbers.

And at 16 hours, we saw a tremendous increase in the efficacy of reducing these bacteria on coupons.

Another product that has been studied heavily by Dr. Sheldon, if you're interested in that, is called HabaGUARD. And he did the work on Salmonella, Listeria, Campylobacter and E. coli 0157. And they were all inhibited anywhere from 3.6 to 7.7 logs.

But the idea here is that these products are incorporated into plastic cutting boards, into surfaces that are used for processing. And the idea is if they can control bacterial growth on those surfaces, then they may be able to control some of the biofilm formation, as well.

So these are some new products out there that you all should be looking for.

Now, another group of scientists have worked with novel edible films. The idea here is that we're using -- on fully cooked or fully processed

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products -- for example, a chicken patty or a whole chicken carcass -- can we take some material and spray or apply it to that carcass, let's say -- let's just use a whole carcass as an example.

Some of these products that can be sprayed on the outside of those carcasses would include lipids and oils, waxes and emulsions, resins like shellac and rosin -- and I don't know about you, but I don't think I'd like to eat shellac, to be honest with you, but -- carbohydrates like Celluloses, pectins, chitin, starches, gums, and then proteins. There are proteinaceous ones.

And someone once asked me the last time I spoke about this, you know, How about allergies? And of course, some of us are allergic to soy or peanuts or whatever. So we'd have to watch that.

But these things are being produced -these films, these edible films -- and you spray these
on the outside of a carcass. The idea here is you can
add antimicrobial substances to these films, spray
them to the surface of a carcass, and it will prevent
the growth of pathogenic bacteria. And it will kill

anything that might have incidentally gotten on there.

Some of the things that antimicrobials would be used for in a coating like that would be organic acids like acetic, benzoic, lactic, proprionic, and so forth, fatty acids, bacteriocins -- that's another one. Bacteriocins would be by far the broadest group there.

And then there are novel packaging films.

And they're really -- these are interesting because these are the films that are used on these packages, and you can incorporate bacteriocins in particular, like nisin, into these things to help control pathogenic bacteria.

Now, there are two ways to go about this. You can incorporate the bacteriocins directly into the plastic itself or you can -- and I have lots of data here that shows that we had good reductions, but we're running out of time. You can also coat the bacteriocins on the surface of these bacterial films and get excellent results, as well.

Another thing that's so important to remember is microbial testing. When you have these

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types of further processing environments, it's important to do rapid microbial and accurate microbial testing. Some of the companies here in Georgia do up to 80 rapid tests per day on all of these organisms: Salmonella, Listeria, Staph, E. coli, APC.

And it's really important that they are able to test the products and release them based on negative results because in the last few years, we've had anywhere from 100 to \$190 million worth of recalls year because of these bacteria, Listeria. per forth -- being found Salmonella and so processed products. So rapid testing is extremely important. Use of rapid methods is going to help this.

So overall, companies that meet the USDA -- they have to meet the USDA FSIS regulations regarding Salmonella and Listeria on fully cooked foods. Electrostatic spraying is an excellent way or an excellent means of applying sanitizers. Clean rooms generally reduce the risk of the incidental contact with environmental Listeria and/or Salmonella.

And employee hygiene is, of course,

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essential to preventing cross-contamination. Novel drain treatments are being developed. Novel biofilm abatement treatments are being developed, and innovative surface materials may have some hope in the future for helping us to control the growth of these biofilms.

In-process sanitation's being used to good effect. Novel packaging materials now exist, but a lot of companies are slow to uptake these types of things because of the cost involved. Those bacteriocins are expensive. And rapid microbiological testing is essential for data-based release programs to prevent recalls in the future.

Thank you all for your attention.

(Applause.)

DR. ENGELJOHN: Well, thank you very much, Scott, for that. And I think, you know, even though some of that information was related to ready-to-eat products, there's no reason why we can't be looking at what we can do in the raw processing areas to see what can in fact be done more there.

With regard to where we're at now, we're

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1	five minutes ahead of schedule. And we're ready for
2	some questions and answers from the panelists who
3	talked this afternoon.
4	And I also just want to remind you all or
5	at least you don't know this yet, but, tomorrow,
6	we're not meeting in this room. We're meeting
7	upstairs in the amphitheater. So I think it's on the
8	second floor, but, in any case, it's right above here,
9	tomorrow, 8:30.
10	So do we have any questions from the
11	audience for any of the panelists?
12	MR. COUGHLIN: Michael Coughlin from
13	Johnson Diversity. A couple of questions, if I could.
14	One is just from a position of ignorance.
15	I'm not familiar with air chilling, so this is
16	directed to Dr. Northcutt.
17	I'm wondering how it is that you can
18	actually get cross-contamination in an air chill,
19	especially if there's no misting. And of course,
20	there's no mystery how that happens in an immersion
21	chiller, but I don't understand how it happens in air.
22	And the other question is directed to Dr.

Byrd.

If you're trying to -- you made the point at the beginning of your talk that pH is extremely important in controlling chlorine. That being the case, if you're using two electrical probes, one being an ORP probe and the other -- I imagine it would be ORP and chlorine -- why would you not have just simply a pH probe and a chlorine probe? Basically, why do you need an ORP probe?

DR. BYRD: Because the ORP actually gives you a better indication of the killing power of what's in the water, the better indication of the sanitation, because, again, it's a conductance type of measurement.

And so you -- what we do -- we measure the pH which or -- monitor the pH, which is fed into a PLC, which then in turn drives an acidifier pump. But also, then by using the ORP probe to monitor the ORP, this goes into the PLC, which in turn drives the chlorine pump.

One of the things that we had to figure out on this was the electrical setup, because we were

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using the 420-millivolt wires. And to make the ORP pump sensitive to what we needed done, we had to reverse that so that it wasn't a 420, but it was a 24, because when your ORP gets lower, then that's when you need more of the ingredient. And so we had some electrical engineering there to do initially.

DR. NORTHCUTT: To back up and answer your air chill question, thank you for asking that because, if you need to ask that, then I'm sure there's other folks that needed to hear that, as well.

All of the large manufacturers of equipment are now making systems for chilling chickens without immersion, and basically in a cold room. And they may or may not incorporate different phases of that room where they have what they might call a stabilization phase with different temperatures that range in the neighborhood of about 33 to about 37 degrees.

And in many cases, the air is going to be blown either across the carcass or in the body cavity.

And blowing the air in the body cavity is -- gives you a much better removal of the heat. And this is --

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1	may or may not be incorporated with a series of
2	misters that will mist the carcass and maintain the
3	humidity at a high level.
4	And so when water is available, then that
5	provides an opportunity for bacteria to survive. And
6	in blowing the air, we have an opportunity to spread
7	bacteria.
8	And so I don't know if that is in some way
9	answering your question, but I will also ask if my
10	colleague down here with Meyn would like to add
11	anything, because I know that they have a system, as
12	well.
13	MR. COUGHLIN: Okay. Are you saying that
14	you're atomizing particulates, basically, the
15	bacteria?
16	DR. NORTHCUTT: Yes. And
17	MR. COUGHLIN: Just the air currents are
18	enough to drive liquid films of bacteria off the
19	carcass?
20	DR. NORTHCUTT: Yes. Plus you also have
21	the potential for the bacteria to become resident in
22	the room if it is not properly cleaned. And

1	obviously, that could provide some cross-
2	contamination.
3	MR. COUGHLIN: So the movement of air must
4	be actually quite violent to strip off the film of
5	water and aerosolize it and transfer it through the
6	room.
7	DR. NORTHCUTT: And it depends on the
8	system. But in the literature that I've read, it will
9	range anywhere from like a half a meter per second up
10	to three meters per second. And it depends on the
11	system, and it depends on the location in the room.
12	MR. COUGHLIN: Okay. Thank you.
13	DR. ENGELJOHN: Any further comments?
14	(Pause.)
15	DR. ENGELJOHN: All right. Any other
16	questions?
17	MS. NESTOR: I'm Felicia Nestor with Food
18	and Water Watch. And I just wanted to say to all of
19	the panelists that as a consumer representative, I
20	don't know very much about the poultry industry at all
21	or how any of this works. And I just found all the
22	presentations really fascinating, and I really feel

1	like I learned a lot today. So thank you all very
2	much. And now I'm going to show my ignorance by
3	asking these questions.
4	I guess the first one is you know, as
5	consumers, we're told to cook the chicken and cook the
6	turkey to 160 degrees inside the meat. So that
7	suggests to me that the contamination is inside the
8	meat and we're not worried about what's on the
9	surface.
10	DR. BAILEY: No.
11	MS. NESTOR: No? Where am I getting it
12	wrong? Why can't we cook it just like a steak, you
13	know, just really fry the outside? And then
14	(Laughter.)
15	DR. BAILEY: It's a safety factor. It's
16	suggesting that it's not out of the realm of
17	possibility with the processing that it conceivably
18	could on a rare occasion have something get down in
19	the muscle. We intact muscle would rarely have any
20	bacteria in it.
21	If you wanted to cook your chicken to
22	where you didn't get an internal temperature of 160

but you got a surface temperature of 160, the overwhelming majority of the time you would be okay. But as a safety factor put in, if you know that every point in your chicken is cooked to 160, then you know that you have eliminated the possibility of survival of vegetative cells of many of these pathogens we're dealing with. And besides, chicken tastes pretty good when it's cooked to 160.

(Laughter.)

MS. NESTOR: Okay. Thank you. I have a couple more.

Dr. Northcutt, in -- the studies went by so quickly. In the one where the chicken gets split and one half gets put in the contaminated tank, was that also the single contaminated tank? Is -- it looked like what you were using there was just a small tank. You're not talking about what's commercially used, you know, a big immersion chiller with thousands of --

DR. NORTHCUTT: Yes. It was the small prototype. And that was the study that Dr. Smith led -- who is sitting across the room. But yes. It

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1	was our we have two of those single prototype
2	tumble chillers that allow us to do things that we
3	couldn't do in a commercial setting with inoculated
4	strains and bacteria. And so yes, it was that small.
5	It holds about 40 gallons.
6	MS. NESTOR: And how contaminated was the
7	water?
8	DR. NORTHCUTT: In Doug's study? I'm
9	not I don't know.
10	Did you look at the water?
11	DR. SMITH: [inaudible due to failure of
12	in-house PA system].
13	DR. NORTHCUTT: Okay. Do you want to
14	share that?
15	DR. SMITH: [inaudible due to failure of
16	in-house PA system].
17	DR. NORTHCUTT: It was not contaminated
18	it was contaminated?
19	DR. SMITH: [inaudible due to failure of
20	in-house PA system].
21	DR. NORTHCUTT: I do have a copy of the
22	manuscript that I will actually give you if you would

1	like it.
2	MS. NESTOR: Okay. I'm just wondering how
3	it compares to some of those immersion the pictures
4	of immersion chillings that we saw, which some of
5	them looked pretty you know, like the color of
6	DR. SMITH: No chlorine.
7	MS. NESTOR: I know there was no chlorine.
8	DR. NORTHCUTT: Yes.
9	MS. NESTOR: Right.
10	(Pause.)
11	MS. NESTOR: Oh. We didn't see any
12	pictures of immersion chillers today? I thought there
13	was at least one.
14	DR. NORTHCUTT: Yes. I showed one at the
15	very beginning of a commercial immersion chiller.
16	MS. NESTOR: Yes. Right. Okay. I've got
L7	one more question.
18	DR. NORTHCUTT: I'm looking for the
19	numbers.
20	(Pause.)
21	MS. NESTOR: Should I wait, or should I
22	ask my one more question?

1	DR. NORTHCUTT: Sure. Go ahead. And I'll
2	keep looking.
3	MS. NESTOR: Okay.
4	Dr. Bailey, you said, if I've got it
5	correctly, that when you've got these chemical rinses,
6	you have to be careful when you take the sample that
7	you deactivate the chemical that's on it so that it
8	doesn't continue to destroy the pathogen in the
9	sample. Does FSIS do that in its Salmonella sampling
10	in any way? Is there any control for that?
11	DR. BAILEY: Yes, they do.
12	MS. NESTOR: And
13	DR. BAILEY: I was pretty sure they did,
14	but I just checked with Dan. And he said they do.
15	MS. NESTOR: And how do they do that?
16	DR. BAILEY: It depends on the chemical
17	you're using in the plant. If it's chlorine, you'd
18	use something like sodium lauryl sulfate. And
19	different chemicals have different things that
20	inactive them. You just have to know what's being
21	used.
22	MS. NESTOR: Oh. So it's something that's

1	actually in the chemical?
2	DR. BAILEY: In the rinse material that
3	you use.
4	MS. NESTOR: Okay. Thank you.
5	DR. ENGELJOHN: If I could? This is
6	Engeljohn with FSIS.
7	Just to follow up on your question,
8	Felicia, we have over time looked at the issue of
9	whether or not the samples are pathogens are
LO	actually dying in the rinse samples that are being
L1	sent to the lab. And so that is something that we
L2	have studied. And so we in working with ARS, we
L3	actually have the answers to those questions. So
L4	but we do have buffers and so forth that we use that
L5	we ensure don't cause a difference in the pathogen
L6	load in the samples we send to the labs.
L7	DR. NORTHCUTT: Just to finish answering
L8	your question and I will be happy to give you a
L9	copy of this if you would like it. What Doug found in
20	the contaminated chiller is for coliforms, he found
21	log 3.2, which is pretty close to what he was

recovering from the carcass. And similarly, for

1	Campylobacter, he found excuse me. For E. coli, he
2	found log 2.7; for Campylobacter, log 2.9, and; for
3	Salmonella, a 1.5 log in the contaminated chiller.
4	MS. NESTOR: Thank you.
5	DR. NORTHCUTT: And you're welcome to
6	this.
7	MR. SANCHEZ: Marcos Sanchez from Texas
8	A&M.
9	Actually, we published a couple of studies
10	on air chilling without a mist. And one of the
11	observations there was the importance of the pre-
12	harvest interventions there, because you don't have
13	this coming-out tank where everything gets mixed up
14	and probably at the same level. So if you have a
15	loaded carcass, it may get loaded until the end of the
16	process, because they're individually hanging.
17	So here brings the concern about the
18	enumeration of the pathogens like Salmonella. That is
19	rarely done, because the MPN process is very time
20	consuming. So I'm just wondering. With the new
21	baselines that are planned on it, what is the opinion

of the panel on becoming -- the importance of these

carcasses. If they have the levels -- if they were in the tank, they were probably similar. But if they were individually hanged, I mean we're going to have some variability there.

DR. BAILEY: I actually was not aware until Dr. Engeljohn told us this morning that they were planning to do MPNs and enumerate the *Salmonella* levels. I knew they were going to enumerate the *Campy*.

I'm not exactly sure of the point you were getting at, except that you probably may see more -there's very -- there's only a couple of air chillers in the country. So that won't be -- there won't be a whole lot of that. There may be a little bit more variability in air chilling because you don't have the washing effect of an immersion chiller.

Immersion chillers historically, a long time ago, when we first started working in the area, were known to potentially cause a fair amount of cross-contamination. But a properly managed chill tank where you control the pH and you use your chlorine or other disinfectant properly is actually a

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significant processing aid to reduce the levels of -whether it's Salmonella, Campylobacter or any of
your -- just total bacteria on the carcass.

So it all goes back to proper -- if you're talking about the chill tank, then you will generally have, as you've seen several slides suggest today, up to 2 logs or more lower coming out of the chill tank than you did going in. And -- but that's pretty much dependent on the quality of the job you're doing managing your chiller. And that would specifically be referring to pH control more than anything else, but it's some other issues, too.

And I would just add from DR. ENGELJOHN: an FSIS perspective on that that that is also dependent on the quality of the birds and the sanitary dressing that go into that chiller. So from our perspective, we don't want to see situations where the chiller is used to clean up the birds. So the case is that we need to pay attention on that slaughter dressing.

MR. SANCHEZ: Okay. Thank you.

MR. McNAUGHTON: James McNaughton with

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1	Solution-BioSciences. We heard several times today
2	about neutralization of the chemicals in the rinse
3	solutions.
4	Stan, you mentioned it numerous times.
5	We're now working with a lot of acids and
6	some caustic compounds. Are we also in our research
7	neutralizing those products in the rinse solution
8	itself?
9	DR. BAILEY: Anybody who knows anything
10	about what they're doing in research is. And that's
11	as reviewers for pretty much all of us sit on
12	review boards for all the journals that we all publish
13	in. I know that's the very first thing that any
14	time we get a paper that has a chemical in it, we make
15	sure that proper neutralization takes place.
16	You know, in the early years, when some of
17	the first things started coming out, there were a few
18	people who weren't paying attention, but I think most
19	of the people who do the research now do.
20	MR. McNAUGHTON: Not particularly the
21	chemical itself, but is pH being neutralized?
22	DR. BAILEY: Yes. I mean certainly, in

1	any research study anybody on this panel I think I
2	know I can speak for has ever done. The very first
3	thing we do before we ever start running our real
4	experiments is run prototype experiments. And we
5	monitor the quality and the condition of the rinse
6	fluid, whether whatever the pH is. That's
7	certainly one of the very first things we do.
8	You'll always take that rinse fluid that's
9	coming off without any kind of study itself, and then
10	you drop some of your test organisms in it to see if
11	they survive or if they die. I mean it's just a given
12	that that's the first thing you're going to do.
13	MR. McNAUGHTON: Does USDA in their rinse
14	solutions adjust pH in their 51-day window?
15	DR. ENGELJOHN: I don't have an answer for
16	you, but we'll have one before you leave tomorrow.
17	DR. ALTEKRUSE: The rinse is a buffered
18	peptone water rinse. So
19	(Pause.)
20	DR. ALTEKRUSE: Oh. I thought this would
21	pick it up.
22	Yes. The rinses that we use are buffered.

1	So it does tend to adjust for that, yes.
2	DR. BAILEY: Sean, I think it that's
3	what we were just talking about here. But I think
4	it's accurate to say that you are using buffered
5	peptone water rinse. So you're getting buffering to
6	the capacity that the buffering capacity of the
7	buffered peptone water is there, but you're not
8	monitoring the pH of those solutions to make sure if
9	you need to do any other adjustments. Is that I
10	think that's a correct statement.
11	DR. ALTEKRUSE: That's correct. But they
12	are placed in buffered peptone water on ice and
13	shipped overnight. And then they're put into
14	[inaudible due to failure of in-house PA system]. So
15	that's the extent of the system, but we think it's
16	pretty good.
17	MR. COUGHLIN: If I may, I Michael
18	Coughlin again. I have a couple other questions, the
19	first one to Dr. Bailey.
20	Assuming that the bacteria on and in the
21	bird are metabolically more active immediately after

slaughter, as opposed to those coming out of the

1	chiller, would it not be more efficacious to have an
2	on-line reprocessing unit pre-chill, as opposed to
3	post-chill?
4	DR. BAILEY: That is where most of them
5	are. It's only recently that there have been some
6	systems put in place where you have a post-chill dip.
7	Almost all reprocessing is done fairly soon after the
8	evisceration step on the processing line prior to
9	chill.
10	MR. COUGHLIN: But can you speak to the
11	efficacy of pre- and post-chill relative to
12	DR. BAILEY: Are you talking about the
13	efficacy of chemicals pre-chill versus post-chill?
14	MR. COUGHLIN: Uh-huh.
15	DR. BAILEY: A little bit. But don't I
16	mean I haven't done an exhaustive evaluation of this.
17	There it depends on the chemical you're using and
18	your ability to get the active ingredient of that
19	chemical to the pathogen of concern, in this case,
20	maybe Salmonella, whether it's in pre-chill or post-
21	chill.
22	If your most of your pre-chill

applications are sprays. And so you can get reasonably good contact with a spray if it's a good quality spray inside/outside, but it's likely that you're getting less contact time with the active ingredient to the potential pathogen in a spray prechill than if you went into a post-chill dip.

Now, if you have a pre-chill dip, you would have for the most part the same likelihood of exposure of your active ingredient to the pathogen. The one thing you do have when you've come out of the chill tank is that you -- with the immersion chillers, you're getting a washing effect.

So you have reduced the organic load, the fat content that kind of washes off a little bit, maybe a little blood and other material that's involved with processing. And you have a somewhat lower level of both total bacteria and in your pathogens if it's in a properly managed chill tank.

And so you have two issues there. It's not quite a straight-forward answer. It's, Are you talking about a dip versus a dip, or a spray versus a spray, before and after the chiller? And then you

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1	probably have a slightly better chance for
2	efficaciousness of the chemical at a post-chill dip
3	because you have reduced the organic load on the
4	chicken and you have reduced the level of bacteria
5	there. So you may get slightly better post-chill.
6	MR. COUGHLIN: Thank you. The other
7	question I have I guess would be for Dr. Buhr or Dr.
8	Cason.
9	I've noticed that carcasses coming out of
10	the chiller the skin seems to be easily detached,
11	and sometimes torn. I'm wondering. Is it possible
12	for the bacteria to actually you know, during the
13	hour's time that the carcass spends in the chiller,
14	can the bacteria be transferred actually underneath
15	the skin? And might that be a reason why it's
16	difficult to get a complete kill?
17	DR. CASON: Well, I think bacteria can be
18	transferred anywhere that water gets to while they're
19	in the chiller. But there are so many locations where
20	the bacteria can stay that it's hard to say that any
21	one location is more important than another.

 ${\tt MR.}$  COUGHLIN: Has that area actually been

1	investigated, though the underside of the skin?
2	DR. BUHR: Mark Berrang has done some work
3	with post-chill skin removal and in looking at parts
4	with or without skin. And relatively the same level
5	of bacteria is recovered. So they are contaminated
6	beneath the skin as related to Campylobacter.
7	MR. COUGHLIN: Thank you.
8	DR. CASON: Huda Lillard did some
9	experiments a long time ago and one other lab, as
10	well where they were doing rinses inside carcasses
11	versus outside the carcasses after chilling. And they
12	found that chilling really redistributes the bacteria
13	so that they're pretty equally distributed all over
14	the carcass inside and out after chilling.
15	DR. ENGELJOHN: I'm going to ask if
16	there's anyone on the phone line that has a question
17	now.
18	(Pause.)
19	DR. ENGELJOHN: Any other questions here
20	in the room while we're waiting for the phone?
21	Yes, Felicia?
22	MS. NESTOR: Felicia Nestor, Food and

Water Watch. I just wanted to ask this. It's actually more pertinent for tomorrow, but I know that some people in the room will be leaving before the discussion tomorrow.

And I'm assuming that the last topic we're going to be talking about is the new <u>Federal Register</u> notice that's out and what the Agency is considering doing with the *Salmonella* results. And one of those things is publishing the results by plant.

And I know that in the consumer group meetings, we're going to be discussing this. So if anybody has any ideas about why you think this is unfair or fair or good or bad, I would really be interested in hearing those ideas so that I can take them back to the consumer group and, you know, consider them as much as we can. Thank you.

DR. ENGELJOHN: Thank you, Felicia, for bringing that in.

We will get into it a bit tomorrow, but the real issue is: We want to hear back from all stake holders on how we can all collectively move to where we want to go, which is a reduction in

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1	pathogens. So those are just what that's just one
2	of the options that we said we would consider and that
3	we prefer that that be considered, anyway.
4	(Pause.)
5	DR. ENGELJOHN: Okay. Any other questions
6	here in the room?
7	If not, then we'll adjourn for today.
8	Again, we will meet at 8:30 tomorrow upstairs in the
9	amphitheater. Thank you.
10	(Whereupon, at 5:30 p.m., the meeting was
11	recessed, to reconvene at 8:30 a.m., February 24,
12	2006.)
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