Towards a US national estimate of the risk of endemic waterborne disease - sero-epidemiologic studies

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ABSTRACT

Worldwide literature on serological methods and sero-surveys on waterborne pathogens has been reviewed. Outbreak investigation and research reports have also been examined to aid understanding of the serological response and transmission dynamics. The aim was to seek an estimate of seroprevalence and to determine if this could inform the US national estimate of risk for endemic waterborne infection associated with public water supplies. Antibody responses indicate infection, both symptomatic and asymptomatic, so probably give a truer indication of prevalence. Outbreak data can probably be regarded as the upper bound for seroprevalence estimations. Antibody is not necessarily protective per se but is a good indicator for at least partial resistance to symptomatic infection; absence of antibody will normally imply susceptibility. Pathogens transmitted by water are commonly transmitted by other routes. However, the fact that other transmission routes are more common does not detract from the potential protective effect of immunity when waterborne transmission occurs. Data indicate that seroprevalence varies widely, reflecting geographic, social and hygiene factors, but is generally greater where surface water sources are used rather than groundwater. Areas of low seroprevalence may expect a high attack rate in the event of contamination of their water supply.

Key words | Campylobacter, Cryptosporidium, ELISA, endemic infection, E. coli 0157, Giardia, molecular studies, noroviruses, outbreaks, sero-epidemiology, seroprevalence, waterborne pathogens, western blot

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ABBREVIATIONS

Ab	antibody
Ag	antigen
AHG	anti-human globulin (combined Ig classes)
BSA	Bovine serum albumin (used as blocking agent)
CMI	cell mediated immunity
ELISA	enzyme-linked immunosorbent assay
EITB	enzyme-linked immuno-electro-transfer blot
	syn. western blot
GW	groundwater
IFAT	immunofluorescent antibody test; also IIFA
	(indirect IFAT), IFA

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Ig	immunoglobulin (syn. antibody); classes IgG,
	IgA, IgM, etc.
mw	molecular weight (mass)
p.i.	post-infection
SRSV	small round structured virus (e.g. Norovirus)
TSW	treated surface water
WB	western blot
MGF	minigel format
LF	large format
OD	optical density

DEFINITIONS AND GENERAL ASPECTS OF IMMUNOLOGY

Epidemiology, at its simplest, is the study of the occurrence (number and frequency) and distribution of cases by person

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(age, gender, etc), place and time, i.e. demographics. It also seeks to describe or statistically analyse the determinants of these (e.g. risk exposures or other causal factors). Cases are usually defined by an agreed set of clinical signs and symptoms although for gastro-enteric infections these are usually not pathognomonic and may be estimated by reference to more broadly defined criteria. Precise diagnosis often requires laboratory findings, including the isolation of the causative organism and/or the detection of specific antibody using methods capable of indicating recent infection. In some cases, this may be further enhanced by the isolation of an identical organism from the putative source of the infection or vehicle of transmission such as food or water (Tillett et al. 1998). Traditionally, epidemiology is associated with the study of epidemics and localized outbreaks (two or more associated cases). Epidemiologic study methods, however, can also be used for the study of sporadic (endemic) and pseudo-sporadic (diffuse, low-level outbreaks only identified as associated by such studies and by typing of isolates, particularly if these are unusual strains) infection that may share common features with outbreaks such as potential routes of transmission, including water (Casemore 1995; Meinhardt et al. 1996; Goh et al. 2004, 2005; Hunter et al. 2004). The occurrence of such background endemic or sporadic infection is of particular importance in the context of waterborne infection, which is often thought, erroneously, to involve only outbreaks. Large outbreaks usually follow a breakdown in, or sub-optimal operation of, water treatment processes. Low-level, intermittent penetration of treatment may lead to low-level intermittent transmission to consumers who may or may not suffer overt disease but who may show an antibody response. Waterborne pathogens are commonly also transmitted by the direct person-to-person route and indirectly via food, recreational water exposure, and other exposures. This makes interpretation of epidemiological findings problematic for estimating drinking water risk because an infected individual may have become infected from a variety of sources. Risk exposures may also differ significantly between different population groups and extrapolation of findings from one population group to another needs to be done with caution.

The outcome of the contamination of water supply and transmission to consumers depends on complex dynamics with various, usually ill characterised, factors—the organism,

the exposed persons (and their levels of immunity) and the environment (Casemore 1994, 1995; Meinhardt et al. 1996). In order to better understand the natural history of waterborne infections and the complexities of the dynamics of transmission (see Figure 1), epidemiological investigations can be enhanced by serologic studies (Craun et al. 2001). The purpose of this paper concerns the national estimate of risk of endemic infection from water. However, reports on immunological studies and on the investigation of outbreaks are fundamental to the understanding of endemic infection and illness. In general terms, prevalence in outbreaks likely represents the upper boundary for endemic infection estimates.

Sero-epidemiology is a form of study in which the demonstration of antibodies to the organism of interest is used as a surrogate or marker for the distribution of infection, with or without overt disease. The samples (usually serum but sometimes other body fluids such as saliva and feces) that are analysed may be from those involved in an outbreak (cases and symptom-free "controls"). Others may be from groups (cohorts) defined by geography or other determinants, or representative sub-sets of whole populations (e.g. informed adult volunteers, anonomized blood donor samples or from patients investigated for other non-infectious conditions, referred to below as convenience samples). The purpose of seroepidemiology is to investigate the frequency of seropositivity in the group studied or, by extrapolation, to estimate the frequency in the population as a whole, and hence estimate the frequency of exposure to the organism. It may also be used to study the transmission or spread of the infecting agent in a defined cohort (e.g. within an outbreak area). Sero-prevalence is a measure of total infection levels at a point in time (point prevalence) or over a defined period (period prevalence), which may be normalized to a rate, e.g. cases per 100 000 population. It is important to recognize that circulating antibodies detected in serologic studies reflect infection per-se, with or without symptoms (i.e. clinical and sub-clinical or asymptomatic infection), at an often ill-defined point in time. Sero-incidence, on the other hand, is a measure of new acute cases in a particular location, at a point in time or over a defined period in time. This is often defined by sero-conversion, a change from sero-negative to sero-positive in a time series of samples.

CRYPTOSPORIDIUM IN WATER: DYNAMICS OF AN OUTBREAK

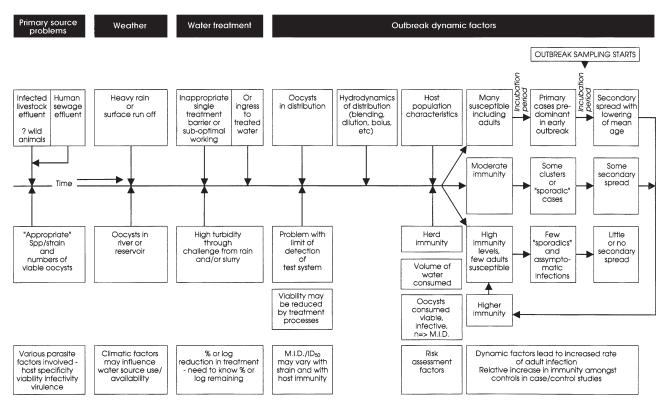


Figure 1 | Cryptosporidium in water: dynamics of outbreaks. Oocysts in water are often derived from a variety of sources and host species, which may be of variable infectivity and pathogenicity for humans. Numbers and viability decline with dilution, age and environmental attrition, including water treatment processes. These will affect the likelihood of an infective dose being ingested. The risks for target populations vary according to several factors including quantities of water consumed and levels of immunity. For a given level of contamination with a pathogenic strain several outcomes are possible: (i) in an area of low seroprevalence the attack rate may be high, primary cases will tend to include an unusual number of adult cases; secondary cases occur more frequently in children so the age-specific rate will decline as the outbreak progresses. Although seroprevalence will be higher following an outbreak, this will decline again unless there is further input of infective oocysts. (ii)Where seroprevalence is moderate, outbreaks will tend to be smaller and secondary transmission less frequent but seroprevalence may be boosted by naive individuals with primary infection and sub-clinically infected exposed persons. Such immune exposed subjects if chosen as controls will tend to reduce apparent relative risk for water consumption. (iii) In areas with high endemicity and seroprevalence, outbreaks will be uncommon but visitors and other at-risk subjects will act as sentinels of high exposure levels. This dynamic compartmentalized model permits change of risk status depending on changes in oocyst input (numbers, viability and infectivity), antibody increase post-exposure and then declining, and recruitment of susceptibles.

Alternatively, the pattern of antibody classes (see below) may be used to indicate the probability of recent or past infection. Positive reactions to some antigens are associated with early or late stages of the immune response. In addition, in some infectious diseases, serum avidity (strength and speed of binding of an antibody with its homologous antigen) is also a useful marker to distinguish recent and past infection (Joynson *et al.* 1990; Roitt & Delves 2001; Garcia *et al.* 2004; Iturriza-Gómara *et al.* 2004).

The nature of antibodies

Immunoglobulins (Ig) are present in the blood and other body fluids as part of the immune system's response to antigens present as components of infecting agents (see below). This may occur because of infection, passive transfer (e.g. from mother to fetus via the placenta), or immunization. Circulating (humoral) antibodies and secretory antibodies (in saliva and other body fluids) have a variable role in recovery and resistance to re-infection that is often poorly understood. Recovery and resistance may depend mainly or in part, according to the infecting agent, on cellular immune mechanisms such as cell-mediated immunity (CMI) or a combination of the two (Steiner & Guerrant 1996; Roitt & Delves 2001; Wyatt & McDonald 2004). However, if immunity (resistance) to re-infection is present then the presence of antibodies may provide a

useful marker for that. Importantly in the context of this report, the absence of antibodies makes it unlikely that there will be immunity and implies lack of exposure. However, it has to be recognized that infrequent exposure may lead to waning of the antibody to levels that are below the threshold of detection of the test(s) used. Such low-level antibody can be boosted by subsequent re-exposure to the same or antigenically related organisms (anamnestic response). Sero-prevalence may thus provide a useful surrogate for estimating the frequency of exposure to the infecting agent, provided that the tests are sufficiently sensitive and specific. Serology may also indicate the frequency of sub-clinical cases, for example in an outbreak, where sero-conversion or increased response (titre) is not associated with symptoms and recovery but nonetheless, does indicate recent infection. This may be of particular value where detection of the infecting agent is difficult.

Antibodies comprise several immunoglobulin classes, IgG, IgA, IgM, IgD and IgE, which have sub-classes and isotypes. Serological tests may detect whole antibody or individual classes or isotypes, depending on the test protocol. The function and significance of each of these differs but for the purposes of this report, the latter two classes, sub-classes, etc, have not been considered. IgM is usually detected early in infection and declines within a few months, often to undetectable levels, although exceptions have been described (see below). IgG often arises a little later but generally lasts for longer; if found in the absence of detectable IgM it is usually taken as evidence of past infection. The presence of IgM, with or without IgG, usually indicates recent infection. However, studies on Cryptosporidium infection, for example, have sometimes indicated prolonged (≥12 months) IgM production, sometimes in the absence of detectable IgG. The reason for this is a subject of debate but it probably indicates ongoing exposure to the stimulating organism. IgA is present in the serum (humoral antibody) and is secreted into the mucosa, during which process its structural form changes to dimeric IgA (IgAs or sIgA) (Roitt & Delves 2001; Riggs 2002). Secretory IgA can be detected in faeces and saliva and behaves differently from the humoral (monomeric) form. It is of particular relevance to parasitic and other infections in the gut mucosa. Antibody studies are sometimes used to help characterise the pathology and pathogenesis of symptoms and recovery, and for diagnosis, rather than to measure sero-prevalence or sero-incidence. Nonetheless, such studies may also yield epidemiologically useful information. The particular importance of antibody studies epidemiologically is that they offer a truer reflection of prevalence than isolation of organisms and/or clinical diagnoses, both of which underestimate true prevalence (Hunter 1997a 2000; Craun et al. 2001; Frost et al. 2003a; Priest et al. 2005).

The nature of antigens

Antigens are substances, usually protein or protein complexes, that when introduced parenterally into the body provoke an immune response, including antibody production. They will subsequently react specifically with the antibodies so produced in some observable way (precipitation, agglutination, etc). In addition, various other molecules such as carbohydrate moieties (sugars) and lipids structurally associated with those proteins may subsequently react in test systems even though not necessarily capable alone of eliciting an immune reaction (partial or incomplete antigens, or haptens). Such molecules may also mask antigenic proteins in intact cells. The antigens traditionally used in laboratory tests often consisted of suspensions of microorganisms, parasites, etc, whole or crudely extracted chemically, by heat or other physical methods. More recently, antigens are, increasingly, welldefined molecular structures or sequences, including those extracted electrophoretically from crude preparations of target organisms or genetically engineered (recombinant) antigens in a microbial cell or virus. Some antigens are associated with structural molecules (e.g. cell wall structures, tubulins, flagellins, etc) or functional molecules (e.g. toxins, intracellular enzymes such as polymerases, etc) that may be biologically well conserved and present in a wide variety of organisms, not just closely related species. These, together with partial antigens, may cause cross-reactions that make interpretation of findings problematic. Other factors that may interfere with specificity of reactions include rheumatoid factor (RF). Those exposed to cattle may react with bovine serum albumin (BSA) which is sometimes used as a blocking agent in the enzyme-linked immunosorbent assay (ELISA) (Chart et al. 1998), and animal-derived sera used as reagents may react inappropriately against target antigens or sera of other animal species used as reagents. Test reagents may also bind non-specifically and variably to the test vessel material, especially some plastics. These problems tend to be greater with ELISA and with methods in which crude target pathogen extracts have been used (Venkatesan & Wakelin 1993; Chart *et al.* 1998). It is less likely a problem if the precise molecular weight and other characteristics of the reacting antigen can be determined and the precise binding reactants identified, such as in western blotting (WB), especially with larger formats (LF WB) that permit the complete resolution of antigen complexes or families.

Test modalities

Various test modalities are used to demonstrate antigen/antibody reactions. These include immunofluorescence (IFAT/IIFT), ELISA, and WB. The results from these different methods may differ both qualitatively and quantitatively from each other and thus may make comparisons of data from different studies difficult. Antibody classes may vary in valency, avidity, etc., and in the optimum conditions required for detection and quantitation. Enzymes used in ELISA differ in their optimum conditions of pH, temperature, and chemical constituents. Batch testing of large numbers of samples may lead to variations in reliability of individual results depending on position in the tray or stack of trays because of varying temperature. Even studies using notionally the same methods may differ depending on the precise antigen used, how the test system has been optimized, and operator variability. Different immunoglobulins having differing valencies need different test conditions to optimize the antigen/antibody reaction. In addition, different test systems may be optimized to maximize either sensitivity or specificity. As a general rule of thumb, maximum sensitivity (the greatest number of true positives) may need to have a broad specificity and may thus pick up more cross-reactions (false positives). Conversely, to maximize specificity (increase precision) these potential cross-reactions have to be excluded and some true (usually low-level) positives may be lost (false negatives). Generally speaking, the cruder the antigen (such as whole cell preparations), the more likely it is to suffer from cross-reactions. ELISA is more subject to

"background noise" from such cross-reactions but has the advantage that it is generally easier to handle large numbers of samples and to automate the process. Sera can be readily titrated to provide an accurate quantitative result and a logical cut-off achieved mathematically (Cox et al. 2005). This approach has been used because of the difficulty of identifying true negative control samples (Cox, personal communication). The development of more precisely defined or genetically engineered antigens for some organisms has overcome some of these problems in recent years. WB is generally more sensitive and specific, at least in its larger format, but is technically demanding and expensive in both reagents and equipment and quantitation is often more subjective. Optical device methods have been developed to semi-quantitate WB reactions by measuring band density and/or size, but this is still likely less accurate than titration-based methods. Visual estimation of reaction strength is too subjective for comparative purposes.

Microorganisms considered in this literature review

This review includes relevant studies in countries other than the United States and United Kingdom. The discussion is arranged according to organism and considers the study designs and results.

A wide variety of pathogens may be transmitted by the water route (Hunter 1997b; Craun et al. 2002; Hrudy & Hrudy 2004). The report is divided according to selected pathogens, which are addressed in order of their importance, in terms of prevalence (endemic and/or epidemic frequency of occurrence), and resistance to amelioration of risk by conventional water treatment, and hence their importance to the estimation of risk. These are Cryptosporidium, Giardia, other parasites transmitted by water in North America; viruses, particularly Norovirus (previously Norwalk virus) and related species known as small round structured viruses (SRSVs); bacteria, particularly Campylobacter jejuni, salmonella and Escherichia coli 0157. Some other species are included for completeness although the significance of these for risk estimation is likely minimal.

CRYPTOSPORIDIUM—REPORTS ON THE SEROLOGICAL RESPONSE AND SERO-EPIDEMIOLOGY

This parasite has been the stimulus for numerous serological and related studies, partly because of its clinical impact and the lack of effective therapy, particularly on the immunocompromised. In addition, it has major impact on public health agencies and the water industry as a common waterborne pathogen, especially given its resistance to chemical disinfection such as chlorination. In addition, its relatively recent emergence has coincided with developments in molecular science that have enabled detailed studies to elucidate the natural history and transmission dynamics of infection, both sporadic (endemic) and epidemic.

Early worldwide studies

Early studies were aimed generally at (i) establishing pathogenicity by demonstrating sero-conversion (part of the Koch-Henle postulates); (ii) studying pathogenesis and understanding the role of immunity in recovery as part of the natural history of the infection; and (iii) finding more widespread evidence of infection including otherwise undetected cases in outbreaks (sero-epidemiology). Implicit in these study reports, however, is the need to report on the development of suitable methodologies, some of which predated the current wider availability of potentially more precise molecular methods. They should therefore be seen as developmental rather than definitive.

The first reported study was by Tzipori & Campbell (1981), who looked at prevalence of antibodies in ten animal species, including humans. Numbers of samples examined were small. The method employed indirect IFAT with cryostat sections from infected lamb intestinal tissue as antigen. The results suggested widespread reactivity for all species tested and in individuals of each species (80–100% positivity). The method has not generally found favour and is unsuitable for more general sero-epidemiology.

Campbell and Current demonstrated antibodies in normal and immuno-deficient persons from North America with confirmed infection, also using indirect IFAT with AHG to detect specific antibody, and with infected mouse gut tissue sections as antigen (Campbell & Current 1983; Current & Bick 1989). Test sera were serially diluted to establish titres and some subjects were sampled over a prolonged period (2–400 days). Sero-conversion was demonstrated and antibody was shown to persist in some cases for more than one year. Two subjects showed a late increase in titre that was thought to result from subsequent re-exposure with a booster effect but showed no sign of clinical illness, an important observation in the context of this report.

Koch *et al.* (1985) studied nosocomial transmission using indirect IFAT to examine sera from 26 subjects exposed to an acute case of severe cryptosporidiosis (a patient with AIDS) and 18 not so exposed. They found evidence of person-to-person transmission in hospital personnel. Thirty-one percent of the exposed had antibody, compared with 17% of the non-exposed.

Using ELISA with sonicated oocysts as antigen, Ungar et al. (1986) demonstrated antibody responses to Cryptosporidium in immunocompetent and immunocompromised persons from North America. Both groups of subjects showed an antibody response, generally IgM first, followed by IgG. The results of serum sets from subjects in an underdeveloped country (21% positive) suggested moderately widespread infection. This study used a panel of sera from "presumed negative" subjects as negative controls but recognised the difficulty of this with a caveat when interpreting results; the antigen used was a crude (polyantigenic) extract. These are important factors in assessing this and some later studies by others. Ungar and Nash (1986) used immuno-blotting (SDS-PAGE) to purify selected antigens, followed by EITB (WB) read with laser densitometry to study responses with IgG and IgM; they also used ELISA. They found that sera from infected individuals reacted with a low molecular weight (23 kDa) and additional high-mw (125-175 kDa) antigens, of which they thought that the 23 kDa antigen would prove useful (see later studies described below). Of particular note is that they found discrepancies in results with the different methods, in particular that ELISA reacted more broadly. Some sera shown to be positive by ELISA failed to show a positive reaction to the 23 kDa antigen by EITB while nine persons who were ELISA-negative and without demonstrable Cryptosporidium oocysts in stools reacted to that

antigen by EITB. Some of the discrepancies were thought to depend on differing antigen preparation methods for the different techniques. The results showed that undiagnosed infection was prevalent. The sero-epidemiology of infection in randomly selected sera from two Latin American populations was studied by Ungar et al. (1988) using ELISA with sonicated, whole, calf-derived oocysts as antigen. Again, "presumed negative" sera were used as controls despite the fact that their study showed that antibody was common in the absence of overt cryptosporidiosis. They found 64% had detectable specific IgG; 32% were also positive for IgM. This increased in the two to three year age group. IgG, and less often IgM, was found to persist for over 12 months. Ungar et al. (1989) looked for serologic evidence of infection in US Peace Corp volunteers serving in Africa, again using sonicated oocysts in ELISA. Sera were collected over a period from six weeks up to two years (time series or kinetic study samples). Positivity was assessed by reference to OD readings of a panel of sera from assumed negative (stool negative) persons. Thirty-two percent were IgG-positive initially, and an additional 5-14% became positive during the study period. Persistence of IgG and/or IgM was noted; in some cases, IgM persisted without IgG. The use of a panel of assumed negative sera would appear to be invalid given the frequency of positive findings in those not apparently suffering from cryptosporidiosis. The results showed that undiagnosed infection was prevalent. The use of the "negative" controls used in these studies will tend to raise the threshold for recognizing positive sera and may therefore reduce reported prevalence.

Casemore et al. (1986) and Casemore (1987) used IFAT with Percol®-purified whole oocysts (animal- and humanderived) fixed on multi-well microscope slides that permit screening serial dilutions to determine titres. This technique permits visualisation of the antibody/antigen binding site(s), in this case associated with the oocyst wall. Sero-conversion was demonstrated in confirmed cases during an outbreak in North Wales. Some high titre single serum responses were detected in previously undiagnosed cases (found to have been symptomatic on further investigation). IgG was found more often in non-outbreak (control) population groups who lived in rural areas when compared with an urban control group. No attempt was made to exclude low titre

responses as there appeared to be no valid evidence to assume, as some others have done, that low titres reflect cross-reactions or non-specific reactions. Some differences were noted in titre and intensity of reaction when sera were tested against oocyst isolates from different host species (calf, lamb, human). A selection of positive test sera was also checked for positivity against Toxoplasma and rheumatoid factor (known to cause false-positive reactions with IgM); all were negative. Patel et al. (1997) detected antibody bands to a variety of molecular weight antigens in convalescent sera from patients with recent cryptosporidiosis. An experimental SDS-PAGE MGF WB phenotypic typing system for oocysts and antibody detection was used to look at isolates and sera from patients associated with waterborne outbreaks in the UK and comparing these with samples from non-outbreak areas (McLaughlin et al. 1998). Antibody to low molecular weight antigens of 6, 14 and 17 kDa was found in 88% of convalescent phase sera from confirmed cases. Nine months later, sera collected for other purposes from subjects not known to have been affected showed antibody to these low mw Ags in 32-49% of residents in the same area, in 15-17% of residents in an adjacent area that received some of the affected water, and \sim 7% in control subjects not resident that area. It was subsequently shown (Patel et al. 1998; Harrison et al. 2002) that this outbreak was predominantly due to C. parvum genotype 1, now C. hominis (Morgan-Ryan et al. 2002) although approximately one third were C. parvum (then genotype 2); some mixed infections were found. There is little published evidence of how responses compare when sera are tested with heterologous and homologous species or strains. However, recent studies in the USA suggest that responses differ quantitatively when sera are tested against the infecting and heterologous isolates and species (C. parvum, C. hominis), and that variable cross-reactions occur to the different isolates (C. Chappell, personal communication; Priest et al. 2006). It is thus important to bear this in mind in interpreting serological studies used for case and/or outbreak investigations if the infecting species has not been identified. It has been shown that the epidemiology, including risk variables, for sporadic infections differs between C. parvum and C. hominis (Hunter et al. 2004). A variety of species, genotypes and hostadapted strains may be present in water, some of which are

of doubtful pathogenicity to humans but may stimulate an immune response (Chalmers *et al.* 2002*a*, *b*; Chalmers & Casemore 2004; Ong *et al.* 2002; Ward *et al.* 2002; Mathieu *et al.* 2004; Zhou *et al.* 2004).

In the USA, Mead *et al.* (1988) identified and defined specific antigens that were recognized by sera from both animals and humans using WB. They used an antigen extracted from sporozoites by SDS-PAGE. The number of antigens recognized by sera increased with time post-infection. The major antigenic determinant appeared to be 20 kDa. The response began to decline in intensity after about five months unless re-exposure occurred. Some of the sera reacted with high mw (96–200 kDa) antigens. Mead *et al.* thought that the 20 kDa Ag corresponded to the 23 kDa Ag of Ungar & Nash (1986), the difference being an artefact of the purification processes used.

A large waterborne outbreak in North America in 1987, at Carrollton, GA, was reported by Hayes *et al.* (1989). In addition to fecal sample examination, paired serum samples and other random single samples were examined for the presence of specific IgG and IgM by the method of Ungar *et al.* (1986). The results suggested that background seroprevalence was higher than shown in other North American studies.

García-Rodríguez *et al.* (1989) used indirect IFAT to detect specific IgG and IgM for a seroepidemiology study in different population groups in Spain. They detected IgG in 6.9% overall but the rate was highest in rural settings and in children (15.6%); Ig M was found in a few cases.

The immune response (IgG, IgA, and IgM) was studied in 15 stool-positive Filipino children by Laxer et al. (1990) using ELISA (modified from Ungar et al. (1986)), with purified oocysts as antigen; they also used IFA. In addition to sera, stool and duodenal fluids were also examined. Serum samples had a high (1/100) primary dilution that will likely reduce sensitivity but increase specificity. Total immunoglobulin levels and CMI markers were also measured. Antibody responses, including IgM, were shown to be marked quantitatively and qualitatively and maintained over time, which were thought to reflect the boosting effect of re-exposure. IgM (which is pentavalent) had the stronger binding capacity to surface antigens as determined by immune-electron microscopy. The relatively high screening dilution may have excluded many lower titre true positives.

A soluble (sonicated) oocyst antigen was used in ELISA by Gomez-Morales *et al.* (1992) to study sero-diagnosis (IgG, IgM) in Italian HIV-positive patients. Sera were diluted 1/50 for screening. Ninety-five percent of oocysts-positive cases were serologically positive (IgG and/or IgM) which persisted for up to a year in some; 5.3% of 300 presumed healthy subjects were also positive for IgG alone, suggesting that infection was common.

Lengerich *et al.* (1993) studied antibody in Wisconsin dairy farmers using an ELISA assay (Ungar *et al.* 1986). Results showed increased sero-prevalence compared with control subjects (44.3% of 24%, RR = 1.9; 95% CI 1.1–3.2). No evidence was given of clinical episodes but in the experience of this author (DPC) over many years, no case of clinical infection has been seen locally in livestock farmers or adults living on farms, suggesting high levels of immunity (resistance) to clinical infection result from such exposure. A note of caution, however, is that the method used requires blocking non-specific binding with BSA, to which dairy farmers may have antibodies (Chart *et al.* 1998).

Cozon and his co-workers in France looked at the humoral and secretory antibody levels in HIV-AIDS patients and the relationship of antibodies to symptomatology using ELISA (Ungar *et al.* 1986) to detect specific IgG, IgA, and IgM in serum and secretory IgA in saliva (Cozon *et al.* 1994); CD4 + lymphocytes were also assayed. The persistence of symptoms in the presence of high titre antibody suggested that these are not sufficient to control infection, an observation of particular relevance to this report.

Newman *et al.* (1994) investigated transmission within 31 households in an urban community in Northeast Brazil using ELISA (modified from Ungar *et al.* (1986)). Tests for sensitivity and specificity were done; sera from "presumed low risk" (North American) children were used as "negative" controls (see the note above). Of 202 persons providing serum samples 191 (94.6%) had antibody (IgG and/or IgM). Interestingly, five (26%) young children developed cryptosporidiosis despite serologic (IgG +) evidence of previous infection. As with Ungar *et al.* (1989), some showed persistence of IgM, sometimes in the absence of IgG, and the significance of this is discussed in this paper. Such persistence has also been noted previously (Casemore 1987; Ungar *et al.* 1989), and in some viral infections (Cox &

Medley 2003). A further report from Newman *et al.* (1999) described fecal studies on children in the same area, sampled over a four-year period, and confirmed evidence of possible recurrent or repeat symptomatic infection as well as asymptomatic cases. It was not possible however to be certain which of these recurrent stool positive cases resulted from persistent carriage, or re-infection with the same or different strains or species of *Cryptosporidium*; in either case, co-pathogens might have been responsible for some clinical episodes. An epidemiologic study of cryptosporidiosis in Peru found evidence that initial infection did not seem to protect against further clinical episodes in childhood although it was rare in adults (Bern *et al.* 2002).

Sero-epidemiology in children in Papua-New Guinea (PNG) was reported on by Groves et al. (1994), who used ELISA with purified, concentrated oocysts as antigen (Luft et al. 1987) to detect specific IgG and IgM. Test sera were diluted 1/100 (see comment re. Laxer et al. (1990)). Controls included known paired samples showing seroconversion, a known negative, a high-titre positive and wells with all reagents except test serum, to allow construction of a standard reaction (OD) curve. A high level of reaction or sero-conversion was found in 24-38% of PNG children, compared with 8% of children and 5% of adults from Melbourne. These authors found IgG and IgM to develop within a few days, reaching a peak within 3-6 weeks, and returning to near baseline levels within 1-6 months. Poor or absent IgG response was found in some cases despite the presence of IgM, as found also by (Casemore 1987; Casemore et al. 1986). Re-exposure of one individual provoked an IgG response as reported by some others above (e.g. Ungar et al. 1986). There was an overlap in antibody levels in confirmed cases and some of the controls, underlining the problem of using sera from healthy persons as so-called negative controls.

Sero-prevalence in three communities in China, Brazil and Virginia, USA, were studied by Zu *et al.* (1994) using ELISA (Ungar *et al.* 1986). Positive rates were 42.3–57.5% in children in China, increasing with age after one year, and 50% in adults. In comparative populations studied, almost all children from a semi-urban population in Brazil became positive by their second year while the rate was 16.9% in a North American population from Virginia. It was thought that the difference between the communities in rates

of acquisition of positive responses reflected differences in weaning age, hygiene practices, water quality and sanitation, family size, and the local environment, which is consistent with epidemiological findings (Casemore 1990; Casemore *et al.* 1997).

Studies by Kuhls et al. (1994) looked at seroprevalence from infancy to adolescence among 803 children of various ethnic groups in Oklahoma. They used ELISA (Ungar et al. 1986) with sonicated, calf-derived oocysts. Thirteen percent of those <5 years were seropositive; the rate was higher for those with a recent history of diarrhea and those attending day-care facilities, a known risk factor (Cordell & Addiss 1994). Thirty-eight percent of children 5–13 yr and 58% of adolescents (14-21 yr) were positive. Rates were higher for Black and Native Americans than white non-Hispanics. Interestingly, there was no significant difference between urban and more rural residents. In a later study but using similar methodology, Leach et al. (2000) looked at seroprevalence in children from various population groups along the Texas-Mexico border using ELISA to measure specific IgG and IgA. Overall, 70.2% were positive; prevalence was higher (82 – 89%) in the colonias and border communities compared with urban, non-border communities (46%). Risk factors identified included, in addition to young age and socioeconomic status, the consumption of municipal water.

In São Paulo, Brazil, Braz *et al.* (1996) used IIF with purified oocysts to measure IgG and IgM. Oocysts were fixed on multi-well microscope slides and these were used to test two-fold test serum dilutions from 1/10; known positive and negative controls were used. Sixty-two percent of children found to be excreting oocysts were positive for both IgG and IgM. Children found negative for oocysts showed 20% positive for IgG and 40% for IgM, indicating likely infection not detected by fecal screening. In a group of HIV-positive adults with positive stools, 57% were positive for IgG and only 2% for IgM. The technique was viewed primarily as an adjunct to fecal study for diagnosis.

In Germany, Petry (1998) used a previously described ELISA test (Ungar *et al.* 1986), with mouse-derived freeze-thawed oocysts as antigen, to study the specific antibody status (IgG, IgM) of 495 persons of all age groups. Positive results were found in 15.4% despite a low incidence (<2%) of fecal positives in diarrheal patients. Fifty percent of sera with high IgG (30 sera) were also positive for IgM,

suggesting recent infection. Sera were tested at 1/100 dilution and the authors suggest that the study may consequently underestimate true seroprevalence.

Miron *et al.* (2000) in Israel studied age-related seroprevalence in children by demonstrating acquisition of specific IgG and IgA by ELISA with sonicated oocysts as antigen, using previously described methods (Ungar *et al.* 1986; Kuhls *et al.* 1994). Serum from a confirmed case was used as a positive control; pooled sera from presumed negative children were used as a negative control. Test sera were diluted 1/100 (see previous comments). Sero-positivity (any Ig) varied from 50.9% to 95.6% with an overall prevalence of 65.6%. Like Casemore and colleagues (Casemore 1987; Casemore *et al.* 1986), they found that IgA was sometimes present in the absence of IgG, also reported by others with IgM. The study indicated high prevalence of the infection and that stool examination results underestimate this.

Steinberg and others studied seroprevalence of several waterborne infections in infants 6–36 months old in Guatemala (Steinberg *et al.* 2004). They used the second generation ELISA (see below, Priest *et al.* 2001) to measure *Cryptosporidium parvum*-specific IgG against the 27 kDa Ag. Antibody levels in 150 (28%) of the cohort samples increased rapidly from 27% at six months (presumably mainly maternal Ab) to 70% at 19–24 months, from when it remained at about that level. The authors expressed the view that serology was a useful tool for estimating prevalence and indicating need for intervention but felt that sero-incidence rates would also be useful.

Four studies by Frost and his co-workers examined sera from overseas locations. High levels of oocysts were reported to have been detected in drinking water in Sydney, Australia but without a concomitant rise in case numbers (Frost *et al.* 2000*d*). Sera were collected from blood donors (convenience samples) from Sydney and Melbourne, and these were assayed by MGF WB for IgG antibodies to 15/17 and 27 kDa Ags. Over half of sera in both groups had antibody with no statistically significant differences between the two groups. This gave further support to the view that the supposed oocyst detection and/or enumeration on Sydney drinking water was flawed. Frost and others looked for serological evidence of infection with *Cryptosporidium* in Italy using the same methodology (Frost *et al.* 2000*a*). The response rate in 100 Italian blood donors was higher

(83% for the 15/17 kDa Ag; 62% for the 27 kDa Ag) than for four United States blood donor populations. The responses for the 15/17 kDa Ag were more pronounced (intensity of reaction has been used as a surrogate for titre). Responses were less pronounced than in sera collected six months after an outbreak in the US but higher than sera collected 2.5 years later. Increased reactivity was associated with increased age. Confirmed diagnosis of sporadic Cryptosporidium infection or of outbreaks is uncommon in the area but the results suggest that exposure is common. The findings suggest that the test may be of low diagnostic specificity for symptomatic infection but of high sensitivity for evidence of exposure. Egorov et al. (2003) examined sera collected in Russia, which were examined by the same methodology. Sixty-eight percent were positive for antibody to the 15/17 kDa Ag and 88% to the 27 kDa Ag. As with the previous study, increased age was associated with increased intensity of the reaction consistent with recurrent exposure. Swimming pool use was associated with increased positivity for both antigens while drinking water from shallow wells was associated with increase in the reaction to the 27 kDa Ag. Both of these practices are recognized risk factors for cryptosporidiosis (Casemore et al. 1997; Meinhardt et al. 1996). Blood donor sera collected from residents of two towns in New Zealand were studied by the same methods and risk exposures estimated by questionnaire responses (Duncanson et al. 2003). Over 60% of sera from both population groups were positive for antibody to one or both antigens and questionnaires gave positive associations for recognized risk factors. These four studies suggest that meaningful comparisons may be made between different population groups, and may help to identify or confirm likely risk factors.

Cox et al. (2005) studied seroprevalence of IgG to the immunodominant 27 kDa antigen in a Brazilian population using a recombinant form of the antigen, cp23 (see below, Priest et al. 1999), in ELISA and MGF WB (method of Frost et al. 1998a). In the ELISA test, a serially diluted high-titre human serum was included in each plate and given an arbitrary unitage. Positivity/negativity of test sera was determined by (i) normal frequency distribution of antibody concentrations (Cox et al. 1998a, b); and (ii) a cut-off point established at 10% of the positive control. Positivity for WB was by reference to the known positive control serum.

Sero-positivity was low in younger infants, increased to $\sim 60\%$ by five years and then 80% by age 10 years, after which the level remained constant. There was also evidence that antibody concentrations (titre) increased with age. The normal frequency method for determining positivity did not reveal an obvious cut-off value. There was a strong correlation between WB and ELISA (r=0.88, P<0.005) but sera with levels near the control cut-off threshold (10% of positive control) were more likely to be found positive when tested by ELISA. Overall, ELISA-positive levels were a higher percentage level (= titre) compared with the positive control. The study suggested that cp23Ag in ELISA may be the more sensitive method when detecting low levels of antibody.

Discussion of early worldwide studies

These early, worldwide studies describe methodologies that were generally useful for further development and refinement. Some were aimed more at understanding the pathology of the infection and characterizing the immune response to it. In addition, however, some do provide useful comparative epidemiological data on epidemic (outbreak) and endemic (sporadic) infection dynamics and prevalence in a variety of settings. Most workers used ELISA, often based on the method described by Ungar and her colleagues (Ungar et al. 1986). Most used calf-derived oocysts, which were *Cryptosporidium parvum* (previously referred to as *C*. parvum genotype 2). However, some of the infections are likely to have been with C. hominis (previously C. parvum genotype 1) and possibly other species (Patel et al. 1998; Ong et al. 1999; Chalmers et al. 2002a, b; Xiao et al. 2004; Priest et al. 2006). Although these appear to share crossreacting antigens, the precise difference in reactivity has not yet been clearly defined. The impact of these related species in inducing secondary or anamnestic responses is also unknown. Tests, such as ELISA, that may rely on crude antigen extracts are known to produce significant crossreactions or "background noise" that makes interpretation difficult, especially of low titres. RF, BSA and other constituents such as reaction vessel plastics, may also interfere with the test (Venkatesan & Wakelin 1993; Chart et al. 1998). The studies by Ungar and Nash (1986) and Mead et al. (1988) used western blotting that enabled

identification of important antigen molecules. This more specific approach has subsequently been greatly enhanced by the more extensive molecular studies reported from various workers described below. Répérant *et al.* (1994) found that antigens of mw 15–17 kDa and 21–23 kDa were major immunogenic molecules in different host species including humans, the larger of which seemed to correspond to the antigen identified by Ungar and Nash (1986).

Recent North American studies

Several highly experienced research groups in North America have studied the immune response and seroprevalence. Some have further developed methodologies, particularly in the use of better-defined antigens, and significant insights have emerged with the use of these methodologies. Some of the reports described below predate these later developments and used previously established methodologies. They are included here to show the context of those developments.

DuPont and colleagues (DuPont et al. 1995; Chappell et al. 1996, 1999) studied experimental infection dynamics and clinical response in a group of healthy young adult volunteers. The antibody pre-exposure status and responses to infection of volunteers were assessed by ELISA adapted from the method of Ungar et al. (1986) using a crude oocyst extract antigen. Some discrepant findings were thought to reflect insensitivity of the ELISA and that low-level antibody in some of the pre-exposure samples may have been missed. This has been the subject of subsequent debate (Frost & Craun 1998; Okhuysen et al. 1998; Chappell et al. 2001; Muller et al. 2001; Frost et al. 2003b; Priest et al. 2003). These wellcharacterized infections were used to study further the immune response (Okhuysen et al. 1998; Moss et al. 1998a, b; Chappell et al. 1999). Thus, Okhuysen and co-workers reported on susceptibility, oocyst excretion and serologic response to attempted re-infection with *C. parvum* one year after the original infectivity studies (Okhuysen et al. 1998). Antibodies (IgG, IgA and IgM) at 0 and +45 days p.i. were detected using ELISA as previously described (DuPont et al. 1995). Rates for diarrhea in this group were about the same, but the diarrhea was generally less severe. IgM responses were also less marked than after primary infection; IgG and IgA seroconversions increased although the responses did

not appear to correlate with the presence or absence of infection. Some of the apparent discrepancies may have been the result of lower oocyst excretion, below the threshold of detection. On the other hand it probably also reflects the low sensitivity of the ELISA used; it would perhaps have been useful had the serum samples been reexamined by WB. The infectivity of C. parvum in healthy adult volunteers with pre-existing specific IgG was also assessed (Chappell et al. 1999). These authors used the ELISA previously described (DuPont et al. 1995). Positive and negative control sera were included but it was not specified how their status was assessed. The positivity of test sera was defined as those giving 1.5 times the OD of the negative controls, and post-challenge change estimated using that baseline value. The authors noted decreased symptoms in these study subjects. They further suggested that a significant proportion of the population might experience protection from low-level exposure although it was accepted that it was not possible to determine whether that was directly related to the antibody or to other immune mechanisms such as CMI. Chappell et al. (1999) estimated that a population with a seroprevalence of 25% might experience a corresponding percentage reduction in clinical cases compared with a sero-negative population. A sensitive and specific and carefully controlled antibody test system is clearly a prerequisite for studies on infection dynamics.

Some of these workers and others (Kjos et al. 2005) have subsequently described the use of a recombinant 41 kDa (rCP41) antigen associated in its native state with the oocyst wall of *C. parvum* but not *C. baileyi* (Jenkins *et al.* 1999). An ELISA test was used for the detection of IgG and IgM, with inclusion of positive and negative control sera. They compared a crude oocyst-derived antigen to test a panel of sera, with highly concordant results (88%, P < 0.0001) for IgG but less well (79%) for IgM, which was more readily detected using native antigen. The sensitivity and specificity were assessed as 94.8% and 77.6% for IgG compared with 48.4% and 93.1% for IgM for the two antigens. The poor concordance for IgM detection did not appear to be related to low-level positive samples. They suggest that this test might provide a reliable and cost-effective method for assessing previous exposure to Cryptosporidium.

Moss et al. (1998a) also used sera from the volunteer infectivity study (DuPont et al. 1995; Chappell et al. 1996) to measure the immune response to 15, 17, and 27 kDa antigens of Cryptosporidium using WB and purified oocyst extracts as previously described. Possible prior exposure in the volunteers was assessed by ELISA (DuPont et al. 1995), which has been the subject of some debate (see above). This study showed that antibody level increase was more marked in symptomatic than asymptomatic subjects. In contrast, volunteers with pre-existing IgG to the 27 kDa Ag excreted fewer oocysts. Asymptomatic infected subjects showed higher initial (day 0) reactivity in IgG to the 17 kDa Ag and for IgM against the 27 kDa Ag. These results suggest that those with pre-existing antibody at the time of exposure may fail to show a change in antibody levels and to be less likely to develop symptomatic infection even though infected. These are critical observations in terms of this report. Some of these antigens, especially in the ~ 15 - $17 \, kDa$ and $\sim 20-23$ and $27 \, kDa$ ranges referred to separately by some authors, represent interrelated groups or families sharing common protein antigens with variable sugar and lipid moieties and with differing cleavage patterns (Mead et al. 1988; Priest et al. 1999, 2001; Riggs 2002). The response to the 27 kDa can be detected using a CP23 recombinant antigen (Priest et al. 2001; Smith et al. 2001). These antigens are associated with the sporozoite surface and probably merozoites, and may thus have potential involvement in any protective effect.

Antibodies have also been detected in fecal fluid, including from experimentally infected volunteers (Kapel *et al.* 1993; Dann *et al.* 2000). It was suggested that this might be more sensitive than detection of humoral antibody but this has been disputed (Muller *et al.* 2001).

Moss *et al.* (1994, 1998*a*) reported on studies related to the massive outbreak in Milwaukee in 1993 (Roy *et al.* 2006), in particular, the serological response of cases that occurred among the crew of a US Coast Guard cutter that had filled its water tanks from the contaminated supply. They used ELISA and EITB (WB) with proteins extracted from calf-derived oocysts that had been sonicated, freezethawed to disrupt them, followed by clarification, and then purified by SDS-PAGE. The method distinguished reactions with 15 kDa, 17 kDa and 27 kDa antigens; IgA was measured against the 17 kDa Ag, IgM to the 27 kDa

Ag and IgG to all three. Changes in intensity of reaction in paired serum samples measured by EITB for these antigens were useful as diagnostic markers when compared with stool results and the clinical picture. Of the 10 confirmed cases, only four showed IgG responses. Results for ELISA did not always identify confirmed cases and did not correlate well with EITB results. EITB gave significantly better correlation with risk (quantity of water consumed) than ELISA, and EITB was considered potentially useful as an epidemiological tool. McDonald and colleagues (McDonald et al. 2001) studied the specific antibody response among children residing in Milwaukee during the outbreak there. They used ELISA to study the response to the 17 and 27 kDa antigens (Priest et al. 1999). The former was partially purified (Triton X-114 treated) native oocystderived antigen while the latter was a recombinant antigen. Controls included WB-confirmed negative samples that were used to calculate OD values indicating a positive result. Hence, the cut-off is more likely accurate than assumed negatives. Sensitivity and specificity were estimated to be >90% relative to WB. The positivity rates increased from 15% to 82% and 17% to 87% for each antibody respectively over a five-week period. Increases noted from children in adjacent areas were smaller but nonetheless suggested more widespread infection than had been thought, as has been noted by others (Pollok et al. 1998).

Priest and his colleagues (Priest et al. 1999) used two antigens, of mw 17 kDa and 27 kDa (McDonald et al. 2001), in an ELISA to detect IgG specific to those antigens. Positive responses to the antigens in the ELISA were re-examined by WB, which showed sensitivity and specificity of ≥90%. These new ELISAs were more sensitive and specific than those using crude oocyst antigens. The IgG levels in sera collected during outbreaks were 2.5-fold higher than non-outbreak sera. This improved test protocol should therefore be of value in epidemiologic studies. The confirmation of the lower sensitivity of the older test format may confirm that this is a source of discrepant findings in some of the earlier studies, including the volunteer infectivity studies. These authors make the point that an ELISA is needed that gives the sensitivity and specificity of LF WB.

Priest *et al.* (2001) extended the above studies to assay IgG in time-series (longitudinal) serum samples from

patients with confirmed cryptosporidiosis. These sera were collected from cases in outbreaks in British Columbia, Canada (see below). The two groups in Atlanta and Vancouver compared results and determined inter- and intra-laboratory consensus on results for comparison of the methods, critical for determining robustness. They compared the two ELISAs previously described with large format (LF) and minigel format (MGF) WB. The LF WB is said to be both sensitive and specific but is generally too cumbersome and expensive for large-scale screening; the MGF is simpler and uses ten-fold less antigen and is therefore cheaper. The MGF compared well with the LF WB for the 17 kDa Ag but failed to detect nearly a third of samples positive for the 27 kDa Ag. Other studies have noted that one third to a half of sera positive by LF WB may have a response only to the larger mw antigen (Frost et al. 1998a; Priest et al. 1999). In addition, MGF is less able to resolve antigens close in size although this may be of little significance in comparing cohort surveys where the same method has been used. The authors conclude, however, that the MGF WB in its current form may significantly underestimate seroprevalence. The ELISAs were generally found to be both sensitive and specific and to provide a convenient test format for large-scale screening. Some discrepancies were found between ELISA and WB. The results for the smaller mw native (Triton-extracted) antigen were somewhat lower than in WB with sequential sera, especially for later specimens (>92 days) but many of these subsequently became negative by WB. Thus, they appeared to result primarily from lower sensitivity with borderline positive samples. The authors feel that the ELISA test using the 17 kDa is still of value for detecting recent infection, while the 27 kDa (Cp23) antigen test is a valuable tool for assessing past exposure. It should be noted that using multiple test formats always results in some discrepant results, as is often also the case for the same test protocol used in different laboratories. ELISA is prone to a variety of inherent errors and great care is required in performance and the use of the necessary quality controls (Venkatesan & Wakelin 1993). The ELISA format described by these authors addresses these problems. In particular, the authors believe that the ELISA tests minimize the effect of cross-reactions from other organisms, many of which are derived from non-protein epitopes. Carbohydrate moieties,

alone or in combination with lipids or proteins, are likely to be important determinants in the immune response to *Cryptosporidium* although they may have a broader reactivity than protein epitopes alone (Luft *et al.* 1987). Antibody to both antigens appeared to have half-lives of about twelve weeks and reach a constant baseline by about one year.

In Canada, Isaac-Renton et al. (1999, 2003) and Ong et al. (2005) reported on sero-prevalence in British Columbia, and Ong et al. (1999) reported on the molecular epidemiology of outbreak isolates. A comparison was made of prevalence of antibodies to Cryptosporidium and Giardia in three communities with different types of water supply (Isaac-Renton et al. 1999). In the first study, Cryptosporidium-specific IgG in 1944 sera were tested by MGF WB to detect reactions to the 15/17 kDa and/or 27 kDa antigens. The overall rate for Cryptosporidium-specific antibody was 50.5%; rates varied by season and between communities/water supply types. In one of the communities that experienced an outbreak of cryptosporidiosis, reaction to both 15/17 kDa and 27 kDa antigens appeared to be the best marker for recent infection. Subsequent re-examination of these sera by the second generation ELISA test (Priest et al. 2001) used in the following study showed that the miniblot significantly under-estimated positive sera (82.6% cf 50.5%). In the later study, Isaac-Renton et al. (2003) and Ong et al. (2005) used ELISA to study the IgG Ab response to the 27 kDa recombinant antigen with arbitrary units to determine cut-off for ascribing positivity. Sera from more than 4000 pregnant women in six communities in British Columbia were collected over a 24-month period. Seroprevalence was high (85% overall, range 77 – 92%) in all of the districts, but with significant differences between communities. Two of the communities had waterborne outbreaks during the study period and showed sharp rises in positive rates that declined over the following 2-3 months. Significantly, in the context of this report, one of these outbreaks was confirmed to be due to C. hominis and the other to *C. parvum*. This test has been shown to be more sensitive and specific than ELISA using the 17 kDa native Ag and is useful in detecting past infection (Priest et al. 1999, 2001; McDonald et al. 2001). The ELISA results remained significantly better than miniblot even if the arbitrary cut-off was increased by 50%. The authors expressed the view that the test was more useful for the type of epidemiologic study described here; it also appeared to be significantly better than

oocyst detection rates for estimating the size of outbreaks. It is well known that detection of oocysts in feces, as with Giardia cysts, is very inefficient (Frost & Craun 1998b). They point out, however, that a proportion of the antibody responders may be asymptomatic; the significance of such infections for potential secondary transmission is unknown. In the experience of the author of this report, such cases excrete only small numbers of oocysts at the limit of detection and probably do not represent a significant risk of transmission in most case where domestic and personal hygiene standards are reasonable (Casemore 1989). The studies were subjected to rigorous control and statistical analysis, which showed that the modified ELISA transferred well between laboratories and was robust. There was no evidence of cross-reactivity with Toxoplasma and Giardia; some studies have suggested a degree of cross-reactivity between Cryptosporidium and Eimeria spp (Ortega-Mora et al. 1992) but this is probably of little significance in human subjects. This is the most comprehensive and robust study to date and confirms the value of the second generation (CDC) ELISA for mass population (seroprevalence) studies. However, discrepant results have been reported using this ELISA, which is a subject of debate (Frost et al. 2003b).

Several groups have used these more advanced methods to study HIV-infected subjects. Caputo and colleagues looked at the antigenic determinants of antibody to Cryptosporidium among gay and bisexual men in Melbourne, Australia, with HIV infection, using MGF WB with 15/17 and 27 kDa Ags (Caputo et al. 1999). Details are given of the way in which the test was calibrated. A positive test indicative of recent infection was indicated when the test serum result was > 35% of the positive control. The 27 kDa Ag response gave the most reliable indication of exposure to risk factors over a two-year period. The same group followed up subjects after one year and found 34% to have seroconverted; seroconversion correlated with patterns of sexual activity (Friedman et al. 2001). Spencer et al. (1997) studied seroprevalence and the prevalence of cryptosporidiosis in HIV-infected persons in New York. They used the ELISA method described by Ungar et al. (1986) to measure specific IgG and stool examination for evidence of previously undetected current infection (patients with previously diagnosed Cryptosporidium infection were excluded from the study). Seroprevalence was 20.3%; evidence by fecal

oocyst detection of newly recognized current infection was found in 3.6% but the authors note that this is likely an underestimate given the poor sensitivity of oocyst detection methods. Eisenberg, together with Priest and colleagues (Eisenberg et al. 2001), used the new ELISA (Priest et al. 1999, 2001) for detecting the serologic response in 11 HIVinfected persons with confirmed cryptosporidiosis. They found that the test was reliably predictive of infection and could provide an effective epidemiologic tool to monitor Cryptosporidium infection in immunocompromised patients. Frost et al. (2005a, b) looked for evidence of protective immunity associated with a strong serologic response among HIV-infected individuals. Such an association was demonstrated between the 27 kDa antigen and a reduced risk of diarrhea in those not exhibiting weight loss; the association was not present in those with diarrhea plus weight loss. The authors suggest that this demonstrated protective immunity against the effects of cryptosporidiosis although it is not evidence that the antibody itself is protective. Their data suggest that the test can be an effective tool to monitor Cryptosporidium infection in immunocompromised patients. These studies suggest that HIV-infected subjects may well respond serologically in much the same way as those with intact immune function, a factor that may well be significant for sero-epidemiologic surveys and risk evaluation. However, the severity and persistence of the infection underlines the critical nature of the CMI response in immunity to Cryptosporidium.

Frost and colleagues conducted a number of studies of seroprevalence in North America. A comparative study (Frost et al. 1998b) estimated seroprevalence to what are described as three antigens (15, 17 and 27 kDa) by both ELISA (Ungar & Nash 1986), and by LF WB (Moss et al. 1994). The WB tested for IgG and IgA, while the ELISA tested for combined IgG, IgA and IgM (AHG). Antigen was SDS-PAGE-purified, sonicated oocyst extract. Sera were from subjects without a known history of cryptosporidiosis, from Talent, in Jackson County, Oregon, four to six months after a waterborne outbreak there. Pooled known negative and positive sera were used as controls in the ELISA; single known positive and negative sera were used as controls in the WB. ELISA tests were read by plate reader and WB strips were scanned and assessed for positivity electronically by reference to the control reactions, thus decreasing subjectivity. Positives in the ELISA were those that were \geq 25% of the positive control, and \geq 35% for WB. The rationale for these limits and other statistical aspects are discussed. The authors report that tests showed poor separation of the 15 kDa antigen group from the 17 kDa antigen group. However, these are now believed to represent the same antigen group. Various potential risk factors were included in the questionnaires used. The numbers of subjects with the main risk exposure histories were small. Both tests detected an almost two-fold increase in positives for those with a history of consumption of Talent water but the differences were statistically significant only for WB. The outbreak investigation was particularly significant in the context of this report in that it appeared to demonstrate protection (negligible attack rate or undetected asymptomatic infection) in the population who had been regularly exposed to surface water-derived supply compared with a nearby population who had not previously been so exposed but who were temporarily supplied from the suspect source. In a further study, Frost et al. (1998b) reported on a two-year follow-up to the same outbreak using paired sera collected at six months and 2.5 years later. The sera were examined by MGF WB to the 15/17 and 27 kDa Ags. Intensity of the former remained largely unchanged, while those to the latter declined to 54% over the follow-up period. Reaction intensity was used as a measure thought to approximate to titre. Increased levels were noted in Talent residents suggesting high prevalence of endemic infection and/or re-infection. The inability of MGF WB to resolve some antigens in the same size range as key antigens may be of little significance in this context where the change over time for the same group is being measured. The 27 kDa Ag response is known to persist somewhat longer than that for the smaller mw antigens but the MGF WB may be less sensitive with this antigen.

The possible effect of enhanced water treatment on seroprevalence was investigated by Frost *et al.* (2000*b*) in an unnamed city in the northeastern USA. The water, which was surface source-derived, had been chlorinated only, but filtration was introduced in spring, 1997. The assay method was MGF WB as used previously by these authors. Serum samples were collected from college student volunteers, one month prior to (107 samples) and five months after (225 samples) the introduction of filtration; most volunteers

were female. Nineteen percent of the first sera showed reaction to the 15/17 kDa Ag, changing to 24% of the second sera. For the 27 kDa Ag the figures were 27% and 41% respectively; the increase in reactions with the larger mw antigen was significant (P = 0.02) and may have been an underestimate given the decreased sensitivity of this test for reactions with this antigen. The 15/17 kDaAg response tends to last for up to six months; the 27 kDaAg reaction tends to persist for more than 12 months. The results suggested an increased rate of exposure over the study period. Statistical analysis of risk factors suggested that swimming and consumption of untreated surface water were predictive for increased intensity of response. The study period covered the summer months when these exposures might be expected to increase. It would have been helpful had the second serum samples been collected one year after the first for the following reasons:

- (i) to avoid this period of confounding risk exposures;
- (ii) to allow for the expected decline in antibody levels resulting from infection prior to the change in water treatment;
- (iii) for the seasonal exposures to be approximately the same, including such factors as the impact of spring period conditions on the source water. Seasonal changes in incidence and risk factors are well recognized (Casemore *et al.* 1997). It is not possible to assess such factors in the study area, other than the likely increased exposure to surface waters during summer, a recognized risk activity for *Cryptosporidium* infection.

Frost and colleagues also conducted a serological study on 89 convenience sera collected from anonymous persons in Collingwood, Ontario, during and following an outbreak there in 1996; 80 sera were also obtained from persons in Toronto for comparison (Frost *et al.* 2000c). Sera were assayed using the same methodology as before. The occurrence of an outbreak had been questioned because of the differing age distribution of cases between affected residents and visitors. A higher proportion of sera from Collingwood had a detectable response and the mean intensity was higher than in the Toronto sera. The mean intensity for Collingwood sera was highest in sera collected during the eight-week period following the initial case

reports. Cases were mainly in visitors, in elderly nursing home residents of Collingwood and in local children but not in other local adults. This is consistent with the findings in a waterborne outbreak in the south-west of England, in which the faulty water supply had been associated with a previous outbreak, almost certainly resulting in increased resistance to symptomatic infection in residents other than young children and those only recently resident, including retirees from other areas (Harrison *et al.* 2002).

Frost (1998) reported on a study of blood donor sera from two cities in the northwest of the USA, one with a surface-derived water supply and the other groundwater. The sera were tested for IgG to the $15/17\,kDa$ and $27\,kDa$ Ags in a MGF WB test format. Sera from the former group had significantly more positive reacting sera (21-31%) compared with the latter (11-23%). Frost et al. (2001) examined 200 convenience (blood donor) sera from each of two cities in southwest USA, which have contrasting water sources expected to lead to differing risk indexes. The sera were tested as in the previous report. There was an increased positive rate and higher levels of reactivity among the population with a surface-derived water supply than among the other population group who had a groundwater-derived supply. Surprisingly, the higher risk group did not show decreased antibody reactions if they consumed bottled water and this requires further study to explore possible confounding risk factors. The epidemiologic differences between the populations represented by the sample donors, including differing rates of reported infection, are discussed.

Frost *et al.* (2002*a*) compared seroprevalence in convenience samples from two study groups from two midwestern US cities; two sets of serum samples were collected nine months apart. One of the cities was supplied with drinking water from surface water and the other from groundwater sources. Sera were examined for specific IgG to the 15/17 kDa and 27 kDa Ags, by MGF WB (Okhuysen *et al.* 1998; Frost *et al.* 2000*c*) for evidence of differing exposure rates. Initial samples showed higher rates of antibody (54% *vs.* 38%, RP = 1.39, CI 1.21–1.60) in those from the surface water supply city. Follow-up sera from those with no baseline response to the 15/17 kDa Ag (indicative of recent infection) showed an increased frequency of seroconversions (33% surface water cf 11% groundwater at 20% of positive control

cut-off). Those with an initial reaction showed increased intensity of responses to both antigens. Although it is not possible to exclude all possible confounding risk exposures, the results strongly suggest that a surface water-derived supply leads to a significantly increased rate of exposure, as might be expected. This may not be expressed clinically and the authors reasonably argue that mild, self-limited or subclinical infection is a more likely outcome of exposure in this setting.

A further study by Frost et al. (2003c) examined differences in serological responses to Cryptosporidium antigens among residents of areas with surface and groundwater sources. Nearly 500 recruits (urban adult blood donors and college students) provided convenience samples, of which 270 also supplied follow-up samples (90-180 days post-initial sample) for kinetic studies. Sera were analysed by MGF WB as previously described by these authors to detect responses to the 15/17 and 27 kDa Ags. Intensity of the reaction was estimated photometrically using pixel density to give a proxy for titre. Participants were questioned about a variety of potential exposures. Participants from the surface water area had a higher seroprevalence to the 15/17 kDa Ag (72.3% vs. 52.4%, P < 0.02) and to the 27 kDa Ag (82.6% vs. 72.5%, P < 0.02). Shorter residence in the higher risk area or consumption of bottled water was associated with lower seropositivity rates. Seroconversion between paired samples to the 15/17 kDa Ag was more common in the surface water area. Use of private wells was also associated with higher seroprevalence to the 15/17 kDa Ag. It is notable that even in the groundwater areas seroprevalence rates were high (>50%) but it is not possible, as with most of the studies described, to draw any conclusions about the likely source of those infections or how often this involved acute symptomatic infection.

Sera from 1356 NHANES III participants were analyzed by Frost *et al.* (2004) for specific IgG. Samples were drawn from seven areas with varying population characteristics, water supplies, etc. They used MGF WB with the antigens and methods previously described by this group. Intensity of reaction was compared by geography, age, sex, race/ethnicity, income, and hepatitis A virus seropositivity. Results showed that Hispanics, blacks and females had higher seropositivity; the first two of these may be proxies

for socio-economic factors, the latter to increase exposure to infants and children. Significant differences were seen with different geographic areas that may reflect differing water sources, including catchment control. Increasing positivity was also noted with increasing age (~ 70% by age 70). The authors suggest that this finding is likely a true reflection of recurrent exposure to *Cryptosporidium* rather than the result of anamnestic boosting from cross-reacting antigens from other species. They suggest that this is more likely true with WB than with ELISA although this is less likely true with the second generation ELISA. While specificity has been less reliable with ELISA using a crude oocyst extract as antigen, the much-improved second generation ELISA appears to be more dependable.

Frost et al. (2005a) conducted a study designed to estimate the importance of protective immunity as an indicator of decreased risk for acquiring cryptosporidiosis. Serum samples and enteric illness records were obtained over a two-year period from 326 people. These were supplied with drinking water from either unfiltered surface sources or a groundwater source; filtration was initiated at one of the groundwater sources during the study period. Analytical methods used were as previously described. Subjects with strong responses to the 15/17 kDa Ag (acute phase reaction) had <65% of the risk of 1-3-day episodes of gastrointestinal illness and <40% of the risk of ≥ 4 -day episodes compared with those who had a moderate response. Water source, treatment intervention, and very weak responses were unrelated to illness events. The authors conclude that endemic Cryptosporidium infections are a common cause of gastrointestinal illness in those with a moderately strong response to the 15/17 kDa Ag. Further, they conclude that users of surface-derived drinking water are more likely to have such antibody and will thus be at lower risk of more severe illness. While this is the finding of a number of studies, the use of serologic findings as an indicator of immunity (i.e. resistance) is a matter of debate. However, it is consistent with epidemiologic findings as a marker for resistance. The difference in responses associated with the different supplies during the two stages of the study and in relation to water treatment change, is unclear. While acute, primary cryptosporidiosis tends to be associated with more extended illness, i.e. ≥ 4 days (Casemore 1989), it is unknown what the cause of illness

was in these subjects. In addition, risk level estimate has to be predicated on infection (asymptomatic as well as symptomatic) and not just clinical illness. In a further study by Frost and colleagues (2005b), serological responses in convenience samples from women in Hungary were analysed according to water source, using MF WB. Results from those consuming riverbank-filtered water indicated protection from infection by this method of natural filtration. It should be noted, however, that some outbreaks in the UK have been attributed to use of bank-side abstraction (Harrison et al. 2002) or migration through a river-associated aquifer possibly due to drought induced fissures (Willocks et al. 1998). The distance between surface water and the well, and the nature of the subsoil or aquifer are thus critical.

Discussion of Cryptosporidium sero-epidemiology

Some of the studies described above were generally designed to investigate infectivity and/or pathogenesis but also yield useful epidemiologic information incidentally. Others were designed specifically to generate epidemiologic data (see Table 1). In both cases, many of the methods used are inherently developmental. The differing test modalities described, the antigens used, the choice of controls, their differing levels of development and optimization make it fundamentally important not to compare too closely the absolute values or levels of antibodies detected. A given test system, if operated in the same way in different times or places, may be used to compare trends for positivity but may not be used reliably to compare quantitative values (titre or strength of reaction, or the precise proportion of the test population found to be positive). Where different modalities are used, or similar tests are optimized differently then especially quantitative, is problematic. comparison. Comparison of trends and epidemiological interpretations may however be possible. The benefits and drawbacks of each method need to be recognized, reflecting the continuously developing nature of the test systems and antigens used. In a useful review, Frost et al. (2003a) have collated information comparing antibody responses according to water sources. This showed that, despite the geographically variable results, surface water-derived supplies tend to be associated with raised seroprevalence and hence increased rates of infection but not necessarily clinical disease. Prior immunity may result in misclassification of infected but asymptomatic persons as uninfected (Casemore 1995; Hunter 1997a, 2000; Craun et al. 2001). When confirmed cases and other symptomatic persons (who may have been true but unconfirmed cases) have been excluded from potential control groups then the remaining cohort will likely contain an increased proportion of subjects who though exposed have not developed symptoms because of pre-existing immunity. Thus, such misclassified cases introduce bias in case-control studies that will tend to reduce the statistical association with water because such immune subjects may have consumed water from the same supply as cases.

Water may contain a variety of different Cryptosporidium species and sub-species or strains (Ong et al. 1999; Chalmers et al. 2002a, b; Ward et al. 2002; Xiao & Ryan 2004; Zhou et al. 2004) that may or may not be pathogenic for humans or may be of low virulence or low viability resulting from environmental attrition, including water treatment processes. By definition, outbreaks must depend upon the presence of more pathogenic species or types. However, less pathogenic isolates may be capable of initiating an antibody response or boosting pre-existing antibody and may be responsible for confusing serological findings. Casemore & Jackson (1984) first noted that there appeared to be an urban (non-zoonotic) cycle of infection that was subsequently borne out by the identification of C. hominis (C. parvum Genotype 2). Hunter et al. (2004) noted that risk factors for acquiring infection with C. parvum and C. hominis differ in addition to the role of animals. However, serology is unable currently to distinguish the infecting species or sub-type. It has been noted in a number of epidemiologic studies that a history of regular consumption of raw vegetables is negatively associated with cryptosporidiosis, a so-called protective effect. However, it is not known if this is the result of such lowlevel exposure resulting in immunity, other non-specific effects, or confounding (Casemore et al. 1997; Hunter et al. 2004; Roy et al. 2004).

Large format WB is generally highly specific and sensitive; however, the test is expensive, technically demanding, labour-intensive and is less useful in quantitation of responses, and is not suitable for mass screening

 Table 1 | Cryptosporidium seroprevalence levels in US studies 1989–2005

Site	Population	Age range	Water supply	Method(s)	n	Antibody frequency	Comments	Reference
Carrollton GA	(i) Residents	Adult	(i) TSW	Ungar ELISA	(i) 86	(i)Ill/well 76%/56%	Outbreak investigation	Hayes et al. (1989)
	(ii) Unexposed controls		(ii) NS		(ii) 20	(ii) 35%	mvesugation	
Wisconsin	(i) Dairy farmers	≥50 yr	NS	Ungar ELISA	(i) 70	(i) 44.3%	Suggests increased infection rate without illness in farmers.	Lengerich <i>et al.</i> (1993)
	(ii) Non-farmers				(ii) 50	(ii) 24.0%	iarmers.	
Oklahoma	Hospital patients Convenience samples	<5 yr - 21 yr	NS	Ungar ELISA	803	13-58%	Rate increased with age.	Kuhls <i>et al.</i> (1994)
Virginia	Hospital patients Convenience samples	<1-29	NS	Ungar ELISA	172	16.90%	Rate increased with age; <1 yr negative.	Zu et al. (1994)
New York	HIV clinic patients	Adult	NS	Ungar ELISA	137	20.30%	3.6% stool positive	Spenser et al. (1997)
Jackson County Oregon	Blood donors	Adult	TSW	(i) Ungar ELISA	380	(i) 15.5%	6 mo. post-outbreak	Frost et al. (1998a)
	Convenience samples			(ii) MGF WB		(ii)21.8-47.8%		
Two cities NW USA	Blood donors	Adult	(i) TSW	MGF WB	500 per site	(i) 21-31%		Frost et al. (1998)
	Convenience samples		(ii) GW			(ii) 11.23%		
NE USA	College students	Young adult	TSW	MGF WB	(i) 107	15-17/27 kDa	(i) Chlorinated only	Frost <i>et al.</i> (2000 <i>c</i>)
			(i) pre-		(ii) 225	(i) 19%/27%	(ii) Plus filtration confounders – see text	
			(ii) post-Filtration			(ii) 24%/41%		
Milwaukee	Convenience samples	Children 6 mo-12 yr	TSW	Second generation ELISA	(i) 218	15 – 17/27 kDa	Evidence of increased infection rate in adjacent area	McDonald et al. (2001)

Table 1 | (continued)

Site	Population	Age range	Water supply	Method(s)	n	Antibody frequency	Comments	Reference
	(i) outbreak area (ii) adjacent area				(ii) 335	(i) 15-82%/ 17-87% (ii) 10-43%/ 22-46%		
SW USA -	Blood donors	Adult	(i) TSW	MGF WB	200 per site	15 – 17/27 kDa	TSW group showed increased intensity of reactions	Frost et al. (2001)
(i)Las Vegas	Convenience samples		(ii) GW			(i) 49%/55%		
(ii)Albuquerque						(ii) 36%/52%		
Two cities	Blood donors	Adult	(i) TSW	MGF WB	(i) 462	(i) 54%	Further detailed analysis of repeat samples and according to mw of Ag in paper.	Frost <i>et al.</i> (2002 <i>a</i>)
Mid-West USA	Convenience samples		(ii) GW		(ii) 503	(ii) 38%	Some evidence of lower illness rates in TSW area	
Four cities in Iowa and Wisconsin	Blood donors & college students	Adult	(i) TSW	MGF WB	(i) 184	15 – 17/27 kDa	Greater frequency of increased intensity of response in repeat samples in group (i).	Frost <i>et al</i> . (2003 <i>a</i> , <i>c</i>)
	Convenience samples		(ii) GW		(ii) 309	(i) 72.3%/82.6%	Higher prevalence in sub-populations users of private wells	
						(ii) 52.4%/72.5%		
USA – 7 sites	NHANES III	11 yr - > 71 yr	Various – see paper cited	MGF WB	1356	(i) 15-17 kDa	Positive trend with age.	Frost et al. (2004)
	Participants					13.4-67.9%	Geographically and socio-economically variable responses.	
						(ii) 27 kDa 21.3 – 81.5%		

ble 1 (continued)	ed)								
e e	Population	Age range	Water supply	Method(s)	u	Antibody frequency Comments	Comments	Reference	
sites in W USA	3 cohorts divided Various by water type and two phases	Various	Cohorts A,B: TSW	MGF WB	522 from 326 persons	All sera	Enteric illness records analysed in relation to water supply suggested surface water use protected against illness.	Frost <i>et al.</i> (2005a, b)	
			Cohort C: GW			(i) 15–17 kDa			
						52%			
						(ii) 27 kDa 73%			

Site

Abbreviations used: NS – not stated; TSW – treated surface water (see reference cited for details); GW – groundwater; Ungar EUSA – see Ungar et al. (1986); MGF WB – minigel format western blot; for details of 15–17/27 kDa

exercises. Minigel WB is easier to use but suffers from some lack of discrimination between antigens of similar mw and relatively poor sensitivity for some antigens that may in part be due to the small amount of antigen used. Nonetheless, trends in rates can be compared where that method has been used. It has yielded very useful epidemiological information and increased understanding of outbreak dynamics, including the impact on risk evaluation of previous exposure and immunity. ELISA often has problems with specificity, background "noise", and robustness, but is relatively cheap, can be automated, readily permits accurate, objective quantitation by titration, and is easy to use for mass screening provided the problem of specificity is adequately addressed. However, it may lack some sensitivity to the 15-17 kDa Ag and produces some falsepositive reactions (Frost et al. 2003b) and the significance of this needs to be assessed. Its more widespread use depends upon the availability and cost of the defined antigens. In both systems, QA, QC and validation for sensitivity (maximum detection of true positives) and specificity (minimum of false positives) are essential. Maximizing sensitivity may lead to inclusion of false positives (crossreactions); maximizing specificity may result in the loss of weak true positives, potentially overlapping with false positive reactions which usually give weak reactions. Evaluation requires a "gold standard", which is usually LF WB using time series samples from confirmed cases with known dates of onset. True negative samples are often difficult to identity for common infections except by testing by the consensus most sensitive method such as LF WB. Establishing a "negative" cut-off for some tests such as ELISA is problematic except by consensus based on experience with the test but this inevitably runs the risk of excluding low-titre true positives. Simply assuming that convenience samples from presumed asymptomatic individuals are negative is unacceptable practice. Methods to be used for epidemiological studies need to be readily transferable between laboratories while maintaining QA/ QC performance.

A note of caution: the studies above uniformly used *C parvum* (previously *C. parvum* genotype 2) as antigen source, often the Iowa strain. It is still unclear how the results are affected where *C. hominis* (previously *C. parvum* genotype 1) or other species have been the etiologic agent.

In some cases, the two species and others were likely present in the water and/or in infections in humans (Patel et al. 1998; Tanriverdi et al. 2003; Mathieu et al. 2004; Priest et al. 2006). It is unclear how important the anamnestic (booster) response is (including from other species or genera with cross-reacting antigens, although some studies have looked at a limited range of related species) in producing high titre responses that might be interpreted as indicating recent infection, especially where subjects are exposed to concurrent or episodic multiple waterborne pathogens. Indeed, it can reasonably be argued that frequent exposure through intermittent low-level contamination, including with different Cryptosporidium species, will lead to infection that is not expressed clinically and that mild, self-limited or sub-clinical infection is more likely the outcome of exposure. In such situations, outbreaks are less common but sporadic cases in the previously unexposed will be common, including infants and visitors; the immunocompromised will be particularly at risk. In a longitudinal study of Cryptosporidium species-specific IgG in Peruvian children, Priest et al. (2006), using C. parvumderived antigens, detected antibody responses during infections with C. parvum, C. felis, C. meleagridis, and four different sub-types of C. hominis. There was however, a reduced serological sensitivity (73% positivity) in C. hominis infections. The significance of this for seroprevalence studies needs to be further evaluated.

The infectivity of different isolates of C. parvum is known to vary (Okhuysen et al. 1999; Teunis et al. 2002). Many oocysts in water may be of low infectivity through poor viability, or low pathogenicity or virulence for man. Some of these may be capable of initiating limited infection and stimulating or boosting immunity. High levels of herd immunity may suppress secondary (person-to-person) transmission (Fine 1993; Casemore 1994, 1995). Whether humoral (circulating) and/or secretory antibodies in the gut are protective is still unclear (Current 1989; Heyworth 1992; Riggs 2002; Wyatt & McDonald 2004). Some antibodies are known to neutralize surface antigens on zoites that are associated with attachment, which would intuitively suggest a potential protective mechanism. Conversely, HIV-positive patients can have chronic disease in the presence of circulating antibody. In terms of risk estimation, it can be said that while antibody per se may not be protective it acts as a marker for increased resistance, which is likely dependent more on CMI although that mechanism may be enhanced by the presence of antibody on surface epitopes.

A further note of caution concerning the test antigens selected for use in antibody assays is that these have been evaluated primarily against adult sera; the response to them in children may be different and this has not been fully evaluated (R. Morris, personal communication).

GIARDIA—REPORTS ON THE SEROLOGICAL RESPONSE AND SERO-EPIDEMIOLOGY

Giardia was first described by Leeuwenhoek (1632–1773), and re-described and named in the 19th century separately by Giard and Lamble. It was generally considered medically insignificant by clinicians until the 1960 s, mainly because of the frequency with which it could be found in apparently asymptomatic subjects. It is the most common enteric protozoan parasite worldwide. It is widely associated with the water route of transmission in the US, and by the food route, as well as in the day-care setting; some 5000 persons are hospitalized annually in the US with giardiasis (Craun 1990; Shandera 1990; US EPA 1998; Gardner & Hill 2001). Curiously, in the UK Giardia is uncommonly associated with the water route although it has been shown to be a risk factor for sporadic infection in a study in the Southwest of England (Stuart et al. 2003). The importance of the water route of transmission in the US became apparent in the 1960s with the recognition of cases among travellers returning from Leningrad in Russia and outbreaks centered on Aspen, CO (Shandera 1990). In an outbreak in New Hampshire in 1977, associated with a contaminated water supply, it was noted that there was a high rate of asymptomatic infection, suggesting prior exposure and immunity to clinical infection (López et al. 1980). The infection has been commonly regarded as a zoonosis but the natural history and etiology is now known to be more complex (Thompson 2004). Infection is generally noninvasive, remaining localised within the small intestine. Innate mechanisms and humoral and cell-mediated immune responses have been identified over many years in humans and in animal models (Farthing 1989, 1990; Janoff

& Smith 1990; Faubert 2000; Wyatt & McDonald 2004). Primary infection in the immunocompetent will usually be symptomatic although this varies with the species or strain involved. Subsequent reinfection is commonly asymptomatic and may persist in the presence of antibody. Infection in the immunocompromised is often symptomatic and very persistent, particularly in those with hypogammaglobulinemia. The latter suggests that humoral antibody has an important role in clearance and recovery. Conversely, chronic infection is found in the presence of circulating antibody. Infection in AIDS patients does not seem to be especially persistent or severe although this view may be a reflection of the availability of specific treatment (Faubert 2000). The diagnosis of giardiasis is usually by detection of cysts (and sometimes trophozoites) in stools but this is known to be insensitive, often requiring multiple samples from known cases to detect the parasite (Farthing et al. 1987). Parasite antigens can also be detected by other methods such as ELISA. Antibodies can be detected by a variety of means, using serum, saliva or fecal fluid. The diagnostic value of this is uncertain but it may be of epidemiological value for indicating exposure.

Giardia antigens

The clinical presentation, severity and immune response differ with the different species of G. lamblia (syn. G. duodenalis; G. intestinalis) capable of infecting humans, their sub-types (assemblages), and other variants, and the frequency of exposure (Nash 1997; Janoff & Smith 1990; Hunt 1999; Faubert 2000; Lane & Lloyd 2002; Thompson 2004). Cysts and trophozoites are thought to contain many antigens, some of which are restricted to one or other life cycle stage; some maybe dependent on in vivo or in vitro exposure of the parasite to immune and non-immune components (e.g. bile) of the gut milieu (Taylor & Wenman 1987; Reiner & Gillin 1992; Udezulu et al. 1992; Palm et al. 2003; Adam & Nash 2004). Some antigens are immunodominant, while others appear not to be directly involved in clearing of or resistance to the parasite. Some antigens detected in tests are not specific to Giardia, i.e. are cross-reacting with antigens from other organisms; false positive reactions appear to be common in unexposed subjects (Moore et al. 1982; Jokipii et al. 1988; US EPA 1998;

Faubert 2000). In addition, *Giardia* undergoes surface antigenic variation that may aid the parasite's persistence (Nash 1997). Analyses of antigens from different isolates has revealed a number of important molecules ranging in size from 14 to 225 kDa in crude extracts of trophozoites and 21 to 49 kDa in cysts. An antigen of mw 31 kDa seems to be common to many isolates and is probably the structural antigen referred to as giardin (Taylor & Wenman 1987). Other antigenic molecules identified include heat shock proteins, lectins, tubulins, etc. (Farthing 1992; Janoff & Smith 1990; Faubert 2000). Antigens of 30 to 34 kDa, 57 and 82 to 88 kDa have been reported to be consistently recognized (Farthing 1989; Char *et al.* 1991; Janoff & Smith 1990), some of which are restricted to either trophozoites or cysts.

Serological study reports

As with Cryptosporidium studies, various methodologies have been used to detect and measure the immune response to Giardia, particularly IFAT and ELISA but also complement fixation, lectin-based tests, immunodiffusion and indirect haemagglutination, etc; WB has been much less frequently used. Some have advocated detection of secretory IgA in saliva by ELISA for seroprevalence studies (Al-Tukhi et al. 1993; Hashkes et al. 1994). Many of the early antibody studies were aimed at characterizing the immune response and/or the pathogenesis of symptoms, and investigate resistance and susceptibility to re-infection (Ridley & Ridley 1976; Farthing 1989, 1990; Janoff & Smith 1990; Faubert 2000; Gardner & Hill 2001). The interpretation of serological findings, the importance of the different responses in producing resistance to infection, and the significance of the various antigens involved continue to be a matter for debate (Farthing 1992; Heyworth 1992; Nash 1993). Some workers have shown that specific antibody levels may indicate more severe infections associated with enteropathy and/or malabsorption (Ridley & Ridley 1976; Farthing 1989, 2003; Faubert 2000; Frost & Craun 1998b), although others have not corroborated this finding (Faubert 2000). Such variable findings may reflect infections with different species and strains. Chronic, asymptomatic or mildly symptomatic infections may persist for months or even years in the presence of circulating antibodies (Faubert

2000). The mechanism of protection afforded by maternal antibody, other non-specific factors in milk, and relative protection from the environment for breast-fed infants is a matter of debate (Miotti et al. 1986; Navak et al. 1987; Sterling et al. 1991; Zu et al. 1992; Walterspeil et al. 1994; Tellez et al. 2003). Some have shown significant benefit while others have suggested that other non-specific mechanisms might be equally important. Mucosal (local gutderived) specific sIgA and/or mucosal secretory IgM may be involved, with the complement pathway and opsonization, probably in concert with CMI mechanisms, in resolution or protective immunity (Heyworth 1990; Palm et al. 2003). Hypogammaglobulinemic patients with IgG and IgA deficiency tend to get persistent infection, implying that antibody may be important in clearance (Palm et al. 2003). However, diagnostically, serum IgA is a less useful indictor of active infection in acute cases as many infected subjects fail to show a humoral IgA response; serum IgM is probably more useful as an indicator of recent infection (Farthing 1989; Janoff & Smith 1990; Nash 1993; Faubert 2000).

Early worldwide studies

Early reports on Giardia serology tend to suffer from the fact that they are based on tests using crude cyst or trophozoite antigens and varied test protocols, often with little or no evaluation of performance. Tests often did not distinguish the Ig classes. Nacapunchai and others found that several of these methods had poor sensitivity and/or specificity (Nacapunchai et al. 1986). However, studies on populations in developing countries in the 1970s and 1980s did indicate that antibodies were highly prevalent (Ridely & Ridely 1976; US EPA 1998; Farthing 1989; Janoff & Smith 1990; Faubert 2000). Exposure to Giardia begins early in life, with rapidly increasing rates in infants and young children; high rates of carriage were found in all age groups in apparently asymptomatic subjects (Gilman et al. 1985, 1988; Miotti et al. 1986; Nacapunchai et al. 1986). Newly acquired infection in infants in these populations is frequently symptomatic, compared with asymptomatic carriage in older family members (Janoff & Smith 1990). As noted with Cryptosporidium, visitors from developed countries often act as sentinels for this high prevalence with high rates of symptomatic infection. The attack rate falls with age in these groups, suggesting a role for the development of immunity (US EPA 1998; Farthing 1989).

Goka et al. (1986) described the sero-diagnosis of 52 patients and control subjects in India and the UK. Testing was by detection of specific IgM and IgG using an ELISA test with frozen trophozoites as antigen; sensitivity and specificity were estimated at 96%. IgM response was consistent with current infection, the response tailing off in as little as 2-3weeks in three patients who provided sequential samples. Tests for IgG were unable to distinguish cases from controls with a history of previous infection. However, Gandhi and others found that the ELISA in their hands was specific but very insensitive for IgM, antibody being detected in only 24/ 73 confirmed cases, one of 37 control patients with other parasitic infections and none of 29 healthy controls (Gandhi et al. 1989). The results suggested that IgM detection might not be suitable for the more prolonged infection seen in their patients, compared with the shorter duration infections in the patients studied by Goka. This finding is consistent with the demonstration by Vinayak & Kumkum (1989), using AHG in an indirect ELISA with a trophozoite plasma membrane and 56 kDa Ags. This showed that lower antibody titres were found in persistent than non-persistent acute infection. Char et al. (1993) also showed a lower IgA response, in an ELISA with a 57 kDa Ag Giardia heat shock antigen, in persistently infected children in The Gambia. This suggests that the more severe symptoms may be immunogenic (Farthing 2003).

Miotti and his co-workers investigated age-related acquisition of IgG sero-positivity in four diverse populations in the USA and South America, using an ELISA test system with trophozoites as antigen (Miotti et al. 1986). The specificity of the test was confirmed by blocking with monoclonal antibody to the giardia antigen. No attempt was made to study stool positivity rates. Rates in South American adult subjects and Apache Indians in Arizona showed rates of 44-48%, while adults living in Baltimore, Maryland, showed only 18% positivity (P < 0.01) and with lower levels of reactivity (P < 0.001). Children living in different countries and settings showed widely varying rates of acquisition, the fastest rate being in Peru. The results are consistent with levels of hygiene and environmental conditions in the respective communities. This was also shown in a study by Gilman and others in Lima, using stool examination, which showed that children rapidly

became re-infected following anti-giardial treatment (Gilman et al. 1988).

Taylor & Wenman (1987) in Canada sought to characterize the immunologic response to *Giardia* in 16 confirmed cases, age range 3 to 76 years, who had acquired their infections in Canada or while travelling abroad; three of the patients had chronic infection associated with common variable hypogammaglobulinemia and no detectable IgA. They also used sera from 10 healthy control subjects. Antigens were obtained from cultivated trophozoites by SDS-PAGE and then used in WB. A majority of the sera reacted with a range of antigens, but particularly with a 31 kDa polypeptide Ag, which was thought to be surface disc-derived protein referred to as "giardin", also described by others.

Char et al. (1991) in India and the UK studied the serum antibody response in children to an immunodominant 57 kDa Ag purified by SDS-PAGE in WB. All sera from confirmed cases showed specific IgG but not IgM although IgM reactivity was found with other mw antigens but these were also often recognized by control sera. Sera from nine of ten patients had detectable IgA to the 57 kDa Ag.

Ljungström & Castor (1992) studied the immune response to Giardia in a waterborne outbreak in Sweden involving 1400 cases in a previously largely unexposed population estimated at ~3000 persons. The outbreak resulted from sewage contamination of the drinking water supply over a defined period of about one week. Serum samples were obtained from 352 exposed persons and tested by IFAT for IgG and IgA. Results were correlated with fecal microscopy; "negative" controls consisted of serum samples from 428 asymptomatic persons. IgG and/or IgA were found in 68% of microscopically confirmed cases and 22% of exposed but microscopy-negative persons, compared with 10% of controls. Antibody titres were generally low and in 32% of microscopy-positive cases antibody could not be detected. A further molecular study by this group looked at the response to non-variable antigens (Palm *et al.* 2003). Background titres in control subjects were low. They suggest that antibody aids clearance and enhances resistance to further clinical infection. Hypogammaglobulinemic patients with decreased levels of IgG and IgA are prone to develop persistent infection (Janoff & Smith 1990).

Soliman et al. (1998) studied serum antibody in symptomatic and asymptomatic Egyptian children using

axenic trophozoites (WB strain) as antigen source with IFAT, ELISA and WB. IFAT showed significant titre differences between the two groups. By ELISA, IgA and IgM levels, but not IgG, were higher in the symptomatic subjects. Total *Giardia*--specific IgG was the same in both groups but isotypes IgG1 and IgG3 levels were higher in the symptomatic group. The significance of different antibody isotypes, which this study measured, is discussed. Results by WB failed to show a clear difference between the groups.

Serological studies in North America

Although giardiasis is the most commonly reported waterborne infection in North America (US EPA 1998; Craun et al. 2002) there are few seroprevalence studies from there. Smith et al. (1981) used an ELISA test to examine sera for antibodies in subjects in Washington, DC, and found 14% to be positive. Lopez et al. (1980) described an outbreak in a community in which those who had a history of exposure to surface water had a significantly higher rate of asymptomatic infection, suggesting acquired immunity to symptomatic disease but not reinfection. Istre et al. (1984) investigated an outbreak of waterborne giardiasis at a mountain resort that also showed evidence of protective immunity in local residents in that local residents were less frequently symptomatic than were visitors. Smith et al. (1981) used intact trophozoites in an ELISA test to examine sera from patients and controls from Colorado and elsewhere in the USA. The study was aimed partly at developing a reliable and reproducible test protocol. They found that 81% of 59 symptomatic patients and 12% of controls had detectable IgG antibody; 11 of 15 patients tested serially had detectable antibody from 2 weeks to 15 months after treatment. Prevalence of antibody in 197 convenience samples from Washington, DC, was 14%. A further study used ELISA to measure IgG, IgM and IgA in sera from 29 AIDS patients with acute giardiasis and other groups, including immunological normal patients with and without giardiasis (Janoff et al. 1988). Responses were further characterised by WB (method of Janoff et al. 1988). This study showed significantly depressed antibody responses (P < 0.0001) in the AIDS patients. All 25 immunocompetent subjects with giardiasis had good IgA and IgM responses. It was not established whether this was related

to IgAs levels (IgA2). Their IgG response was less marked than in AIDS patients with more persistent Giardia infection whose symptoms were not related to the parasite, although the latter group had a poor IgM response. WB showed that, although there were many reactive bands, all positive sera reacted to antigens between 30 and 34 kDa also identified by other workers as important immunodominant antigenic molecules. Of particular interest to this report, Birkhead et al. (1989) studied serum IgG, IgA and IgM levels in an ELISA with trophozoite-derived soluble protein antigen(s). Subjects included 24 convalescent persons and 20 non-residents following a waterborne outbreak in a rural trailer park in Vermont. Residents showed higher levels of IgG and IgA but not IgM. Nine residents showed higher mean levels of IgA, which increased relative to their consumption of tap water. This study indicated that anti-giardial IgA might be useful in the investigation of outbreaks.

Miotti *et al.* (1986) used ELISA to look at age-related antibody acquisition in subjects in an inner city area of Baltimore, Maryland, and three other locations described above. They found 18% of Baltimore subjects had detectable IgG antibodies to *Giardia*, significantly less frequent than in subjects from other groups studied from areas with poorer hygiene and environmental conditions. This is broadly comparable with the rate in the previously described study. Stool isolation rate for the same area was 0.9% over a five-year period, underlining the poor sensitivity of microscopy for diagnosis and prevalence studies.

Sullivan *et al.* (1987) conducted serological studies on patients with giardiasis and others in a Midwestern city in the USA. Using IFAT, they showed that symptomatic patients tended to have higher titres and titres remained high for a prolonged period in those chronically infected, suggesting that antibody was not associated with clearance of the infection. Indochinese refugees, HIV-positive patients and other symptomatic individuals generally had higher mean titres than healthy controls but there was a broad overlap.

The anti-Giardia-specific antibody response was evaluated as a diagnostic tool in children with suspected infection (Sullivan *et al.* 1991) and compared with stool microscopy and jejunal mucosal biopsy. High titre (≥1/800) specific IgM had a sensitivity of 63%, specificity

of 93%, and predictive value of 85% and correlated better with active infection than IgG or IgA. Sensitivity may have been improved by using a lower cut-off but this might be offset by decreased specificity.

Several sero-epidemiologic studies have been reported from Canada. Isaac-Renton et al. (1994) investigated the use of serology in an outbreak in a locality in British Columbia that had had another outbreak five years before. They used an ELISA with a soluble antigen derived from axenized, cultured trophozoites from the strain recovered from water samples and an ATTC reference strain (WB). Sera were collected from 51 cases, 21 non-cases in the affected area, and from 35 controls from two different groups. Full demographic details of study subjects are described. Sera were analyzed for Giardia-specific IgG, IgA and IgM. Forty-three (84%) of the cases gave significant positive antibody reactions in one or more Ab class. In contrast to the study by Birkhead et al. (1989), levels of IgG were more elevated than IgA; IgM levels were elevated but less frequently, as reported by other workers. Use of the heterologous isolate did not affect the results. The study, including isolation rates, confirmed that those resident at the time of the previous outbreak were significantly less likely (P < 0.001) to suffer symptoms on reexposure. The authors point out that serologic studies can be useful in non-endemic communities, giving a better estimate of case numbers than stool examination, especially where there is delay in investigation but are likely to be of limited value in endemic areas. Isaac-Renton et al. (1996) conducted a 24-month longitudinal study of two discrete water catchment communities in British Columbia with Giardia cysts in their water supplies. There had been a waterborne outbreak in one of the two communities some five years before this study. Over the two-year period, there were no outbreaks but evidence suggested an endemic background level of infection but with amelioration in the area that had previously experienced an outbreak. Serologic studies, using the previously described ELISA, on 1122 samples showed 37% had IgG levels above the cut-off value; some had raised IgM levels. Differences were noted in serologic rates reflecting water treatment differences and the previous history of an outbreak in one area. Sera from the latter group showed that 64% had no detectable antibody and this community was thought to be at risk should further significant contamination occur. Isaac-Renton et al. (1999) studied three communities

with different water source types [(i) deep wells, (ii) protected watershed surface water, (iii) a surface water known to be contaminated] by estimating sero-prevalence rates against *Cryptosporidium* (see above) and *Giardia*. Using the previously described ELISA, seroprevalence for *Giardia* antibodies was 30.3% overall (590/1944). Curiously, group (i) had a rate of 19.1%, group (ii) 34.7%, and group (iii) 16.0%; the rate for (ii) dropped significantly over the study period but not in the other two areas (P < 0.001) although the reason for this difference was not known, it suggests perhaps that group (ii) had experienced an unrecognized outbreak prior to the study period.

Discussion of Giardia serology

Given the high prevalence of this parasite, surprisingly little has been reported on prevalence of *Giardia*-specific antibodies in the USA. However, the studies described above suggest that useful information:

- can be gained from studies on the natural history and dynamics of transmission of giardiasis;
- can help to identify areas of high prevalence that may not be apparent from clinical diagnostic rates and which may be attributable to water;
- can identify communities at risk from occasional water contamination events.

With reference to risk estimation, the evidence generally supports the view that while antibody may not protect against infection *per se*, reinfection in an antibody-positive person is less likely to be symptomatic (US EPA 1998). The potential for risk from zoonotic sources has perhaps been overestimated as research indicates that many animal types are not infective for humans other than those belonging to assemblages A or B (Lane & Lloyd 2002; Thompson 2004).

SEROLOGIC ASPECTS OF OTHER WATERBORNE PROTOZOAN PARASITES

Entamoeba histolytica

Although waterborne outbreaks have occurred in the USA, these are very uncommon (Marshall *et al.* 1997; Tarleton &

Petri 2004). Many older studies have been invalidated by the separation of the so-called non-pathogenic variant as a species, *Ent. dispar* (Clark 2004). Little is known of the prevalence of antibodies to these and other classic species in the USA and such data would be of little relevance to the risk estimate.

Emerging parasites

Several parasite species have emerged in recent years as waterborne protozoan pathogens, although all have other routes of transmission. These include:

- Toxoplasma. Toxoplasmosis is occasionally waterborne in North America but this is uncommon (Benenson et al. 1982; Bowie et al. 1997; Dubey 2004). Infection is usually transmitted by means of oocysts from infected cat feces, directly or via the soil or water, and from tissue stages via consumption of raw meat. Seroprevalence in the USA has been estimated at ≥20% in NHANES and other studies and appears to be stable at that level (Bowie et al. 1997; Jones et al. 2001 2003). Methodology and interpretation are well defined in the USA (Garcia et al. 2004). In Brazil, widespread infection, estimated through seroepidemiology, is associated with consumption of contaminated water and commonly results in ocular toxoplasmosis (Bahia-Oliveira et al. 2003).
- Cyclospora This parasite has emerged as an enteric pathogen in recent years and is thought to be commonly waterborne in some developing countries. The requirement for maturation of oocysts in the environment means that direct person-to-person spread is unlikely. Evidence was found in a study in Peru to suggest that primary infection protects against further clinical episodes (Bern et al. 2002). Age-specific rates supported an environmental route for the infection. In the USA, it appears to be primarily foodborne and is uncommon in children although waterborne transmission has been reported in the USA occasionally (Herwaldt 2000; Sterling & Ortega 2004). The failure to recover and purify oocysts in large numbers from human feces or to develop an animal model means that serological studies have not yet been developed.
- Microsporidia This large and ubiquitous group of parasites has emerged in recent years as a cause of

infection, mainly in the immunocompromised. Concerns have been expressed about its potential role as a waterborne pathogen but little is known about this yet (Dowd *et al.* 1998; Franzen & Müller 1999). Serological tests have been developed but little is yet known about the prevalence of antibodies in the USA (Hollister *et al.* 1991; Van Gool *et al.* 2004).

While antibody studies to these parasites may be of general epidemiologic interest, it would be of doubtful value to the national risk estimate.

SEROLOGIC ASPECTS OF WATERBORNE VIRUSES

Introduction

It has long been recognized that a variety of viruses may be transmitted by the water route, including noro-, entero-, adeno- and rotaviruses, and some hepatitis viruses. Waterborne outbreaks of what later became known as Norwalk virus have been recognised since the 1920s (Spenser's disease), long before the etiologic agent was identified. In the USA, work in the 1970s and 1980s by Kapikian, Greenberg, Kaplan and others led to the identification of the Norwalk virus (now norovirus) as one of the most important waterborne infections. Various other members of a group of related viruses, known as Norwalk-like or small round structured viruses (SRSVs), were also identified. The group is now known to be a complex of genotypic and antigenic subtypes classified as members of the Caliciviridae (Fankhauser et al. 1998; Atmar & Estes 2001; Von Bonsdorff & Maunula 2003; Gallimore et al. 2003). Other viruses sharing similar epidemiology include the sapoviruses and astroviruses. These may account for as little as 3% of outbreaks in which the etiologic agent and route of transmission is identified (Glass et al. 2000). However, many more outbreaks are reported in which an etiologic agent is not identified, but with SRSV-like symptoms and which are associated with water (Frost et al. 2002b).

The perceived epidemiology of rotavirus has been skewed by the concentration on infection in infants and young children but it is more widespread in adults than is usually recognized, in developed as well as developing countries (Casemore 1987, unpublished data; Cox et al.

1998*a*, *b*; Cox & Medley 2003; Iturriza-Gomara *et al.* 2004). Transmission is usually assumed person-to-person and other potential routes have been little explored.

All of these viruses are more commonly transmitted by the direct person-to-person route and via food, environmental/recreational water exposure, etc. (Appleton 2000; Lopman et al. 2003; Widdowson et al. 2005a). The multiple routes make it difficult to estimate the proportion due to water as the only positive indication of source or route is in well-defined outbreaks that are investigated epidemiologically using case-control studies. As with the parasites described above, endemic areas may mask high rates of environmental transmission and this reduces the apparent contribution of waterborne transmission or level of association (relative risk) in epidemiological studies. Visitors from non-endemic areas act as sentinels, thus suggesting the importance of immunity in resistance to symptomatic infection. However, primary infection leads to short-term immunity lasting only weeks or months and little or no immunity to re-infection with homologous as well as heterologous types although there is considerable crossreactivity in testing (Appleton 2000; Matsui & Greenberg 2000; Schaub & Oshiro 2000).

Serological studies with the *Caliciviridae* (norovirus and associated SRSVs)

As with the parasite studies described above, many studies are developmental and this makes comparison and interpretation difficult. More recently, the introduction of molecular methods in particular has enhanced the ability to distinguish morphologically similar species and sub-types, while the development of recombinant capsid-protein antigen (baculovirus-expressed norovirus-like particles) has made serology more precise (Atmar & Estes 2001). As with parasites, detection of fecal (secretory) antibodies may be useful diagnostically although not much used for seroepidemiology (Okhuysen et al. 1995). However, their use has also shown the antigenic as well as genetic diversity, which makes interpretation of findings problematic unless studies are done using homologous virus antigens (Hale et al. 1998; Pelosi et al. 1999; Lopman et al. 2002). For the purposes of this report, no attempt is made to distinguish

the members of this group, which share similar epidemiologic features, except for Hepatitis E virus (HEV) which is dealt with separately (see below). Volunteer and other studies have shown a mixed response in SRSV-specific antibody-positive subjects, some being resistant to reinfection while others have had symptoms on reinfection, thus indicating a variable degree of protective immunity and that several exposures might be required for protection (Matsui & Greenberg 2000; Schaub & Oshiro 2000; Lopman *et al.* 2002). Gray and others in the UK estimated seroprevalence at nearly three quarters in >3000 subjects (Gray *et al.* 1993); age-related rates were 75% in infants <6 months, followed by a dip to 25% (suggesting the effect of maternal antibody) and then progressively increasing, reaching 74% in the teens, rising to 80–94% in adults.

Gray et al. (1994) conducted volunteer studies with norovirus. They found little difference in initial IgG titres between those who were symptomatic following challenge, and those who were not. An IgM response was subsequently found in all infected volunteers. In those who were symptomatic, there was a marked increase in IgA and IgG titres but this was variable in those without symptoms. This study suggests that antibody is often not protective but the immunological difference between those who did and those who did not develop symptoms is not clear.

In Europe, seroprevalence has been found to be high in all age groups. For example, a study of > 1000 sera in France using NVL (recombinant baculovirus-derived) antigen found high prevalence in all age groups, similar to those in the UK; >60% in infants <6 months, followed by a dip and then progressively increasing, reaching a peak of >80% in the teens; overall rate 74.1% (Nicollier-Jamot *et al.* 2003). Lopman and others reported that the test system tended to be restrictive in indicating infection with heterologous strains. A similar study in Italy found a seroprevalence of 91% (Pelosi *et al.* 1999). Others have reported, however, that antibodies have been found in human subjects to bovine strains, the significance of which is not yet clear (Widdowson *et al.* 2005*b*).

Seroepidemiologic studies have been reported from developing countries. For example, O'Ryan *et al.* (1998) studied risk factors for acquisition of antibody to Norwalk and Mexico viruses in Chilean individuals in two geographic locations. The main determinants were lower socioeconomic

status and increasing age; other factors varied with location and setting. Smit *et al.* (1999) in South Africa conducted a seroepidemiologic study of genogroup I and II infections; seropositivity ranged from 81-99%, indicating the high prevalence of these infections. Steinberg *et al.* (2004) and others in Guatemala found a similar age-related pattern but, as might be expected, with a more rapid progression from 27% at 6-12 months to 94% at 25-30 months.

In North America, Payment *et al.* (1994) studied seroprevalence and the possible impact of point-of-use filtration of subjects' drinking water. They used a native (stool-derived whole virus) antigen in a blocking (indirect) ELISA test system. Seropositivity increased with age from 55% in those aged 9-19 years, increasing steadily to 100% at 60+ years. Changes in serum titre (seroincidence) were seen in 8.7-24.11% in different study seasons, but these changes did not appear to be related to waterborne infection.

Noel et al. (1997) reported on the correlation of patients' immune response to four different genetically characterised SRSV antigens in 23 outbreaks in the US, 1990–95, to estimate the distribution of these viruses. Recombinant-expressed capsid proteins were used in a direct ELISA to determine whether the responses were strain-specific. The results suggested that significant differences were present in responses to heterologous and homologous antigens, resulting from the distinct antigenic diversity of different SRSVs, and suggested that additional expressed antigens were needed.

Other viruses

A variety of other viruses may be transmitted by the water route, including astroviruses, rotaviruses, Polioviruses, Coxsackieviruses, Echoviruses, and hepatitis A (HAV), and E (HEV), and they may be found in surface and groundwaters along with animal-derived species or types. HAV is transmitted fecal-orally by the water route as well as directly from person to person. HAV is of relatively low endemicity in the US with the exception of certain socially deprived groups (Poovorawan *et al.* 2002). Improved knowledge of hepatitis viruses has enabled the recognition of further types for which water is an important route of transmission, especially HEV. HEV is often asymptomatic but can be fatal in pregnant women (Poovorawan *et al.*

2002). Such cases are uncommon in the US but antibody has been found in up to 28% of healthy subjects in the US in some surveys (Meng et al. 2002). Highest rates have been found in veterinarians; a closely related virus is known to be present in pigs in the US, and that may have been transmitted to humans (Meng et al. 1997; Schlauder et al. 1998). HEV may be more common in developed countries than is generally appreciated (Aggarwal & Krawczynski 2000; Clemente-Casares et al. 2003). Little is known of its seroprevalence in the USA. Enteroviruses are generally ubiquitous, even in areas with high levels of hygiene and water quality, although sub-types vary in frequency of reporting geographically and temporally. Antibody studies generally reflect this ubiquity (Payment 1991; Cox & Medley 2003). Data from a study by Jiang and others supports a protective role for antibody to rotavirus, or as a correlate of protective immunity (Jiang et al. 2002). Some rotaviruses may be zoonotic (Iturriza-Gómara et al. 2004). Groundwater as well as surface water supplies have been associated with viral infection in the USA (Craun et al. 2002).

Discussion of virus seroprevalence

Fecal contamination of water sources, both surface and groundwater, may lead to transmission of viruses as well as other pathogens and through 1971-1998 there were 51 outbreaks in the US involving recognized viruses and 347 of undetermined etiology, many of which will have been viral (Craun et al. 2002; Frost et al. 2002b). In Europe, Norovirus was responsible for >85% of all non-bacterial gastroenteritis outbreaks in 1995–2000 (Lopman et al. 2003). Serology has provided considerable help in better understanding epidemiologic aspects of these infections. Progress in molecular-based studies has enhanced the validity of serologic findings but also highlight the problem for serology, and interpretation of results, because of the multiplicity of enteroviruses, SRSVs and small round nonstructured viruses (SRVs), including animal types that are of doubtful significance in humans. Serologic studies do indicate, however, the very high prevalence of these virus infections, both symptomatic and asymptomatic, in industrialized as well as developing countries. Common antigens may result in cross-reactivity and anamnestic responses, which complicates interpretation of findings. The protective

effect of antibody is generally short-lived and antibody requires boosting by re-infection to be protective. It has to be noted that water, although undoubtedly implicated in some sporadic cases and outbreaks, is probably a minor route of transmission compared with person-to-person spread, directly or via food, in industrialized communities. It is difficult to see how serologic data can be reliably applied to estimating risk from this route.

SERO-EPIDEMIOLOGIC STUDIES OF BACTERIAL INFECTIONS

A variety of bacterial pathogens may be transmitted by the water route. Species that have been reported in the USA with some frequency include campylobacter species (mainly Campylobacter jejuni), salmonella, pathogenic Escherichia coli (e.g. E. coli 0157), Helicobacter (Hunter 1997b; Craun et al. 2002; Hrudey & Hrudey 2004). Species reported as occasional or localized waterborne pathogens (e.g. Plesiomonas, Aeromonas), water-associated species (e.g. Leptospira, Legionella), and the classical waterborne pathogens that cause typhoid and cholera, occur much less frequently in industrial countries including the USA and thus will not be addressed in this report. Most bacterial pathogens that are addressed here are ubiquitous and more commonly associated with foodborne transmission. Standard treatment, particularly chlorination, of potable supplies mitigates the risk from bacteria, given their relative susceptibility to disinfection. Infection with these species tends to be more commonly associated with untreated waters, including consumption of well water and exposure to environmental or recreational water. Given the relative ease of isolation of the etiologic agents in clinical specimens, sero-epidemiology is not commonly used. However, epidemiological studies have shown that regular consumption of water from at-risk supplies (e.g. private wells) is associated with increased resistance to illness that masks the risk to the immunologically naive (Raina et al. 1999; Strauss et al. 2001).

Selected study reports

The source and transmission routes for most *Campylobacter* infections are not clearly known; infection is widely blamed

on poultry although definitive evidence that poultry is the primary source of infection is generally lacking (Adak et al. 1995; Rodrigues et al. 2001; Michaud et al. 2004; Meldrum et al. 2005; Nichols 2005). It has sometimes been associated with the water route, mainly recreational use and consumption of untreated supplies but occasionally resulting from contamination of and/or operational problems with treated municipal supplies (Anon 2000; Said et al. 2003; Michaud et al. 2004; M. Evans, personal communication). General population sero-surveys for antibodies to Campylobacter are problematic for several reasons, including the ubiquity of the infection, complex antigenic structure (including cross-reacting antigens), the difficulty of typing outbreak strains and identifying key genotypic markers, etc. The development of SDS-PAGE purified flagellar antigen has been used in a WB test that seems to offer some benefits but the use of this antigen in an ELISA test leads to some loss of sensitivity (J. Cheesbrough, personal communication). Rural inhabitants regularly exposed to livestock may have increased antibody titres to pathogens such as E. coli 0157 and Campylobacter and reduced likelihood of symptomatic infection, presumably through recurrent boosting of immunity (Adak et al. 1995; Evans et al. 2000; Belongia et al. 2003). Salmonella has been the cause of waterborne outbreaks in the USA but is much more commonly attributed to food. Waterborne E. coli 0157 infection is much less common than either of the preceding two species. In the USA, a survey revealed that 9% of outbreaks from 1982-2002 were waterborne but only 10 (3%; 15% of reported cases) were associated with drinking water. Of these, only three involved municipal supplies, two of which did not use chlorination and the other system had a malfunction (Rangel et al. 2005). Sero-diagnostic tests for E. coli 0157 have been developed but not widely used for seroprevalence studies, other than in the limited case of studying close contacts of microbiologically confirmed cases. Asymptomatic infected subjects do not appear to produce a serologic response to the 0157 lipopolysaccharide (LPS) antigen used (H. Chart, personal communication). Helicobacter pylori infection of the gastric mucosa is very common and may result in chronic gastric inflammation that may lead to ulceration and other disorders. It has been postulated that this infection may be acquired from water, as well as other sources but its

ecology is poorly understood (Hopkins *et al.* 1993). The organism has been detected in water in the US but serological studies may be problematic (Khanna *et al.* 1998; Hegarty *et al.* 1999).

Discussion of bacterial serology

While bacterial infections are associated with transmission by the water route from time to time, this is only a small fraction of infections transmitted from infected food animals or via contaminated food, for example by infected food handlers. The majority of incidents of waterborne transmission tend to be associated with untreated supplies. Regular users of untreated supplies may have reduced risk of IID, due to the effects of acquired immunity. However, such supplies that may appear safe to those consumers represent a particular risk to the previously unexposed such as infants and visitors (Strauss *et al.* 2001).

It is difficult to envisage sero-epidemiologic data providing meaningful information for the risk estimation although undoubtedly it sometimes provides valuable insights in conventional epidemiological outbreak investigation.

DISCUSSION OF THE ROLE OF SEROPREVALENCE IN RISK ESTIMATION

Immunology is primarily the study of observable phenomena associated with reactions between antibodies and/or cellular immune mechanisms and their homologous antigens (not whole organisms per se). These reactions are evidence for exposure to the source of that antigen but not necessarily evidence of disease and recovery; nor do they necessarily denote resistance to re-infection. The presence of high or moderately high levels of IgM and/or IgA especially in the absence of IgG, or weak IgG avidity, is usually interpreted as evidence of recent infection; the presence of IgG alone, and/or of high avidity, is generally taken to denote past infection. Similarly, molecular studies indicate that reactions with particular specific mw antigens may provide similar temporal associations. While the presence of antibody cannot generally be taken to imply that the individual may be resistant to reinfection, or be asymptomatic on re-infection, it may denote

the likely presence of other functional protective mechanisms (e.g. CMI). A factor perhaps not taken sufficiently into account in many studies is the significance of the intensity of an antibody response, whether expressed as titre, size and/or intensity of a WB band, OD value, etc., and the impact that this may have on the level of immunity (resistance). It is generally understood that multiple exposure to an antigen given in a course of vaccine shots is required to produce adequate (high titre) immunity and prolong the effect (prolonged half-life) associated with ongoing protection.

Various studies cited above showed seroprevalence against a number of the agents tested tends to be high in young infants (<6 months) reflecting maternal antibody levels; levels then dip and subsequently increase with age, reaching a plateau at a variable age. The rate at which this occurs generally reflects hygiene levels in the communities studied. This is consistent with the reciprocal age-related decline in frequency of symptomatic cases resulting from recurrent exposure and is an argument in favour of assuming that antibody is linked to or correlated with immunity.

An additional consideration in the risk equation is that of herd immunity, in which high rates of individual immunity act as an inhibitor of secondary (person-to-person) spread. Such secondary spread may amplify the impact of waterborne transmission in low- or non-endemic settings and will often result in a fall in mean age of cases as the outbreak progresses (Casemore 1995; Meinhardt et al. 1996; Harrison et al. 2002). More importantly perhaps, the absence of detectable antibody can generally be interpreted as indicating likely susceptibility. This is important in populations generally protected from waterborne infection by the provision of good quality, safe drinking water. Penetration of pathogens through exceptional challenge and/or breakdown in treatment will likely result in a much increased attack rate in such communities. This is increasingly likely with the climatic instability and increase in exceptional weather associated with global change (Rose et al. 2001). Such communities can be identified by the evidence of seroprevalence studies and improved protective measures then initiated where indicated. Where there are high levels indicating frequent exposure, the greatest concern is for immunocompromised persons, especially those in whom that condition has not been diagnosed. Again, seroprevalence studies can help identify these at-risk communities.

Public water suppliers are required to provide potable water that does not represent a risk to public health. However, investigations of outbreaks of waterborne disease affecting such supplies have usually revealed treatment deficiencies or operational failings. Poor quality source water may result in frequent, usually low level, penetration that may enhance seroprevalence and result in lower attack rates except in young children, visitors and the immunocompromised. Improved water treatment in areas with a challenged supply may lead to a fall in seroprevalence. When source waters are of high quality and in the absence of other exposures, seroprevalence is likely to be low or very low. Breakdown in treatment is then likely to result in a high attack rate in the exposed population. Small local and private supplies, including those in many rural areas, many of them untreated, are often inherently unable to meet public health quality requirements. Seroprevalence studies may indicate the frequency of transmission resulting in sub-clinical infection in the immune and the potential risk to others.

SUMMARY AND CONCLUSIONS

This review of published serologic and seroprevalence studies on a wide range of waterborne pathogens concentrates on information relevant to estimating risk of waterborne infection. A number of conclusions can be drawn concerning the organisms and the antibody detection methods that are of importance.

In assessing the importance of different potentially waterborne pathogens in relation to seroprevalence, it needs to be recognized that:

- Most species have multiple sources and routes of transmission. Nonetheless, antibody from infection acquired by other routes (e.g. person-to-person, food) will also impact susceptibility when transmission occurs via water.
- Many have multiple types or sub-types that make it difficult to be sure from serologic findings of the precise organism or strain of organism involved.
- Some organisms (e.g. vegetative bacteria) are readily controlled in drinking water by simple disinfection. Priority has therefore been given in this report to *Cryptosporidium* and *Giardia*, both of which are frequently waterborne and

relatively resistant to disinfection. The most extensive literature available relates to *Cryptosporidium*. The relevant features of papers on seroprevalence for *Cryptosporidium* in North America are shown in Table 1. Findings are highly variable in place and time, between different population groups, and by different methods.

- The immune response is complex and interpretation of laboratory findings requires caution and considerable expertise.
- A serological response indicates that the subject has been infected with the organism concerned or an antigenically related species. It does not necessarily imply that the subject has suffered acute clinical (symptomatic) infection.
- Primary infection usually results in clinical (symptomatic) infection and a typical pattern of antibody responses that can be used to estimate the likely period of infection. Such kinetic studies may be based on the different antibody classes, seroconversion in time series samples, and/or the precise antigen for which there is a measurable response. In *Cryptosporidium* infection, there has been debate about the pattern of the kinetic response, but this may reflect the relative sensitivity of the methodologies used.
- Some infections, for example those occurring noninvasively in the gut (e.g. *Giardia*), may produce little or no measurable antibody response. Conversely, invasive infection is usually associated with a measurable antibody response.
- Continuing chronic infection may result in persistence of IgM, which is often associated with recent primary infection.
- After primary infection, antibody declines and usually returns to baseline generally in less than a year.
 However, subsequent reinfection with the same or antigenically related species may boost the level and may result in a different pattern of antibody response.
- Reinfection may occur in the presence of antibody. Such infection may or may not be symptomatic but will produce a boost to the antibody response. Although reinfection has been demonstrated in antibody-positive subjects, symptoms are usually less severe.
- Antigenically related organisms may also boost antibodies produced against the earlier infecting organism

- (anamnestic response). Subjects may not be protected against the effects of infection with an antigenically related but distinct species or sub-species. Whether this is true for *Cryptosporidium* species is not definitively known.
- Antibodies are variably protective but may indicate likelihood of the presence of other mechanisms (e.g. CMI) more closely associated with protection (resistance) in some infections such as *Cryptosporidium*.
- High rates of antibody in a population reflect frequency
 of exposure but not necessarily of overt disease. Such
 antibody prevalence may produce indirect protection to
 individuals from person-to-person (secondary) transmission through the mechanism of herd immunity. In
 the event of contamination of drinking water, the impact
 (attack rate) will likely be lower than in a similar event in
 areas of low seroprevalence.
- In the case of organisms with multiple routes of transmission in communities where there is high prevalence of antibody, the immunocompromised, visitors and young children may act as sentinels of the high endemicity and are at particular risk.
- Seroprevalence rates are generally higher in areas supplied by surface water sources, compared with groundwater. However, incidence (disease) rates may be lower than expected and this may be due to natural immunization of the resident population to symptomatic infection. Not all strains present will necessarily be pathogenic or virulent in humans but may be capable of provoking or boosting immunity.
- The multiple routes of transmissions (food, water, personto-person, etc.) of many of the pathogens described above make it difficult to estimate the proportion due to water. The only positive indication of source or route is in well-defined outbreaks that are investigated epidemiologically using case-control studies. In some of the studies described, there is a clear increase in prevalence of infection (symptomatic and asymptomatic) in areas supplied with water from surface water supplies compared with groundwater sources. It is reasonable to ascribe the excess, but not the total contribution, to water, if other potentially confounding risks can be excluded.
- Antibody resulting from infection acquired from other sources and routes (generally more common) will

contribute to moderating the impact of exposure derived from water.

- Antibody studies are valuable in the investigation of waterborne and other outbreaks as they are a more sensitive indicator of exposure rates than detection or isolation of the putative causal organism. This is especially true of the detection of parasites by stool microscopy, which is very insensitive (excretion of parasites is often intermittent; PCR detection can be used but is expensive and technically more demanding).
- Seroprevalence found in outbreak investigations can be used as an upper boundary for estimation of endemic prevalence.
- Absence of antibody indicates likely susceptibility to infection. In the event of widespread exposure through contamination of the water supply in a population with a low rate of seroprevalence, the attack rate is likely to be high. There is often a marked increase in primary cases amongst adults, followed by increased secondary transmission, mainly through children.
- Whether or not antibody *per se* is protective is not necessarily of importance if it acts as a correlate for immunity (resistance). Although subjects with antibody can often be reinfected (naturally and experimentally), and sometimes have symptoms, these are often reduced in severity. In addition, the age-related increase in seroprevalence correlates well with the reciprocal decline in incidence of acute clinical (symptomatic) infection.

Antibody measurement can be achieved by many different methods, some more suitable than others for epidemiologic purposes. Each method may have specific benefits or shortcomings. The results obtained from various studies using different methods or variations of methods cannot be compared in absolute terms but can be useful for illustrating general trends in relative frequency (prevalence). All tests used need to be rigorously controlled (QA/QC), including the use of known positive and negative samples. The main methods for use are western blot and ELISA, as follows:

 Large format western blot is both sensitive and specific. It provides a "gold standard" for identifying antibody positive and negative control samples and assessing

- equivocal results. However, it is cumbersome and expensive, which makes it unsuitable for epidemiological surveys.
- Minigel format western blot is economic and convenient
 to use. It has provided much useful epidemiologic
 information in relation to waterborne infection. It has a
 reduced sensitivity to a key antigen (27 kDa) and the
 significance of this needs to be investigated. Other
 workers have reported poorer results than those who
 developed the method and the robustness of the method
 for transfer needs to be established.
- The ELISA test is relatively cheap and easy to use. It is open to criticism if crude antigens are used but the use of clearly defined antigens in the CDC second-generation test makes an improved epidemiologic tool for estimating seroprevalence in different populations. However, false-positive and false-negative results have been reported and this needs to be further investigated. Although it has been demonstrated that it is robust and capable of transfer to other laboratories, the more widespread and economic availability of the antigens is crucial.

Certain caveats apply to methods using well-defined antigens:

- Two *Cryptosporidium* antigens are used in the current tests as important marker antigens but others have been identified and these need to be further investigated.
- The evaluation of such antigens has been with sera from adults. Their performance for sera from children needs to be evaluated.
- Some tests have been done to exclude cross-reacting antigens (e.g. *Toxoplasma*) but others may need to be tested. It is known, for example, that *Eimeria* may cross-react (Ortega-Mora *et al.* 1992), but this is thought to be of little significance in humans.
- The current tests have been developed with, and use, a particular strain of bovine-derived *C. parvum* and not *C. hominis* and or any of the other species known to infect humans. The relative sensitivity and specificity of these antigens in tests for antibodies in those who have been infected with species other than *C. parvum* is not currently clear and needs to be further evaluated.
- The significance of the intensity of an antibody response, whether expressed as titre, size and/or intensity of a WB

band, OD value, etc., and the impact that this may have on the assessment of the level of immunity (resistance) needs to be further investigated.

The current information is of value in assessing the approximate seroprevalence in certain populations of North America but it is unclear if these are sufficiently representative or robust enough to be used yet in the calculation of a national risk estimate.

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DISCLAIMER

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REFERENCES

- Adak, G., Cowden, J., Nichols, G. & Evans, H. 1995 The Public Health Laboratory Service national case-control study of primary indigenous sporadic cases of campylobacter infection. *Epidemiol. Infect.* **115**(1), 15–22.
- Adam, R. & Nash, T. 2004 Antigenic variation of the VSP genes of Giardia lamblia. In The Pathogenic Enteric Protozoa: Giardia, Entamoeba, Cryptosporidium and Cyclospora (ed. C. Sterling & R. Adam), pp. 59–73. Kluwer Academic, Boston.
- Aggarwal, R. & Krawczynski, K. 2000 Hepatitis E: an overview and recent advances in clinical and laboratory research. *J. Gastroenterol. Hepatol.* 15, 9–20.
- Al-Tukhi, M., Al-Ahdal, M., Ackers, J. & Peters, W. 1993 Enzymelinked immunosorbent assay for the detection of anti *Giardia* specific salivary immunoglobulin A. *Med. Sci. Res.* 21, 579–580.
- Anon 2000 Waterborne outbreak of gastroenteritis associated with a contaminated water supply. *Can. Commun. Dis. Rep.* **26**, 170–173.
- Appleton, H. 2000 Norwalk Virus and the small round viruses causing foodborne gastroenteritis. In *Foodborne Disease*

- *Handbook* (ed. K. Murrell, P. Stanfield, N. Wai-Kit, Y. Hui & S. Sattar), pp. 77–97. Marcel Dekker, New York.
- Atmar, R. & Estes, M. 2001 Diagnosis of noncultivable gastroenteritis viruses, and human Calicivirus. *Clin. Microbiol. Rev.* **14**(1), 15–37.
- Bahia-Olivera, L., Jones, J., Azavedo-Silva, J., Alves, C., Oréfice, F. & Addiss, D. 2003 Highly endemic, waterborne toxoplasmosis in North Rio de Janeiro State, Brazil. *Emerging Infect. Dis.* 9(1), 55-62.
- Belongia, E., Chyou, P., Greenelee, R., Perez-Perez, G., Bibb, W. & DeVries, E. 2003 Diarrhea incidence and farm-related risk factors for *Escherichia coli* 0157:H7 and *Campylobacter jejuni* antibodies among rural children. *J. Infect. Dis.* 187, 1460–1468.
- Beneson, M., Takafuji, E., Lemon, S., Greenup, R. & Sulzer, A. 1982 Oocyst-transmitted toxoplasmosis associated with ingestion of contaminated water. *New Engl. J. Med.* **307**, 666–669.
- Bern, C., Ortega, Y., Checkley, W., Roberts, J., Lescano, A., Cabera, L., Verastegui, M., Black, R., Sterling, C. & Gilman, R. 2002 Epidemiologic differences between cyclosporiasis and cryptosporidiosis in Peruvian children. *Emerging Infect. Dis.* 8(6), 581–585.
- Birkhead, G., Janoff, E., Voget, R. & Smith, P. 1989 Elevated levels of immunoglobulin A to *Giardia lamblia* during a waterborne outbreak of gastroenteritis. *J. Clin. Microbiol.* 27(8), 1707–1710.
- Bowie, W., King, A., Werker, D., Isaac-Renton, J., Bell, A., Eng, S. & Marion, S. 1997 Outbreak of toxoplasmosis associated with municipal drinking water. *The Lancet* **350**, 173–177.
- Braz, L., Neto, V., Ferrari, C., Palhares, M., Amato, V., Santos, M., Marques, H., Vallada, M., Nakanishi, L. & Andrade, H. 1996 Human cryptosporidiosis: detection of specific antibodies in the serum by an indirect immunofluorescence. *Rev. Saúde. Pública.* 30(5), 395-402.
- Campbell, P. & Current, W. 1983 Demonstration of serum antibodies to *Cryptosporidium* sp in normal and immunodeficient humans with confirmed infections. *J. Clin. Microbiol.* 18(1), 165–169.
- Caputo, C., Forbes, A. & Frost, F. 1999 Determinants of antibodies to *Cryptosporidium* infection among gay and bisexual men with HIV infection. *Epidemiol. Infect.* 122, 291–297.
- Casemore, D. 1987 The antibody response to cryptosporidium: development of a serological test and its use in a study of immunologically normal persons. *J. Infect.* 14, 125–134.
- Casemore, D. 1989 Human cryptosporidiosis. In *Recent Advances in Infection*, No. 3. (ed. D. Reeves & A. Geddes), pp. 209–236. Churchill Livingstone, Edinburgh.
- Casemore, D. 1990 Epidemiological aspects of human cryptosporidiosis. *Epidemiol. Infect.* **104**, 1–28.
- Casemore, D. 1994 Enteric protozoa and the water route of transmission epidemiology and dynamics. In *Water and Public Health* (ed. A. Golding, N. Noah & R. Stanwell-Smith), pp. 123–136. Smith-Gordon, Nishimura, London.
- Casemore, D. 1995 The problem with protozoan parasites. In *Protozoan Parasites and Water* (ed. W. Betts, D. Casemore, C.

- Fricker, H. Smith & J. Watkins), pp. 10-18. Royal Society of Chemistry, Cambridge.
- Casemore, D. & Jackson, F. 1984 Hypothesis: cryptosporidiosis in humans is not primarily a zoonosis. *J. Infect.* 9, 153-156.
- Casemore, D., Jessop, E., Douce, D. & Jackson, F. 1986 *Cryptosporidium* plus campylobacter: an outbreak in a semirural population. *J. Hygiene Camb.* **96**, 95–105.
- Casemore, D., Wright, S. & Coop, R. 1997 Cryptosporidiosis human and animal epidemiology. In *Cryptosporidium and Cryptosporidiosis* (ed. R. Fayer), pp. 65–92. CRC Press, Boca Raton, FL.
- Chalmers, R. & Casemore, D. 2004 Epidemiology and strain variation if Cryptosporidium. In The Pathogenic Enteric Protozoa: Giardia, Entamoeba, Cryptosporidium and Cyclospora (ed. C. Sterling & R. Adam), pp. 27–42. Kluwer Academic, Boston.
- Chalmers, R., Elwin, K., Reilly, W., Irvine, H., Thomas, A. & Hunter, P. 2002a Cryptosporidium in farmed animals: the detection of a novel isolate in sheep. Int. J. Parasitol. 32, 21–26.
- Chalmers, R., Elwin, K., Thomas, A. & Joynson, H. 2002b Infection with unusual types of *Cryptosporidium* is not restricted to immunocompromised patients. *J. Infect. Dis.* **185**, 270–271.
- Chappell, C., Okhuysen, P. & Dann, S. 2001 Serological response to *Cryptosporidium* infection. *Infect. Immun.* **69**(3), 1974–1975.
- Chappell, C., Okhuysen, P., Sterling, C. & DuPont, H. 1996 Cryptosporidium parvum: intensity of infection and oocyst excretion patterns in healthy volunteers. J. Infect. Dis. 173, 232–236.
- Chappell, C., Okhuysen, P., Sterling, C., Wang, C., Jakubowski, W. J. & Dupont, H. L. 1999 Infectivity of *Cryptosporidium parvum* in healthy adults with pre-existing anti-*C. parvum* serum immunoglobulin G. *Am. J. Trop. Med. Hygiene* **60**(1), 157–164.
- Char, S., Cevallos, A., Yamson, P., Sullivan, P. & Farthing, M. 1993 Impaired IgA response to *Giardia* heat shock antigen in children with persistent diarrhoea and giardiasis. *Gut* 34, 38–40.
- Char, S., Shetty, N., Narishman, M., Elliott, E., Macaden, R. & Farthing, M. 1991 Serum antibody response in children with *Giardia lamblia* infection and identification of an immunodominant 57-kilodalton antigen. *Parasit. Immunol.* 13, 329–337.
- Chart, H., Evans, J., Chalmers, R. & Salmon, R. 1998 Escherichia coli 0157 serology: false-positive ELISA results caused by human antibodies to bovine serum albumin. Lett. Appl. Microbiol. 27, 76–78.
- Clark, C. 2004 Entamoeba histolytica and Entamoeba dispar, the non-identical twins. In The Pathogenic Enteric Protozoa: Giardia, Entamoeba, Cryptosporidium and Cyclospora (ed. C. Sterling & R. Adam), pp. 15–26. Kluwer Academic, Boston.
- Clemente-Cesares, P., Pina, S., Buti, M., Jardi, R., Martin, M., Bofil-Mas, S. & Girones, R. 2003 Hepatitis E virus epidemiology in industrialized countries. *Emerging Infect. Dis.* **9**(4), 448–454.
- Cordell, R. & Addiss, D. 1994 Cryptosporidiosis in child care settings: a review of the literature and recommendations

- for prevention and control. *Pediatr. Infect. Dis. J.* **13**(4), 310-317.
- Cozon, G., Biron, F., Jeannin, M., Cannella, D. & Revillard, J-P. 1994 Secretory IgA antibodies to *Cryptosporidium parvum* in AIDS patients with chronic cryptosporidiosis. *J. Infect. Dis.* 169, 696–699
- Cox, M., Azavedo, R., Nokes, Beards, G., McCrae, M., Massad, E. & Medley, G. 1998a Seroepidemiology of group A rotavirus in suburban São Paulo. Brazil. *Epidemiol. Infect.* 120, 327 334.
- Cox, M., James, V., Azavedo, R., Massad, E. & Medley, G. 1998b Infection with group C rotavirus in a suburban community in Brazil. *Trop. Med. Int. Health* **3**(11), 891–895.
- Cox, M., Elwin, K., Massad, E. & Azavedo, R. 2005 Age-specific seroprevalence to an immunodominant *Cryptosporidium* sporozoite antigen in a Brazilian population. *Epidemiol. Infect.* 133, 951–956.
- Cox, M. & Medley, G. 2003 Serological survey of anti-group A rotavirus IgM in UK adults. *Epidemiol. Infect.* **131**, 719–726.
- Craun, G. 1990 Waterborne giardiasis. In *Giardiasis; Human*Parasitic Diseases (ed. E. Meyer), pp. 267–293. Elsevier,

 Amsterdam.
- Craun, G., Calderon, R. & Nwachuku, N. 2002 Causes of reported waterborne outbreaks in the United States, 1991-1998. In *Drinking Water and Infectious Disease: Establishing the Links* (ed. P. Hunter, M. Waite & E. Ronchi), pp. 105–117. CRC Press, Boca Raton, FL.
- Craun, G., Frost, F., Calderon, R., Hilborne, H., Fox, K., Reasoner, D., Poole, C., Rexing, D., Hubbs, S. & Dufour, A. 2001 Improving waterborne disease outbreak investigations. *Int. J. Environ. Health Res.* 11, 229–243.
- Current, W. 1989 Immunobiology of *Cryptosporidium* spp. *Pathol. Immunopathol.* **8**, 141–160.
- Current, W. & Bick, P. 1989 Immunobiology of Cryptosporidium spp. Pathol. Immunopathol. Res. 8, 141–160.
- Dann, S., Okhuysen, P., Salameh, B., DuPont, H. & Chappell, C. 2000 Fecal antibodies to *Cryptosporidium parvum* in healthy volunteers. *Infect. Immun.* 68, 5068–5074.
- Dowd, S., Gerba, C. & Pepper, I. 1998 Confirmation of the humanpathogenic Microsporidia *Enterocytozoon bieneusi*, *Encephalitozoon intestinalis*, and *Vittaforma cornea* in water. *Appl. Environ. Microbiol.* **64**(9), 3332–3335.
- Dubey, J. 2004 Toxoplasmosis a waterborne zoonosis. *Vet Parasitol.* **126**, 57–72.
- Duncanson, M., Chang, W., Frost, F., Muller, T. B. & Weinstein, P. 2003 Ubiquitous risk factor exposure and high prevalence of antibodies to *Cryptosporidium parvum* in two New Zealand communities. *Appl. Environ. Sci.*, *Public Health* 1(2), 111–117.
- DuPont, H., Chappell, C. & Sterling, C. 1995 The infectivity of Cryptosporidium parvum in health volunteers. New Engl. J. Med. 332, 855–859.
- Egorov, A., Frost, F., Muller, T., Naumova, E., Tereschenko, A. & Ford, T. 2003 Serological evidence of *Cryptosporidium* infections in a Russian city and evaluation of risk factors for infection. *Ann. Epidemiol.* **14**, 129–136.

- Eisenberg, J., Priest, J., Lammie, P. & Colford, J. 2001 The serological response to *Cryptosporidium* in HIV-infected persons: implications for epidemiologic research. *Emerging Infect. Dis.* 7(6), 1004–1009.
- Evans, J., Chalmers, R., Chart, R., Salmon, R., Kench, S., Coleman, T., Meadows, D., Morgan-Capner, P., Softly, P., Sillis, M. & Thomas, D. 2000 Evidence of persisting serum antibodies to *Escherichia coli* 0157 lipopolysaccharide and Verocytotoxin in members of rural communities in England. *Eur. J. Epidemiol.* 16, 885–889.
- Fankhauser, R., Noel, J., Monroe, S., Ando, T. & Glass, R. 1998 Molecular epidemiology of "Norwalk-like Viruses" in outbreaks of gastroenteritis in the United States. J. Infect. Dis. 178, 1571–1578.
- Farthing, M. 1989 Host-parasite interactions in human giardiasis. Quart. J. Med., New Series 70(263), 191–204.
- Farthing, M. 1990 Immunopathology of giardiasis. *Springer Semin. Immunopathol.* **12**, 269–282.
- Farthing, M. 1992 Giardia comes of age: progress in epidemiology, immunology and chemotherapy. J. Antimicro. Chemother. 30, 563-569.
- Farthing, M. 2003 Immune response-mediated pathology in human parasitic infection. *Parasit. Immunol.* 25, 247–257.
- Farthing, M., Goka, A., Butcher, P. & Aravind, A. 1987 Serodiagnosis of giardiasis. Serodiagn. Immun. 1, 133–238.
- Faubert, G. 2000 Immune response to Giardia duodenalis. Clin. Microbiol. Rev. 13(1), 35-54.
- Fine, P. 1993 Herd immunity: history, theory, practice. *Epidemiol. Rev.* **15**(2), 265–302.
- Franzen, C. & Müller, A. 1999 Molecular techniques for detection, species differentiation, and phylogenetic analysis of Microsporidia. Clin. Microbiol. Rev. 12(2), 243–285.
- Friedman, N., Frost, F., Caputo, C., Horrocks, M. & Fairley, C. 2001 One year followup of antibodies to *Cryptosporidium* among individuals with HIV infection. *Venerol.* **14**, 21–24.
- Frost, F. 1998 Two-city Cryptosporidium study. *Drink. Wat. Res.* **8**, 2–5. Frost, F. & Craun, G. 1998 Serologic response to human
 - Cryptosporidium infections. Infect. Immun. **66**(8), 4008.
- Frost, F. & Craun, G. 1998b The importance of acquired immunity in the epidemiology of cryptosporidiosis and giardiasis. In *OECD Workshop Molecular Methods for Safe Drinking Water*. Interlaken, pp. 1–12.
- Frost, F., de la Cruz, A., Moss, D., Curry, M. & Calderon, R. 1998a Comparison of ELISA and western blot assays for detection of *Cryptosporidium* antibody. *Epidemiol. Infect.* 121, 205–211
- Frost, F., Calderon, R., Muller, T., Curry, M., Rodman, J., Moss, D. & de la Cruz, A. 1998b A two-year follow-up survey of antibody to *Cryptosporidium* in Jackson County. Oregon following an outbreak of waterborne disease. *Epidemiol. Infect.* **121**, 213–217.
- Frost, F., Fea, E., Gilli, G., Biorci, F., Muller, T., Craun, G. & Calderon, R. 2000*a* Serological evidence of *Cryptosporidium* infections in southern Europe. *Eur. J. Epidemiol.* **16**, 385–390.

- Frost, F., Muller, T., Calderon, R. & Craun, G. 2000b A serological survey of college students for antibody to *Cryptosporidium* before and after the introduction of a new filtration plant. *Epidemiol. Infect.* **125**, 87–92.
- Frost, F., Muller, T., Craun, G., Fraser, D., Thompson, D., Notenboom, R. & Calderon, R. 2000c Serological analysis of a cryptosporidiosis epidemic. *Int. J. Epidemiol.* **29**, 376–379.
- Frost, F., Muller, T., Fairley, C., Hurley, J., Craun, G. & Calderon, R. 2000d Serological evaluation of *Cryptosporidium* oocyst findings in the water supply for Sydney, Australia. *Int. J. Environ. Health Res.* 10, 35–40.
- Frost, F., Muller, T., Craun, G., Calderon, R. & Roeffer, P. 2001 Paired city *Cryptosporidium* serosurvey in the southwest USA. *Epidemiol. Infect.* **126**, 301–307.
- Frost, F., Muller, T., Craun, G., Lockwood, W. & Calderon, R. 2002a Serological evidence of endemic waterborne *Cryptosporidium* infections. *Ann. Epidemiol.* **12**, 222–227.
- Frost, F., Kunde, T. & Craun, G. 2002b Is contaminated groundwater an important cause of viral gastroenteritis in the United States? *I. Environ. Health* **65**(3), 9–14.
- Frost, F., Muller, T., Kunde, T., Craun, G. & Calderon, R. 2003*a*Seroepidemiology. *Drinking Water and Infectious Disease* (ed. in P. Hunter, M. Waite & E. Ronchi), pp. 165–173. CRC
 Press, Boca Raton, FL.
- Frost, F., Muller, T., Kunde, T. & Craun, G. 2003b Quality assurance considerations in *Cryptosporidium* antibody tests. *Clin. Diagn. Lab. Immunol.* **10**(1), 193.
- Frost, F., Kunde, T., Muller, T., Craun, G., Katz, L., Hibbard, A. & Calderon, R. 2003c Serological responses to *Cryptosporidium* antigens among users of surface- vs. ground-water sources. *Epidemiol. Infect.* **131**, 1131–1138.
- Frost, F., Muller, T., Calderon, R. & Craun, G. 2004 Analysis of serological responses to *Cryptosporidium* antigen among NHANES III participants. *Ann. Epidemiol.* **14**, 473–478.
- Frost, F., Roberts, M., Kunde, T., Craun, G., Tollestrup, K., Harter, L. & Muller, T. 2005*a* How clean must our drinking water be: the importance of protective immunity. *J. Infect. Dis.* **191**, 809–814.
- Frost, F., Craun, G., Mihály, K., György, B., Calderon, R. & Muller, T. 2005*b* Serological responses to *Cryptosporidium* antigens among women using riverbank-filtered water, conventionally filtered surface water and groundwater in Hungary. *J. Wat. Health* 3, 77–82.
- Gallimore, C., Richards, A. & Gray, J. 2003 Molecular diversity of noroviruses associated with outbreaks on cruise ships: comparison with strains circulating within the UK. *Commun. Dis. Public Health* 6(4), 285–293.
- Gandhi, B., Buch, P., Sharma, M., Irshad, M. & Samantray, S. 1989 ELISA for anti-Giardia IgM. (letter) The Lancet 2, 685.
- Garcia, L., Fritsche, T., Grady, K., Healy, G., McAuley, J., Rocha, A., Wilson, M. & Wong, J. 2004 Clinical Use and Interpretation of Serologic Tests for Toxoplasma gondii; Approved Guideline. NCCLS document M36-A, Pennsylvania. 24(6), 1–21.
- García-Rodriguez, J., Sánchez, A., Canut, A. & García-Luis, E. 1989 The seroepidemiology of *Cryptosporidium* species in

- different groups in Spain. Serodiag. Immunother. Infect. Dis. 3, 367–373.
- Gardner, T. & Hill, D. 2001 Treatment of giardiasis. *Clin. Microbiol. Rev.* 14(1), 114–128.
- Gilman, R., Brown, K., Visvesvara, G., Mondal, G., Greenberg, B., Sack, B., Brandt, F. & Khan, M. 1985 Epidemiology and serology of *Giardia lamblia* in a developing country: Bangladesh. *Trans. Roy, Soc. Trop. Med. Hygiene* 79, 469–473.
- Gilman, R., Marquis, G., Miranda, E., Vestegui, M. & Martinez, H. 1988 Rapid reinfection by *Giardia lamblia* after treatment in a hyperendemic third world community. *The Lancet i*, 343–345.
- Glass, R., Noel, J., Ando, T., Fankhauser, R., Belliot, G., Mounts, A., Parashar, U., Bresee, J. & Monroe, S. 2000 The epidemiology of enteric Caliciviruses from humans: a reassessment using new diagnostics. J. Infect. Dis. 181(Suppl 2), S254–S261.
- Goh, S., Reacher, M., Casemore, D., Verlander, N., Chalmers, R., Knowles, M., Williams, J., Osborn, K. & Richards, S. 2004 Sporadic cryptosporidiosis, North Cumbria, England, 1996-2000. Emerging Infect. Dis. 10(6), 1007-1015.
- Goh, S., Reacher, M., Casemore, D., Verlander, N., Charlett, A., Chlamers, R., Knowles, M., Pennington, A., Williams, J., Osborn, K. & Richards, S. 2005 Sporadic cryptosporidiosis. Decline after membrane filtration of public water supplies, England, 1996–2002. *J. Emerging Infect. Dis.* 11, 251–259.
- Goka, A., Rolston, D., Mathan, V. & Farthing, M. 1986 Diagnosis of giardiasis by specific IgM antibody enzyme-linked immunosorbent assay. *The Lancet ii*, 184–186.
- Gomez-Morales, M., Pozio, E. & Croppo, G. 1992 Serodiagnosis of cryptosporidiosis in Italian HIV-positive patients by means of an oocyst soluble antigen in an ELISA. *J. Infect.* **25**, 229–236.
- Gray, J., Cunliffe, C., Ball, J., Graham, D., Dessulberger, U. & Estes, M. 1994 Detection of immunoglobulin M (IgM). IgA, and IgG Norwalk virus-specific antibodies by enzyme-linked immunosorbent assay using baculovirus-expressed Norwalk virus capsid antigen in adult volunteers challenged with Norwalk virus. J. Clin. Microbiol. 32(12), 3059–3063.
- Gray, J., Jiang, X., Morgan-Capner, P., Dessulberger, U. & Estes, M. 1993 Prevalence of antibodies to Norwalk virus in England: detection by enzyme-linked immunosorbent assay using baculovirus-expressed Norwalk virus capsid antigen. *J. Clin. Microbiol.* 31(4), 1022–1025.
- Groves, V., Lehman, D. & Gilbert, G. 1994 Seroepidemiology of cryptosporidiosis in children in Papua New Guinea and Australia. *Epidemiol. Infect.* **113**, 491–499.
- Hale, A., Lewis, D., Jiang, X. & Brown, D. 1998 Homotypic and heterotypic IgG and IgM antibody responses in adults infected with small round structured viruses. J. Med. Virol. 54, 305–312.
- Harrison, S., Nelder, R., Hayek, L., Mackenzie, I., Casemore, D. & Dance, D. 2002 Managing a large outbreak of cryptosporidiosis: how to investigate and when to decide to lift a 'boil water' notice. *Commun. Dis. Public Health* 5, 230-239.
- Hashkes, P., Spira, D., Decklebaum, R. & Granot, E. 1994 Salivary IgA antibodies to *Giardia lamblia* in day care center children. *Pediatr. Infect. Dis.* **13**, 953–958.

- Hayes, E., Matte, T., O'Brien, T., McKinley, T., Logsdon, G., Rose, J., Ungar, B., Word, D., Pinsky, P., Cummings, M., Wilson, M., Long, E., Hurwitz, E. & Juranek, D. 1989 Large community outbreak of cryptosporidiosis due to contamination of a filtered public water supply. *New Engl. J. Med.* 320, 1372–1376.
- Hegarty, J., Dowd, M. & Baker, K. 1999 Occurrence of *Helicobacter pylori* in surface water in the United States. *J. Appl. Microbiol.* 87(5), 687-701.
- Herwaldt, B. 2000 *Cyclospora cayetanensis*: a review, focusing on the outbreaks of cyclosporiasis in the 1990s. *Clin. Infect. Dis.* **31**, 1040–1057.
- Heyworth, M. 1990 Biological significance of *Giardia*-specific antibodies. *West. J. Med.* **153**(3), 293-295.
- Heyworth, M. 1992 Immunology of *Giardia* and *Cryptosporidium* infections. *J. Infect. Dis.* **166**, 465–472.
- Hollister, W., Canning, E. & Willcox, A. 1991 Evidence for widespread occurrence of antibodies to *Encephalitozoon cuniculi* (Microspora) in man provided by ELISA and other serological tests. *Parasitology* **102**, 33–43.
- Hopkins, R., Vial, P., Ferreccio, C., Ovalle, J., Prado, P., Sotomayer, V., Russel, R., Wasserman, S. & Morris, J. 1993 Seroprevalence of *Helicobacter pylori* in Chile: vegetables may serve as a route of transmission. *J. Infect. Dis.* 168, 222-226.
- Hrudy, S. & Hrudy, E. 2004 Safe Drinking Water: Lessons Learned From Recent Outbreaks in Affluent Nations. IWA Publishing, London.
- Hunt, C. 1999 Differentiation of human and animal isolates of Giardia intestinalis. MSc Thesis. Massey University Palmerston North, New Zealand.
- Hunter, P. 1997a Community study of infectious intestinal disease in England. *Brit. Med. J.* **319**, 258–259.
- Hunter, P. 1997b Waterborne Disease: Epidemiology and Ecology. Wiley, Chichester.
- Hunter, P. 2000 Modelling the impact of prior immunity, case misclassification and bias on case-control studies in the investigation of outbreaks of cryptosporidiosis. *Epidemiol. Infect.* **125**, 713–718.
- Hunter, P., Hughes, S., Woodhouse, S., Syed, Q., Verlander, N.,
 Chalmers, R., Morgan, K., Nichols, G., Beaching, N. &
 Osborn, K. 2004 Sporadic cryptosporidiosis case-control study with genotyping. *Emerging Infect. Dis.* 10(7), 1241–1249.
- Isaac-Renton, J., Blatherwick, J., Bowie, W., Fyfe, M., Khan, M., Li, A., King, A., McLean, M., Medd, L., Moorhead, W., Ong, C. & Robertson, W. 1999 Epidemic and endemic seroprevalence of antibodies to *Cryptosporidium* and *Giardia* in residents of three communities with different drinking water supplies. *Am. J. Trop. Med. Hygiene* 60(4), 578–583.
- Isaac-Renton, J., Lewis, L., Ong, C. & Nulsen, M. 1994 A second community outbreak of waterborne giardiasis in Canada and serological investigation of patients. *Trans. Roy. Soc. Med. Hygiene* 88, 395–399.
- Isaac-Renton, J., Moorhead, W. & Ross, A. 1996 Longitudinal studies of *Giardia* contamination in two community drinking water supplies: cyst levels, parasite viability, and health implications. *Appl. Environ, Microbiol.* 62(1), 47–54.

- Isaac-Renton, J., Ong, C., Bowie, W., Lammie, P. & Priest, J. 2003 *Cryptosporidium Serology in Human Populations*. AWWA Research Foundation, Denver, CO.
- Istre, G., Dunlop, T., Gaspard, B. & Hopkins, R. 1984 Waterborne giardiasis at a mountain resort: evidence for acquired immunity. *Am. J. Public Health* **74**(6), 602–604.
- Iturriza-Gómara, M., Clarke, I., Desselberger, U., Brown, D., Thomas, D. & Gray, J. 2004 Seroepidemiology of group C rotavirus infection in England and Wales. Eur. J. Epidemiol. 19, 589-595
- Janoff, E. & Smith, P. 1990 The role of immunity in *Giardia* infections. In *Giardiasis* (ed. E. Meyer), pp. 215-233. Elsevier Science, Amsterdam.
- Janoff, E., Smith, P. & Blaser, M. 1988 Acute antibody responses to Giardia lamblia are depressed in patients with AIDS. J. Infect. Dis. 157(4), 798-804.
- Jenkins, M., Trout, J., Murphy, C., Harp, J., Higgins, J., Wergin, W. & Fayer, R. 1999 Cloning and expression of a DNA sequence encoding a 41-kilodalton *Cryptosporidium parvum* oocyst wall protein. *Clin. Diagn. Lab. Immunol.* 6(6), 912–920.
- Jiang, B., Gentsch, J. & Glass, R. 2002 The role of serum antibodies in the protection against rotavirus disease: an overview. Clin. Infect. Dis. 34, 1351–1361.
- Jokipii, L., Miettinen, A. & Jokipii, A. 1988 Antibody to cysts of Giardia lamblia in primary giardiasis and in absence of giardiasis. J. Clin. Microbiol. 26(1), 121-125.
- Jones, J., Kruszon-Moran, D. & Wilson, M. 2003 Toxoplasma gondii infection in the United States, 1999-2000. Emerging. Infect. Dis. 9(11), 1371-1374.
- Jones, J., Kruszon-Moran, D., Wilson, M., McQuillan, G., Navin, T. & McAuley, J. 2001 *Toxoplasma gondii* infection in the United States: seroprevalence and risk factors. *Am. J. Epidemiol.* 154(4), 357-365.
- Joynson, D., Payne, R. & Rawal, B. 1990 Potential role of IgG avidity for diagnosing toxoplasmosis. J. Clin. Pathol. 43, 1032-1033.
- Kapel, N., Meillet, D., Buraud, M., Favennec, L., Magne, D. & Gobert, J. 1993 Determination of anti-Cryptosporidium coproantibodies by time-resolved immunofluorometric assay. Trans. Roy. Soc. Trop. Med. Hygiene 87, 330–332.
- Khanna, B., Cutler, A., Israel, N., Perry, M., Lastovica, A., Fields, P. & Gold, B. 1998 Use caution with serologic testing for *Helicobacter pylori* infection in children. *J. Infect. Dis.* **178**, 460–465.
- Kjos, S., Jenkins, M., Okhuysen, P. & Chappell, C. 2005 Evaluation of recombinant oocyst protein CP41 for detection of *Cryptosporidium*-specific antibodies. *Clin. Diagn. Lab. Immunol.* 12(2), 268–272.
- Koch, K., Phillips, D., Aber, R. & Current, W. 1985 Cryptosporidiosis in hospital personnel. Ann. Int. Med. 102, 593-596.
- Kuhls, T., Mosier, D., Crawford, D. & Griffis, J. 1994 Seroprevalence of cryptosporidial antibodies during infancy, childhood, and adolescence. Clin. Infect. Dis. 18, 731-735.
- Lane, S. & Lloyd, D. 2002 Current trends in research into the waterborne parasite *Giardia*. Crit. Rev. Microbiol. 28(2), 123-147.

- Laxer, M., Alcantara, A., Javato-Laxer, M., Menorca, D., Fernando, M. & Ranoa, C. 1990 Immune response to cryptosporidiosis in Philippine children. Am. J. Trop. Med. Hygiene 42(2), 131–139.
- Leach, C., Koo, F., Kuhls, T., Hilsenback, S. & Jenson, H. 2000 Prevalence of *Cryptosporidium parvum* infection in children along the Texas-Mexico border and associated risk factors. *Am. J. Trop. Med.* **62**(5), 656–661.
- Lengerich, E., Addiss, D., Marx, J., Ungar, B. & Juranek, D. 1993 Increased exposure to cryptosporidia among dairy farmers in Wisconsin. J. Infect. Dis. 167, 1252-1255.
- Ljungström, I. & Castor, B. 1992 Immune response to *Giardia lamblia* in a waterborne outbreak of giardiasis in Sweden. *J. Med. Microbiol.* **36**, 347–352.
- Lopez, C., Dykes, A., Juranek, D., Sinclair, S., Conn, J., Christie, R., Lippy, E., Schultz, M. & Mires, M. 1980 Waterborne giardiasis: a communitywide outbreak of disease and a high rate of asymptomatic infection. *Am. J. Epidemiol* 112, 495–507.
- Lopman, B., Brown, D. & Koopmans, M. 2002 Human caliciviruses in Europe. *J. Clin. Virol.* **24**, 137–160.
- Lopman, B., Reacher, M., Duijnhoven, Y., Hanon, F-X., Brown, D. & Koopman, M. 2003 Viral gastroenteritis outbreaks in Europe, 1995-2000. *Emerging Infect. Dis.* **9**(1), 90–96.
- Luft, B., Payne, D., Woodmansee, D. & Kim, C. 1987 Characterisation of the *Cryptosporidium* antigens from sporulated oocysts of *Cryptosporidium parvum*. *Infect. Immun*. 55(10), 2436–2441.
- Marshall, M., Naumovitz, D., Ortega, Y. & Sterling, C. 1997 Waterborne protozoan pathogens. *Clin. Microbiol. Rev.* **10**(1), 67–85.
- Mathieu, E., Levy, D., Veverka, F., Parrish, M-K., Sarisky, J., Shapiro, N., Johnston, S., Handzel, T., Hightower, A., Xiao, L., Lee, Y-M., York, S., Arrowood, M., Lee, R. & Jones, J. 2004 Epidemiologic and environmental investigation of a recreational water outbreak caused by two genotypes of *Cryptosporidium parvum* in Ohio in 2000. *Am. J. Trop. Med.* 71(5), 582–589.
- Matsui, S. & Greenberg, H. 2000 Immunity to Calicivirus infection. *J. Infect. Dis.* **181**(Suppl 2), S331–S335.
- McDonald, A., MacKenzie, W., Addiss, D., Gradus, M., Linke, G., Zembrowski, E., Hurd, M., Arrowood, M., Lammie, P. & Priest, J. 2001 *Cryptosporidium parvum*-specific antibody among children residing in Milwaukee during the 1993 waterborne outbreak. *J. Infect. Dis.* 183, 1373–1379.
- McLaughlin, J., Casemore, D., Moran, S. & Patel, S. 1998 The epidemiology of cryptosporidiosis: application of experimental sub-typing and antibody detection systems to the investigation of water-borne outbreaks. *Folia Parasitol.* **45**, 83–92.
- Mead, J., Arrowood, M. & Sterling, C. 1988 Antigens of *Cryptosporidium* sporozoites recognized by immune sera of infected animals and humans. *J. Parasitol.* **74**(1), 135–143.
- Meinhardt, P., Casemore, D. & Miller, K. 1996 Epidemiological aspects of human cryptosporidiosis and the role of waterborne transmission. *Epidemiol. Rev.* 18, 118–136.

- Meldrum, R., Griffiths, J., Smith, R. & Evans, M. 2005 The seasonality of human campylobacter infections and *Campylobacter* isolates from fresh, retail chickens in Wales. *Epidemiol. Infect.* **133**, 49–52.
- Meng, X., Purcell, R., Halbur, P., Lehman, J., Webb, D., Tsareva, T., Haynes, J., Thacker, B. & Emerson, S. 1997 A novel virus in swine is closely related to the human hepatitis E virus. *Proc. Natl. Acad. Sci. USA* 94, 9860–9865.
- Meng, X., Wiseman, B., Elvinger, F., Guenette, D., Toth, T., Engle, R., Emerson, S. & Purcell, R. 2002 Prevalence of antibodies to hepatitis E virus in veterinarians working with swine and in normal blood donors in the United States and other countries. *J. Clin. Microbiol.* 40(1), 117-122.
- Michaud, S., Ménard, S. & Arbeit, R. 2004 Campylobacteriosis, Eastern Township, Québec. *Emerging Infect. Dis.* 10(10), 1844–1847.
- Miotti, P., Gilman, R., Santosham, M., Ryder, R. & Yolken, R. 1986
 Age-related rate of seropositivity of antibody to *Giardia lamblia* in four diverse populations. *J. Clin. Microbiol.* 24(6), 972–975.
- Miron, D., Colodner, R. & Kenes, Y. 2000 Age-related seroprevalence of *Cryptosporidium* in northern Israel. *Israel. Med. Assoc. J.* 2, 343–345.
- Moore, G., Sogandares-Bernal, F., Dennis, M., Dennis, M., Root, D., Beckwith, D. & van Voorhis, D. 1982 Characterization of *Giardia lamblia* trophozoite antigens using polyacrylamide gel electrophoresis, high performance liquid chromatography, and enzyme-labelled immunosorbent assay. *Vet. Parasitol.* 10, 229–237.
- Morgan-Ryan, U., Fall, A., Ward, L., Hijjawi, I., Sulaiman, I., Fayer, R., Thompson, R., Olson, M., Lal, A. & Xiao, L. 2002 Cryptosporidium hominis n.sp (Apicomplexa: Cryptosporidiidae) from Homo sapiens. J. Euk. Microbiol. 49(6), 433-440.
- Moss, D., Bennett, S., Arrowood, M., Hurd, M., Lammie, P. & Wahlquist, S. 1994 Kinetic and isotypic analysis of specific immunoglobulins from crew members with cryptosporidiosis on a US Coast Guard cutter. J. Euk. Microbiol. 41(5), 52S-55S.
- Moss, D., Bennett, S., Arrowood, M., Wahlquist, S. & Lammie, P. 1998a Enzyme-linked immunoelectrotransfer blot analysis of a cryptosporidiosis outbreak on a US Coast Guard cutter. *Am. J. Trop. Med. Hygiene* **58**(1), 110–118.
- Moss, D., Chappell, C., Okhuysen, P., DuPont, H., Arrowood, M., Hightower, A. & Lammie, P. 1998b The antibody response to 27-, 17-, and 15-kDa *Cryptosporidium* antigens following experimental infection in humans. *J. Infect. Dis.* 178, 827–833.
- Muller, T., Frost, F., Craun, G. & Calderon, R. 2001 Serological responses to *Cryptosporidium* infection. *Infect. Immun.* 69(3), 1974
- Nacapunchai, D., Tepmongkol, M., Tharavanij, S., Thammapalerd, N. & Subchareon, A. 1986 A comparative study of four methods for detecting antibody in asymptomatic giardiasis. *Southern Asian J. Trop. Med. Hygiene* 17(1), 96–100.
- Nash, T. 1993 *Giardia lamblia* and giardiasis. In *Immunology and Molecular Biology of Parasitic Infections* (ed. in R. Warren & N. Agabian), pp. 157–169. Blackwell Scientific, New York.

- Nash, T. 1997 Antigenic variation in *Giardia lamblia* and the host's immune response. *Phil. Trans: Biol. Sci.* **352**, 1369–1375.
- Nayak, N., Ganguly, N., Walia, B., Wahi, V., Kanwar, S. & Mahajan, R. 1987 Specific secretory IgA in the milk of *Giardia lamblia*-infected and uninfected women. *J. Infect. Dis.* **155**(4), 724–727
- Newman, R., Sears, C., Moore, S., Nataro, J., Wuhib, T., Agnew, D., Guerrant, R. & Lima, A. 1999 Longitudinal study of *Cryptosporidium* infection in children in northeastern Brazil. *J. Infect. Dis.* 180, 167–175.
- Newman, R., Shu-Xian, Z., Wuhib, T., Lima, A., Guerrant, R. & Sears, C. 1994 Household epidemiology of *Cryptosporidium parvum* in an urban community in northeast Brazil. *Ann. Int. Med.* **120**(6), 500–505.
- Nichols, G. 2005 Fly transmission of *Campylobacter*. *Emerging Infect. Dis.* **11**(3), 361–364.
- Nicollier-Jamot, B., Pico, V., Pothier, P. & Kohli, E. 2003 Molecular cloning, expression, self-assembly, antigenicity, and seroepidemiology of a genotype II Norovirus isolated in France. J. Clin. Microbiol. 41(8), 3901–3904.
- Noel, J., Ando, T., Leite, J., Green, K., Dingle, K., Estes, M., Seto, Y., Monroe, S. & Glass, R. 1997 Correlation of patient immune responses with genetically characterized small round-structured viruses involved in outbreaks of nonbacterial acute gastroenteritis in the United States. *J. Med. Virol.* 53, 372–383.
- Okhuysen, P., Chappell, C., Crabb, J., Sterling, C. & DuPont, H. 1999 *Virulence of three distinct Cryptosporidium parvum* isolates for healthy adults. *J. Infect. Dis.* **180**, 1275–1281.
- Okhuysen, P., Chappell, C., Sterling, C., Jakubowski, W. & DuPont, H. L. 1998 Susceptibility and serologic findings of healthy adults to reinfection with *Cryptosporidium parvum*. *Infect. Immun*. 66(1), 441–443.
- Okhuysen, P., Jiang, X., Ye, L., Johnson, P. & Estes, M. 1995 Viral shedding and fecal IgA response after Norwalk virus infection. *J. Infect. Dis.* 171, 566–569.
- Ong, C., Eisler, D., Alikhani, A., Fung, V., Tomblin, J., Bowie, W. & Isaac-Renton, J. 2002 Novel *Cryptosporidium* genotypes in sporadic cryptosporidiosis cases: first report of human infections with a cervine genotype. *Emerging Infect. Dis.* 8(3), 263–268.
- Ong, C., Eisler, D., Goh, S., Tomblin, J., Awad-el-Kariem, F., Beard, C., Xiao, L., Sulaiman, I., Lal, A., Fyfe, M., King, A., Bowie, W. & Isaac-Renton, J. 1999 Molecular epidemiology of cryptosporidiosis outbreaks and transmission in British Columbia. Canada. Am. J. Trop. Med. Hygiene 61(1), 63–69.
- Ong, C., Li, A., Priest, J., Copes, R., Khan, M., Fyfe, M., Marion, S., Roberts, J., Lammie, P. & Isaac-Renton, J. 2005 Enzyme immunoassay of *Cryptosporidium*-specific immunoglobulin G antibodies to assess longitudinal infection trends in six communities in British Columbia, Canada. *Am. J. Trop. Med. Hygiene* 73(2), 288–295.
- Ortega-Mora, L., Tronsco, J., Rojo-Vasquesz, F. & Gomez-Bautista, M. 1992 Cross-reactivity of polyclonal serum antibodies generated against *Cryptosporidium parvum* oocysts. *Infect. Immun.* 60(8), 3442–3445.

- O'Ryan, M., Vial, P., Mamani, N., Jiang, X., Estes, M., Ferrocio, C., Lakkis, H. & Matson, D. 1998 Seroprevalence of Norwalk virus and Mexico virus in Chilean individuals: assessment of independent risk factors for antibody acquisition. *Clin. Infect. Dis.* 27, 789–795.
- Palm, J., Weiland, M., Griffiths, W., Griffiths, W., Lungström, J. & Svärd, S. 2003 Identification of immunoreactive proteins during acute human giardiasis. *J. Infect. Dis.* 187, 1849–1859.
- Patel, S., McLauchlin, J. & Casemore, D. 1997 A simple SDS-PAGE immunoblotting technique using an enhanced chemiluminescence detection system to identify polyclonal antibody responses to complex cryptosporidial antigen preparation following a monoclonal test and image overlay technique. *J. Immunol. Meth.* 205, 157 161.
- Patel, S., Pedraza-Diaz, S., McLauchlin, J. & Casemore, D. 1998 Molecular characterisation of *Cryptosporidium parvum* from two large suspected waterborne outbreaks. *Commun. Dis. Public Health* 1(4), 231–233.
- Payment, P. 1991 Antibody levels to selected enteric viruses in a French-Canadian population in the Province of Quebec. *Immunol. Infect. Dis.* 1, 317–322.
- Payment, P., Franco, E. & Fout, G. 1994 Incidence of Norwalk virus infections during a prospective epidemiological study of drinking water related gastrointestinal illness. *Can. J. Microbiol.* 40, 805–809.
- Pelosi, E., Lambden, P., Caul, E., Liu, B., Dingle, K., Deng, Y. & Clarke, I. 1999 The seroepidemiology of genogroup 1 and genogroup 2 Norwalk-like viruses in Italy. *J. Med. Virol.* 58, 93–99.
- Petry, F. 1998 Epidemiological study of *Cryptosporidium parvum* in sera of persons from Germany. *Infect.* **26**(1), 7–10.
- Pollok, R., Aljenaibi, M., McLauchlin, J., Kelly, M. & Farthing, M. 1998 Comparison of enzyme linked immunosorbent assay and western blotting in the detection of anti-cryptosporidial antibodies in the investigation of a waterborne outbreak of cryptosporidiosis. Gut. 42(Suppl 1), A85.
- Poovorawan, Y., Chatchatee, P. & Chongsrisawat, V. 2002 Epidemiology and prophylaxis if viral hepatitis: a global perspective. *J. Gastroenterol. Hepatol.* 17, S155–S166.
- Priest, J., Kwon, J., Moss, D., Roberts, M., Arrowood, M., Dworkin, M., Juranek, D. & Lammie, P. 1999 Detection by immunoassay of serum immunoglobulin G antibodies that recognize Cryptosporidium parvum antigens. J. Clin. Microbiol. 37(5), 1385-1392.
- Priest, J., Lammie, P., Li, A., Khan, M., Ong, C. & Isaac-Renton, J. 2003 Quality assurance considerations in *Cryptosporidium* antibody tests (Authors' reply). *Clin. Diagnost. Lab. Immun.* **10**(1), 193–194.
- Priest, J., Li, A., Khan, M., Arrowood, M., Lammie, P., Ong, C., Roberts, M. & Isaac-Renton, J. 2001 Enzyme immunoassay of antigen-specific immunoglobulin G antibodies in longitudinal serum samples from patients with cryptosporidiosis. *Clin. Diagnost. Lab. Immunol.* 8(2), 415-423.

- Priest, J., Bern, C., Roberts, J., Kwon, J., Lescano, A., Checkley, W., Cabrera, L., Moss, D., Arrowood, M., Sterling, C., Gilman, R. & Lammie, P. 2005 Changes in serum immunoglobulin G levels as a marker for *Cryptosporidium* sp. infection in Peruvian children. *J. Clin. Microbiol.* 43, 5298-5300.
- Priest, J., Bern, C., Xiao, L., Roberts, J., Kwon, J., Lescano, A.,
 Checkley, W., Cabrera, L., Moss, D., Arrowood, M., Sterling,
 C., Gilman, R. & Lammie, P. 2006a Longitudinal analysis of
 Cryptosporidium species-specific immunoglobulin G antibody
 responses in Peruvian children. Clin. Vacc. Immunol. 13(1),
 123-131.
- Raina, P., Pollari, F., Teare, G., Gross, M. J., Barry, A. J. & Wilson, J. B. 1999 The relationship between E. coli indicator bacteria in well-water and gastrointestinal illness in rural families. Rev. Canad. De Santé Publ. 90(3), 172-175.
- Rangel, J., Sparling, P., Crowe, C., Griffin, P. & Swerdlow, D. 2005 Epidemiology of *Escherichia coli* 0157:H7 outbreaks, United States, 1982-2002. *Emerging Infect. Dis.* 11(4), 603–609.
- Reiner, D. & Gillin, F. 1992 human secretory and serum antibodies recognize environmentally induced antigens of *Giardia lamblia*. *Infect. Immun.* **60**(2), 637–643.
- Répérant, J-M., Naciri, M., Iochmann, S., Tilley, M. & Bout, D. 1994 Major antigens of *Cryptosporidium parvum* recognized by serum antibodies from different infected animal species and man. *Vet. Parasitol.* **55**, 1–13.
- Ridley, M. & Ridley, D. 1976 Serum antibodies and jejunal histology in giardiasis associated malabsorption. *J. Clin. Pathol.* **29**, 30–34.
- Riggs, M. 2002 Recent advances in cryptosporidiosis: the immune response. *Microbes Infection* 4, 1067–1080.
- Rodrigues, L., Cowden, J., Wheeler, J., Sethi, D., Wall, P., Cumberland, P., Tompkins, D., Hudson, M., Roberts, J. & Roderick, P. 2001 The study of infectious intestinal disease in England: risk factors for cases of infectious intestinal disease with *Campylobacter jejuni* infection. *Epidemiol. Infect.* 127, 185–193.
- Roitt, I. & Delves, P. 2001 Essential Immunology. Blackwell Science, Oxford.
- Rose, J., Epstein, P., Lipp, E., Sherman, B., Bernard, S. & Patz, J. 2001 Climate variability and change in the United States: potential impacts on water- and foodborne disease caused by microbiologic agents. *Environ. Health Perspect.* 109(Suppl 2), 211–221.
- Roy, S., DeLong, S., Stenzel, S., Shiferaw, B., Roberts, J., Khalakdina, A., Marcus, R., Segler, S., Shah, D., Thomas, S., Virgia, D., Zansky, S., Dietz, V. & Beach, M. 2004 Risk factors for sporadic cryptosporidiosis among immunocompetent persons in the United States from 1999 to 2001. *J. Clin. Microbiol.* 42(7), 2944–2951.
- Roy, S. L., Scallan, E. & Beach, M. J. 2006 The rate of acute gastrointestinal illness in developed countries. *J. Wat. Health* **4**(Suppl. 2), 31–70.
- Said, B., Wright, F. & Nichols, G. 2003 Outbreaks of infectious disease associated with private drinking water supplies in

- England and Wales, 1970-2000. *Epidemiol. Infect.* **130**, 469–479.
- Schaub, S. & Oshiro, R. 2000 Public health concerns about Caliciviruses as waterborne contaminants. *J. Infect. Dis.* **181**(Suppl. 2), S374–S380.
- Schlauder, G., Dawson, G., Erker, J., Kwo, P., Knigge, M., Smalley, D., Rosenblatt, J., Desai, S. & Mushahwar, I. 1998 The sequence and phylogenetic analysis of a novel hepatitis E virus isolated from patient with a cute hepatitis reported in the United States. *J. Gen. Virol.* 79, 447–456.
- Shandera, W. 1990 From Leningrad to the day-care center. The ubiquitous Giardia lamblia. *West. J. Med.* **153**, 154–159.
- Smit, T., Bos, P., Peenze, I., Jiang, X., Estes, M. & Steele, A. 1999 Seroepidemiological study of genogroup I and II Calicivirus infections in South and Southern Africa. *J. Med. Virol.* 59, 227–231.
- Smith, L., Priest, J., Lammie, P. & Mead, J. 2001 Human T and B cell immunoreactivity to a recombinant 23-kDa Cryptosporidium parvum antigen. J. Parasitol. 87(3), 704-707.
- Smith, P., Gillin, F., Brown, W. & Nash, T. 1981 IgG antibody to *Giardia lamblia* detected by enzyme-linked immunosorbent assay. *Gastroenterology* **80**, 1476–1480.
- Soliman, M., Taghi-Kilani, R., Abou-Shady, A., El-Mageid, S., Handousa, A., Hegazi, M. & Belesovic, M. 1998 Comparison of serum responses to *Giardia lamblia* in symptomatic and asymptomatic patients. *Am. J. Trop. Med. Hygiene* 58(2), 232–239.
- Spenser, K., Soave, R., Acosta, A., Gellin, B., Prince, A., Ramos, L. & Jacobs, J. 1997 Cryptosporidiosis in HIV-infected persons: prevalence in a New York City population. *Int. J. Infect. Dis.* 1, 217–221.
- Steinberg, E., Mendoza, C., Glass, R., Arana, B., Lopez, B., Mejia, M., Gold, B., Priest, J., Bibb, W., Monroe, S., Bern, C., Bell, B., Hoekstra, R., Klein, R., Mintz, E. & Luby, S. 2004 Prevalence of infection with waterborne pathogens: a seroepidemiologic study in children 6-36 months old in San Juan Sacatepequez, Guatemala. Am. J. Trop. Med. Hygiene 70(1), 83-88.
- Steiner, T. & Guerrant, R. 1996 The pathogenesis of and host response to *Cryptosporidium parvum*. *Curr. Opin. Infect. Dis.* 9, 156-160.
- Sterling, C., Gilman, R., Sinclair, N., Cama, V., Castillo, R. & Diaz, F. 1991 The role of breast milk in protecting urban Peruvian children against cryptosporidiosis. *J. Protozool.* 38(6), 23S-25S.
- Sterling, C. & Ortega, Y. 2004 Cyclospora cayetanensis: an emergent and still perplexing coccidian parasite. In The Pathogenic Enteric Protozoa: Giardia, Entamoeba, Cryptosporidium and Cyclospora (ed. C. Sterling & R. Adam), pp. 43–57. Kluwer, Boston.
- Strauss, B., King, W., Ley, A. & Hoey, J. 2001 A prospective study of rural drinking water quality and acute gastrointestinal illness. *Bio. Med. Centr. Public Health* 1(8) Available at: http://www.biomedcentral.com/1471-2458/1/8.
- Stuart, J., Orr, H., Warburton, F., Jeyakanth, S., Pugh, C., Morris, I., Sarangi, J. & Nichols, G. 2003 Risk factors for sporadic

- giardiasis: a case-control study in Southwestern England. *Emerging Infect. Dis.* **9**(2), 229–233.
- Sullivan, P., Neale, G., Cevallos, A. & Farthing, M. 1991 Evaluation of specific serum anti-Giardia IgM antibody response in diagnosis of giardiasis in children. Trans. Roy. Soc. Trop. Med. Hygiene 85, 748–749.
- Sullivan, R., Linneman, C., Clark, C. & Walzer, P. 1987 Seroepidemiologic study of giardiasis patients and high-risk groups in a Midwestern city in the United States. *Am. J. Public Health* 77(8), 960–963.
- Tanriverdi, S., Arslan, M., Akiyoshi, D., Tzipori, S. & Widmer, G. 2003 Identification of genotypically mixed *Cryptosporidium* parvum populations in humans and calves. *Mol. Biochem.* Parasitol. 130, 13–22.
- Tarleton, J. & Petri, W. 2004 Pathogenesis and immunity to Entamoeba histolytica. In The Pathogenic Enteric Protozoa: Giardia, Entamoeba, Cryptosporidium and Cyclospora (ed. C. Sterling & R. Adam), pp. 75–89. Kluwer, Boston.
- Taylor, G. & Wenman, W. 1987 Human immune response to *Giardia lamblia* infection. *J. Infect. Dis.* **155**(1), 137–140.
- Tellez, A., Winiecka-Krusnell, J., Paniagua, M. & Linder, E. 2003 Antibodies in mother's milk protect children against giardiasis. *Scand. J. Infect. Dis.* **35**, 322–325.
- Teunis, P., Chappell, C. & Okhuysen, P. 2002 *Cryptosporidium* dose response studies: variation between isolates. *Risk Analysis* **22**, 175–183.
- Thompson, R. 2004 Epidemiology and zoonotic potential of *Giardia* infections. In *The Pathogenic Enteric Protozoa: Giardia*, *Entamoeba, Cryptosporidium and Cyclospora* (ed. C. Sterling & R. Adam), pp. 1–13. Kluwer, Boston.
- Tillett, H., De Louvois, J. & Wall, P. 1998 Surveillance of outbreaks of waterborne infectious disease: categorizing levels of evidence. *Epidemiol. Infect.* **120**, 37-42.
- Tzipori, S. & Campbell, I. 1981 Prevalence of *Cryptosporidium* antibodies in 10 animal species. *J. Clin. Microbiol.* **34**, 455–456.
- Udezulu, I., Visvesvara, G., Moss, D. & Leitch, G. 1992 Isolation of two Giardia lamblia (WB strain) clones with distinct protein and antigenic profiles and differing infectivity and virulence. Infect. Immun. 60(6), 2274–2280.
- Ungar, B., Gilman, R., Lanata, C. & Perez-Schael, I. 1988 Seroepidemiology of *Cryptosporidium* infection in two Latin American populations. *J. Infect. Dis.* 157(3), 551–556.
- Ungar, B., Mulligan, M. & Nutman, T. 1989 Serologic evidence of Cryptosporidium infection in US volunteers before and during Peace Corps service in Africa. Arch. Intern. Med. 149, 804 – 807
- Ungar, B. & Nash, T. 1986 Quantification of specific antibody response to *Cryptosporidium* antigens by laser densitometry. *Infect. Immun.* 53(1), 124-128.
- Ungar, B., Soave, R., Fayer, R. & Nash, T. 1986 Enzyme immunoassay detection of immunoglobulin M and G antibodies to *Cryptosporidium* in immunocompetent and immunocompromised persons. *J. Infect. Dis.* 153(3), 570–578.

- US EPA 1998 Giardia: Human Health Criteria Document. Report EPA-823-R-002. United States Environmental Protection Agency, Office of Water, Washington, DC.
- Van Gool, T., Biderre, C., Delbac, F., Wentink-Bonnema, E., Peek, R. & Vivarès, C. 2004 Serodiagnostic studies in an immunocompetent individual infected with *Encephalitozoon* cuniculi. J. Infect. Dis. 189, 2243–2249.
- Venkatesan, P. & Wakelin, D. 1993 ELISAs for parasitologists: lies, damned lies and ELISAs. *Parasitol. Today.* 9(6), 228-232.
- Vinayak, V. & Kumkum, K. 1989 Serum antibodies to giardial surface antigens: lower titres in persistent than in non-persistent giardiasis. *J. Med. Microbiol.* **30**, 207–212.
- Von Bonsdorff, C-H. & Maunula, L. 2003 Microbiology and the investigation of waterborne outbreaks: typing of Norwalk-like virus. In *Drinking Water and Infectious Disease: Establishing* the Links (ed. P. Hunter, M. Waite & E. Ronchi), pp. 79–85. CRC Press, Boca Raton, FL.
- Walterspiel, J., Morrow, A., Guerrero, M., Ruiz-Palacios, G. & Pickering, L. 1994 Secretory anti- *Giardia lamblia* antibodies in human milk: protective effect against diarrhea. *Pediatr*. 93(1), 28-31.
- Ward, P., Deplazes, P., Regli, W., Rinder, H. & Mathis, A. 2002 Detection of eight *Cryptosporidium* genotypes in surface and waste waters in Europe. *Parasitology* **124**, 359–368.
- Widdowson, M., Sulka, A., Bulens, S., Beard, R., Chaves, S., Hammond, R., Salehi, E., Swanson, E., Totaro, J., Woron, R., Mead, P., Bresee, J., Monroe, S. & Glass, R. 2005a Norovirus and foodborne disease. United States, 1991-2000. Emerging Infect. Dis. 11(1), 95-102.

- Widdowson, M., Monroe, S. & Glass, R. 2005b Are Noroviruses emerging? *Emerging Infect. Dis.* 11(5), 735-737.
- Willocks, L., Crampin, L., milne, C., Seng, C., Susman, M., Gair, R., Moulsdale, M., Shafi, S., Wall, R. & Lightfoot, N. 1998 A large outbreak of cryptosporidiosis associated with a public water supply from a deep chalk borehole. *Commun. Dis. Pub. Hlth*. 1, 239–243.
- Wyatt, C. & McDonald, V. 2004 Innate and T cell-mediated immune responses in cryptosporidiosis. In *The Pathogenic Enteric Protozoa: Giardia, Entamoeba, Cryptosporidium and Cyclospora* (ed. C. Sterling & R. Adam), pp. 91–101. Kluwer, Boston.
- Xiao, L., Fayer, R., Ryan, U. & Upton, S. 2004 Cryptosporidium taxonomy: recent advances and implications for public health. Clin. Microbiol. Rev. 17(1), 72-97.
- Xiao, L. & Ryan, U. 2004 Cryptosporidiosis: an update in molecular epidemiology. Curr. Opin. Infect. Dis. 17, 483-490.
- Zhou, L., Fayer, R., Trout, J., Ryan, U., Schaeffer, F., III & Xiao, L. 2004 Genotypes of *Cryptosporidium* species infecting furbearing mammals differ from those of species infecting humans. *Appl. Environ. Microbiol.* 70(12), 7574–7577.
- Zu, S-X., Fang, G-D., Fayer, R. & Guerrant, R. 1992 Cryptosporidiosis: pathogenesis and immunology. *Parasitol. Today* 8(1), 24-27.
- Zu, S-X., Li, J., Barrett, L., Fayer, R., Shu, S., McAuliffe, J., Roche, J. & Guerrant, R. 1994 Seroepidemiologic study of *Cryptosporidium* infection in children from rural communities of Anhui. *Am. J. Trop. Med.* 51(1), 1–10.