

Vector Competence of California Mosquitoes for West Nile virus

Laura B. Goddard,* Amy E. Roth,* William K. Reisen,* and Thomas W. Scott*

To identify the mosquito species competent for *West Nile virus* (WNV) transmission, we evaluated 10 California species that are known vectors of other arboviruses or major pests: *Culex tarsalis*, *Cx. pipiens pipiens*, *Cx. p. quinquefasciatus*, *Cx. stigmatosoma*, *Cx. erythrothorax*, *Ochlerotatus dorsalis*, *Oc. melanimon*, *Oc. sierrensis*, *Aedes vexans*, and *Culiseta inornata*. All 10 became infected and were able to transmit WNV at some level. *Ochlerotatus*, *Culiseta*, and *Aedes* were low to moderately efficient vectors. They feed primarily on mammals and could play a secondary role in transmission. *Oc. sierrensis*, a major pest species, and *Cx. p. quinquefasciatus* from southern California were the least efficient laboratory vectors. *Cx. tarsalis*, *Cx. stigmatosoma*, *Cx. erythrothorax*, and other populations of *Cx. pipiens* complex were the most efficient laboratory vectors. *Culex* species are likely to play the primary role in the enzootic maintenance and transmission of WNV in California.

Three years since its 1999 introduction into North America, *West Nile virus* (WNV) has spread rapidly from New York to the Rocky Mountains and to the Gulf of Mexico. As of September 2002, over 1,900 human cases of WNV encephalitis have been confirmed with 94 deaths; >6,000 equine cases also occurred during 2002 (1). The imminent spread of this virus culminated in the establishment of WNV surveillance programs in 48 states. Surveillance programs include testing mosquito pools for virus, sentinel chickens for seroconversion, wild birds for virus and seroconversion, and equine and human cases (2).

WNV is a geographically widespread arbovirus in the family *Flaviviridae*, genus *Flavivirus* (3). The virus, first isolated from the blood of a woman in the West Nile district of Uganda in 1937 (4), historically has been endemic to Africa, Western Asia, and the Middle East. Recently, WNV has expanded its distribution and caused epidemics in Russia, Romania, France, and Israel (5,6).

WNV is maintained in an enzootic transmission cycle among *Culex* mosquitoes and wild birds. In Africa and the Middle East, WNV has been most frequently isolated from *Cx. univittatus* (7,8). In Asia, members of the *Cx. vishnui* complex have been implicated as the primary vectors (9). *Cx. modestus* was identified as a principal vector during a 1960s epidemic in France (3). During the North American outbreak, members of the *Cx. pipiens* complex were considered the primary epizootic vectors (10). Since the New York outbreak in 1999, WNV has been recovered from 26 North American mosquito species, including *Cx. pipiens*, *Cx. salinarius*, *Cx. restuans*, *Ochlerotatus canadensis*, *Oc. japonicus*, *Aedes vexans*, and *Culiseta melanura* (11,12). Recent vector competence studies indicate that some North American *Culex* and *Ochlerotatus* species are relatively efficient laboratory vectors (13–15).

As WNV expands its range westward across North America, examining the vector competence of the different mosquito species will help to anticipate patterns of transmission and the relative contribution of different vector species to virus amplification and persistence. The enzootic transmission cycles of WNV, *Saint Louis encephalitis virus* (SLEV), and *Western equine encephalomyelitis virus* (WEEV) in North America are conceptually identical, with *Culex* vectors transmitting virus among passerine avian hosts. In the western United States, SLEV and WEEV share a common mosquito host, *Cx. tarsalis*, which will presumably also support WNV transmission. Moreover, WNV and SLEV are closely related viruses in the *Japanese encephalitis virus* (JEV) serocomplex (3), and *Cx. tarsalis* has been shown to be an efficient vector of both SLEV (16) and JEV (17). Current WNV control strategies are based largely on vector control (18); therefore, identifying which species have the greatest potential for transmission is essential in formulating and focusing a prevention plan (19). We evaluated 10 California vector and pest mosquito species' for their ability to become infected with and transmit WNV.

Materials and Methods

Mosquitoes

We assessed the vector competence for WNV of 10 California mosquito species from 14 different geographic locations (Table 1). Vector competence refers to the intrinsic permissiveness of an arthropod for the infection, replication, and transmission of a virus (20,21). Voucher specimens for each species were deposited at the Bohart Museum of Entomology at the University of California, Davis, California. *Cx. tarsalis* is the principal enzootic vector of WEEV and SLEV in California (22). Members of the *Cx. pipiens* complex have been primary vectors of WNV in New York (10) and could potentially play a

*University of California, Davis, California, USA

RESEARCH

Table 1. California mosquito species tested for vector competence for WNV^a

Species	Source ^b	Generation
<i>Culex tarsalis</i>	Yolo Co.	F ₁
	Bakersfield, Kern Co.	F ₁
	Coachella Valley, Riverside Co.	F ₁
<i>Cx. pipiens quinquefasciatus</i>	Bakersfield, Kern Co.	F ₀
	Coachella Valley, Riverside Co.	F ₁
	Orange Co.	Wild adults
<i>Cx. p. pipiens</i>	Shasta Co.	F ₁
<i>Cx. stigmatosoma</i>	Chino, San Bernardino Co	Wild adults
<i>Cx. erythrothorax</i>	San Joaquin Marsh, Orange Co.	Wild adults
	Coachella Valley, Riverside Co.	Wild adults
<i>Ochlerotatus dorsalis</i>	Morro Bay, San Luis Obispo Co.	F ₀
<i>Oc. melanimon</i>	Lost Hills, Kern Co.	Wild adults
<i>Oc. sierrensis</i>	Lake Co.	F ₀
<i>Aedes vexans</i>	Coachella Valley, Riverside Co.	F ₁
<i>Culiseta inornata</i>	Lost Hills, Kern Co.	F ₀

^aWNV, West Nile virus, Co., County; F₀, adults reared from wild-caught larvae or eggs; F₁, progeny from wild-caught adults reared in the laboratory; wild adults, wild-caught adults of unknown age.

^bAll mosquitoes were collected during 2001 except *Cx. p. quinquefasciatus* (Orange Co.), *Cx. stigmatosoma*, *Cx. erythrothorax*, and *Cx. inornata*, which were collected during 2002.

similar role in California, especially in urban environments. We defined members of the *Cx. pipiens* complex on the basis of the geographic location of collection and on previously described hybrid zones in California (23). Consequently, we considered members of the complex collected from northern California to be *Cx. p. pipiens*, and those collected from central and southern California to be *Cx. p. quinquefasciatus*. *Cx. stigmatosoma*, an abundant species in California, is naturally infected with WEEV and SLEV (24) and is an efficient laboratory vector of SLEV (16). *Cx. erythrothorax*, another widespread species, typically inhabits marshlands and is an opportunistic feeder (25). *Oc. dorsalis* and *Oc. melanimon*, involved in the transmission of WEEV among small mammals, are laboratory-confirmed vectors of WEEV (26,27). *Oc. sierrensis* is a major pest in California that frequently bites humans and other mammals and transmits dog heartworm, *Dirofilaria immitis* (28). We tested *Ae. vexans* because it feeds readily on mammals (29) and was found to be naturally infected with WNV during the 1999 New York outbreak (11,12). *Cs. inornata* is a mosquito that is active during the winter; this species could potentially maintain WNV amplification and transmission during winter months when *Culex* species are inactive (30).

Virus and Virus Assay

We used WNV strain 35211 AAF 9/23/99, which was isolated from a flamingo during the 1999 New York outbreak and passaged twice in Vero (African green monkey kidney) cell cultures. All artificial blood meal, transmission, and mosquito

body samples were examined for virus by plaque assay in six-well tissue culture plates (Costar, Corning, NY) containing monolayers of Vero cells. Mosquito bodies were ground individually in 0.5 mL of mosquito diluent (phosphate-buffered saline [PBS], 20% fetal bovine serum [FBS], antibiotics). Plaque assays were conducted by adding 100 μ L of each sample to confluent cell monolayers and incubating inoculated cells at 37°C for 1.5 h to allow for virus to attach and enter cells. After incubation, cells were covered with a 2% agarose overlay containing 0.005% neutral red. After 96 h and 120 h of incubation at 37°C, in a 5% CO₂ atmosphere, plaques were counted, and virus concentrations were calculated as PFUs per 1.0 mL.

Mosquito Infection

Mosquitoes were infected orally by feeding on hanging blood droplets (defibrinated rabbit blood [Microbiological Media, San Ramon, CA]) containing 2.5% sugar and 10^{7.1±0.1} or 10^{4.9±0.1} WNV PFUs/1.0 mL of blood. Infectious blood was diluted in bovine albumin-PBS and stored at -80°C until examined by plaque assay to determine the titer. Engorged mosquitoes were held at 28°C, during a 16:8 light:dark photoperiod, and provided a 10% sucrose solution in cotton wicks.

Experimental Transmission

Mosquitoes were deprived of sucrose for 24 h before transmission attempts. On days 7 and 14 after infection, mosquitoes were immobilized by exposure to triethylamine and their proboscises were inserted into a capillary tube containing a 1:1 FBS and 10% sucrose solution for 10 min (31). Transmission fluid was expelled into 250 μ L of mosquito diluent and frozen at -80°C until assayed. Individual mosquito bodies were similarly frozen at -80°C before being thawed, ground, and assayed.

Statistical Analysis

Infection and transmission rates were compared at day 7 and day 14 data for each dose by the Fisher exact test using SAS 8.2 (SAS Institute, Inc., Cary, NC). Differences were considered statistically significant at alpha \geq 0.05 and adjusted for multiple comparisons.

Results

All mosquito species tested were susceptible to infection, and WNV was detected, to some extent, in their salivary secretions. Infection rates were generally higher after 7 days' incubation than 14 days. Transmission rates were generally highest for females infected with the high dose of 10^{7.1±0.1} PFU/mL and incubated for 14 days (Table 2).

Infection rates varied markedly among species but were consistently highest after infection with the high dose of WNV. Infection rates of *Culex* species and *Cs. inornata* tested 14 days after imbibing the high virus dose ranged from 58% to 100%, except for *Cx. p. quinquefasciatus* from the Coachella Valley and Orange County. *Oc. dorsalis* and *Oc. melanimon*

Table 2. Infection and transmission rates for California mosquito species orally infected with $10^{7.1\pm 0.1}$ PFU/mL of *West Nile virus* (WNV)

Species	Source by county	Day transmission attempted	No. tested	Infection rate ^a	Transmission rate ^b
<i>Culex tarsalis</i>	Yolo	7	30	87	60
		14	1	100	100
	Kern	7	15	93	40
		14	35	74	60
	Riverside	7	49	94	10
		14	55	85	62
<i>Cx. pipiens quinquefasciatus</i>	Kern	7	50	86	4
		14	50	58	52
	Riverside	7	60	8	0
		7	60	13	2
		14	58	28	19
		14	45	80	9
	Orange	7	45	80	9
		14	50	66	36
<i>Cx. p. pipiens</i>	Shasta	7	17	100	0
		14	31	100	71
<i>Cx. stigmatosoma</i>	San Bernardino	7	15	67	0
		14	48	77	19
<i>Cx. erythrothorax</i>	Orange	7	15	100	33
		14	25	100	64
<i>Ochlerotatus dorsalis</i>	Kern	7	30	50	13
		14	29	41	34
<i>Oc. melanimon</i>	San Luis Obispo	7	50	46	18
		14	60	48	20
<i>Oc. sierrensis</i>	Lake	7	40	5	3
		14	50	14	6
<i>Aedes vexans</i>	Riverside	14	22	32	23
<i>Culiseta inornata</i>	Kern	14	28	75	21

^aPercent of mosquito bodies positive for WNV.

^bPercent of transmission attempts positive for WNV.

infection rates ranged from 41% to 48%. *Ae. vexans* had a moderate infection rate of 32%, whereas *Oc. sierrensis* and *Cx. p. quinquefasciatus* from the Coachella Valley had infection rates <15%. The last two infection rates are significantly lower than the day-14 high-dose infection rates for all species tested, except for *Cx. tarsalis* (Yolo County), *Cx. p. quinquefasciatus* (Orange County), *Oc. dorsalis*, and *Ae. vexans* ($p < 0.0009$). Despite the high susceptibility of *Cx. tarsalis* (Yolo County), its day-14 infection rates are not statistically significant, which may be attributed to the small sample size.

Culex species, excluding *Cx. p. quinquefasciatus* from the Coachella Valley and Orange County, were most efficient at transmitting virus after exposure to the high dose and 14-day incubation period. *Cx. tarsalis* (Yolo County) was the most efficient laboratory vector; 60% of expectorate samples contained virus after only 7 days of incubation. These *Cx. tarsalis* (Yolo County) transmission results were significantly higher than all other day-7 high-dose transmission rates ($p < 0.001$), except for *Cx. tarsalis* (Bakersfield) and *Cx. erythrothorax*

(Coachella Valley). Only one *Cx. tarsalis* (Yolo County) was tested on day 14 because of excessive mortality beginning on day 10. After 14 days of incubation, $\geq 60\%$ of the *Cx. tarsalis* from all three regions in California, *Cx. stigmatosoma* and *Cx. erythrothorax* (Coachella Valley) transmitted virus. *Cx. p. quinquefasciatus* (Bakersfield) followed closely with a 52% transmission rate. *Cx. p. quinquefasciatus* (Orange County), *Cx. p. pipiens*, *Cx. erythrothorax* (Orange County), *Oc. dorsalis*, *Oc. melanimon*, *Ae. vexans*, and *Cs. inornata* had moderate transmission rates ranging from 19% to 36%. *Oc. sierrensis* and *Cx. p. quinquefasciatus* from the Coachella Valley were poor vectors, transmitting virus at rates $\leq 6\%$. Transmission rates for *Cx. p. quinquefasciatus* (Coachella Valley) were significantly lower than those of *Cx. tarsalis* (Coachella Valley, Bakersfield), *Cx. p. quinquefasciatus* (Bakersfield), *Cx. p. pipiens*, *Cx. stigmatosoma*, *Cx. erythrothorax* (Coachella Valley), and *Oc. dorsalis* ($p < 0.0009$). *Oc. sierrensis* transmission rates were significantly lower than the same six species except for *Oc. dorsalis* ($p < 0.0009$).

RESEARCH

Table 3. Infection and transmission rates for California mosquito species orally infected with $10^{4.9\pm 0.1}$ PFU/mL of *West Nile virus* (WNV)

Species	Source by county	Day transmission attempted	No. tested	Infection rate ^a	Transmission rate ^b
<i>Culex tarsalis</i>	Yolo	7	25	8	0
		14	11	36	82
	Kern	7	10	30	10
		14	45	7	0
	Riverside	7	40	13	0
		14	10	0	0
<i>Cx. pipiens quinquefasciatus</i>	Kern	7	50	58	0
		14	50	10	0
	Riverside	7	50	0	0
		14	55	0	0
<i>Cx. p. pipiens</i>	Shasta	7	25	36	0
		14	35	23	60
<i>Cx. stigmatosoma</i>	San Bernardino	14	29	69	34
<i>Cx. erythrothorax</i>	Orange	7	47	15	0
	Riverside	7	12	67	0
		14	20	65	30
<i>Ochlerotatus dorsalis</i>	San Luis Obispo	7	29	3	0
		14	25	4	4
<i>Oc. melanimon</i>	Kern	7	50	0	0
		14	60	3	2
<i>Oc. sierrensis</i>	Lake	7	25	4	0
		14	30	0	0

^aPercent of mosquito bodies positive for WNV.

^bPercent of transmission attempts positive for WNV.

Infection and transmission rates were lower for mosquitoes exposed to $10^{4.9\pm 0.1}$ PFU/mL of WNV (Table 3) than to the higher dose after both 7 and 14 days of incubation. After 7 and 14 days of incubation, *Culex* species had a wide range of infection rates, except for *Cx. tarsalis* (Coachella Valley) on day 14 and *Cx. p. quinquefasciatus* (Coachella Valley) on days 7 and 14, for which infection was not detectable. *Cx. stigmatosoma* infection rates on day 14 for the low dose were significantly higher than all other day-14 low-dose infection rates, except for *Cx. erythrothorax* (Coachella Valley) and *Cx. tarsalis* (Yolo County) ($p < 0.001$). *Cx. erythrothorax* (Coachella Valley) infection rates on day 7 were significantly higher than all day-7 infection rates, except for *Cx. tarsalis* (Bakersfield), *Cx. p. quinquefasciatus* (Bakersfield), and *Cx. p. pipiens* ($p < 0.001$). Infection rates for *Cx. erythrothorax* (Coachella Valley) on day 14 also were significantly higher than all day-14 low-dose infection rates except for *Cx. tarsalis* (Yolo County, Coachella Valley), *Cx. p. pipiens*, and *Cx. stigmatosoma* ($p < 0.001$). Infection rates for *Ochlerotatus* species were $< 5\%$ at 7 and 14 days of incubation.

After imbibing a low dose of virus and undergoing 7 days of incubation, positive transmissions were not detected except for *Cx. tarsalis* (Bakersfield). Transmission rates were highest after 14 days of incubation for *Cx. tarsalis* (Yolo County), *Cx.*

p. pipiens, *Cx. stigmatosoma*, and *Cx. erythrothorax* (Coachella Valley), although transmission rates for *Cx. tarsalis* (Yolo County) and *Cx. p. pipiens* were higher than the infection rates. Their transmission rates were significantly higher than all others ($p < 0.001$). Transmission rates were $\leq 4\%$ for *Oc. dorsalis* and *Oc. melanimon* after 14 days. WNV transmission was not detected for *Cx. tarsalis* (Coachella Valley), *Cx. p. quinquefasciatus* (Bakersfield and Coachella Valley), *Cx. erythrothorax* (Orange County), and *Oc. sierrensis*. *Ae. vexans* and *Cs. inornata* were not tested at the low dose of virus.

Discussion

All 10 California mosquito species were competent laboratory vectors of WNV, although infection rates varied by species, dose, and incubation period. The amount of virus we used for infection was comparable to published natural WNV avian viremias in Egypt (32) but less than reported for North American birds infected with the NY strain of WNV (33). In addition, artificial blood meals with defibrinated blood may be less infectious by ~ 2 logs of virus/mL (34), although recent comparisons among *Cx. tarsalis* (infected with WEEV by feeding on viremic chickens or heparinized viremic chicken blood presented by hanging blood droplets, pledgets, or solutions through a biomembrane) did not show significant differences

in infection rates or titers in infected female mosquitoes (F. Mahmood et al., unpub. data). Regardless, all mosquito species became infected and transmitted WNV at some level.

Cx. tarsalis is one of the most efficient laboratory vectors of WNV tested from North America (10,13–15). This species is abundant in California and much of western North America, where it is involved in the maintenance and amplification of WEEV and SLEV (22). Considering its central role in the transmission of arboviruses in avian hosts and its susceptibility to WNV infection in the laboratory, *Cx. tarsalis* has the greatest potential of the species we studied to amplify and maintain WNV in California.

Mosquitoes in the *Cx. pipiens* species complex also may be an important enzootic mosquito host in California. *Cx. p. pipiens* was identified as a primary WNV vector during the 1999 New York outbreak (10) and has been suggested as a host for overwintering flaviviruses such as WNV and SLEV (35–38). This species could play a similar role in WNV transmission in California. *Cx. p. pipiens* is mainly ornithophilic (39), but *Cx. p. quinquefasciatus* feeds readily on mammals (25,40), potentially transferring WNV from birds to humans and horses.

Cx. p. quinquefasciatus from Coachella Valley and Orange County were significantly less susceptible to infection than those collected from Bakersfield in the southern Central Valley. Differences in infection and transmission rates indicated that geographic differences may exist in the vector competence for WNV of mosquitoes within this species complex, which could relate to the introgression of *Cx. p. pipiens* genes into the Bakersfield population (23). The extent to which differences in infection and transmission are caused by the genetic structure of mosquito populations throughout the state and the impact of these differences on WNV transmission require additional study.

Results for *Cx. tarsalis* (Yolo County) and *Cx. p. pipiens* exposed to the low dose of virus and incubated for 14 days were unexpected. Infection rates for both species were consistent with results for most *Culex* species, but transmission rates were high and exceeded infection rates (i.e., some positive expectorate samples were not associated with positive results for the associated mosquito bodies, even after retesting). These incongruous results may be attributed to experimenter error. Additional replicates of these experiments may be needed to verify our results.

Infection with WNV may have increased death rates in infected female mosquitoes. In most groups, infection rates after 14 days were less than infection rates after 7 days, perhaps indicating that susceptible females died more rapidly than less susceptible or uninfected females. Most noticeable were the synchronous deaths of *Cx. tarsalis* in both the high- and low-dose groups from the highly susceptible Yolo County population after 10 days of incubation.

Cx. stigmatosoma and *Cx. erythrothorax* are widely distributed species in California and were highly susceptible to WNV infection. *Cx. stigmatosoma* preferentially feeds on

birds and may play a role as an enzootic vector. Conversely, *Cx. erythrothorax* behaves as an opportunistic feeder, potentially bridging WNV transmission between birds and mammals (25).

In California, *Oc. dorsalis* and *Oc. melanimon* are involved in the transmission of WEEV among small mammals and are both laboratory-confirmed vectors of WEEV (26,27). Both species have a similar ecology and can be found in fresh water; however, *Oc. dorsalis* also develops in saline and alkaline habitats in coastal and southeastern California, respectively (26,41,42). *Oc. melanimon* plays a secondary role in the maintenance of WEEV in lagomorphs during the late summer in the Central Valley of California (43). WEEV and California encephalitis viruses have been isolated from *Oc. melanimon* (44,45). *Oc. melanimon* is an abundant pest species in the Central Valley that readily bites humans, other mammals, and (occasionally) birds (29,46). With moderate WNV transmission rates and a preference for mammalian hosts, these species have little potential to act as secondary or bridge vectors from birds to mammals.

Oc. sierrensis, a widely distributed tree hole mosquito, is a major pest in California that frequently bites humans and other mammals (28,46). However, arboviruses have not been isolated from this species to date, and its infection and transmission rates for WNV were low in the current study. Mammalian feeding preferences coupled with low vector competence for WNV indicate that this species probably would not be an enzootic or bridge vector of WNV in California.

WNV was isolated from wild *Ae. vexans* collected from the eastern United States during 2001 (11). Arboviruses rarely have been isolated from *Ae. vexans* in California (24), even though this species has been found infected with WEEV during epizootics in the central United States (47) and has been shown capable of laboratory transmission of WEEV (48) and SLEV (49) at high infectious doses. In a single trial during the current study, *Ae. vexans* exhibited moderate infection and transmission rates for WNV. Mammalian feeding preferences (29,45) decrease its potential as an enzootic vector for WNV in California.

Cs. inornata is a widely distributed winter mosquito in California with relatively high infection and moderate transmission rates for WNV. The species is a laboratory-confirmed vector of WEEV and SLEV viruses (49,50) and a primary horizontal and vertical vector of some bunyaviruses (51,52). We tested this species because of its potential to extend the transmission season of WNV in California beyond the November–January diapause of *Cx. tarsalis* (53,54). *Cs. inornata* primarily feeds on livestock and occasionally on birds (46,55,56) and may play a minor role in the amplification and transmission of WNV in California.

Because WNV was recently introduced into North America, little is known about the vector competence of New World mosquitoes for this invading strain of virus. Assessing the vector competence of California mosquitoes provides arbovirus surveillance and mosquito control programs with valuable

information concerning the possible roles of different species in the transmission and maintenance of WNV. Our results indicated that, similar to other parts of the world, mosquitoes in the genus *Culex* are anticipated to be the principal enzootic mosquito hosts of WNV in California. On the basis of their vector competence and host-feeding patterns, *Cx. tarsalis* may be the principal vector in rural agricultural ecosystems; in addition, members of the *Cx. pipiens* complex and perhaps *Cx. stigmatosoma* will be important vectors in urban settings. If WNV becomes established in a *Cx. tarsalis*–passerine transmission cycle, the effect of sharing a common vector on the evolution of two closely related flaviviruses, WNV and SLEV, will be determined. The variation in WNV vector competence and other components of vectorial capacity within single mosquito species will need to be studied. *Cx. erythrothorax* and species in the genera *Ochlerotatus* and *Culiseta* are likely to serve as secondary or bridge vectors. Our results for *Cx. p. quinquefasciatus* collected in different geographic locations, however, indicate that not all mosquitoes in a single taxonomic unit will contribute equally to WNV transmission.

Acknowledgments

We thank Laura D. Kramer for providing the NY flamingo 35211 AAF 9/23/99 strain of WNV; the Sacramento-Yolo Mosquito and Vector Control District for providing *Cx. tarsalis*; Laura Gore for providing the *Cx. p. pipiens*; Hugh Lothrop for providing *Cx. p. quinquefasciatus*, *Cx. tarsalis*, *Cx. erythrothorax*, and *Ae. vexans* from the Coachella Valley; David Woodward for providing *Ochlerotatus sierrensis*; Richard Meyer and Robert Cummings for providing *Cx. erythrothorax* and *Cx. p. quinquefasciatus*; Min-Lee Cheng for providing the *Culex stigmatosoma*; and Mitchell Watnik for helping with the statistical analysis.

This research was supported by a grant from the University of California Mosquito Research Program.

Ms. Goddard is a graduate student in the Entomology Department at the University of California, Davis. Her research focuses on the evolution of *western equine encephalomyelitis virus* (WEEV) as it is serially passaged in different insect vectors and the correlation between phenotypic and genotypic changes in WEEV as it replicates in different vector species.

References

- Centers for Disease Control and Prevention. West Nile virus activity—United States, September 5–11, and Texas, January 1–September 9, 2002. *MMWR Morb Mortal Wkly Rep* 2002;51:812–23.
- Centers for Disease Control and Prevention. Update: West Nile virus activity—eastern United States, 2000. *MMWR Morb Mortal Wkly Rep* 2000;49:1044–7.
- Hayes CG. West Nile fever. In: Monath TP, editor. *The arboviruses: epidemiology and ecology*. Vol 5. Boca Raton (FL): CRC Press; 1989. p. 59–88.
- Smithburn KC, Hughes TP, Burke AW, Paul JH. A neurotropic virus isolated from the blood of a native of Uganda. *Am J Med Hyg* 1940;20:471–92.
- Hubalek Z, Halouzka J. West Nile fever—a reemerging mosquito-borne viral disease in Europe. *Emerg Infect Dis* 1999;5:643–50.
- Petersen LR, Roehrig JT. West Nile virus: a reemerging global pathogen. *Emerg Infect Dis* 2001;7:611–4.
- McIntosh BM, Jupp PG, Dos Santos I, Meenehan GM. Epidemics of West Nile and Sindbis viruses in South Africa with *Culex (Culex) univittatus* Theobald as vector. *S Afr J Sci* 1976;72:295.
- Nir Y, Goldwasser R, Lasowski Y, Margalit J. Isolation of West Nile virus strains from mosquitoes in Israel. *Am J Epidemiol* 1968;87:496–501.
- Akhter R, Hayes CG, Baqar S, Reisen WK. West Nile virus in Pakistan. III. Comparative vector capability of *Culex tritaeniorhynchus* and eight other species of mosquitoes. *Trans Roy Soc Trop Med Hyg* 1982;76:449–53.
- Turell MJ, O'Guinn M, Oliver J. Potential for New York mosquitoes to transmit West Nile virus. *Am J Trop Med Hyg* 2000;62:413–4.
- Centers for Disease Control and Prevention. West Nile virus activity—eastern United States, 2001. *MMWR Morb Mortal Wkly Rep* 2001;50:617–9.
- Centers for Disease Control and Prevention. Weekly Update: West Nile virus activity—northeastern United States, 2000. *MMWR Morb Mortal Wkly Rep* 2000;49:820–2.
- Sardelis MR, Turell MJ, Dohm DJ, O'Guinn ML. Vector competence of selected North American *Culex* and *Coquillettidia* mosquitoes for West Nile virus. *Emerg Infect Dis* 2001;7:1018–22.
- Turell MJ, O'Guinn ML, Dohm DJ, Jones JW. Vector competence of North American mosquitoes (Diptera: Culicidae) for West Nile virus. *J Med Entomol* 2001;38:130–4.
- Sardelis MR, Turell MJ. *Ochlerotatus j. japonicus* in Frederick County, Maryland: discovery, distribution, and vector competence for West Nile virus. *J Am Mosq Control Assoc* 2002 17:137–41.
- Hardy JL, Reeves WC. Experimental studies on infection in vectors. In: Reeves WC, editor. *Epidemiology and control of mosquito-borne arboviruses in California, 1943–1987* Sacramento, CA. Sacramento (CA): California Mosquito Vector Control Association; 1990. p. 145–250.
- Reeves WC, Hammon W McD. Laboratory transmission of Japanese B encephalitis virus by seven species (three genera) of North American mosquitoes. *J Exp Med* 1946;83:185–94.
- Centers for Disease Control and Prevention. Epidemic/epizootic West Nile virus in the United States: revised guidelines for surveillance, prevention, and control. Atlanta: The Centers; 2001.
- Kramer VL, editor. California State mosquito-borne virus surveillance and response plan. 2001. Available from: URL: <http://westnile.ca.gov/CA%20Mosquito%20Response%20Plan%202006-02.doc>
- Hardy JL. Susceptibility and resistance of vector mosquitoes. In: Monath TP, editor. *The arboviruses: epidemiology and ecology*. Vol. 2. Boca Raton (FL): CRC Press; 1988. p. 87–126.
- Woodring JL, Higgs S, Beaty BJ. Natural cycles of vector borne pathogens. In: Marquardt WC, Beaty BJ, editors. *Biology of disease vectors*. Boulder (CO): University Press of Colorado; 1996. p. 51–72.
- Reeves WC, Hammon W McD. *Epidemiology of the arthropod-borne viral encephalitides in Kern County, California 1943–1952*. University of California Publications in Public Health. Vol. 4. Berkeley (CA): University of California Press; 1962. p. 75–108.
- Urbanelli S, Silverstrini F, Reisen WK, De Vito E, Bullini L. California hybrid zone between *Culex pipiens pipiens* and *Cx. p. quinquefasciatus* revisited (Diptera: Culicidae). *J Med Entomol* 1997;34:116–27.
- Milby MM, Reeves WC. Natural infection in arthropod vectors. In: Reeves WC, editor. *Epidemiology and control of mosquito-borne arboviruses in California, 1953–1987*. Sacramento (CA): California Mosquito and Vector Control Association; 1990. p. 128–144.
- Reisen WK, Reeves WC. Bionomics and ecology of *Culex tarsalis* and other potential mosquito vector species. In: Reeves WC, editor. *Epidemiology and control of mosquito-borne arboviruses in California, 1943–1987*. Sacramento (CA): California Mosquito and Vector Control Assoc; 1990. p. 254–329.
- Kramer LD, Reisen WK, Chiles RE. Vector competence of *Aedes dorsalis* (Diptera: Culicidae) from Morro Bay, California for western equine encephalomyelitis virus. *J Med Entomol* 1998;35:1020–4.

27. Hardy JL, Bruen JP. *Aedes melanimon* as a vector of WEE virus in California. *Proc Calif Mosq Cont Assoc* 1974;42:36.
28. Bohart RM, Washino RK. Mosquitoes of California. Berkley (CA): University of California Division of Agricultural Sciences; 1978. p. 153.
29. Gunstream SE, Chew RM, Hagstrum W, Tempelis CH. Feeding patterns of six species of mosquito in arid southeastern California. *Mosq News* 1971;31:97-101.
30. Reisen WK, Meyer RP, Milby MM. Studies on the seasonality of *Culiseta inornata* in Kern County, California. *J Am Mosq Control Assoc* 1989;5:183-95.
31. Aitken TH. An in vitro feeding technique for artificially demonstrating virus transmission by mosquitoes. *Mosq News* 1977;23:130-3.
32. Work TH, Hurlbut HS, Taylor RM. Indigenous wild birds of the Nile Delta as potential West Nile virus circulating reservoirs. *Am J Trop Med Hyg* 1955;4:872-88.
33. Komar N, Davis BS, Bunning M, Hettler D. Experimental infection of wild birds with West Nile virus (New York 1999 strain) [Abstract]. *Am J Trop Med Hyg* 2001;62(suppl): 229-30.
34. Weaver SC, Lorenz LH, Scott TW. Distribution of western equine encephalomyelitis virus in the alimentary tract of *Culex tarsalis* (Diptera: Culicidae) after natural and artificial blood meals. *J Med Entomol* 1993;30:391-7.
35. Nasci RS, Savage HM, White DJ, Miller JR, Cropp CB, Godsey MS, et al. West Nile virus in overwintering *Culex* mosquitoes, New York City, 2000. *Emerg Infect Dis* 2001;7:742-4.
36. Dohm DJ, Sardelis MR, Turell MJ. Experimental transmission of West Nile virus by *Cx. pipiens* (Diptera: Culicidae). *J Med Entomol* 2002;39:640-4.
37. Hardy JL, Rosen L, Reeves WC, Scrivani RP, Presser SB. Experimental transovarial transmission of St. Louis encephalitis virus by *Culex* and *Aedes* mosquitoes. *Am J Trop Med Hyg* 1984;33:166-75.
38. Bailey, CL, Eldridge BF, Hayes DE, Watts DM, Tammariello RF, Dalrymple JM. Isolation of St. Louis encephalitis virus from overwintering *Culex pipiens* mosquitoes. *Science* 1978;129:1346-9.
39. Tempelis CH. Host preferences of mosquitoes. *Proc Calif Mosq Cont Assoc* 1970;38:25-8.
40. Reisen WK, Meyer RP, Tempelis CH, Spoehel JJ. Mosquito abundance and bionomics in residential communities in Orange and Los Angeles Counties, California. *J Med Entomol* 1990;27:356-67.
41. Reisen W K, Hardy JL, Chiles RE, Kramer LD, Martinez VM, Presser SB. Ecology of mosquitoes and lack of arbovirus activity at Morro Bay, San Luis Obispo County, California. *J Am Mosq Control Assoc* 1996;12:679-87.
42. Telford AD. The pasture *Aedes* of central and northern California. Seasonal history. *Ann Entomol Soc Am* 1958;51:360-5.
43. Hardy JL. The ecology of western equine encephalomyelitis virus in the Central Valley of California, 1945-1985. *Am J Trop Med Hyg* 1987;37(suppl):18s-32s.
44. Hammon W McD, Reeves WC, Galindo P. Epidemiologic studies of encephalitis in the San Joaquin Valley of California, 1943, with isolation of viruses from mosquitoes. *Am J Hyg* 1945;42:299-306.
45. Hammon W McD, Reeves WC, Sather GE. California encephalitis virus, a newly described agent. Isolation and attempts to identify the agent. *J Immunol* 1952;69:493-510.
46. Tempelis CH, Washino RK. Host-feeding patterns of *Culex tarsalis* in the Sacramento Valley, California, with notes on other species. *J Med Entomol* 1967;4:315-8.
47. Reisen WK, Monath TP. Western equine encephalomyelitis. In: Monath TP, editor. *The arboviruses: epidemiology and ecology*. Vol 4. Boca Raton (FL): CRC Press; 1988. p. 89-137.
48. Meyer RP, Hardy JL, Presser SB, Reisen WK. Preliminary evaluation of the vector competence of some southern California mosquitoes to western equine encephalomyelitis (WEE) and St. Louis encephalitis (SLE) viruses. *Proc Calif Mosq Vector Control Assoc* 1988;56:42-8.
49. Hammon W McD, Reeves WC. Laboratory transmission of St. Louis encephalitis virus by three genera of mosquitoes. *J Exp Med* 1943;78:241-53.
50. Hammon W McD, Reeves WC. Laboratory transmission of western equine encephalomyelitis virus by mosquitoes of the genera *Culex* and *Culiseta*. *J Exp Med* 1943;78:425-34.
51. Reeves, WC. Jerry Slough. In: Karabatsos N, editor. *International catalogue of arboviruses*. San Antonio (TX): American Society of Tropical Medicine and Hygiene for the Subcommittee on Information Exchange of the American Committee on Arthropod-borne Viruses; 1985. p. 575-513.
52. Schopen S, Laubuda M, Beaty B. Vertical and venereal transmission of California group viruses by *Aedes triseriatus* and *Culiseta inornata* mosquitoes. *Acta Virol* 1991;35:373-82.
53. Reisen WK, Meyer RP, Milby MM. Overwintering studies on *Culex tarsalis* (Diptera: Culicidae) from Kern County, California: temporal changes in female abundance and reproductive status with comparative observations on *C. quinquefasciatus* (Diptera: Culicidae). 1986;79:677-85.
54. Nelson RT. Parity in winter populations of *Culex tarsalis* Coquillett in Kern County, California. *Am J Hyg* 1964;80:242-53.
55. Anderson RA, Gallaway WJ. The host preferences of *Culiseta inornata* in southwestern Manitoba. *J Am Mosq Cont Assoc* 1987;3:219-21.
56. Tempelis CH. Current knowledge of feeding habits of California mosquitoes. *Proc Calif Mosq Cont Assoc* 1964;32:39-42.

Address for correspondence: Laura B. Goddard; Department of Entomology, University of California, Davis, One Shields Ave, Davis, CA 95616, USA; fax: 530-752-1537; e-mail: lgoddard@ucdavis.edu

OPPORTUNITIES FOR PEER REVIEWERS

The editors of *Emerging Infectious Diseases* seek to increase the roster of reviewers for manuscripts submitted by authors all over the world for publication in the journal. If you are interested in reviewing articles on emerging infectious disease topics, please e-mail your name, address, qualifications or curriculum vitae, and areas of expertise to eideditor@cdc.gov

At *Emerging Infectious Diseases*, we always request reviewers' consent before sending manuscripts, limit review requests to three or four per year, and allow 2-4 weeks for completion of reviews. We consider reviewers invaluable in the process of selecting and publishing high-quality scientific articles and acknowledge their contributions in the journal once a year.

Even though it brings no financial compensation, participation in the peer-review process is not without rewards. Manuscript review provides scientists at all stages of their career opportunities for professional growth by familiarizing them with research trends and the latest work in the field of infectious diseases and by improving their own skills for presenting scientific information through constructive criticism of those of their peers. To view the spectrum of articles we publish, information for authors, and our extensive style guide, visit the journal web site at www.cdc.gov/eid.

For more information on participating in the peer-review process of *Emerging Infectious Diseases*, e-mail eideditor@cdc.gov or call the journal office at 404-371-5329.

Instructions for Emerging Infectious Diseases Authors

Research Studies. Articles should be under 3,500 words and should include references, not to exceed 40. Use of subheadings in the main body of the text is recommended. Photographs and illustrations are encouraged. Provide a short abstract (150 words) and a brief biographical sketch of first author—both authors if only two. Report laboratory and epidemiologic results within a public health perspective. Although these reports may be written in the style of traditional research articles, they should explain the value of the research in public health terms and place the findings in a larger perspective (i.e., "Here is what we found, and here is what the findings mean").