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Title: LC/MS/MS Screen for the Presence of Melamine in swine and poultry tissues				
Revision: Original	Effective: (5-07)			
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FERN Method Coordination Committee				

<u>Note</u>: This method has been approved for Emergency Response use and is posted following the FERN guidelines for urgent usage /level 1 validation. This validation level indicates that it meets minimal validation criteria. Emergency Response/Urgent Use methods will be removed from eLEXNET six months after the emergency has ended if there has been no validation data submitted to the Method Coordination Committee for consideration by a Technical Review Committee.

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### A. INTRODUCTION

1. Theory

Melamine is extracted from muscle tissue with a mixture of acetonitrile and water. The extract is cleaned up by liquid/liquid extraction with methylene chloride followed by solid phase extraction (SPE). Melamine is eluted from the SPE cartridge with 5% ammonium hydroxide / methanol, evaporated to dryness, reconstituted with internal standard and 50% acetonitrile / water, and analyzed by LC/MS/MS.

2. Applicability

This method is applicable for screening of melamine in swine and poultry muscle at  $\geq$  50 ppb.

3. Structure



### B. EQUIPMENT

Note: Equivalent equipment may be substituted for the following.

#### 1. Apparatus

- a. Balance analytical, 0.1 mg sensitivity, Model No. A120S, Sartorious.
- b. Balance top loading, 0.01 g sensitivity, PJ3600 Delta Range, Mettler.
- c. Centrifuge with 15 mL tube carriers, centrifuge: Model HN-S, rotor: Cat. No. 809, International Equipment Company.
- d. Centrifuge with 50 mL tube carriers, centrifuge: Model PR-7000, rotor, Cat. No. 276, International Equipment Company.
- e. Centrifuge tubes 50 mL, Pyrex® round-bottom with Teflon lined screw-cap, 29 mm, Cat. No. 21023-401, VWR.
- f. Centrifuge tubes 15 mL, glass, disposable, screw top, Cat. No. 73785, Kimble.
- g. Culture tubes glass, disposable, 15 mL, 16 x 100 mm, Cat. No. 73500 16100,

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Kimble.

- h. Graduated cylinders class A, 25 mL, 100, 500 mL.
- i. Homogenizer Omni 7H, Omni International.
- j. Micropipetters Adjustable, 100 µL to 10 mL, Eppendorf.
- k. Monoject tuberculin syringe, sterile 1 mL (VWR Part No.: MJ8881-501178)
- I. Nitrogen evaporator N-Evap, Model No. 111, Organomation Associates Inc.
- m. Pasteur pipettes glass, disposable, 9 inch, Cat. No. 14672-380, VWR.
- n. Repipetters bottle top dispensers, 0 25 mL and 1 10 mL Dispensettes, Brinkmann.
- o. Shaker horizontal flatbed, two speed, Cat. No. 511105, Eberbach.
- p. Solid Phase Extraction columns Strata X-C, 60 mg, 3mL, Phenomenex.
- q. Spatula stainless steel.
- r. Squirt Bottle Teflon, Cat. No. 8-250, Nalgene.
- s. Syringe 3 mL, disposable, Cat. No. BD301077, Becton Dickinson.
- t. Syringe Filters Millex, HV, 0.45 µm, Cat. No. SLHVR04NL, Millipore.
- u. Test tube racks for 15 and 50 mL tubes.
- v. Vacuum manifold for solid phase extraction, Cat. No. 5-7030, Supelco.
- w. Volumetric flask class A, 50 mL, 1L.
- x. Vortex mixer variable speed, Cat. No. S8223-1, American Scientific Products.
- 2. Instrumentation
  - a. HPLC Waters Alliance 2695.
  - b. Mass Spectrometer Waters Micromass Quattro micro API.
  - c. Column Phenomenex Synergi<sup>™</sup> Polar RP (150 x 4.6mm; 4 µm particle size).

# C. REAGENTS AND SOLUTIONS

Note: Equivalent reagents and solutions may be substituted for the following.

- 1. Reagents
  - a. Acetic Acid MS grade, Cat. No. 49199 Fluka / Sigma Aldrich.
  - b. Acetonitrile (ACN) HPLC grade, Cat. No. 015-4, Burdick & Jackson.
  - c. Ammonium Acetate MS grade, Cat. No. 73594 Fluka / Sigma Aldrich.

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- d. Ammonium Hydroxide (NH<sub>4</sub>OH) 30%, Cat. No. 9721-33, J.T. Baker.
- e. Hydrochloric acid (HCI) Concentrated, Cat. No. 9535-05, J.T. Baker.
- f. Methanol HPLC grade, Cat. No. 230-4, Burdick & Jackson.
- g. Methylene Chloride HPLC grade, Cat. No. D151-4, Fisher.
- h. 18 M $\Omega$  deionized (DI) H<sub>2</sub>O from Millipore Rx system.

### 2. Solutions

a. ACN:H<sub>2</sub>O (1:1, v/v):

Mix 500 mL of ACN with 500 mL of DI water.

b. Rinse Solution:

Fill a jug with about half ACN and half water. No measurement required.

c. 1N HCI:

Measure 83 mL of concentrated HCI using a 100 mL graduated cylinder. Add to a 1L volumetric flask. Bring to volume with DI water.

d. 0.1N HCI:

Pipette 8.33 mL of concentrated HCl into a 1L volumetric flask. Bring to volume with DI water.

e. 5% Ammonium hydroxide in methanol:

Using class A graduated cylinders, measure 25 mL ammonium hydroxide and 475 mL methanol. Combine in a glass bottle.

f. Mobile Phase A (25 mM ammonium acetate / 25 mM acetic acid in water):

Weigh 1.93 g of ammonium acetate and add to a 1L volumetric flask. Add 1430  $\mu$ L of acetic acid. Bring to 1L with DI water.

### D. STANDARDS

Note: Equivalent standards and solutions may be substituted for the following.

- 1. Source
  - a. Melamine Cat. No. M2659, Aldrich.
  - b. <sup>15</sup>N<sub>3</sub> Melamine (for internal standard) Cat. No. IN 5451, ICON Services. Cat. No. 592889, Aldrich. (no longer available from either source 5/17/07)
- 2. Preparation
  - a. Melamine stock standard (100  $\mu$ g/mL):

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Weigh 5 mg of melamine into a 50 mL volumetric flask. Fill flask about half way with 50% ACN:H<sub>2</sub>O. Sonicate to dissolve. Bring to volume with ACN:H<sub>2</sub>O (1:1, v/v).

b. Melamine working standard (1  $\mu$ g/mL):

Pipette 500  $\mu$ L of the melamine stock solution into a 50 mL volumetric flask. Bring to volume with ACN:H<sub>2</sub>O (1:1, v/v).

c.  ${}^{15}N_3$  Melamine internal standard (IS) (1  $\mu$ g/mL):

Prepare 1  $\mu$ g/mL solution of <sup>15</sup>N<sub>3</sub> Melamine in ACN:H<sub>2</sub>O (1:1, v/v).

d. Calibration curve standards:

Concentration Tissue equivalent		Volume melamine working std. (1 μg/mL)	Volume IS (1 μg/mL)	Volume ACN:H <sub>2</sub> O (1:1, v/v)
0 ng/mL	0 ppb	0 μL	500 μL	500 μL
125 ng/mL	25 ppb	125 μL	500 μL	375 μL
250 ng/mL 50 ppb		250 μL	500 μL	250 μL
500 ng/mL 100 ppb		500 μL	500 μL	0 μL

# 3. Storage and Stability

Store refrigerated at 2 - 8 °C. Stability has not been tested.

# E. SAMPLE PREPARATION

After removing excessive fat from muscle sample, cut it into smaller pieces and homogenize with a mechanical food processor. Transfer homogenized sample into plastic bags and store in a freezer at  $\leq$  -20°C. Let the sample partially thaw prior to analysis.

# F. ANALYTICAL PROCEDURE

Note: avoid plastic - it may leach melamine and contaminate samples.

Melamine is very sticky. Rinse all re-usable glassware with acetonitrile/water rinse solution before use.

- 1. Meat Sample Preparation
  - a. Weigh  $5.00 \pm 0.05$  g of sample into a 50 mL glass centrifuge tube. Also weigh two known blank tissues for use as positive and negative controls.





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- b. Fortify a blank tissue with 250  $\mu$ L of 1 $\mu$ g/mL melamine working standard for use as a 50 ppb positive control.
- c. Add 25 mL of ACN: $H_2O$  (1:1, v/v).
- d. Tissuemize to homogenize at medium to high speed for 30 seconds to 1 minute.

Between each sample, clean the homogenizer probe 5 times with rinse solution. Fill a 50 mL glass tube with sufficient volume to clean a probe (~ 40 - 45 mL). Put the probe in the tube and tissuemize on high speed for about 5 seconds. Repeat 4 more times, each time with a fresh volume of rinse solution.

- e. Centrifuge at 3500 g for 10 minutes, or sufficient to pack tissue.
- f. Remove 5 mL aliquot of the extract into a glass 50 mL glass centrifuge tube.
- g. Add 110 µL of 1 N HCl and vortex to mix.
- h. Add 10 mL of methylene chloride.
- i. Shake for 2 minutes.
- j. Centrifuge at 3500 g for 10 minutes, or sufficient to separate layers.
- k. Remove the aqueous layer (top) into a disposable, screw cap, glass centrifuge tube.
- I. Add 2.5 mL of water to the original 50 mL tube.
- m. Shake for 2 minutes.
- n. Centrifuge at 3500 g for 10 minutes, or sufficient to separate layers.
- o. Carefully remove the top layer; combine aqueous layers.
- p. Cap and vortex for 10 seconds.
- q. Centrifuge at 3500 g for 10 minutes if necessary.
- r. Thoroughly clean SPE manifold and fittings with rinse solution.
- s. Pre-wash the SPE column with 5 mL of methanol followed by 5 mL of water (vacuum optional).
- t. Load the sample onto the column (gravity)
- u. Wash the column with 2 mL of 0.1N HCl, followed by 1 mL of methanol, discard washes. (gravity)
- v. Aspirate the column for 1 minute. (vacuum)
- w. Elute melamine with 5 mL of 5% ammonium hydroxide/methanol into a clean, glass, disposable tube. (gravity)
- x. Thoroughly clean the N-evap tips by spraying with rinse solution from a Teflon squirt bottle and collecting the rinse in a beaker for proper disposal.





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- y. Evaporate the eluant to dryness using an N-evap set to 50 60 °C.
- z. Add 100  $\mu$ L of ACN:H<sub>2</sub>O (1:1, v/v), and 100  $\mu$ L of IS (<sup>15</sup>N<sub>3</sub> Melamine, 1  $\mu$ g/mL).
- aa. Vortex the sample and sonicate for at least one minute to re-dissolve the residue.
- bb. Filter through a 0.45 µm HV filter into an autosampler vial. Transfer the filtered extract into a vial insert using a glass Pasteur pipette. Put the insert into the original vial, cap, and submit for LC-MS/MS analysis.
- 2. Instrumental Settings

Note: The following instrument parameters may be optimized.

- a. HPLC Conditions
  - i. Mobile Phase A: 25 mM Ammonium Acetate / 25 mM Acetic Acid in H<sub>2</sub>O

25°C

- ii. Mobile Phase B: Acetonitrile
- iii. Mobile Phase C: Methanol
- iv. Flow Rate: 0.5 mL/min
- v. Gradient:

Time (min)	%A	%В	%C
0.0	80	15	5
5.0	30	65	5
5.1	5	90	5
7.0	5	90	5
7.1	80	15	5
12.0	80	15	5
vi. Injection	Volume:	50 µL	

- vii. Column Temperature:
- b. Interface Conditions:

i.	Polarity:	Positive
ii.	Ion Source:	Electro Spray
iii.	Source Temperature :	125
iv.	Desolvatiion Temperature :	450
۷.	Cone Gas Flow :	20 L/hour





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	vi.	Desol	lvation Gas Flow :	900 L/hour		
С.	Multi	ple Rea	action Monitoring (MR	M) Parameters	:	
	i.	Reso	lution Q1 & Q3	Unit		
	ii.	Cone Voltage :		35V		
	iii.	Dwell Time :		0.2 sec		
	iv.	Melar	nine Transitions:			
		(a)	m/z 127→ 85	27eV collisio	on voltage	(quant. lon)
		(b)	m/z 127→ 68	17eV collisio	on voltage	
	V.	<sup>15</sup> N <sub>3</sub> N	Aelamine Transition:			
(a) m/z 130→ 87		m/z 130→ 87	17eV collisio	on voltage		

- 3. Sample Chromatograms Examples of typical chromatograms found with this method are shown below.
  - a. 50 ppb External Standard



#### b. Blank Poultry Muscle



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### c. 50 ppb Poultry Muscle Recovery



### G. SCREENING CRITERIA

- 1. The retention time must match that of an external standard within 5%.
- 2. The melamine product ion abundance ratios must match that of an external standard within 20% relative.

Example calculation:

Melamine ion ratio = (area of m/z 127 to 85 peak) ÷ (area of m/z 127 to 68 peak)

Note: Most mass spectrometers calculate the reciprocal of the above and state it as a percentage, which is equally valid.

**Passing ion ratio of a sample =** ion ratio of a standard  $\pm$  (0.2 \* ion ratio of a standard)

- 3. The solvent blank, ACN:H<sub>2</sub>O (1:1, v/v), injected after the standard curve must be negative for melamine (no peak within the retention time window with passing ion ratio), demonstrating no carryover.
- 4. The positive tissue control must be positive for melamine (meeting retention time and ion ratio criteria).
- 5. The negative tissue control must not contain melamine at levels higher than 10% of the positive tissue control. Melamine peaks meeting the retention time and ion ratio criteria must be compared to the positive tissue control.

The Internal standard is used, the melamine quantification peak area (m/z 127 to 85) divided by the internal standard peak area (m/z 130 to 87) must be less than or equal to



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10% of that of the positive tissue control.

6. For a sample to be screen positive, it must be found to contain melamine at or above the 50ppb minimum proficiency level of this analytical method.

The internal standard is used, the melamine quantification peak area (m/z 127 to 85) divided by the internal standard peak area (m/z 130 to 87) must be greater than or equal to that of the positive tissue control.

# H. SAFETY INFORMATION AND PRECAUTIONS

1. Required Personal Protective Equipment (PPE) - safety glasses, disposable gloves, lab coats.

. .

2. Hazards

3.

Reagents	Hazard	Recommended Safe Procedures
Acetonitrile, Methanol	Flammable, poisonous; inhalation will cause headache, fatigue, nausea	Wear gloves and work in the hood. Use protective eyewear. Avoid contact with skin, eyes.
Methylene Chloride	Flammable, poisonous, carcinogenic. Inhalation may cause dizziness, sleepiness, nausea, and respiratory problems. In large dosages can cause unconsciousness or death.	Wear <b>neoprene</b> gloves when pouring or working with large volumes. Methylene chloride will eat through nitrile gloves! Work in the hood. Use protective eyewear. Avoid contact with skin, eyes.
Hydrochloric acid, Ammonium hydroxide	Corrosive, burns	Wear PPE, avoid skin contact, work in a fume hood.
Disposal Procedures		
Procedure Step	Hazard	Recommended Safe Procedures
Acetonitrile, Methanol	Flammable, poisonous; inhalation will cause headache, fatigue, nausea	Store waste in a tightly sealed container away from non- compatibles in a cool, well ventilated, flammable liquid storage area/cabinet for disposal in accordance with local, state and federal regulations.





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Methylene Chloride	Flammable, poisonous, carcinogenic. Inhalation may cause dizziness, sleepiness, nausea, and respiratory problems. In large dosages can cause unconsciousness or death.	Store waste in a tightly sealed container away from non- compatibles in a cool, well ventilated, flammable liquid storage area/cabinet for disposal in accordance with local, state and federal regulations.
Hydrochloric acid, Ammonium hydroxide	Corrosive, burns	Collect waste and store in a tightly sealed container. Store away from non-compatibles in a cool, well ventilated, acid liquid storage area/cabinet for disposal in accordance with local, state, and federal regulations, or neutralize and dispose in accordance with local, state and federal regulations.

### I. QUALITY ASSURANCE PLAN

1. Performance Standard

Positive control is positive for melamine. Negative control is negative for melamine.

2. Critical Control Points and Specifications

Record Acceptable Control

- a. Weight of muscle sample  $5.00 \pm 0.05$  g
- 3. Readiness To Perform
  - a. Familiarization
    - i. Phase I: Standards Duplicate standard curve (D.2.d.) on each of 3 days, which will include the following:
      - (a) 0 ppb
      - (b) 25 ppb
      - (c) 50 ppb
      - (d) 100 ppb
    - ii. Phase II: Analyst Fortified samples 3 replicates at 0ppb and 3 replicates Issuing Authority: Food Emergency Response Network (FERN)

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at 50 ppb over a period of 3 different days.

NOTE: Phase I and Phase II may be performed concurrently.

- iii. Phase III: Check samples for analyst accreditation.
  - (a) 8 blind check samples: at least one of which must be negative, others shall be fortified at 50 ppb.
  - (b) Report analytical findings to supervisor / Quality Assurance Manager (QAM).
  - (c) Letter from QAM is required to commence official analysis.
- b. Acceptability criteria.

Refer to I. 1.

- 4. Intralaboratory Check Samples
  - a. System, minimum contents.
    - i. Frequency: One per week per analyst when samples are analyzed.
    - ii. Records are to be maintained for review.
  - b. Acceptability criteria.
    - Refer to I. 1.

If unacceptable values are obtained, then:

- i. Stop all official analyses by that analyst.
- ii. Take corrective action.
- 5. Sample Acceptability and Stability
  - a. Matrix: swine and poultry muscle
  - b. Sample size: approximately 500 g
  - c. Condition upon receipt: cold, not spoiled
  - d. Sample storage:
    - i. Time: 2 months
    - ii. Condition: frozen < -10°C
- 6. Sample Set
  - a. Negative control (tissue blank).
  - b. Positive control (tissue blank fortified with 50 ppb melamine).
  - c. Test samples to be analyzed.

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# 7. Sensitivity

a. Minimum proficiency level (MPL): 50 ppb.

# J. WORKSHEET

Following worksheet is an example.





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Start Date	
End Date	
Notebook	
Data Folder	

Analyst	
Peer Review	
Supervisor	

Standards	ID
0 x	
1/2 x	
Х	
2x	
fortification	
internal standard	

Reagents	ID	DISP/MIPP	Volume
ACN:H2O 1:1 v/v			
1N HCI			
methylene chloride			
Water	NA		
methanol			
0.1 N HCI			
5% NH₄OH/methanol			
aq. mobile phase		NA	NA
ACN mobile phase		NA	NA
MeOH mobile phase		NA	NA

Equipment	ID
balance	
tissuemizer	
shaker	
centrifuge	
micropipette	
N-evap	
LC/MS/MS	

### Comments

Sample	Tissue	Weight	Ion Ratio	RT	Ppb	Spike	Recovery
		4.95 - 5.05 g		min.			
1							
2							
3							
4							
5							
6							
7							
8							
9							
10							
11							
12							

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### K. APPENDIX

Reference: LC-MS/MS Method for the Analysis of Melamine in Porcine Meat Tissue (Not for Publication), California Animal Health and Food Safety Laboratory System, University of California, Davis, April 30, 2007.

### L. Approvals and Authorities