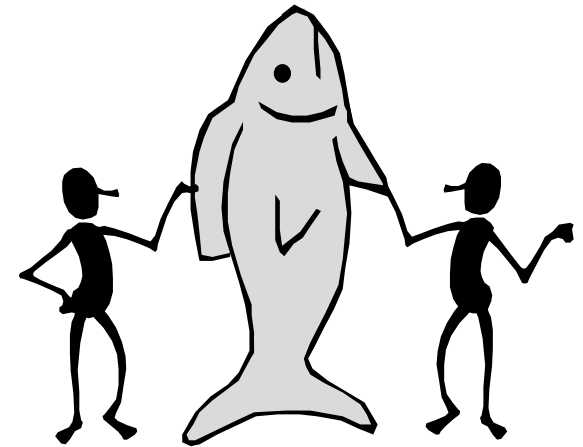





Coeur d'Alene, Idaho
31 March – 4 April, 2003

Section 4e: Methods for Sampling Fish in Large Rivers

Presented by
Joseph E. Flotemersch, USEPA,
Office of Research & Development



A photograph of a person holding a large, light-colored catfish. The fish's mouth is wide open, and a speech bubble is superimposed on the image, containing the text "I'm healthy, I'm healthy, let me go...". The person is wearing a dark blue t-shirt and a black harness. The background is a blurred outdoor setting with green foliage.

I'm healthy,
I'm healthy,
let me go...

Fish are a widely identifiable

component of aquatic systems

Many are valued for their recreational uses

Most species, however, are obscure

And comprise the second most endangered group of animals



Characteristics of Vertebrates (e.g., Fish) that make them useful indicators

- 1) **Accurate environmental assessment of health**
- 2) **Visibility**
- 3) **Standardized use and interpretation**
- 4) **Extensively used in large river programs around the world**
- 5) **Long history of development and use in assessment; thus a strong body of literature from which to draw**
- 6) **Historical knowledge of distribution**

Ref: Simon 1999

Fish (Vertebrates)

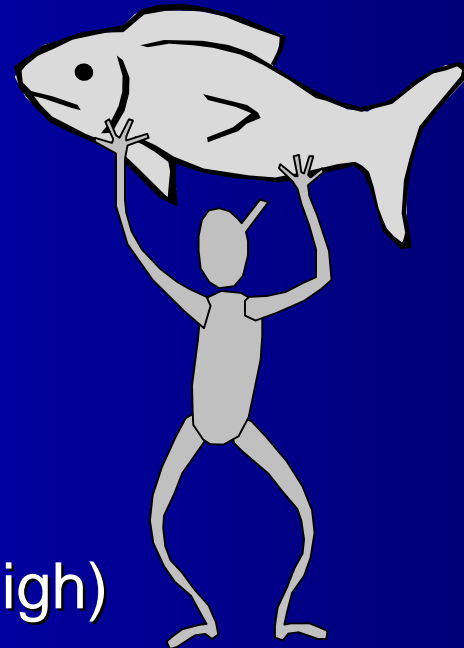
Important program development questions

- **Which sub-habitats**
- **What reach length**
- **What time of day**
- **Which methods (single vs. multiple gear)**
- **Field identification (knowing what to take back to the lab)**
- **What is the final indicator**

Fish (Vertebrates)

Common Sampling Approaches

- **Active sampling methods**
 - Electrofishing
 - Seining
- **Passive sampling methods**
 - Nets (hoop, fyke, gill, trap, etc.)
 - Specific applications
 - Electrofishing prohibited
 - Target Species
 - Prohibitive conductivity (low and high)



Fish (Vertebrates)

Active Sampling Methods



Electrofishing – Widely considered the most comprehensive and effective *single* method for collecting river fishes

Electrofishing Examples

Wisconsin



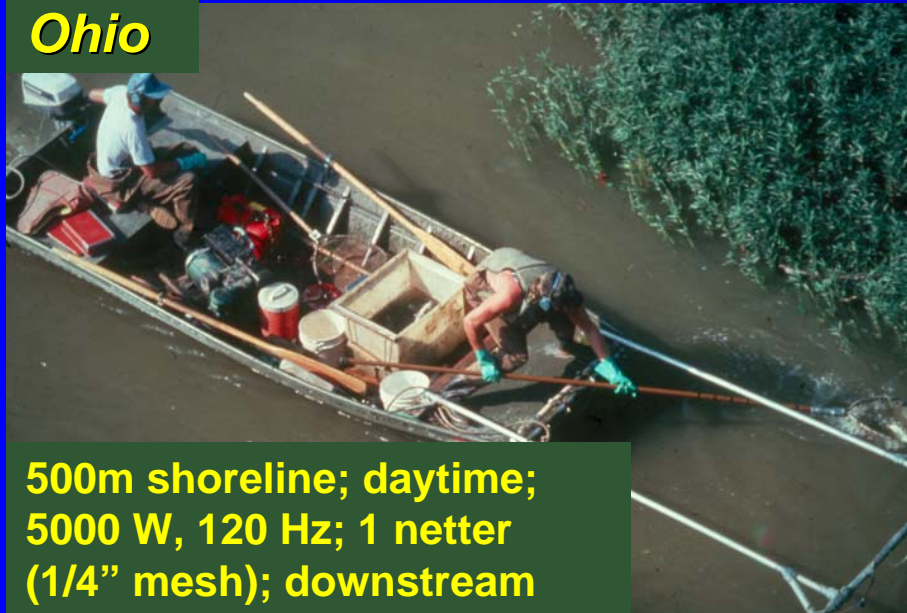
1 mile shoreline; daytime;
3000 W, 60 Hz; 1 netter (17
mm mesh); downstream

EPA – EMAP (Western Rivers)



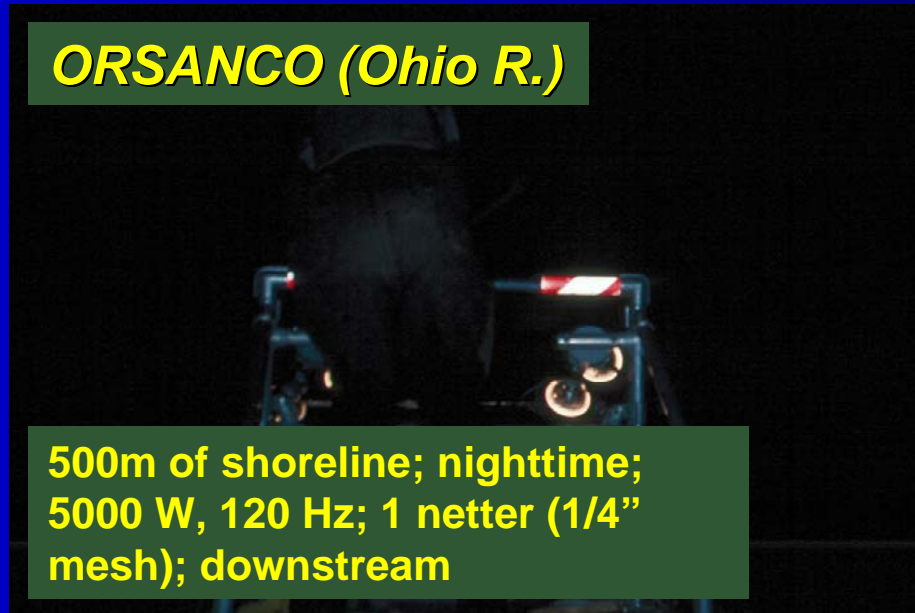
80X width shoreline; daytime;
2500 W, 120 Hz; 1 netter (1/4"
mesh); downstream

Ohio



500m shoreline; daytime;
5000 W, 120 Hz; 1 netter
(1/4" mesh); downstream

ORSANCO (Ohio R.)



500m of shoreline; nighttime;
5000 W, 120 Hz; 1 netter (1/4"
mesh); downstream

May require an array of equipment to cover all encountered systems.



Human factors

influencing electrofishing performance

✓ Equipment

✓ Configuration

- ✓ Boat size
- ✓ Electrode array
- ✓ Setting
- ✓ Equipment condition

✓ Crew experience

- ✓ Especially crew leader
- ✓ Skill of boat driver
- ✓ Historical focus

✓ Physical skill and capacity

✓ Attention to detail

✓ Skill in fish identification

✓ Training

Environmental factors influencing electrofishing performance

- ✓ Recent weather patterns
- ✓ Time of day
- ✓ Wind

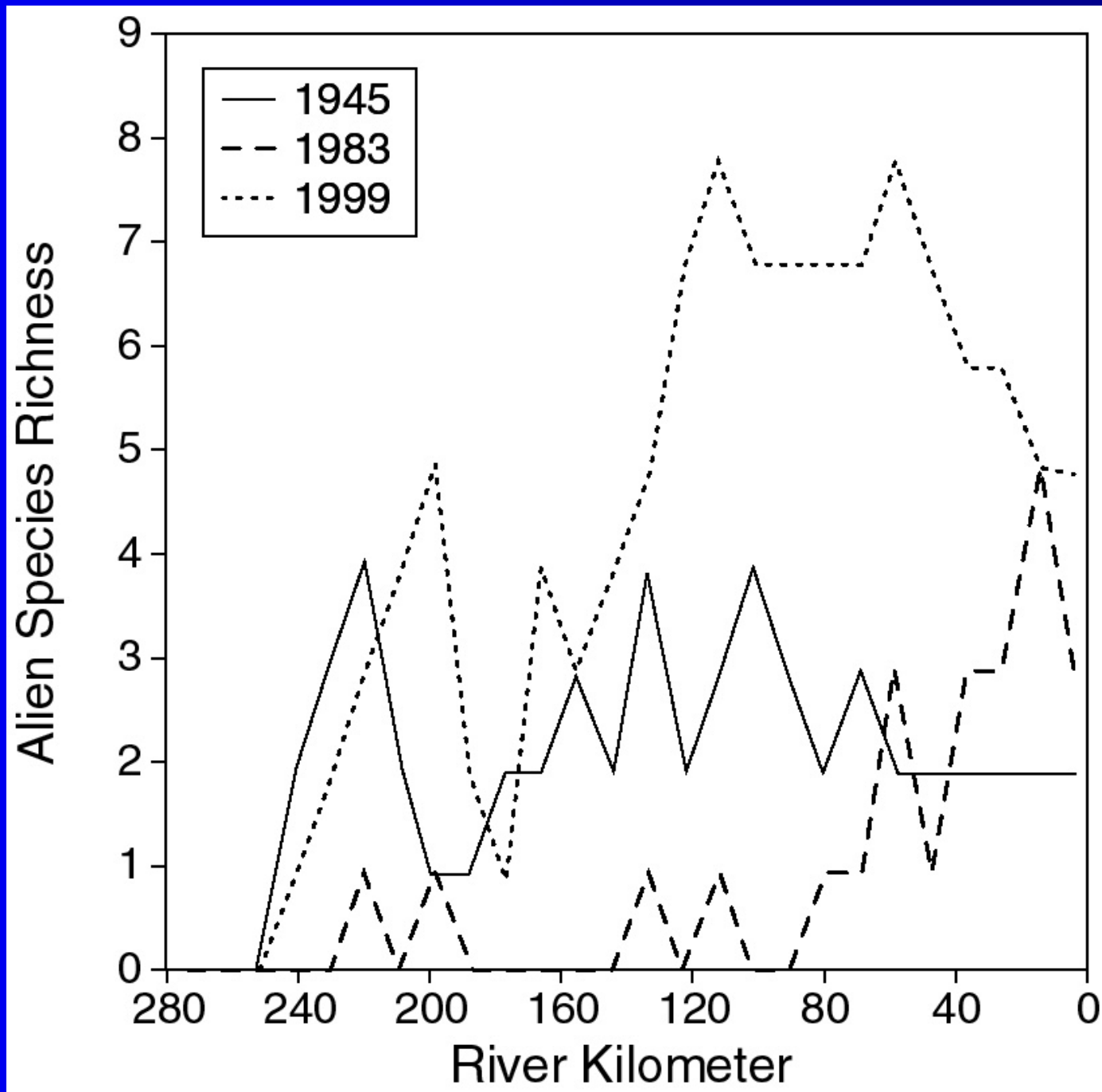
- ✓ Departures from normal summer (low flow) water conditions
 - ✓ Flow rate
 - ✓ Water level
 - ✓ Conductivity
 - ✓ Clarity of water

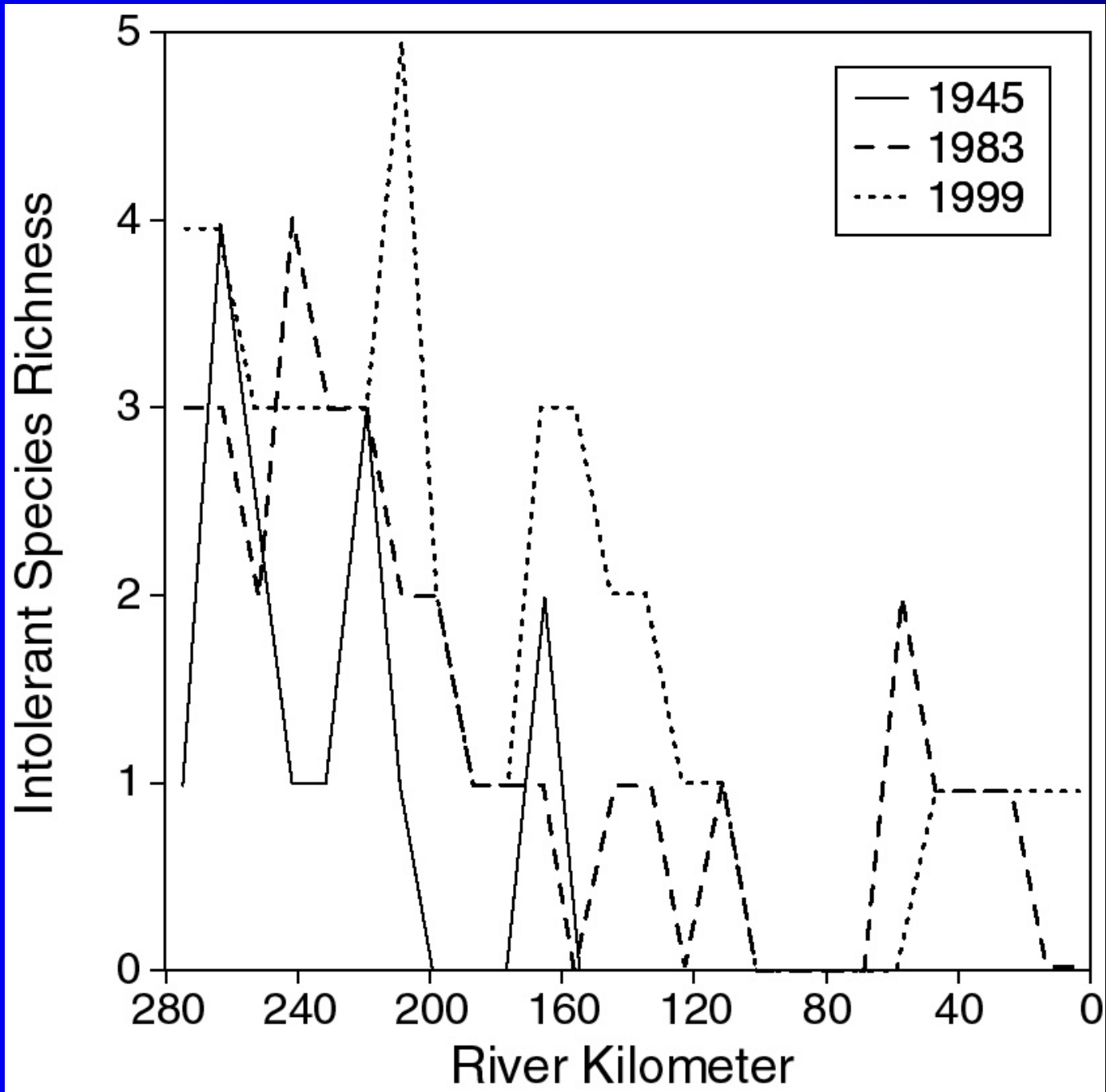
Recent Electrofishing Sample Design Research

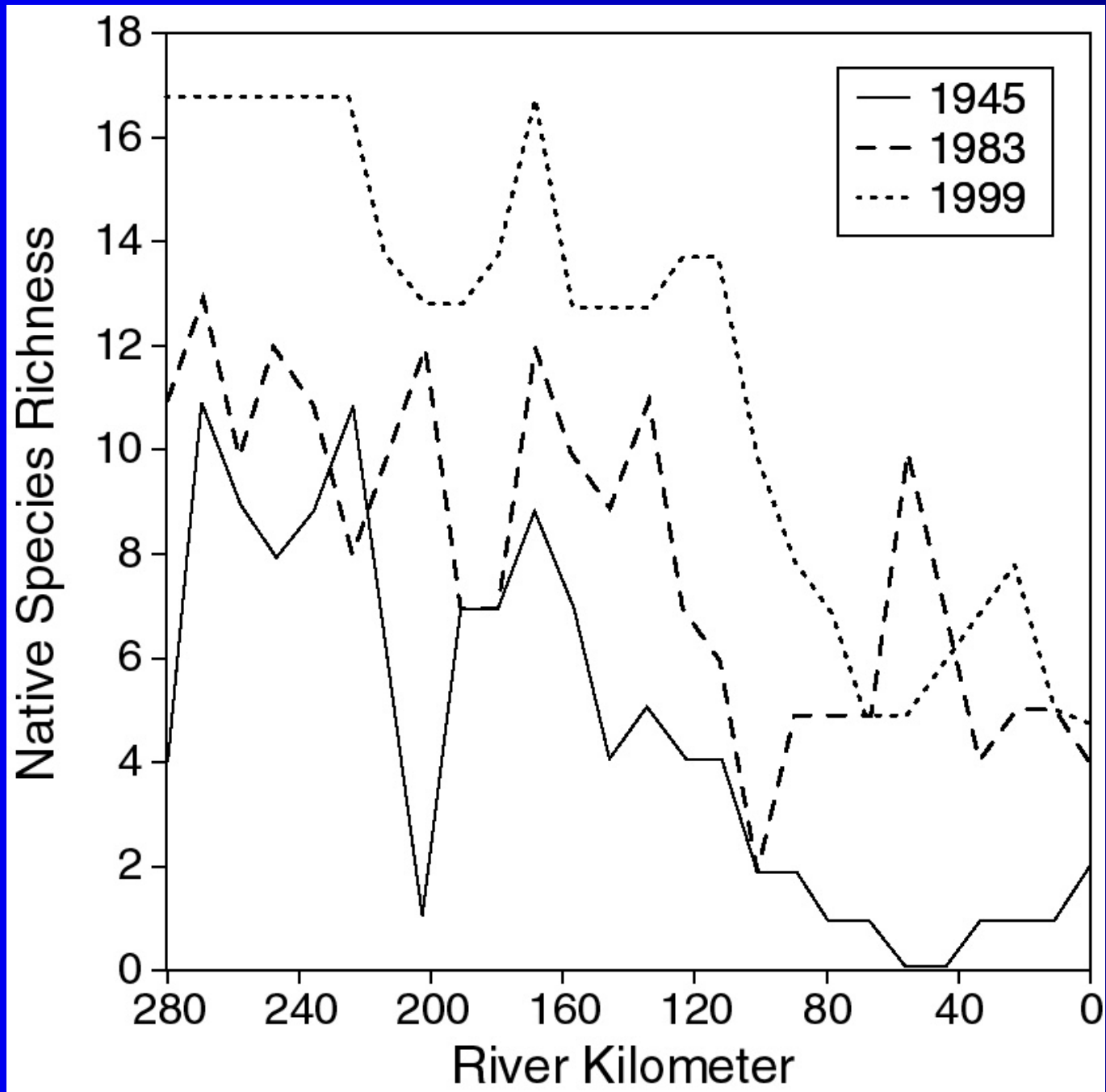
Western Rivers

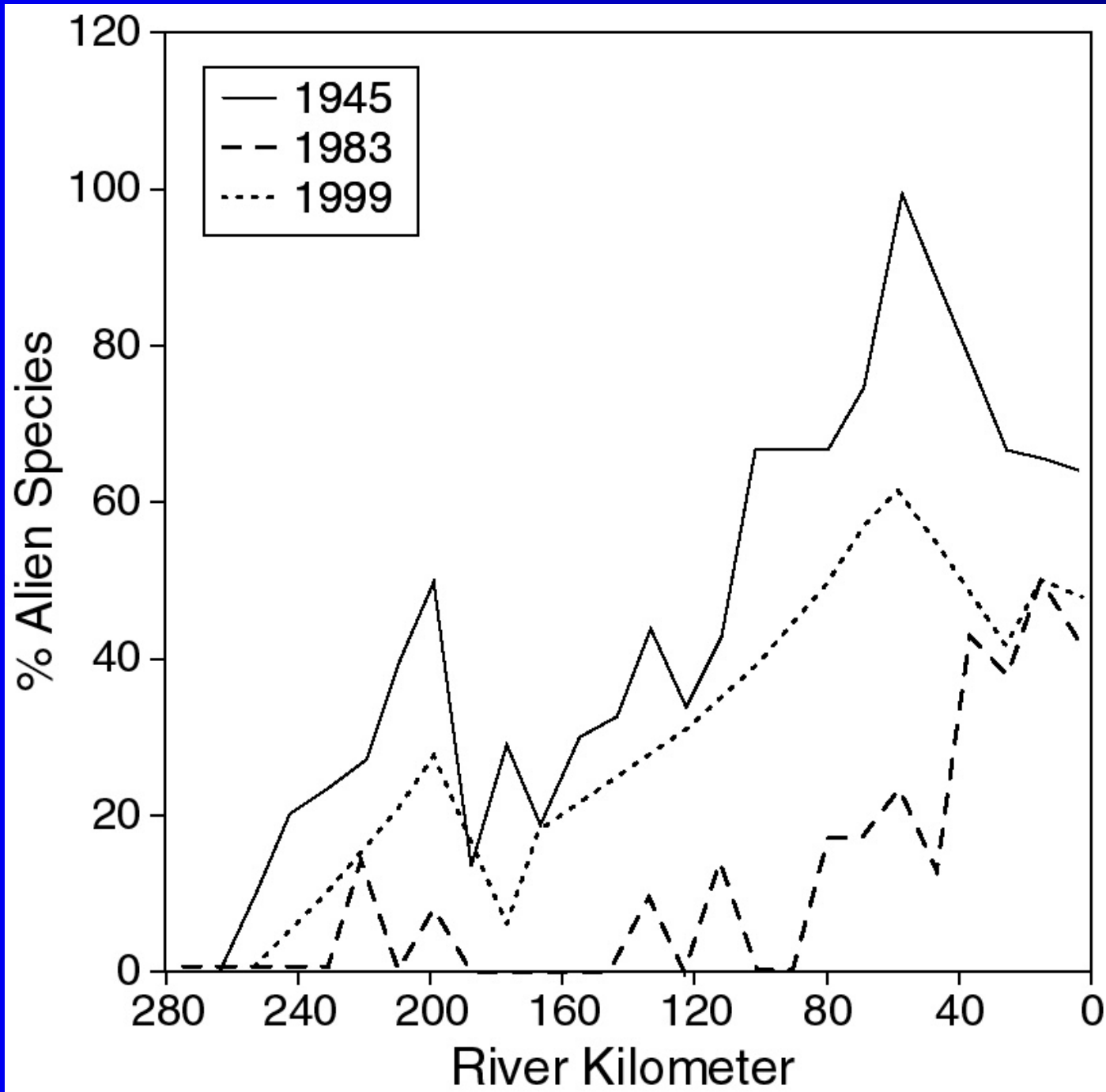


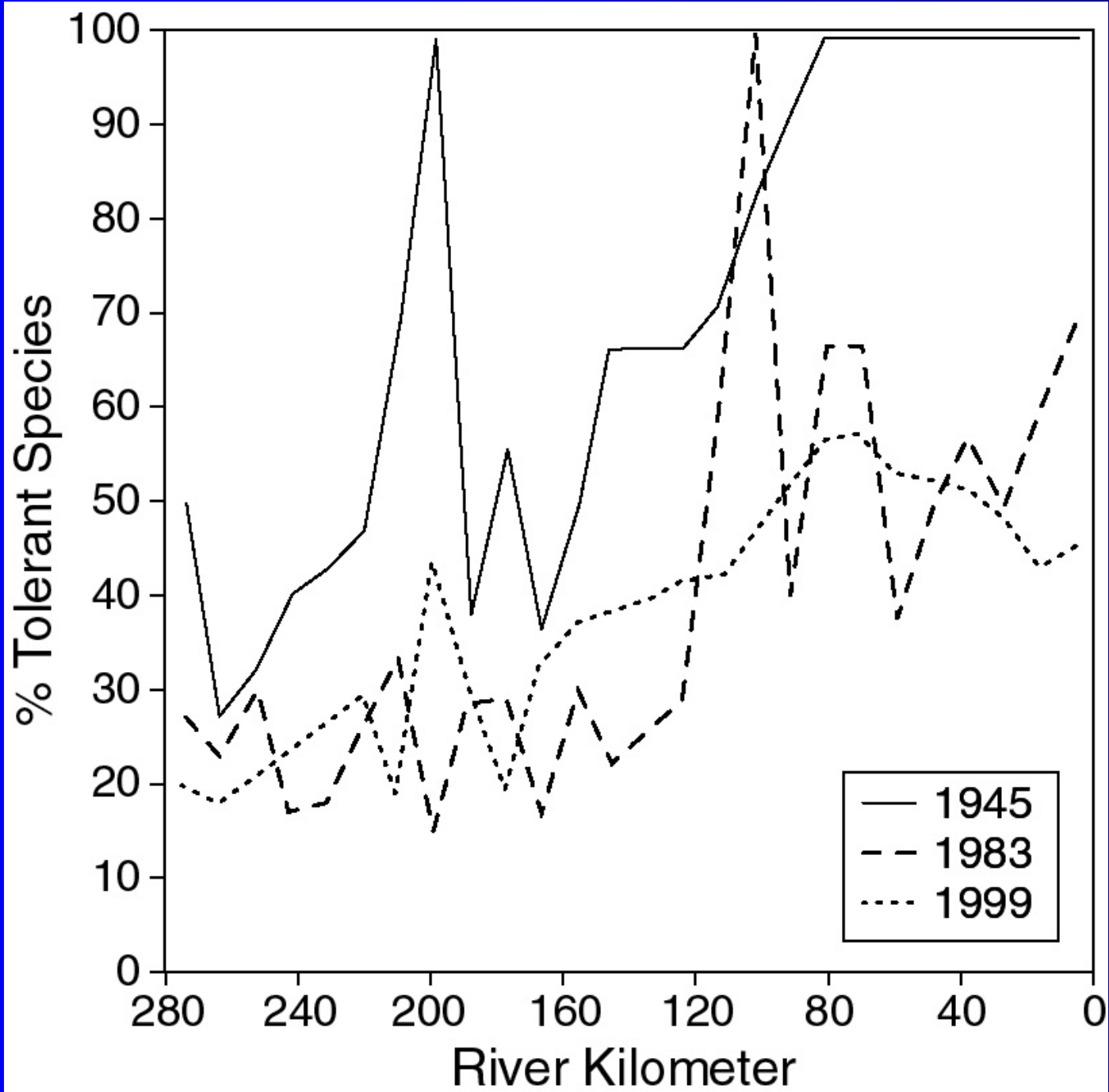
Phil Kaufmann, USEPA, Corvallis, OR.
Bob Hughes, Dynamac, Corvallis, OR.

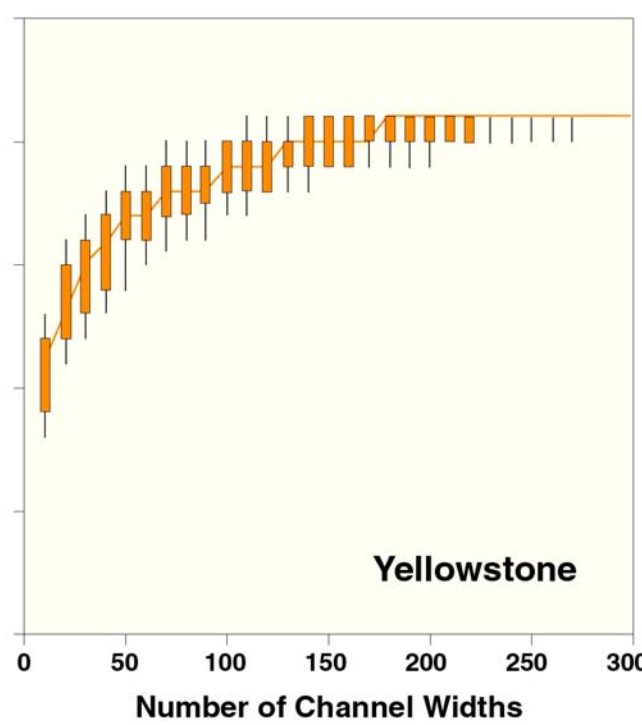
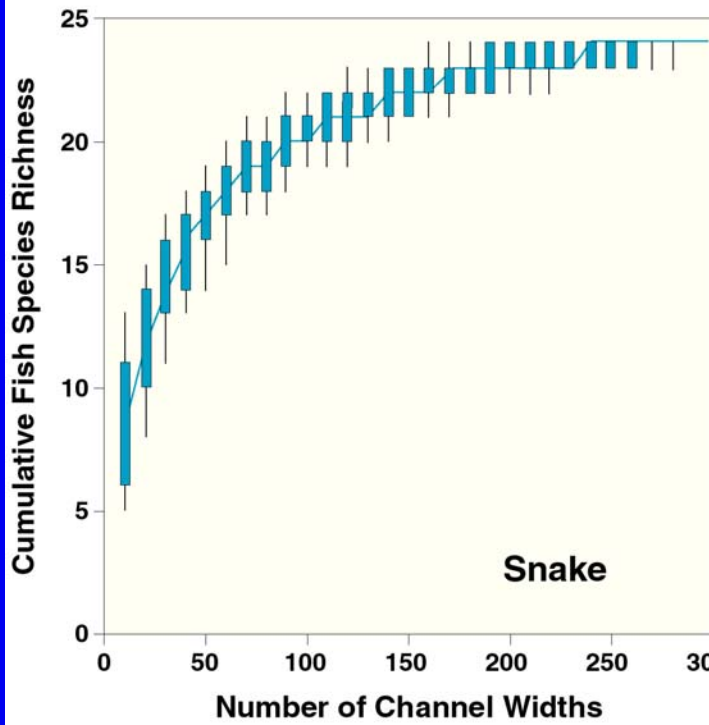
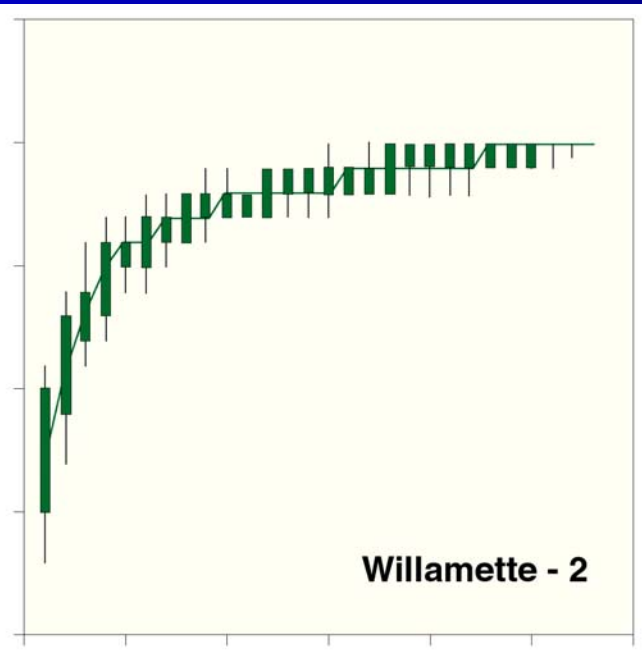
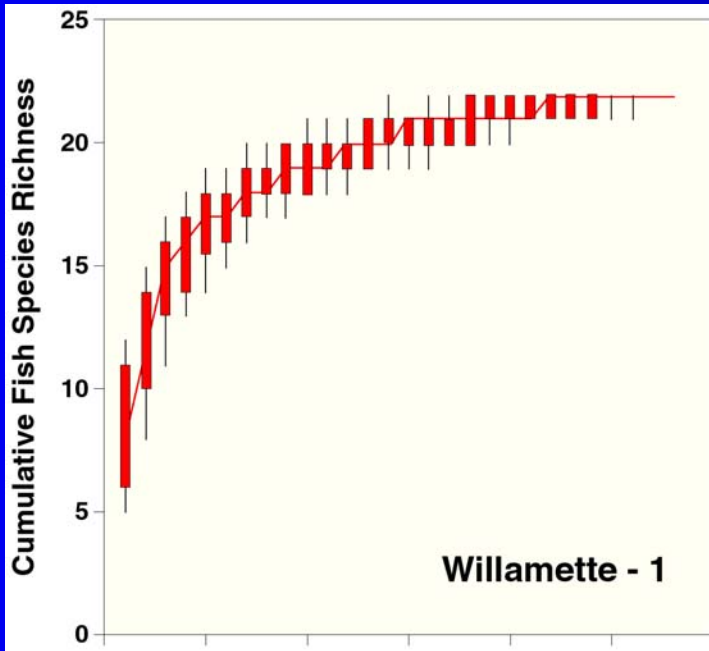












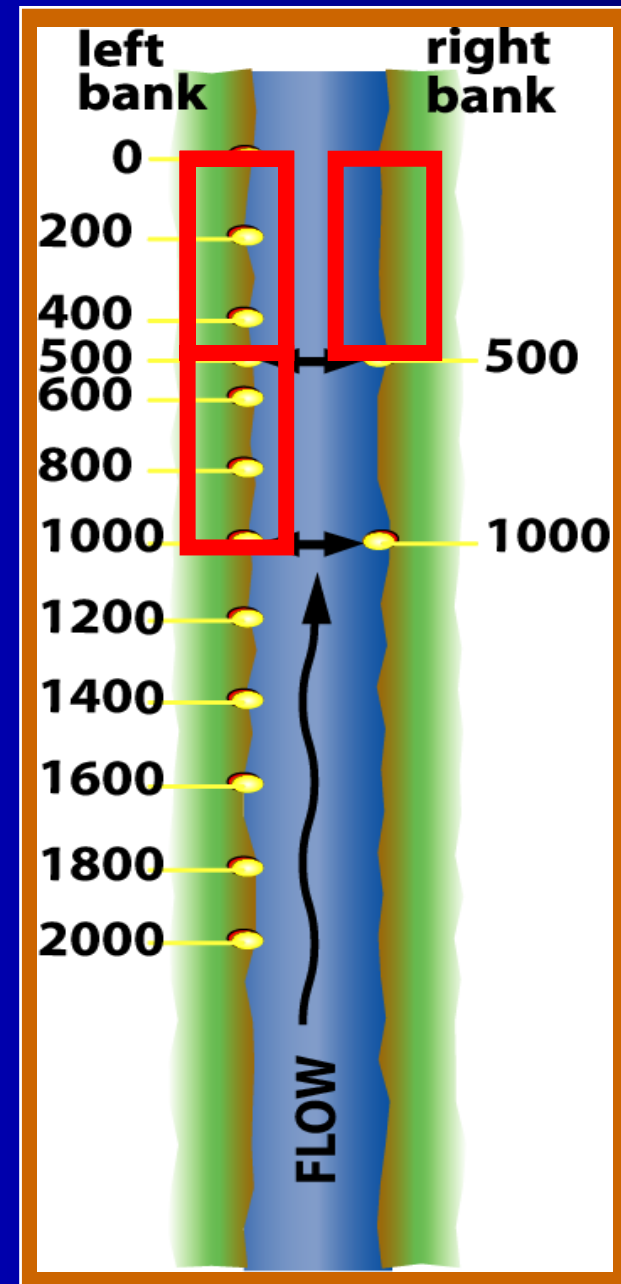
	Willamette-1	Willamette-2	Snake	Yellowstone
Species Observed	22	20	24	21
Number of Individuals	470	445	580	564
No. Species Occurring Once	2	2	2	2
No. Species Occurring Twice	2	2	2	1
True Species Richness (TSR)	23	23	25	22
Channel-widths for 80% TSR	92	77	105	79
Channel-widths for 90% TSR	164	138	182	166
Channel-widths for 95% TSR	220	186	240	240
Channel-widths for 100% TSR	294	250	316	348

Recent Electrofishing Sample Design Research

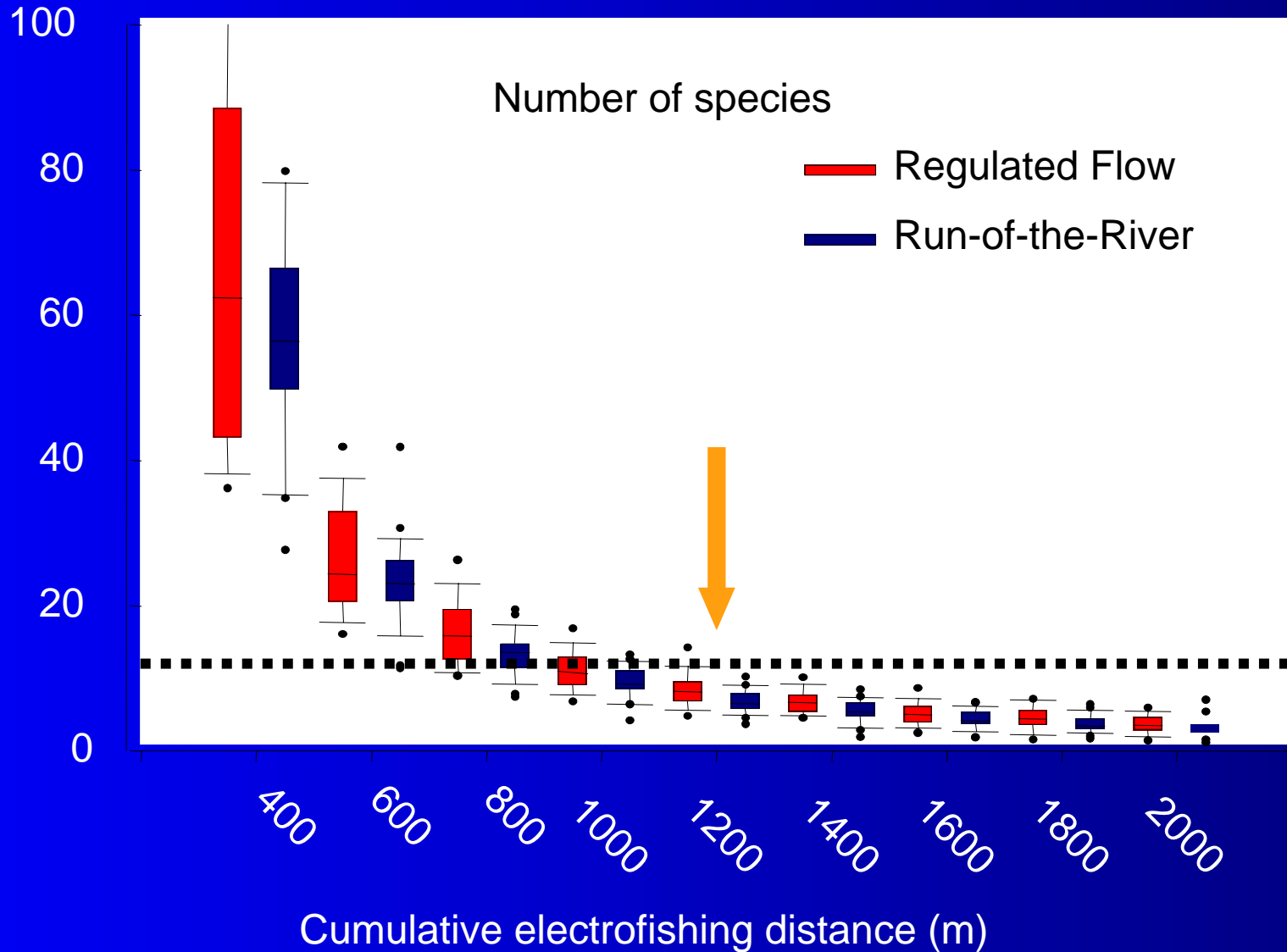
Field Sampling Methods Comparison Notes
(East-Central Rivers)

Joseph E. Flotemersch and Karen A. Blocksom,
USEPA, Office of Research & Development,
Cincinnati, OH.

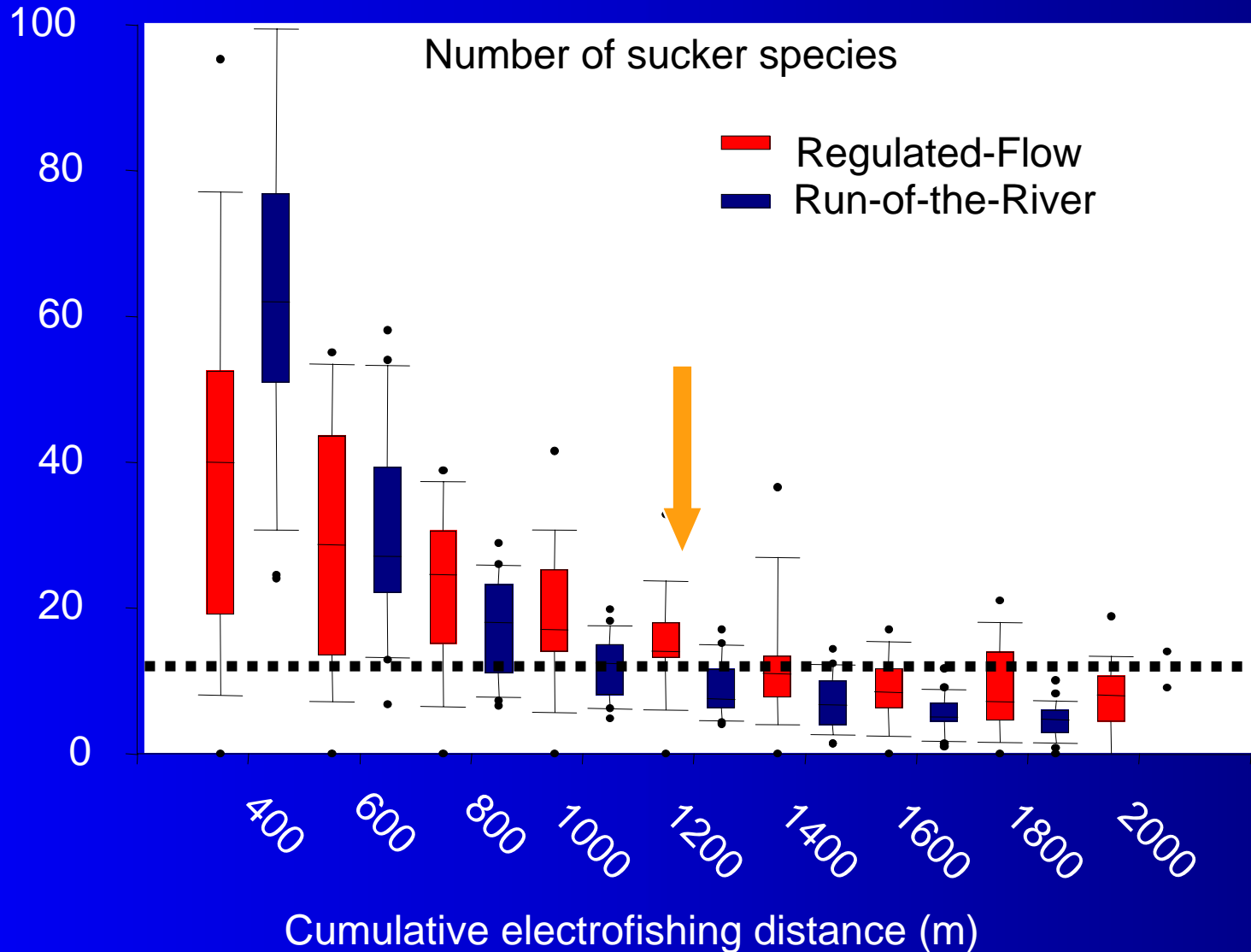
- **Single experimental design**
- **Testing of multiple designs**
- **Testing of distance effects on metrics**
- **Collected >28,000**
- **Electrofished 180 km**



Monte Carlo Simulations



Monte Carlo Simulations



Overview of Conclusions...

- Degree of impoundment plays a critical role in characterizing sites.
 - Metrics did not perform the same across sites of differing impoundment status (e.g., free-flowing vs. impounded).
 - May categorize by degree of impoundment
 - Different designs may be required to adequately describe different categories of systems.
 - Shallow systems – daytime electrofishing
 - Deeper, impounded systems – night electrofishing
 - Distance required may also vary

Ref: Flotemersch & Blocksom, submitted

Active Sampling Methods: Seining

- ✓ In places where electrofishing is prohibited
- ✓ Difficult boat access
- ✓ Low conductivity
- ✓ Low equipment cost
- ✓ Per-capita cost may be higher

Active Sampling Methods: Seining

- Selective
 - Small (species and juveniles)
 - Schooling (normally inhabit shallow water areas)
 - Slower



Horse seining, Columbia River, Oregon

Passive Fish Sampling Methods

Nets: Hoop, Fyke, Trap, Gill, Etc.



- **Advantages**
 - Simple in design and construction
 - No electrical equipment to fail
 - Require little specialized training
 - Yield fairly precise data (relative abundance)
 - **Disadvantages**
 - Selective (species, size, sex)
 - Require multiple trips to a site
 - Cannot pull fish out of cover
 - Spatial coverage is limited
- (Ref: Hubert 1992)

Field and Laboratory Processing of Fish

- **Be humane to collected specimens**
- **Be cognizant of who is watching**
 - Public relations
- **Identification**
 - Vouchers
 - Length or size classes
 - Weight
 - * Recording anomalies
 - * Tissue samples
- **Other issues**

External Anomalies:

Deformities, Erosions, Lesions, Tumors (DELT) anomalies

- Effective communicator of degraded quality
- Useful in sites degraded by multiple and cumulative stresses
- Reliable indicator condition
- Occurrence may be part of the recovery
- Important diagnostic tool
- Includes parasites (Ref: Sanders et al. 1999)



Fish Tissue Sampling

- Fish Tissue
 - Commonly used indicator of contaminant risk
 - Strong connection to resource use and exposure
 - Standard methods exist
- Important questions
 - How to sample?
 - What to sample?
 - Which analytes to consider?