

N O R T H W E S T  R E G I O N A L

# Oregon Practitioner's Manual



N E W B O R N S C R E E N I N G P R O G R A M  
8th Edition, 2008



---

# The Northwest Regional Newborn Screening Program

## **Oregon Practitioner's Manual**

### **Oregon Health & Science University**

Cary Harding, MD

Stephen L. LaFranchi, MD

Gregory Thomas, MD

Michael Wall, MD

### **Oregon Department of Human Services**

Michael R. Skeels, PhD, MPH

Cheryl A. Hermerath, MBA, DLM (ASCP), RM (NRM)

Leanne Rien, RN

Judith M. Tuerck, RN, MS

8th Edition, 2008

## Table of Contents

---

<b>Medical program consultants</b> . . . . .	<b>vi</b>
<b>Laboratory and follow-up team</b> . . . . .	<b>vii</b>
<b>Introduction</b> . . . . .	<b>1</b>
<b>Newborn screening essentials</b> . . . . .	<b>3</b>
<b>Conditions included in the screening panel</b> . . . . .	<b>4</b>
<b>Table I summary of conditions on the screening panel</b> . . . . .	<b>5</b>
<b>Table II normal values and criteria for requesting follow-up specimens</b> . . . . .	<b>8</b>
<b>Screening practices</b> . . . . .	<b>10</b>
Definition . . . . .	10
Who is responsible for ensuring that the screening test is performed? . . . . .	10
Parent refusal to have the infant screened . . . . .	10
Screening before discharge . . . . .	10
Proper time for specimen collection . . . . .	10
Diagnostic laboratories for screening older children and/or adults . . . . .	12
Specimen collection before transfer of infant to another facility . . . . .	12
Patient demographic information . . . . .	12
Specimen transport . . . . .	12
Special considerations . . . . .	13
Clinical signs or family history . . . . .	14
<b>Recommendations for specimen collection</b> . . . . .	<b>15</b>
Recommendation for heel puncture site in newborns . . . . .	16
<b>Unsatisfactory specimens</b> . . . . .	<b>17</b>
<b>Reporting of results</b> . . . . .	<b>18</b>
Web-Rad . . . . .	18
Normal results . . . . .	18
“Significant” abnormal results . . . . .	18
Other abnormalities and repeats . . . . .	18
Practitioner responsibility for documentation . . . . .	18
<b>Problems in screening practices</b> . . . . .	<b>19</b>
Infants who are never tested . . . . .	19
Parental refusal to have infant tested . . . . .	19
Common misconceptions of practitioners . . . . .	19
Timing of the tests . . . . .	20
Specimen inadequacy . . . . .	20
Inadequate demographic information . . . . .	21
Problems related to specimen transport to the laboratory . . . . .	21
<b>Educational services</b> . . . . .	<b>22</b>
Screening practice surveillance . . . . .	22
Birth facilities, health departments and community practitioners . . . . .	22
Additional education resources . . . . .	23

## Table of Contents

<b>Fees and screening kit information</b> .....	<b>24</b>
Type of kits .....	24
Cost .....	24
Storage of filter paper kits .....	24
Cost of diagnostic tests for confirmation of abnormal screening results .....	24
Order form for newborn screening kits .....	25
<b>Exemptions</b> .....	<b>26</b>
Fees .....	26
Statement of fee exemption .....	26
Religious objections .....	26
Parental refusal .....	26
Newborn screening test refusal form .....	27
<b>Core conditions</b> .....	<b>28</b>
<b>Cystic fibrosis</b> .....	<b>29</b>
<b>Endocrine conditions</b> .....	<b>31</b>
Congenital adrenal hyperplasia .....	31
Congenital hypothyroidism .....	33
<b>Hemoglobin conditions</b> .....	<b>35</b>
Sickle cell disease and other hemoglobinopathies .....	35
<b>Metabolic conditions</b> .....	<b>38</b>
Amino acid conditions: hypermethioninemia .....	38
Homocystinuria .....	38
Phenylketonuria (PKU) and hyperphenylalaninemia .....	40
Tyrosinemia, type I, II and transient .....	42
Fatty acid oxidation (FAO) conditions .....	44
Organic acid conditions .....	47
Urea cycle conditions .....	50
Other metabolic conditions .....	53
Galactosemia .....	53
Biotinidase deficiency .....	55
<b>References</b> .....	<b>56</b>
<b>Appendices</b> .....	<b>60</b>

### **Acknowledgment:**

We are indebted to the screening coordinators, consultants and practitioners in each of the regional states for their assistance and advice and to Leanne Rien, RN, for the many hours dedicated to this manual.

### **Recommended Citation:**

NW Regional Newborn Screening Program: Oregon Practitioner Manual, 8th Edition, 2008

## Medical Program Consultants

Medical consultants are available to provide consultation for the follow-up, evaluation and long-term management of infants detected in this program. Oregon patients are enrolled in a long-term management program at the Oregon Health & Science University. Questions concerning medical aspects of the program should be directed as below:

### **Metabolic Consultants:**

(biotinidase deficiency, galactosemia, organic acidemias, urea cycle, amino acid, fatty acid oxidation disorders)

Cary Harding, MD, [hardingc@ohsu.edu](mailto:hardingc@ohsu.edu)  
David Koeller, MD, [koellerd@ohsu.edu](mailto:koellerd@ohsu.edu)  
Robert Steiner, MD, [steinerr@ohsu.edu](mailto:steinerr@ohsu.edu)  
Department of Pediatrics  
Oregon Health & Science University, CDRC-F  
3181 SW Sam Jackson Park Road  
Portland OR 97201  
503-494-7859 • 503-494-9000 (paging)  
503-494-2781 (fax)

### **Endocrinology Consultants:**

(hypothyroidism, congenital adrenal hyperplasia)

Stephen LaFranchi, MD, [lafranchs@ohsu.edu](mailto:lafranchs@ohsu.edu)  
Cheryl Hanna, MD  
Bruce Boston, MD  
Dan Marks, MD PhD  
Department of Pediatrics  
Oregon Health & Science University  
3181 SW Sam Jackson Park Road, CDRC-P  
Portland OR 97201  
503-494-1927 • 503-494-9000 (paging)  
503-494-1933 (fax)

### **Hemoglobinopathy Consultant:**

(sickle cell disease)

Gregory Thomas, MD, [thomasgr@ohsu.edu](mailto:thomasgr@ohsu.edu)  
Department of Pediatrics  
Oregon Health & Science University  
3181 SW Sam Jackson Park Road, CHEMO  
Portland OR 97201  
503-494-8716 • 503-494-9000 (paging)  
503-494-2721 (fax)

### **Pulmonary Consultants:**

(cystic fibrosis)

Michael Wall, MD  
Department of Pediatrics  
Oregon Health & Science University  
3181 SW Sam Jackson Park Road, CHEMO  
Portland OR 97201  
503-494-8023 • 503-494-9000 (paging)  
503-494-8898 (fax)

Richard Cohen, MD  
Kaiser Permanente, NW Region  
3550 N. Interstate  
Portland, OR 97227  
503-904-7943 (paging)

## Oregon State Public Health Laboratory and Follow-up Team

**Laboratory Director:**

Michael R. Skeels, PhD, MPH  
Director  
Oregon State Public Health Laboratory  
3150 NW 229th Ave., Suite 100  
Hillsboro, OR 97124  
503-693-4100  
503-693-5602 (fax)  
michael.r.skeels@state.or.us

**Newborn Screening Laboratory Manager:**

Cheryl Hermerath, MBA, DLM (ASCP), RM (NRM)  
Newborn Screening Section Manager  
Oregon State Public Health Laboratory  
3150 NW 229th Ave., Suite 100  
Hillsboro, OR 97124  
503-693-4172  
503-693-5601 (fax)  
cheryl.a.hermerath@state.or.us

**Newborn Screening Nurse Follow-up Coordinator:**

Leanne Rien, RN  
Newborn Screening Nurse Coordinator  
Oregon State Public Health Laboratory  
3150 NW 229th Ave., Suite 100  
Hillsboro, OR 97124  
503-693-4173  
503-693-5601 (fax)  
leanne.c.rien@state.or.us

**Newborn Screening Education Coordinator:**

Judi Tuerck, RN, MS  
c/o Oregon State Public Health Laboratory  
3150 NW 229th Ave., Suite 100  
Hillsboro, OR 97124  
503-693-5601 (fax)  
tuerckj@msn.com





## Introduction

There has been a sea of change in newborn screening (NBS) programs in the last 5-10 years, both in terms of how NBS programs are viewed and the ability to screen for an increasing number of conditions.

### **Newborn screening is a public health system**

comprised of screening, follow-up, diagnosis, management, evaluation and education.<sup>1</sup> A systems approach recognizes the complexity and the multitude of stakeholders involved in NBS programs, including affected infants, children, adults, their families, practitioners, birthing facilities, mail and courier groups, treatment centers, schools, public health officials, legislators as well as the general public and the importance of effective communication between them. It also recognizes the great responsibility NBS programs have to identify affected infants in time to prevent damage and to ensure a seamless coordination of efforts of many people to that end. It is recognized that appropriate treatment must be available to every affected infant into adulthood and that long-term follow-up of the children is necessary regardless of geographic location. Finally, a systems approach emphasizes the importance of education and evaluation for everyone involved in NBS.

There is an **increasing number of conditions** for which NBS tests are now available. In the late 1990s, new technology, specifically tandem mass spectrometry (MS/MS), was adapted for use in NBS laboratories. It became possible to screen for more than 30 conditions from one small disc of blood. Previously, one bloodspot was needed for each separate test/condition and screening programs were limited as to the number of conditions that could be included on the screening panel. Multiplex platforms, such as MS/MS and others under development, open the possibility of screening for hundreds of conditions. Once it is technically possible, we as a society must decide whether we should screen, and if so for what conditions.

Traditionally, screening programs in the United States have been administered and carried out by individual states' public health laboratories and/or health divisions.

Screening panels and program amenities have been determined by the population risk for disorders and the resources available to each state. As a result, some states screen for as few as four disorders, while others screen for more than 30.<sup>2</sup> This disparity has galvanized both public and private groups to demand more uniform and expanded screening panels and to ensure a "standard of care" for all infants in the United States. Most other countries have a national NBS program, rather than state/province driven programs.

Parents of children affected with conditions detectable through NBS have lobbied state and national legislatures to provide funding and authority for state screening programs to implement the new technology and expand the number of conditions included, despite high start up costs and unproven sensitivity and specificity for many of the conditions. There is increasing pressure from advocacy groups to screen for conditions that have unproven treatments or that do not meet the classical criteria outlined by the World Health Organization in the 1960s.<sup>3</sup>

In 2005 a report from the American College of Medical Genetics (ACMG), commissioned by the Maternal and Child Health Bureau of the Health Resources and Services Administration (HRSA), recommended that all United States screening programs include a core panel of 29 conditions.<sup>2</sup> The National March of Dimes (MOD) organization not only supported the ACMG report, but began to issue "report cards" of state screening panels on their Web site.

As of July 2007, 90 percent of all United States infants are screened for the 29 recommended conditions.<sup>4</sup> The Oregon State Public Health Laboratory (OSPHL) implemented MS/MS screening in all the states in its regional program from 2002–2003 and added screening for cystic fibrosis in 2006, thereby including all 29 conditions and another 25 secondary conditions usually found in the course of screening for core conditions.

While each of these conditions is rare, collectively they affect about one in 900 infants, so the chance that any single infant will be affected is remote. The cost of not detecting one of these conditions is immense, both in human suffering and financial terms, because early

diagnosis and treatment can result in normal growth and development. Infants with these conditions appear normal at birth. It is only with time that the biochemical abnormality affects the infant's health and development.

By the time clinical symptoms appear the damage may be permanent or the infant may die, sometimes without a diagnosis having been made. However, NBS is a screening test, not a diagnostic test, and not all affected infants will be identified even with two routine screening tests, as is recommended or mandated in the majority of states.

The goal of the Northwest Regional Newborn Screening Program (NWRNSP), which includes Oregon, Idaho, Nevada, Alaska, Hawaii and New Mexico, is to identify as many affected infants as possible before damage can occur. To do so, every infant must be screened. This requires coordinated efforts from three groups of health care providers:

**Practitioners:** Responsible for the collection and handling of screening specimens, for providing parents with correct and current information and for prompt follow-up in the event of an abnormal result.

**Oregon State Public Health Laboratory (OSPHL) and Follow-up Team:** Responsible for testing, record keeping, quality control of laboratory methods, notification of results and tracking of abnormal and unresolved results.

**State Health Divisions and Oregon Health & Science University (OHSU) Subspecialty Programs:** Responsible for ensuring confirmatory testing of infants with abnormal results, for the management of confirmed cases, collection of long-term follow-up (LTFU) data and for education of practitioners.

Practitioner's responsibilities to the program are termed Newborn Screening Practice (see page 10 - 27).<sup>5</sup> A practitioner is defined as the person(s) responsible for supervising the birth and/or the early neonatal care of an infant. Inadequate screening practices can greatly affect the quality of screening tests and increase the chances that an affected infant may be missed or diagnosed late. At least one-third of cases missed by screening programs in the United States have been caused by errors in screening practice.<sup>6</sup>

The OSPHL, the follow-up team, together with regional state health agencies, have developed a quality assurance program to assist practitioners in improving their screening practices. Components of this program include ongoing education for practitioners and parents, computerized monitoring of measurable screening practices, and an examination of communication channels between practitioners, the laboratory, the follow-up team and the treatment centers.

This manual describes the conditions currently covered by the program as well as the standards and common problems for certain screening practices, and provides general program information as well as bulleted "NBS Essentials" summaries at the beginning of each chapter. We invite practitioners to contact us with any questions or concerns that may arise.

# Newborn Screening Essentials:

## 10 important points to remember about NBS

- 1 Please use the term **“Newborn Screening”** or **“Newborn Blood Spot Screening”**

The term, “PKU test” is outdated and confusing to parents. The screening panel now includes markers for approximately 30+ separate disorders, the most recently added being cystic fibrosis.

- 2 **Incidence** of all the blood spot conditions is now **One infant in 900** or 45–50 new cases each year in Oregon (~1 infant/week identified)
- 3 **Screen every normal infant TWICE**
  - 1st screen: 24–48 hours of age or before discharge, whichever comes first
  - 2nd screen: 10–14 days of age
- 4 **Screen every NICU infant TWICE or THRICE**
  - 1st: on admission (regardless of age)
  - 2nd: 5–7 days of age (regardless of treatments)
  - 3rd: 3–4 weeks of age (for premature or low birth weight only)
- 5 **Validity of screening specimens:**
  - 1st screen identifies 90% of all conditions (1st & 2nd for NICU)
  - 2nd/3rd screen identifies 10% of all conditions
- 6 **Goal of NBS: Diagnose and treat in the first week of life since:**
  - ~ 20 disorders can kill or maim in the first week or two of life
  - 10–20% of infants will be symptomatic in the first week
  - 5–10% may/will die in the first week
  - Infants with PKU and hypothyroidism lose significant IQ points if thyroid stimulating hormone (TSH) and phenylalanine are not under control by 2 weeks of age

- 7 **Maintain high index of suspicion for early presentation of emergent conditions**

- Symptoms: lethargy, poor feeding, weight loss or sudden cardio/pulmonary arrest
- Four infants have died before 10 days of age in our region even with NBS
- None of the diagnoses were suspected despite symptoms
- 1–2% of affected infants may have false negative results
- Screening tests are not diagnostic and affected infants may be missed. Practitioners should remain alert for signs of these conditions in infants and children regardless of the screening result

- 8 **Primary blood markers in emergent conditions:**

- Hypoketotic **hypoglycemia**, sudden cardio/pulmonary arrest (fatty acid oxidation disorders)
- **Hyperammonemia** (urea cycle disorders)
- **Acidosis** (organic acidemia)
- Abnormal **electrolytes** and **sodium** (congenital adrenal hyperplasia)
- **Reducing substances** in urine, liver dysfunction (galactosemia)

- 9 **If parents refuse testing:**

- Complete Newborn Screening Test Refusal form and counsel parents (page 27)

- 10 **Visit the Oregon State Public Health Laboratory Web site:** [www.oregon.gov/DHS/ph/nbs/index.shtml](http://www.oregon.gov/DHS/ph/nbs/index.shtml)

**Contact the NBS coordinator for your state.  
Oregon: Leanne Rien, RN: 503-693-4173**

## Conditions Included in the Screening Panel

Oregon newborns are screened for all core and secondary conditions recommended by the College of Medical Genetics and the March of Dimes. A description of these conditions start on page 28 and complete summaries are available on the OSPHL Web site: [www.oregon.gov/DHS/ph/nbs/index.shtm](http://www.oregon.gov/DHS/ph/nbs/index.shtm).

### **CYSTIC FIBROSIS\***

### **ENDOCRINE CONDITIONS:**

- Congenital adrenal hyperplasia (CAH)\*\*
- Congenital hypothyroidism\*

### **HEMOGLOBIN CONDITIONS:**

- Sickle cell disease and other hemoglobinopathies\*

### **METABOLIC CONDITIONS:**

#### AMINO ACID CONDITIONS:

- Homocystinuria\*
- Hyperphenylalanemia, including phenylketonuria (PKU)
- Tyrosinemia\*

#### FATTY ACID OXIDATION CONDITIONS:

- Carnitine uptake defect
- Carnitine palmitoyl transferase I deficiency (CPT I)\*
- Carnitine palmitoyl transferase II deficiency (CPT II)
- Multiple acyl-CoA dehydrogenase deficiency (MADD)
- Short chain acyl-CoA dehydrogenase deficiency (SCAD)
- Medium chain acyl-CoA dehydrogenase deficiency (MCAD)±
- Long chain 3 hydroxyacyl-CoA dehydrogenase deficiency (LCHAD)\*\*
- Very long chain acyl-CoA dehydrogenase deficiency (VLCAD)\*\*

#### ORGANIC ACID CONDITIONS:

- Beta-ketothiolase deficiency (BKD)±
- Glutaric acidemia, Type I (GA I)\*
- Isobutyryl CoA dehydrogenase deficiency (IBD)±
- Isovaleric acidemia (IVA)\* ±
- Malonic aciduria
- Maple syrup urine disease (MSUD)±
- Methylmalonic acidemias (MMA/8 types)±
- Propionic acidemia (PA)\* ±
- 3-Hydroxy-3-methylglutaryl CoA lyase deficiency (HMG)\*
- 2-Methyl-3-hydroxybutyryl CoA dehydrogenase deficiency (MBHD)\*
- 2-Methylbutyryl CoA dehydrogenase deficiency (2MBC)\*
- 3-Methylcrotonyl CoA carboxylase deficiency (3MCC)
- 3-Methylglutaconyl CoA hydratase deficiency (3MGH)
- Multiple carboxylase deficiency

#### UREA CYCLE CONDITIONS:

- Arginase deficiency
- Argininosuccinate lyase deficiency (ASA)±
- Citrullinemia±

#### OTHER CONDITIONS:

- Biotinidase deficiency
- Galactosemia±

\*The screening test will not detect 100 percent of affected infants.

± Represent emergent conditions. Infants are at risk of illness or death in the first week or two of life.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***

## Table I Summary of Conditions on the Screening Panel

**Table I** summarizes the conditions on the Oregon screening panel, including the incidence, symptoms, and treatment.

Disorders	Analyte Tested For	Incidence in NW Region	Symptoms if Not Treated	Treatment
Cystic fibrosis	Immunotrypsinogen (IRT)		Lung disease; growth failure	Pulmonary enzyme replacement therapy
Congenital adrenal hyperplasia (CAH)*	17-OH-Progesterone	1:12,000 1:300 in Yupik Eskimos	Addisonian crisis/salt wasting in 2/3 infants; dehydration; shock; hyperkalemia; virilization of females	Glucocorticoid and/or mineralocorticoid (Florinef)
Congenital hypothyroidism	Thyroid hormones (T4 with TSH confirmation)	1:3,000	Mental retardation, other brain damage; growth delay	Thyroid hormone (L-Thyroxine)
Hemoglobinopathies including sickle cell anemia	Hemoglobin patterns	1:15,000 (1:400 in African Americans)	In sickle cell disease: death by sepsis or splenic sequestration anemia; sickling crisis	Penicillin & comprehensive care
Biotinidase deficiency	Biotinidase	1:60,000	Mental retardation; seizures; skin rash; alopecia; hearing loss; death	Biotin
Galactosemia	Galactosemia enzyme (GALT)	1:60,000	Severe brain damage; liver disease; cataracts; death	Galactose-restricted diet
Argininase deficiency	Arginine	1:350,000	Irritability; developmental delay; spastic tetraplegia	Low protein diet, medication
Arginosuccinate lyase deficiency (ASA)*	ASA/Citrulline	1:88,000	Hyperammonemia; mental retardation; seizure; death	Low protein diet, medication
Arginosuccinate synthetase deficiency (Citrullinemia)*	Citrulline	1:71,000	Hyperammonemia; mental retardation; seizure; death	Low protein diet, medication
Homocystinuria	Methionine	1:118,000	Mental retardation; dislocation of lenses; marfanoid body habitus	Pyridoxine; methionine restricted, cysteine supplemented diet
Hyperphenylalaninemia, including phenylketonuria (PKU)	Phenylalanine	1:13,600	Profound mental retardation; seizures	Low phenylalanine diet
Tyrosinemia	Tyrosine	1:350,000	Vomiting, lethargy; liver disease; coagulopathy; renal tubular acidosis	Medication; low phenylalanine and/or low tyrosine diet

(continued on next page)

## Table I (continued)

Disorders	Analyte Tested For	Incidence in NW Region	Symptoms if Not Treated	Treatment
Beta-ketothiolase deficiency (BKD)	C5:1, C5OH	>1:300,000	Severe bouts of acidosis possibly resulting in mental retardation, death	IV support during episodes, bicarbonate supplement
Glutaric acidemia, Type I (GA I)	C5DC	1:71,000	Metabolic crisis; damages basal ganglia	IV support for brain illness
Isobutyryl CoA dehydrogenase deficiency (IBD)	C4	>1:300,000	None to cardiomyopathy	Carnitine therapy, protein restriction, avoid fasting
Isovaleric acidemia (IVA)	C5	1:120,000	Vomiting; lethargy; acidosis possibly resulting in coma, death	Protein restriction
Malonic aciduria	C3DC	>1:300,000	Mental retardation	Carnitine therapy, protein restriction
Maple syrup urine disease (MSUD)*	Leucine	1:350,000	Vomiting; lethargy; acidosis possibly resulting in death	Protein restriction
Methylmalonic acidemias (MMA/8 types)*	C3, C3/C2	1:50,000	Vomiting; lethargy; acidosis possibly resulting in death	Protein restriction; carnitine therapy, B-12
Multiple Carboxylase Deficiency (MCD)	C3, C5OH	>1:300,000	Hypotonia, seizures, skin rash, alopecia, lactic acidosis, brain damage. Average age at presentation: birth-18 months	Biotin
Propionic acidemia (PA)*	C3, C3/C2	>1:300,000	Vomiting; lethargy; acidosis possibly resulting in death	Protein restriction; carnitine therapy
3-hydroxy-3-methylglutaryl CoA lyase deficiency (HMG)	C5OH	>1:300,000	Hypoglycemia; acidosis possibly resulting in death; may be asymptomatic	Protein restriction
2-methyl-3-hydroxybutyryl CoA dehydrogenase deficiency (MHBD)	C5:1, C5OH	1:350,000	Asymptomatic 9–14 mos. Then severe mental and motor skill loss	Protein restriction
2-methylbutyryl CoA dehydrogenase deficiency (2MBC)	C5	>1:300,000	Hypoglycemia; mental retardation; may be asymptomatic	Avoid fasting, protein restriction
3-methylcrotonyl CoA carboxylase deficiency (3MCC)	C5OH	1:51,000	Most have been asymptomatic	None or protein restriction

(continued on next page)

## Table I (continued)

Disorders	Analyte Tested For	Incidence in NW Region	Symptoms if Not Treated	Treatment
3-methylglutaconyl CoA hydratase deficiency (3MGH)	C5OH	>1:300,000	Hypoglycemia; acidosis; may be asymptomatic	Protein restriction, avoid fasting
Short chain acyl-CoA dehydrogenase deficiency (SCAD)	C4	1:350,000	Most asymptomatic; hypotonia, mental retardation	Avoid fasting
Medium chain acyl-CoA dehydrogenase (MCAD)*	C6, C8, C10, C8/C10	1:11,000	Hypoglycemia possibly resulting in coma, death; may be asymptomatic	Avoid fasting, carnitine supplement
Long chain 3-hydroxyacyl-CoA dehydrogenase deficiency (LCHAD)	C14:1, C16, C16OH, C18, C18OH,	1:177,000	Hepatic dysfunction; hypoglycemia; failure to thrive	Long chain fatty acid restriction, Medium Chain Triglycerides (MCT) oil, carnitine, avoid fasting
Very long chain acyl-CoA dehydrogenase deficiency (VLCAD)	C14, C14:1, C16, C16:1, C18, C18:1,	1:71,000	Hypoglycemia with or without cardiomyopathy; muscle fatigue	Avoid fasting, low fat diet with MCT oil
Carnitine uptake/transport defects	C0, C16, C18	1:118,000	Hypoglycemia; cardiomyopathy	Avoid fasting, low fat diet, carnitine supplement
Carnitine palmitoyl transferase I deficiency (CPT I)	C0/C16+C18	Increased in Hutterite and Alaska native populations	Hypoketotic hypoglycemia, brought on by fasting or intercurrent illness. Average age at presentation: birth-18 months	Avoid fasting and long chain fatty acids; MCT oil supplement
Carnitine palmitoyl transferase II deficiency (CPT II)	C0, C4, C5, C6, C14, C16, C16:1, C18, C18:1	>1:300,00	Muscle weakness, pain and myoglobinuria leading to renal failure in 25%. Average age at presentation: 15-30 yrs; also a severe neonatal form usually lethal with multiple congenital anomalies	Avoid fasting and severe exercise; MCT oil supplement
Multiple acyl-CoA dehydrogenase deficiency (MADD)	C4, C5, C6, C8, C10, C14, C16, C18:1	>1:300,000	Multiple congenital abnormalities; acidosis, hypoglycemia	Low fat diet, avoid fasting, not effective in severe neonatal form

\* Infants may have severe neonatal presentation.



## Table II Normal Values And Criteria For Requesting Follow-up Specimens

**Table II** summarizes normal laboratory values and laboratory criteria used for follow-up. Phone follow-up (with mail confirmation) is provided for significantly abnormal test results. Mail follow-up is provided for normal and mildly abnormal results. More information on these conditions is available on Oregon Department of Human Services Web site: [www.oregon.gov/DHS/ph/nbs/expand.shtml](http://www.oregon.gov/DHS/ph/nbs/expand.shtml). See additional information on MS/MS conditions in Appendix II.

Analyte	Normal***	Follow Up by Medical Consultant/NBS Nurse**	Follow Up by Mail
Thyroxine	Upper 90% of T4 determinations	T4 in lower 10% and TSH >100 µU/mL	T4<5.0 µmg/dL, TSH normal (low birth weight infant)
TSH	<60 µU/mL if 0–11 hrs old <35 µU/mL if 12–48 hrs old <25 µU/mL if >48 hrs old	>200 µU/mL if 0–11 hrs old >100 µU/mL if 12–23 hrs old >60 µU/mL if 24–48 hrs old >35 µU/mL if >48 hrs old	60.1–100 µU/mL if 0–11 hrs old 35.1–100 µU/mL if 12–23 hrs old 35.1–60 µU/mL if 24–48 hrs old 25.1–35 µU/mL if >48 hrs old
17-OH-Progesterone*	<125 ng/mL if <24 hrs old <125 ng/mL if 24 hrs to ≤10 days old, BW ≤2500 g <75 ng/mL if 24 hrs to <10 days old, BW >2500 g <60 if >10 days old	>150.1 ng/mL if <24 hrs old >150.1 ng/mL if 24 hrs to ≤10 days old, BW ≤2500 g >100.1 ng/mL if 24 hrs to <10 days old, BW >2500 g >80.1 if >10 days old	125.1–150 ng/mL if <24 hrs old 125.1–150 ng/mL if 24 hrs to ≤10 days old, BW ≤2500 g 75.1–100 ng/mL if 24 hrs to <10 days old, BW >2500 g >60.1–80.0 if >10 days old
Hemoglobin IEF/HPLC	Hgb F and A	Probable homozygote	Probable heterozygote
Biotinidase	Activity present	Activity absent	Partial activity
Galactosemia enzymes (Gal-1-PO4 uridyl transferase, GALT)	Fluorescence present	No or reduced fluorescence AND galactose ≥20 mg/dL OR galactose <20 mg/dL if <48 hrs old	No or reduced fluorescence AND galactose <20 mg/dL age ≥48 hrs old
Arginine	<130 µM/L	≥200 µM/L	≥130 & <200 µM/L
ASA*	<6.0 µM/L	≥6.0 µM/L	
Citrulline*	<100 µM/L 1st NBS <200 µM/L 2nd NBS	≥100 µM/L 1st NBS	≥200 µM/L & ASA <6.0 µM/L 2nd NBS
Leucine*	<200 µM/L & Leu/Ala <1.30 1st NBS <330 µM/L & Leu/Ala <1.30 2nd NBS	≥200 µM/L & Leu/Ala ≥1.30 1st NBS ≥330 µM/L & Leu/Ala ≥1.30 2nd NBS	≥200 µM/L & Leu/Ala ≤1.30 1st NBS ≥330 µM/L & Leu/Ala ≤1.30 2nd NBS
Methionine	<90 µM/L	≥210 µM/L 1st & 2nd NBS ≥90 µM/L 2nd NBS with previous elevation on 1st NBS	≥90 µM/L & ≤210 µM/L
Phenylalanine	<190 µM/L	≥190 µM/L and Phe/Tyr ≥3.0	≥190 µM/L & Phe/Tyr <3.0
Tyrosine	<400 µM/L	≥600 µM/L 1st NBS ≥400 µM/L & <600 µM/L 1st NBS & ≥400 µM/L 2nd NBS ≥600 µM/L 2nd NBS	>400 µM/L & <600 µM/L
C0	>15.0 µM/L & <300 µM/L	<15.0 µM/L 1st & 2nd NBS, BW >2500 grams or not on hyperal ≥300 on 1st & 2nd NBS	<15 µM/L 1st NBS, BW >2500 gram or not on hyperal <15 µM/L 1st & 2nd NBS, BW <2500 gr or on hyperal ≥300 µM/L 1st NBS
C0/C16+18	<130	≥130	
C3*	<7.5 µM/L & C3/C2=<0.30	≥7.5 µM/L & C3/C2=≥0.30	≥7.5 µM/L & C3/C2=<0.30

(continued on next page)



Table II (continued)

Analyte	Normal***	Follow Up by Medical Consultant/NBS Nurse**	Follow Up by Mail
C3DC	<0.50 µM/L	≥0.50 µM/L	
C4	<1.80 µM/L	≥2.50 µM/L	≥1.80 µM/L & <2.50 µM/L
C5	<0.70 µM/L 1st NBS <1.10 µM/L 2nd NBS	≥0.70 µM/L 1st NBS	≥1.10 µM/L 2nd NBS if on hyperal & 1st NBS normal
C5:1	<0.80 µM/L	≥0.80 µM/L	
C5OH	<1.50 µM/L	≥1.50 µM/L	
C5DC	<0.50 µM/L	≥0.50 µM/L	
C6, C8, C10*	<0.80 µM/L, <1.0 µM/L, <0.70 µM/L	≥0.80 µM/L, ≥1.00 µM/L, ≥0.70 µM/L	
C14	<0.80 µM/L	≥0.80 µM/L if other long chain analytes elevated	≥0.80 µM/L only elevation
C14:1	<0.80 µM/L	≥0.6 µM/L	
C16	<9.70 µM/L	≥9.70 µM/L if other long chain analytes elevated	≥9.70 µM/L only elevation
C16:1	<0.80 µM/L	≥0.80 µM/L if other long chain analytes elevated	≥0.80 µM/L only elevation
C16OH	<0.50 µM/L	≥0.50 µM/L	
C18	<3.10 µM/L	≥3.10 µM/L if other long chain analytes are elevated	≥3.10 µM/L only elevation
C18:1	<4.60 µM/L	≥4.60 µM/L if other long chain analytes are elevated	≥4.60 µM/L only elevation
C18:1OH	<0.30 µM/L	≥0.30 µM/L if other long chain analytes are elevated	≥0.30 µM/L only elevation
C18OH	<0.30 µM/L	≥0.30 µM/L	
Immunotrypsinogen (IRT)	<100 ng/mL 1st NBS <80 ng/mL 2nd NBS	≥100 ng/mL 1st NBS & ≥80 ng/mL 2nd NBS	≥100 ng/mL 1st NBS

\* Infants may have severe neonatal presentation.

\*\*All phoned results are followed by mailed confirmation. All of these tests are screening tests. Abnormal results need full evaluation/discussion with a consultant before a diagnosis is confirmed or treatment is started.

\*\*\*Normal and abnormal values are subject to change based on continuing statistical evaluation.

## Screening Practices

### Definition

SCREENING PRACTICES are the actions and decisions of practitioners regarding the collection, handling and follow-up of newborn screening specimens. Practitioners are integral to the newborn screening in that they are responsible for collection and handling of specimens for every infant in their care, parent education and prompt action on incomplete or abnormal results referred to them. In the event an infant is affected, the practitioner's actions and decisions to ensure rapid evaluation and appropriate treatment can have life long implications for the infant and the family.

### Who Is Responsible For Ensuring That The Screening Test Is Performed?

Oregon statutes require that every infant be tested, and designates practitioners as being responsible for specimen collection. The definition of "practitioner" includes physicians, nurses, and midwives who deliver or care for infants in hospitals, birth centers or at home. Oregon law also specifies that parents are responsible to ensure that their infants are tested.

### Parent Refusal To Have The Infant Screened

A parent may refuse screening for personal and/or religious beliefs. Parents may be influenced by the attitude of their practitioner toward newborn screening, and the majority will agree to screening if properly counseled about the importance of early detection. Infants have been harmed as a result of practitioners who have advised against screening as the "conditions were too rare," or "we don't want to poke the infant," or "the second test is not really necessary," etc.

In the event that parents are adamantly opposed to screening, it is the responsibility of the practitioner to fully inform them about the process and all screening conditions and to obtain a signed Newborn Screening Test Refusal (informed dissent) document. This should be placed in the infant's medical record and a copy sent to the OSPHL or the state coordinator. Failure to do so may result in significant practitioner and facility liability if an affected infant is not screened and subsequently harmed. Unfortunately, informed dissent does not confer protection from liability, but can limit damage in

some cases. Sample statements of parental refusal and

**TABLE III: VALIDITY OF 1ST AND 2ND NBS TESTS**

Disorder	Percent detected on first test	Percent detected on second test
Hemoglobinopathies	>99	NA
Hypothyroidism	90	10
Congenital Adrenal Hyperplasia	95	5
Phenylketonuria	>99	1
MSUD	>99	1
Other Amino Acidopathies	50–90	10–50
Galactosemia	>99	1
Biotinidase Deficiency	100	0
Fatty Acid Oxidation Disorders	90	10
MCAD	95	5
CPT1 (Alaska Native)	55	45
Carnitine uptake	40	60
LCHAD/MLCAD	85	15

religious exemption are given on pages 26 and 27 or the hospital's form may be used.

### Screening Before Discharge

All birthing facilities, including birth centers in Oregon, are required to practice uniform discharge screening regardless of the age or feeding status. This is done because some infants do not return for routine post-natal care and most of the tests are valid at any age. Failure to collect a specimen before discharge may result in a significant liability on both the facility and responsible practitioner if an affected infant is missed.

### Proper Time For Specimen Collection

There are over 20 conditions on the screening panel that can kill or maim in the first week or two of life; however, another three to four may not show any abnormal results until the second or third week of life. Improved technology in the laboratory makes the issue of milk ingestion less important than it was in the past. All states in our region require infants to have specimens collected prior to discharge from the birth facility.

Specimens obtained prior to 48 hours of age are valid for most of the conditions covered on the newborn screening panel. There is, however, a statistical chance that certain amino acid and endocrine disorders, urea

cycle defects and FAO disorders may be missed. In the NWRNSP, 10–12 percent of those with hypothyroidism, CAH and certain disorders detected via MS/MS are found only on the second specimen as outlined in Table III.

#### NORMAL NEWBORNS:

In Oregon, all infants must have two specimens. For normal, non-premature infants, the first specimen should be collected before discharge, ideally between 24–48 hours and no later than five days of age as shown in the table below. These recommendations are earlier than the Oregon Administrative Rules currently specify, but due to the number and severity of emergency metabolic conditions and CAH it is felt an earlier specimen is preferable. A second specimen must be collected between 10–15 days of age on all infants. Infants born at home should also be screened at these times. A parental refusal form should be signed if screening is refused.

#### PREMATURE, LOW BIRTH WEIGHT OR SICK INFANTS:

All Neonatal Intensive Care Units (NICU) in our region are strongly urged to practice “admission screening” for all infants admitted to their units. In some cases the infants may only be minutes or a few hours old. This is followed by a second specimen between 5–7 days of age (see Appendix I).

#### DYING INFANTS:

If an infant is likely to die, especially if there is no apparent cause for death, it is appropriate to collect a newborn screening specimen. In addition, collect a urine specimen, which should be frozen immediately. While dying infants may have abnormal amino acids

as a normal response to organ failure, the blood may also provide a diagnosis of an early onset screening disorder or be a source of DNA for future diagnostic studies. In many cases, the blood spots have been the only source of DNA available for that infant and have been a valuable resource for families.

#### OLDER INFANTS

The American Academy of Pediatrics recommends that physicians know the screening status of every child in their care. Older infants and children may enter the practice without evidence of screening, or the physician may be suspicious of a screenable condition. Unfortunately, OSPHL has established standards and cutoffs for newborns and infants only up to 6 months of age. They are not applicable nor reliable for older infants and children. OSPHL therefore, cannot accept specimens for children older than 6 months of age.

We highly recommend proceeding directly to diagnostic testing if there are signs or symptoms of a metabolic disorder present in the child (see below).

Please see page 12 for a listing of metabolic laboratory who can provide Tandem Mass Spectrometry (MS/MS) testing on older children. Also listed, are recommendations for further diagnostic testing for the other disorders on the newborn screening battery. These laboratories are listed for your convenience only. You may send your specimen to any accredited laboratory that provides the appropriate testing.

**TABLE IV: AGE OF INFANT AT SPECIMEN COLLECTION**

Infant Type	1st Specimen	2nd Specimen
NORMAL INFANT (hospital, birth center and home births)	Ideal: 24-48 hours of age* Acceptable: 0-23 hours; 49-120 hours	Ideal: 10-15 days Acceptable: 16-183 days*
PREMATURE OR LOW BIRTH WEIGHT INFANTS	Upon admission (any age)* Before red cell transfusion	5-7 days; if <2000 gms or still in NICU, collect third specimen
OLDER INFANTS ( >6 months)	Diagnostic tests only Blood spots may be used	Not needed
DYING INFANTS (of unknown cause)	Four full blood spots (any age) Urine frozen ASAP	n/a

\*These recommendations are earlier than those recommended in Oregon Administrative Rules.

### **Diagnostic Laboratories for Screening Older Children and/or Adults**

Also listed are recommendations for further diagnostic testing for the other disorders in the newborn screening battery. *These laboratories are listed for your convenience only. You may send your specimen to any accredited laboratory that provides the appropriate testing.*

- **MS/MS TESTING (AKA EXPANDED OR SUPPLEMENTAL NEWBORN SCREENING):**

This includes testing for selected amino acid, fatty acid, organic acid and urea cycle disorders.

Baylor University Medical Center at Dallas  
200 First Street SW  
Dallas, Texas 75226  
Telephone: 214-820-2822

- **BIOTINIDASE DEFICIENCY:**

Biotinidase enzyme assay

Biochemical Genetics Laboratory  
University of Maryland, School of Medicine  
Division of Human Genetics  
BRB 11-047, 665 West Baltimore Street  
Baltimore, Maryland 21201  
Telephone: 410-706-2810

- **CONGENITAL HYPOTHYROIDISM:**

Serum – Free T4, TSH can be done at most local laboratories.

- **SICKLE CELL DISEASE:**

Isoelectric Focusing/High Performance Liquid Chromatography can be done at most local laboratories.  
Never use Sickledex.

- **Galactosemia:**

Galactose-1-Phosphate  
Galactose-1-Phosphate Uridyl Transferase

University Children's Genetic Laboratory  
116 East Broadway  
Glendale, CA 91205  
Telephone: 818-548-0999  
Fax: 818-548-1555

- **CONGENITAL ADRENAL HYPERPLASIA:**

Serum 17-OH-Progesterone can be done at most local laboratories.

### **Specimen Collection Before Transfer of Infant to Another Facility**

Oregon Administrative Rule (OAR 333-024-0215) specifies that infants who are transferred to another facility within 48 hours of birth should be tested by the receiving facility. (See timing of specimen collection on page 11.) If an NBS kit has been issued by the birth hospital to the infant, it can be sent with the infant ensuring that both facilities are notified of the results.

### **Patient Demographic Information**

Accurate patient information is critical for rapid follow-up in the event of abnormal results. Be sure to use the correct part of the kit: PART 1 for the first specimen and PART 2 for the second specimen. Failure to do so makes it difficult to track the infant's results. In addition, hemoglobin and cystic fibrosis testing is done only on the first specimen. The laboratory form should be completed with the information requested. This information is required by Clinical Laboratory Improvement Amendments of 1988 (CLIA) and is part of the legal record and must be legible. Infant's name, birth date and time, specimen collection date and time and physician's name are particularly important. Please name the physician who will be responsible for this infant's care after discharge, not the resident or on-call physician who may have seen the infant in the hospital. In the event of an abnormal test result, the physician and/or facility named on the card is legally obligated to locate the infant. Do not use plastic imprint cards, as they often produce unreadable information and cause compression damage to the filter paper. All demographic data should be completed before the blood is drawn to avoid contamination of the specimen. The person collecting the specimen should initial the form.

### **Specimen Transport**

If lives are to be saved, it is critically important that the OSPHL receives newborn screening specimens as soon as possible after collection. Ideally, specimens should be mailed or transported as soon as they are dried (2–4 hours) and no later than 24 hours after collection. Many of the conditions on the screening battery kill or maim infants in the first week of life. To prevent this, diagnosis and treatment must occur rapidly. Consideration should be given to the use of overnight courier or mail services if specimens are taking longer than two or three days

to arrive at the laboratory. Significant degradation of hemoglobin and some analytes also occurs in specimens older than one week.

If a courier is used, it is advisable to establish a list of specimens sent and to document the time of specimen pickup and delivery. This protects the submitter if specimens are lost in transit.

### Special Considerations

Newborn screening for premature, low birth weight (LBW) or sick infants can be complex and sometimes frustrating as they generate more false positive results than normal infants, are more likely to have the screening test forgotten, and generally require serial specimens. The infant's immaturity and/or illness and the critical nature of their care conspire to interfere both with the collection of the specimens and interpretation of results. In addition to the timing of specimen collection discussed earlier, some of the problems unique to these infants are outlined below.

**TRANSIENT HYPOTHYROXINEMIA OF PREMATURITY (THOP):** The hypothalamic-pituitary-thyroid (HPT) axis is not functioning properly in premature infants and it is positively correlated with gestational age. In addition, sick and premature infants have a greater need for thyroxine and topical iodine skin cleansers depress T4. As a result T4 levels can be quite low for a week or more; TSH levels are usually normal. The significance of THOP on later development is still unknown due to the lack of well controlled long term outcome studies, but is thought to be somewhat benign. While a low T4 and an elevated TSH are the classic hallmarks of congenital hypothyroidism, some CH affected infants have a delayed rise in their TSH, so practitioners cannot assume a premature or sick infant with a low T4 only has THOP and not CH. Serial screening specimens for T4/TSH are required until the T4 normalizes.

**ANTIBIOTIC THERAPY:** Antibiotics containing pivalic acid given to mothers during labor or to newborns may cause false elevation of isovaleryl/2-methyl butyryl carnitine. Be sure to specify this antibiotic on the request slip if it has been used.

**HYPERALIMENTATION AND CARNITINE THERAPY:** These are not contraindications to screening, but samples

should not be taken from the line that is used to deliver the alimentation or drugs. High levels of several amino acids can occur during hyperalimentation and are the most common reason for "mixed elevations." Carnitine, added to intravenous solutions or formulas, may mask the biochemical markers for carnitine acylcarnitine translocase deficiency or carnitine palmitoyl transferase deficiency, type II. Screening specimens must be repeated after these therapies are discontinued. Be sure to specify these therapies on the request slip.

**RED CELL TRANSFUSIONS:** A specimen should be obtained before the transfusion as donor cells provide normal levels of enzymes and hemoglobins and may temporarily normalize certain metabolites. Metabolite tests (e.g., phenylalanine, carnitine, galactosemia) will become abnormal after a few days regardless of the transfusion. It may take as long as 120 days for infants to develop abnormal enzyme levels and hemoglobins after a transfusion, significantly delaying diagnosis and treatment. This is the rationale for admission screening for every NICU infant.

An early screening test, before these interventions are begun, has been life saving for affected infants in the past. While only a small percentage of infants receive red cell transfusions, it has been impossible to establish policies and procedures in NICUs that ensure specimen collection before transfusion. Furthermore, even with aggressive follow up and the assistance of local physicians we are able to obtain a valid screening test on less than 40 percent of the transfused infants three months after the last transfusion. For affected infants, damage will have accrued during this time, increasing the liability to the physician, hospital, state and program.

**TRANSIENT ABNORMALITIES:** While NICU admission screening is strongly recommended, some premature/sick infants tested in the first few hours of life may have an increased chance of false positive results from elevated thyroid stimulating hormone (TSH) and/or 17-OH-Progesterone. These are generally elevated in the first 24 hours after birth as a reaction to stress, falling into normal range a day or two later. A recent study of infants in NICUs who were tested on admission showed that an additional 4 percent of NICU infants had elevated TSH on these early specimens, but 99 percent resolved

on the second test. Also, a very early specimen is not valid for many of the amino acids, such as phenylalanine or leucine, but these metabolites rise independent of transfusions and should be identified on the second test. We believe NICU admission screening improves screening for infants who will receive transfusions without creating an undue number of false positives for TSH.

### **Clinical Signs or Family History**

A number of clinical situations will modify the usual approach of obtaining a newborn screening specimen and waiting for the result. The following are suggested guidelines for particular situations that may arise.

When in doubt about the course of management for any of the conditions on the screening test, consultation with a specialist is advised. The directory on page vi offers consultants by disease to assist with screening and confirmation testing.

- **INFANTS WHO EXHIBIT CLINICAL SIGNS AND SYMPTOMS:** The newborn screening test, like any laboratory test, may have false positives and false negatives. If signs and symptoms of one of the newborn screening conditions are clinically evident, the physician should proceed to diagnostic testing, pending the results of the screen or in spite of the results of the screen. It may be necessary to treat as if the infant has the condition. Medical consultation is available 24-hours, 7 days a week for assistance for rapid diagnosis and institution of treatment for infants suspected to have disease. Common laboratory tests available locally may help distinguish conditions. These include:
  - CAH: electrolytes
  - FAO disorders: glucose
  - OA disorders: ph, acidosis
  - Urea cycle disorders: ammonia, glucose
- **IF THE RESULTS OF THE NEWBORN SCREEN ARE PENDING:** For any of the screened conditions, but especially those in which the metabolite accumulation can be life threatening such as adrenal hyperplasia or many of the metabolic conditions, contact a consultant specialist for instructions on further evaluation of the patient.
- **IF THE NEWBORN SCREENING TEST RESULT WAS "NORMAL":** If clinical symptoms suggest one of the screened conditions despite a "normal" screening result, the physician should proceed as if the patient has the condition and immediately contact a consultant specialist for instructions on further evaluation of the patient. If the infant was found to be affected, notify the OSPHL as soon as possible.
- **NEWBORN SCREENING OF AN INFANT WITH AN AFFECTED SIBLING OR OTHER CLOSE RELATIVE:** As many of the conditions tested for by newborn screening are genetic, it is possible that multiple members of a family may be affected. Prenatal diagnosis is possible for many of these conditions; if prenatal diagnosis determines that the infant is affected, any appropriate treatment (e.g., special diet) should be initiated immediately after birth. If prenatal diagnosis predicts an unaffected infant, practitioners should bear in mind that no prenatal diagnostic test is 100 percent accurate. Neonates who are siblings or close relatives of an affected individual have a 25 percent risk of having the condition themselves. They are, therefore, not part of the "general population" for whom newborn screening is designed. For any infant with a positive family history, providers should contact appropriate consultant specialists, ideally prenatally, or immediately at birth, to determine the proper diagnostic tests and proper timing of those tests and whether supportive therapy is needed.
- **SCREENING SIBLINGS OR AN INFANT OLDER THAN 6 MONTHS OF AGE:** The OSPHL does not screen infants older than 6 months due to a lack of age appropriate cutoffs. Infants and children older than 6 months can be "screened" using a filter paper specimen, but it should be sent to a diagnostic laboratory with appropriate cutoffs as listed below. (See page 12 for information on screening older infants and children.)



## Recommendations for Specimen Collection\*

1. To prevent specimen contamination, do not touch any part of the filter paper circles before, during or after collection. Multiple agents can contaminate filter paper (see Unsatisfactory Specimens chart on page 17).
2. Identify infant and match with correct screening kit. Make sure to select the correct kit part (1 or 2) depending on which specimen is being collected.
3. Complete all demographic data before proceeding to collection.
4. Observe universal precautions.
5. Capillary blood obtained from a heel lance is the preferred specimen. Cord blood is not a satisfactory specimen as the infant's biochemistry will not be reflected. Specimens obtained from peripheral or central lines are acceptable if they are flushed of hyperalimentation or antibiotics. Blood from an intravenous stick is acceptable as long as it does not clot and can be applied to the filter paper directly from the syringe (needle must be removed).
6. It is essential to open a capillary bed in order to obtain sufficient blood. The most effective method is to use scalpel bladed lancets. Pointed lancets are painful to the infant and make a hole rather than a small slit, greatly reducing blood flow. Under no circumstances should a lancet longer than 2.4 mm be used on infants weighing less than 2500 grams.
7. Heat infant's foot if necessary in warm water, towel, or chemical pack. Heat source should not exceed 42 degrees centigrade and should not be left in contact with skin for a prolonged period.
8. Select a lance site on the infant's heel (see diagram), cleanse with alcohol and air dry. Hold infant's limb lower than heart.
9. Lance the heel with the sterile scalpel bladed lancet. Wipe away the first drop of blood to remove tissue fluids. Do not milk the heel. If blood flow is insufficient it is better to stop and re-lance the heel.
10. Allow sufficient blood to collect on the heel to fill each circle by a single application of blood to the filter paper. Do not use capillary tubes or other collection devices. Apply blood only to one side of the filter paper (it doesn't matter which side is used). Blood should soak all the way through the filter paper so that the blood spots look similar on both sides. Complete, even saturation of the entire circle is essential for accurate testing. Neatness doesn't count.
11. It is important not to superimpose blood drops on top of each other. Let each drop touch the paper about 1/8 inch away from each other. This may prevent layering and uneven saturation, one cause of false results.
12. Collect the blood in all four circles. A minimum of three circles is necessary to complete the screening battery. If there are problems with sufficient blood flow, it is better to fill three circles completely than to fill four circles inadequately (see #9 above).
13. After circles are filled, the foot should be elevated above the body and a sterile gauze pad or cotton swab pressed against the puncture site until the bleeding stops. Bandages should be avoided as they may irritate sensitive skin.
14. Air dry specimens at room temperature for 2–4 hours only in a horizontal position with the blood spots exposed. (A CD holder works well.) Hanging wet specimens will cause heavier red cells to migrate to the dependent end of the circle resulting in uneven saturation.
15. Do not expose the specimen to heat or humidity at any time. Do not dry on a heater, in a microwave, with a hair dryer or in sunlight. Do not place in plastic bags, leave in a hot mailbox or in a hot car; proteins and enzymes will be destroyed. Ensure that the specimen is completely dry before mailing. In hot weather, desiccant packets may be added to reduce humidity during transit.

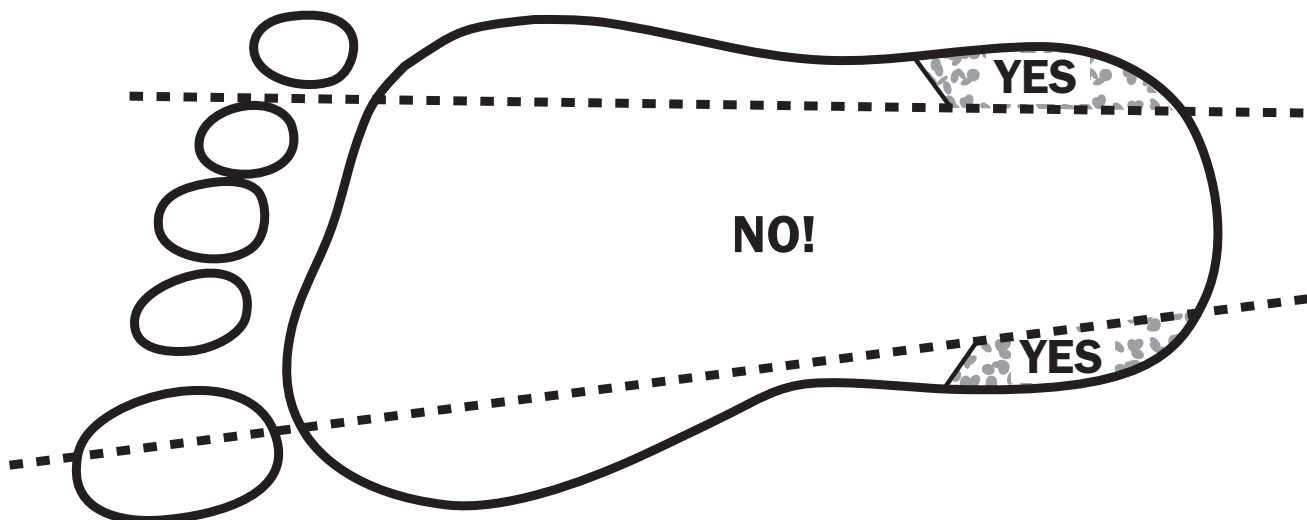
16. Do not batch specimens collected over several days as infants affected with emergent disorders may die before results can be made available. Specimens can be sent in the same package, but there must be a daily mail or courier pick up for all specimens.
17. Insert dried specimens into envelope (do not use plastic), seal and mail within 4–12 hours of collection and no later than 24 hours after collection. Weekend and holiday specimens should be stored at room temperature and sent by overnight mail or courier at the earliest opportunity. All specimens should be sent by first class or overnight mail or by courier. Specimens should be received by the OSPHL within 12–48 hours of collection.
18. Specimens should be documented as sent. If by courier, a packing list should be kept and the courier should sign for pick up and delivery of specimens. This protects the hospital from liability in the event the specimen is lost in transit.

**Recommendation for Heel Puncture  
Site in Newborns**

Perform punctures on the most medial or most lateral portion of the plantar surface of the heel (see diagram).

A full-color chart illustrating proper specimen collection, “Neonatal Screening Blood Specimen Collection and Handling Procedure,” may be obtained at no charge from Whatman, Inc., Telephone: 1-800-whatman or 1-973-245-8300, Fax: 1-973-245-8329, E-mail: info@whatman.com.

These recommendations conform to CLSI publication LA4-A4. See page 23.





## Unsatisfactory Specimens

Newborn screening laboratories receive many specimens that are unacceptable for testing. If the specimen is improperly collected, the accuracy of the screening test results is compromised, and the laboratory must reject them. This requires that the submitter locate the infant

and repeat the collection procedure and delays the screening of the newborn. The following table outlines the most common errors in specimen collection.

Consult the OSPHL for additional information and assistance with specimen collection.

UNSATISFACTORY SPECIMEN	POSSIBLE CAUSES
<p><b>Uneven Saturation:</b> Blood spot appears layered and/or scratched or abraded. DO NOT use a capillary tube for application of the blood, layer successive drops of blood or apply blood more than once in the same circle.</p>	<ul style="list-style-type: none"> <li>• Application with a capillary tube or needled syringe.</li> <li>• Applying blood to both sides of the filter paper.</li> <li>• Touching the filter paper with fingers or gloves.</li> <li>• Layering several blood drops on top of each other.</li> <li>• Hanging filter paper to dry.</li> </ul>
<p><b>Insufficient Blood (QNS):</b> Blood spots should look the same from both sides of the filter paper.</p>	<ul style="list-style-type: none"> <li>• Failing to open a large enough capillary bed resulting in insufficient blood flow from the infant's heel to allow complete absorbency through to the second side.</li> <li>• Removing the filter paper before blood has completely soaked through to the second side.</li> </ul>
<p><b>Contaminated:</b> Blood spots appear to be contaminated.</p>	<ul style="list-style-type: none"> <li>• Not wiping the alcohol from the puncture site before lancing the skin.</li> <li>• Not wiping away the first drop of blood to collect on the heel.</li> <li>• Allowing the filter paper to come in contact with alcohol, formula, bodily fluids, water, etc., before or after the blood specimen collection.</li> <li>• Squeezing or milking the area around the puncture site may cause contamination or dilution with tissue fluids.</li> <li>• Using a non-isopropyl alcohol swab (betadine or non-sting skin prep swabs) may destroy the proteins in the blood, alter absorbency or give false positive/negative results.</li> </ul>
<p><b>Blood Received Wet:</b> Blood spots appears to be "wet" when received.</p>	<ul style="list-style-type: none"> <li>• Mailing the specimen before drying a minimum of two hours.</li> <li>• Placing in a plastic bag even after minimum drying time.</li> </ul>
<p><b>Pale Blood:</b> Blood spot appears very pale or "washed out" in color.</p>	<ul style="list-style-type: none"> <li>• Low hematocrit in a normal newborn.</li> <li>• Infant is very ill or of very low birth weight.</li> <li>• Contamination of the filter paper before or after application of the blood.</li> </ul>
<p><b>Specimen Too Old:</b> Specimen received in the laboratory more than 14 days after collection date.</p>	<ul style="list-style-type: none"> <li>• Specimen was not mailed within 24 hours after collection.</li> </ul>

## Reporting of Results

Before discharge, hospital and birth unit personnel may be designated and listed on the request slip as the physician-of-record. In the event of an abnormal test result, the screening laboratory will refer to the physician-of-record although the infant is no longer under his or her care and may not even be known to him or her. Responsibility for follow-up remains with the physician-of-record until another practitioner actively accepts it. It is essential to list the practitioner who will be providing direct care after discharge.

### Web-Rad

Results for all infants born in your facility or under care in your practice can now be accessed online through the OSPHL's Web site. Contact Oregon State Public Health Laboratory, Newborn Screening Section by phone (503-693-4174) or e-mail to sign up and obtain a user ID and password.

### Normal Results

Normal results are mailed daily to hospitals and to the physician-of-record. The complete test battery is usually completed in 2–10 working days after receipt by the laboratory.

### “Significant” Abnormal Results

All results considered urgent are reported to the medical consultants who will phone the submitting hospital or practitioner with recommendations for further action. Such action is followed by mail or fax confirmation.

### Other Abnormals and Repeats

Lesser abnormalities or inadequate samples are reported immediately by phone or fax to the submitting hospital and/or practitioner with a request for retesting.

It is the practitioner's responsibility to ensure that all infants with abnormal results are retested; however all such infants are also tracked by the laboratory and/or the medical consultants until a resolution or diagnosis is confirmed.

State health agencies are notified for assistance when there are problems in obtaining repeat tests for infants with abnormal results.

## Practitioner Responsibility for Documentation

It is the responsibility of the practitioner to ensure every infant is tested and that a result is received and filed in the medical record. Practitioners should know the screening status of every infant in their care, regardless of age. Specific care should be paid to infants/children adopted from overseas as not all countries have newborn screening programs.

Specimen collection must be documented in the infant's chart and if preferred in a separate logbook. Information should include name of infant, hospital ID number, screening kit ID number, date collected, date mailed and the name of the person who collected the specimen. When screening results are returned to the submitter they should be noted in the logbook and the report filed in the medical records. Community practitioners must also ensure that the second specimen is obtained at the appropriate time and that documentation is completed.

In the event the results of an infant's screening tests are not received from the screening laboratory within two weeks of collection, the hospital and practitioner should assume responsibility for follow-up. We recommend the following procedure:

1. Contact the Oregon State Public Health Laboratory, Newborn Screening Section, 503-693-4174, or WebRad, to determine if specimen was received and to request a report to be mailed or faxed.
2. If the specimen was not received, it must be presumed lost. Notify the infant's private physician, local health department nurse and/or parents by phone and letter that the specimen may have been lost and that another should be obtained without delay.
3. Document these actions in the infant's medical record.
4. Request a copy of screening results be sent to your facility for the medical record.
5. If these steps do not result in the infant being screened, notify Leanne Rien, RN, 503-693-4174, the infant's health care provider and consider a public health nurse referral.

## Problems in Screening Practice

### Infants Who Are Never Tested

Mandatory state screening laws in the United States were passed in the 1960s not because parents were opposed to screening but because hospitals and practitioners were slow to provide screening services. While screening programs are now universal in the United States, it is estimated that approximately 1–2 percent of births are not screened either because of parental refusal or practitioner oversight or omission. For every one percent of newborns not screened in the United States, approximately 45–50 infants will be missed each year. These infants represent a major medical and legal liability to the program and to practitioners involved in their care. The legal awards for missed cases have been as high as \$30 million per case. Hospitals must develop a fail-safe system to ensure screening of every infant before discharge. In addition, community practitioners must pay special attention to older infants not born in the United States (i.e., adopted and/or migrant), infants born at home and/or attended by a lay midwife or older infants or children with symptoms of one of these conditions.

### Parents' Refusal To Have The Infant Tested

See page 10 for guidelines, and page 27 for an example of form to use in this situation.

### Common Misconceptions of Practitioners

Failure to test an infant is often due to misconceptions about the program or the tests. For example:

- **The diseases are so rare practitioners may advise parents not to have screening done.**

FACT: NBS identifies 1:900 infants with one of the conditions included on the screening panel. Nationally this represents 5,000 infants/year who would otherwise die or become irreversibly harmed by a late or missed diagnosis. In Oregon, 45–50 infants are identified each year, or about one infant per week.

- **Practitioners may believe the first test is of no value if obtained before 24–48 hours of age or before the infant has ingested protein or milk.**

FACT: 90 percent of affected infants have abnormal results on the first test regardless of age or food

ingestion. The ability to identify infants relates to a combination of the severity of their genetic lesion, physiologic and environmental factors, the quality of their screening specimen and the rapidity of the entire screening system to find them before they become symptomatic. There is nothing magic about a specific age cutoff as each disorder has its own “best screening window” and they are all different. Dietary intake does make a difference in galactosemia, in that a galactosemic infant on a soy based or other non-lactose formula would not be ingesting galactose and may have false negative results. Dietary information is helpful to evaluate the significance of certain abnormal results.

- **Practitioners may believe the second screen is unnecessary.**

FACT: Approximately eight states mandate a second screen for every infant and virtually all states recommend it if the first screen is drawn early. In Oregon and other states with repeat screens, 10 percent of all infants are routinely found only on the second screen.

Approximately four of the conditions on the screening panel may take a week or more to manifest themselves, long after an infant has been discharged and screened the first time. The efficacy of a second screen is controversial and under study by the U.S. Centers for Disease Control and Prevention.

- **Practitioners may believe the second screen is unnecessary if the first screen is collected when the infant is 5–7 days of age.**

FACT: More than 20 conditions on the screening panel can kill or maim in the first week to ten days of life. To delay screening would mean almost certain death or disability for these infants and a test at 5–7 days is too early to pick up many of the later onset disorders. A test at the end of the first week of life is better than no test, but is probably the worst time to collect it in terms of saving infant's lives and mental capacity.

- **Some practitioners believe the pain of specimen collection is so great for the infant that screening is not “worth it,” or that the infant will suffer permanent psychological damage.**

FACT: The pain of a heel stick can be minimized if not eliminated by using a scalpel bladed lancet device and by feeding the infant a small amount of sucrose water or breast milk before and during the procedure. Many families can attest that it is far more painful to have the infant die or to spend one’s life permanently affected because a few drops of blood were not collected at birth.

- **Practitioners may believe newborn screening is “just a piece of paperwork” to be gotten out of the way.**

FACT: Newborn screening represents one of the most successful public health programs ever undertaken in terms of prevention and cost savings, surpassing even immunizations. It is true that most programs do not do enough to alert practitioners to their importance and value to the screening process and to the number of infants identified.

- **Some practitioners tell parents that newborn screening is a test for “mental retardation.”**

FACT: Newborn screening tests for certain conditions that in some cases can lead to mental retardation. It is by no means a test for all sources of mental retardation and a normal screening result does not mean an infant will not be retarded due to other causes.

- **Many practitioners use the antiquated term “PKU test” which implies to parents that PKU is the only condition screened for.**

FACT: Over 30 separate disorders (including PKU) can now be screened for in the newborn period. PKU has not been the only disorder on most screening batteries since the mid-1970s when hypothyroidism was added. Newborn screening for PKU started in Oregon in 1961, and tests for galactose and leucine were added in 1966. The correct term is newborn screening (NBS).

### Timing of the Tests

In Oregon in 2006, 86 percent of all first samples were obtained before 48 hours of age, and 94 percent of

infants were screened twice. Most infants were initially screened between 24–48 hours of age, providing the best chance to pick up early onset disorders. Only a small number (0.9 percent) were screened at five days or later.

Early discharge and specimen collection at less than 48 hours is no longer the issue it once was thanks to improved technology in the OSPHL and increased compliance with second screens.

### Specimen Inadequacy

For the last couple of years, unsatisfactory specimens, particularly uneven saturation, have been increasing in the region and in 2007 represented 1.5 percent of all first specimens. This is triple the number of unsatisfactory specimens that usually occur in this region. These specimens generate an automatic request for a retest and delay screening, exposing affected infants to unnecessary delay in treatment and possible death or harm from their conditions.

This dramatic increase in unsatisfactory specimens is due to a number of factors. New technology in the OSPHL is very sensitive and requires a precise amount of blood to give reliable and correct results. As a result, the OSPHL staff have had to examine specimens more carefully than in the past and have had to reject more specimens. In the hospitals and birth centers many are still using capillary tubes or needles to collect specimens which is the primary reason for uneven saturation. New, inexperienced or temporary staff in hospitals are more likely to collect unsatisfactory specimens as hospitals cope with nursing shortages. Finally, educational efforts over the last several years have not been focused on specimen collection per se, as it has not been recognized as a large problem. Fortunately, by working together, the unsatisfactory rate for the region can be greatly reduced.

Hospitals and practitioners are immediately notified of inadequate specimens by e-mail and fax.

### Inadequate Demographic Information

In Oregon three percent of specimens are missing key patient data. For example, the specimen does not list the infant’s name, or it is unreadable. In the event of an abnormal result, these specimens are difficult and time consuming to match to the correct infant, especially in

large facilities. The most common cause for unreadable specimens is illegible handwriting. It is important to write clearly and neatly. Data which are critical include: name, birthdate/hour, sample date/hour, feeding status, mother's name, and practitioner's name.

### Problems Related to Specimen Transport to the Laboratory

All specimens should be transported as soon as they are dry and no later than 24 hours after collection. The screening laboratory should receive them within 24–48 hours and no later than five days after collection. About 5 percent of Oregon's specimens take longer than five days to reach the laboratory. While some are due to unavoidable postal delays such as holidays, most are caused by failure to mail the samples promptly. Samples may be delayed because of sluggish in-house mail, inefficient courier services or simple forgetfulness, but perhaps the most dangerous practice is "batch mailing," when samples are held up to a week so postage costs can be reduced.

Specimens collected on Sundays or Monday holidays are best stored in a cool room and sent express mail at the first opportunity.

The importance of early sample collection and prompt transit is illustrated by the fact that infants with galactosemia, organic acidemias and fatty acid oxidation disorders may die within a week or two of birth. Enzyme activity and hemoglobin may be destroyed or diminished in specimens which are older than 10 days or which have been exposed to heat and humidity.

### Educational Services

The charge for testing Oregon infants includes a specific amount for educational activities to improve the quality of the screening practices within the state. The educational activities include a quality assurance surveillance program, educational sessions at your facility regarding any or all aspects of screening and comprehensive review of your facility's screening system by the education coordinator/consultant. In addition, educational materials are provided to all practitioners and parents. These include this manual, parent brochures and disease fact sheets. All of the educational services are available at NO CHARGE to your facility. Additional educational

videotapes and/or DVDs listed below are approved by the Oregon NBS Program and can be purchased by individuals or facilities. Newborn screening is changing very rapidly and will continue to do so in the foreseeable future. Recommendations from a few years ago may be obsolete today. Practitioners moving into Oregon or one of the regional states must reorient themselves to newborn screening rules and regulations that pertain to that state. While states are making an effort to develop consensus about screening recommendations, there is still a lot of variation from program to program and practitioners must be aware of them.

**Screening Practice Surveillance:** A quality assurance program for birth facilities and home birth practitioners.

In an effort to assist hospitals, birth facilities and individual practitioners, the laboratory monitors some screening practices that can be ascertained from the screening cards (transit time, inadequate specimens, demographics omissions and timing errors). Screening Practice Profiles are provided on a monthly basis to hospitals and birth facilities in Oregon. A 2006 summary of the screening practices for Oregon is shown below.

<b>Screening Practice Profile 1st Specimens: Oregon Summary 2006</b>		
Number of specimens/month	3894	RANGE (3321-4435)
Percent error free	94.3%	(91.0-96.3)
Error profile		
Transit delays	3.0%	(1.6-5.1)
Timing errors	0.8%	(0.5-1.7)
Inadequate specimens	0.9%	(0.5-1.6)
Demographic errors	1.6%	(0.0-2.8)

*Practitioners' Manual and Condition Specific Fact Sheets can be downloaded online at: [www.oregon.gov/DHS/ph/nbs/index.shtm](http://www.oregon.gov/DHS/ph/nbs/index.shtm).*

## Additional Education Resources

CLINICAL LABORATORY STANDARDS INSTITUTE (CLSI, FORMERLY NCCLS)

### **LA04-A3-V** (PAL)

#### **Making a Difference Through Newborn Screening: Blood Collection on Filter Paper (PAL format)**

This video provides a visualization of each step in the blood specimen collection process and depicts the standard of practice, as defined by the CLSI consensus process, for collecting specimens on filter paper. It explains how to select and prepare the safest puncture site; choose the appropriate equipment; puncture the skin and apply blood to filter paper; care for the puncture site; identify and verify a valid specimen; and handle and mail the specimen to the laboratory. LA4-A4 accompanies the videotape along with laminated summary sheets. For more information, see this entry in the Immunology and Ligand Assay section. (25 minutes).

Member price: \$ 95.00

Non-member price: \$ 175.00

CustomerService@clsi.org

### **LA4-A4** (paper document or electronic document)

#### **Blood Collection on Filter Paper for Newborn Screening Programs; approved standard – Fourth Edition**

This document addresses the issues associated with specimen collection, the filter paper collection device, and the transfer of blood onto filter paper, and provides uniform techniques for collecting the best possible specimen for use in newborn screening programs. (Member price: \$60.00; Non-member price: \$120.00)

**Blood Specimen Collection and Handling** (poster): Demonstration in words and pictures, suitable for hanging in specimen collection area. Available at no charge from: Whatman, Inc.; 200 Park Ave, Suite 210; Florham Park, New Jersey 07932; Phone: 973-245-8300 or toll free 1-800-whatman; Fax: 973-245-8301; E-mail: info@whatman.com.

**Simple Spot Check** (poster): Demonstration with words and pictures examples of adequate and inadequate blood spots. Suitable for hanging in specimen collection area. Available at no charge from: Whatman, Inc.; 200 Park Ave, Suite 210; Florham Park, New Jersey 07932; Phone: 973-245-8300 or toll free 1-800-whatman; Fax: 973-245-8301; E-mail: info@whatman.com.



### Birth Facilities, Health Departments and Community Practitioners

1. In-service or medical/nursing rounds can be provided at your facility by newborn screening program staff. These presentations cover any (or all) aspects of the screening program and can be tailored to meet your needs and time frame. For example, physicians can request disease specific updates or hospital laboratory staff may want the specifics of specimen collection. It is recommended that facilities provide orientation sessions about newborn screening to all new employees, which should include review of this manual and have regular in-services from NBS personnel. These sessions are eligible for CEU credits.
2. Assistance with specific screening practice problems is available from the NBS educator or any program staff. These usually entail single-issue problems, such as a large number of unsatisfactory specimens or specimens delayed in transit. These problems are generally handled by phone and/or e-mail and are readily solved.
3. Policy development and/or review. We are available to assist administrators and/or practitioners in the drafting of sound screening policies and procedures for their facility or practice based on state laws, administrative rules as well as recommendations of the American Academy of Pediatrics and the Clinical Laboratory Standards Institute. It is recommended that all hospitals and birth centers have similar policies and procedures for NBS so that standards of care exist for all infants born in Oregon. This can be done in conjunction with in-service sessions or by e-mail or phone.
4. Newborn Screening System Review for Hospitals. This entails the formation of an ad hoc Quality Assurance (QA) Committee for NBS and includes representation from QA or risk management, nursing, medicine, laboratory, medical records, transportation, courier service, prenatal education and in some cases, community practitioners as well as the NBS educator as a consultant and/or

committee member. The purpose of the committee is to review the NBS system within a facility to ensure uniformity and best practices. It generally results in the rewriting of policies and procedures and reeducation of all staff. This is particularly useful for large facilities with a large number of employees and numerous hospital departments involved in NBS. Depending on the problems and the ease of implementing solutions, this process can take 4–12 months. The NBS Educator can attend meetings in person or by conference call.

#### **For additional information or to schedule a session please contact:**

Judi Tuerck, RN, MS  
 c/o Oregon State Public Health Laboratory  
 3150 NW 229th Ave., Suite 100  
 Hillsboro, OR 97124  
 E-mail: tuerckj@msn.com

## **Fees and Screening Kit Information**

Only a standardized, quality tested filter paper can be used for specimen submission (Whatman). Requests for kits must be made using a kit request form (see page 25) showing the quantity requested and must include a check for prepayment.

### **Orders and payment should be submitted to:**

Receipting Office  
DHS Public Health Division  
PO Box 14260,  
Portland, OR 97293-0260

Please allow 2–3 weeks for preparation and shipping. Kit orders and specimens are not to be sent by collect mail.

### **Type of Kits**

Hospitals, midwives and birthing facilities receive double kits with identical kit numbers, so that the first and second specimens can be matched easily. Health departments, clinics, and private practitioners may order double or single kits. Single kits should only be used when the second part of a double kit has been lost, damaged or infant is born out of state. Please note: all kits are precoded for the specific individual/facility; they must not be loaned to, or borrowed from, other facilities.

To obtain uncoded kits in emergency situations, contact Leanne Rien, RN at 503-693-4173.

### **Cost**

Double kits are \$54.00 per kit (two specimens) and single kits cost \$27.00. There is no charge to local health departments. Fee exemption is available for infants of impoverished parents.

### **Storage of Filter Paper Kits**

Store unused filter papers on their sides. The filter paper used for NBS is classified as a medical device. Filter paper can be compressed if stored lying flat and other objects are placed on top. Compressed filter paper will not absorb blood in a normal manner and will result in specimen rejection.

### **Cost Of Diagnostic Tests For Confirmation Of Abnormal Screening Results**

When diagnostic quantitative tests are requested by the laboratory or by the medical consultant for full evaluation of an infant considered likely to have one of the disorders, the costs of performing these will be absorbed by the program if the family has no insurance and provided that the samples are properly handled and submitted to a laboratory approved by this program. Fees for tests which are not specifically requested, or which are sent to other laboratories, will not be reimbursed.

The program does not pay for family studies.



## Oregon Request for Newborn Screening Kits

Per Oregon Administrative Rule 333-024-0240(4)(b), prepayment of newborn screening kits is required.



- 1. Completely fill out this form to ensure you will get the correct kits.**
- 2. Mail Request to:  
Department Of Human Services  
Fiscal Services  
PO Box 14260, Portland, OR 97293-0260**

ATTN: Facility Name: \_\_\_\_\_

Submitter Code: \_\_\_\_\_

Street Address: \_\_\_\_\_  
(NOT P.O. BOX)

City \_\_\_\_\_ State \_\_\_\_\_ Zip Code: \_\_\_\_\_

Telephone Number: \_\_\_\_\_ POrder #: \_\_\_\_\_ Ordered by \_\_\_\_\_

EFFECTIVE OCTOBER 14, 2002

***(Allow two weeks for delivery.)***

Number of Double Kits: \_\_\_\_\_ X \$54.00 = \$ \_\_\_\_\_

Number of Single Kits: \_\_\_\_\_ X \$27.00 = \$ \_\_\_\_\_

REVENUE CODE 2395  
FUND CODE: 76400 72636

Place Bar Code Here Verified \_\_\_\_\_  
(OSPHL USE ONLY)

**May be obtained at any time - No Charge:**

	Amount	Verified
English / Spanish Pamphlet	_____	_____
Manila Envelopes	_____	_____
Striped envelopes	_____	_____
Practitioner's Manual	_____	_____
Other	_____	_____

**OSPHL USE ONLY** Kit Numbers:

\_\_\_\_\_

\_\_\_\_\_

\_\_\_\_\_

\_\_\_\_\_

Date \_\_\_\_\_ By: \_\_\_\_\_ Reviewed by \_\_\_\_\_

**Questions? Call (503) 693-4100**

In compliance with the Americans with Disabilities Act (ADA), if you need this information in an alternate format, please call: Oregon State Public Health Laboratory at 503-693-4100. (Rev. 4/20/07)

## Exemptions

### Fees

In Oregon, no person is refused service because of the inability to pay the fee for testing. A practitioner or parent requesting exemption from fees must complete a statement on the back of the specimen form similar to the one shown below:

### Statement of Fee Exemption

The undersigned states that the parents of \_\_\_\_\_ are unable to pay the fee for testing for Newborn Screening disorders because of lack of sufficient funds.

\_\_\_\_\_  
**Parent or Practitioner**                      **Date**

Alternatively the practitioner must provide a list of exempt infants that includes each infant's name, birth date, the name of the parent or practitioner and date. The hospital/practitioner will be reimbursed for the cost of the screening kit.

### Religious Objections

A religious exemption can be claimed from the requirement for the newborn screening tests. In this event, the person otherwise responsible for submitting the specimen for testing is responsible for submitting a copy of the Informed Dissent form to the state laboratory signed by the infant's parent; see page 27.

### Parental Refusal

In the event a parent refuses the testing, the program strongly urges practitioners to obtain an "Informed Dissent" signed by the parent and placed in the infant's medical record. Suggested wording is provided on the next page.

**Newborn Screening Test Refusal**

Name of Infant \_\_\_\_\_

Birth Date \_\_\_\_\_

Medical Record Number \_\_\_\_\_

Hospital of Birth \_\_\_\_\_

Street Address \_\_\_\_\_

City/State/Zip \_\_\_\_\_

I have read the Department of Human Services brochure entitled Newborn Screening could save your Infant's Life. This brochure explains newborn screening for cystic fibrosis, metabolic, endocrine and hemoglobin disorders.

I have been told and I understand that state law requires screening for all infants born in Oregon because of the benefit to the infant and family of early detection and treatment of disorders on the screening panel.

I have been told and I understand that NBS detects over 30 disorders whose symptoms may not appear for several weeks or months.

I have been told and I understand that the risk of my infant having one of these conditions is approximately 1:900.

I have been told and I understand that untreated, these conditions may cause permanent damage to my child. If affected and not treated, my infant may suffer serious mental retardation, growth failure and in some cases death.

I have discussed the testing with \_\_\_\_\_ MD / RN.

He/She has explained and I understand all the risks involved if my child is not screened.

I have been informed and I understand the nature of the screening and how the screening sample is collected.

I object to newborn screening and I do not want \_\_\_\_\_ screened for these conditions.

I have freely made my decision without force or encouragement from my doctor, hospital personnel, or state officials.

Signed \_\_\_\_\_ Relationship \_\_\_\_\_

Witnessed by \_\_\_\_\_ Date \_\_\_\_\_

CC: OSPHL  
 Medical Records  
 Pediatrician / Primary Practitioner

# Core Conditions

## Cystic Fibrosis (CF)

### CF Essentials

- **Incidence:** 1:3,700
- **Screening Test:** Trypsinogen >60 meq/L on two filter paper specimens. (Trypsinogen is elevated in pancreatic insufficient neonates but disappears from the blood by ~2-3 months of age.)
- **Confirmatory test:** Sweat chloride test and/or DNA mutation analysis
- **Validity:** Approximately 5% of cases will be missed due to pancreatic sufficiency. All other cases should be abnormal on the first screen. Trypsinogen may be falsely abnormal in premature or sick infants.
- **Treatment:** Comprehensive care, enzyme and vitamin replacement, high calorie diet and pulmonary care
- **Outcome:** Early diagnosis decreases morbidity and need for hospitalization; improves growth and development; provides a pulmonary benefit to age 15; survival to third decade. Refer to pediatric pulmonologist.

Cystic Fibrosis (CF) is a recessively inherited defect of the cystic fibrosis transmembrane conductance regulator (CFTR) protein. Over 1,000 mutations of the CFTR protein have been identified, but a single mutation ( $\Delta F508$ ), accounts for two-thirds of all the mutations worldwide. The incidence of CF in the United States is approximately 1:3,700, being more frequent in non-Hispanic whites (1:2,500) and less so in non-Hispanic blacks (1:15,000). Newborn screening for CF is currently done for all Oregon, Alaska, Hawaii, Idaho, New Mexico and military infants.

### Clinical Features

Mutations in the CFTR gene alter the structure, function or production of the transmembrane chloride channel protein that is critical to the normal functioning of multiple organs. These include the lungs and upper respiratory track, pancreas, liver, sweat glands and genitourinary track.

The first symptom for 15–20 percent of infants with CF is meconium ileus, an intestinal obstruction that may require surgical correction and is also diagnostic of CF. Other symptoms of CF develop over time.

For infants without meconium ileus, the first symptoms include recurrent cough, wheezing, abdominal pain, loose stools and/or failure to thrive. Pancreatic insufficiency is present in 95 percent of CF individuals, and can lead to severe nutritional deficiencies and malnutrition. Recent studies suggest severely malnourished CF patients can suffer cognitive impairment as well.<sup>(7,8)</sup> Respiratory symptoms may be absent in the neonatal period but develop in most individuals by the end of the first year of life. Unfortunately most of the symptoms are not specific to CF and in the absence of newborn screening, meconium ileus or prenatal diagnosis, the average age at diagnosis is 14.5 months. Most patients suffer progressive lung damage. Survival has improved dramatically over the years and patients now live into their 30s. Like most inherited disorders there are milder variants with proportionally fewer symptoms and a longer course.

### Causes of CF

CF is a recessively inherited defect in the CFTR protein. CFTR deficiency results in abnormal chloride transport and the formation of thick sticky mucus, which in turn leads to organ dysfunction and failure.

### Laboratory Tests

The screening test measures trypsinogen, an enzyme produced in the pancreas that is transiently elevated in the blood of pancreatic insufficient CF infants at birth. This is detected by immunoreactive trypsinogen testing (IRT) test obtained from neonatal dried blood spots<sup>(9)</sup>.

An elevated IRT on two screening specimens is an indication for diagnostic workup.

### There are several issues to keep in mind regarding elevated IRT tests:

- Elevated IRT is not diagnostic of CF. False positives occur and are found in two out of three infants with high IRTs.

# Cystic Fibrosis

## Northwest Regional Newborn Screening Program

- Infants with meconium ileus may not have an elevated IRT, so this diagnosis should lead to definitive testing for CF regardless of the IRT result. All infants with meconium ileus, however, should have a newborn screening specimen collected even if CF is suspected, as they should be screened for the other conditions on the screening panel.
- About 5 percent of infants with CF, who are pancreatic sufficient, may not have an elevated IRT. Thus a normal IRT at birth does not completely rule out CF. Children with recurrent respiratory problems, failure to thrive, etc., may still need a CF work-up, and sweat chloride testing.

### Confirmation

CF can be diagnosed by two different methods. The gold standard remains the sweat chloride test in a certified CF center and is recommended for all infants with persistently elevated IRT. A chloride value in the sweat of >50 or 60 meq/L makes the diagnosis, while a value of <30 meq/L effectively rules out CF. For a minority of infants, chloride values will fall in an intermediate range (30–60 meq/L) and will need follow-up.

DNA mutation analysis of the CFTR gene is another diagnostic method; however this may be problematic in patients with atypical mutations. Approximately 70 percent of CF is caused by a single mutation,  $\Delta F508$ , but

there are over 1,000 other mutations associated with CF, and most are not included in standard multi-array DNA analyses<sup>(10,11)</sup>. Confirmation of two deleterious mutations confirm the diagnosis, but only one may indicate a carrier state or an affected individual with a private mutation on the second allele.

### Treatment

Treatment aims to ensure adequate nutrition and growth by supplementing pancreatic enzymes and vitamins and providing a high calorie diet. Daily chest physiotherapy and medications are required by most patients to loosen secretions. People with CF need rapid treatment of any chest infection with antibiotics. Routine immunizations plus inoculation against the flu and pneumococcus are recommended to help prevent chest infections. Infants should be referred to a CF center.

### Screening Practice Considerations

- CF infants with meconium ileus or who are pancreatic sufficient may have normal IRT levels.
- IRT levels in affected infants will decline and be in the normal range by 3 months. Older infants or children suspected to have CF should have a sweat chloride test.
- IRT may be falsely elevated in premature or sick infants.

Results	Likely Causes	Actions
Two elevated IRTs on filter paper specimens	* Cystic fibrosis probable * Possible false positive	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***

## Congenital Adrenal Hyperplasia (CAH)

### CAH Essentials

- **Neonatal Emergency:** 2/3 will develop salt wasting crisis, which can be fatal, in the first week to month of life.
- **Incidence:** 1:12,000 newborns; 1:300 Alaskan Natives (Yupik Eskimo)
- **Screening test:** 17-OH-Progesterone
- **Validity:** 90% identified on 1st screen, 10% on 2nd screen
- **Causes:** 21-hydroxylase deficiency or other inborn error of cortisol synthesis; recessive inheritance
- **Treatment:** Hydrocortisone and mineralocorticoids
- **False positives:** Occur more frequently in premature, low birth weight or sick infants
- **Outcome:** Early detection and treatment can be lifesaving. Chromosome analysis in infants with ambiguous genitalia will prevent gender mis-assignment. Ultimate outcome depends on severity of defect, days to treatment and adherence. Refer to pediatric endocrinologist.

CAH is an inherited defect of cortisol synthesis. The adrenal gland cannot make cortisol and overproduces male hormones. Without cortisol, infants are at risk for adrenal crisis and may be unable to regulate salt and fluids, and can die. The most common disorder is 21-hydroxylase deficiency. The incidence is 1:12,000 live births. The incidence is 1:300 in certain Yupik Eskimo populations. CAH is screened for in all infants in our region.

### Clinical Features<sup>(12)</sup>

Infants may be symptomatic at birth. By 4 to 5 months' gestation, diminished cortisol production stimulates the fetal pituitary gland to produce ACTH and excessive adrenal androgens. The androgens virilize female

external genitalia, but ovaries and uterus are unaffected. Male infants may have increased scrotal pigmentation or may be asymptomatic.

In two-thirds of cases, the 21-hydroxylase deficiency causes reduced production of mineralocorticoids. This leads to a hypotensive, hyperkalemic, salt-losing crisis with rapid onset of adrenocortical failure within 7–28 days of birth. This can be fatal. In one-third of cases, the infant has a “non-salt losing” or “simple virilizing form.” If untreated, these children have mild postnatal virilization, rapid growth with advanced skeletal age, early puberty, and short stature as adults. In adulthood, there is hirsutism and acne. Women have irregular menses and infertility.

### Causes of CAH

The term “congenital adrenal hyperplasia” or “adrenogenital syndrome” covers a group of disorders. All are due to an inborn error of steroid hormone synthesis, which blocks the production of cortisol. The low level of cortisol stimulates ACTH, causing adrenal hyperplasia and increased secretion of steroid precursors. Different enzyme defects block the metabolic pathway at different sites and result in different clinical features. There are variants to this disorder, which have later onset. All forms of CAH are inherited as autosomal recessive disorders.

### Laboratory Tests

Screening is based on an immunoassay for a precursor steroid, 17-hydroxyprogesterone (17-OHP). Affected infants have high levels of 17-OHP. Infants with milder disorders have intermediate levels. False positives may occur in preterm, low birth weight and sick infants.

### Confirmation

Confirmation is by measurement of serum 17-OHP and if salt wasting is suspected, sodium, potassium and plasma rennin activity. Chromosome analysis to confirm gender if genitalia are ambiguous.

### Treatment

Infants should be treated with hydrocortisone and mineralocorticoids in consultation with a pediatric endocrinologist.

# Endocrine Conditions

## Northwest Regional Newborn Screening Program

### Screening Practice Considerations

- This disorder kills quickly and is a neonatal emergency. In both sexes, salt wasting and shock may develop rapidly within 7–28 days of birth. Collect specimens between 24–48 hours of life. Transport all specimens 4–12 hours after collection and no later than 24 hours.
- Female infants who are virilized or infants with ambiguous genitalia should be considered at risk for this condition, tested at birth, and monitored for electrolyte abnormalities until the diagnosis is excluded.
- Male infants are not usually recognized at birth.
- About 10 percent of infants will be detected only on a second screen<sup>(13,14,15)</sup>.

Results	Likely Causes	Actions
17-OHP >200 ng/mL, 24 hrs–≤10 days: BW ≤2500 gms. 17-OHP >150 ng/mL, 24 hrs–≤10 days: BW >2500 gms. 17-OHP >125 ng/mL, >10 days: any BW	* CAH probable * False positive	Neonatal emergency; NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
17-OHP >200 ng/mL, <24 hrs: any BW	* CAH possible * False positive due to early collection and/or stress.	Medical consultant reviews, NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations, if needed.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***



## Congenital Hypothyroidism

### CH Essentials

- **Incidence:** 1:3,000 newborns
- **Screening test:** T4 (thyroxine) and TSH (thyroid stimulating hormone)
- **Validity:** 90% identified on 1st screen, 10% on 2nd screen
- **Causes:** Thyroid dysgenesis: 85%; hereditary inborn error of thyroid hormone biosynthesis: 15%.
- **Treatment:** normalize T4 by 2 weeks of age; TSH by 1 month
- **False positives:** Use of iodine on infant and/or mother who is breast-feeding; collection within 24 hours of birth; premature or ill infants
- **Outcome:** Can be normal, but depends on severity of thyroid deficit, days to treatment and adherence to treatment. Infants with just a 2-week delay in reaching a serum T4 >10 ug/dL may have up to a 10 point drop in IQ.<sup>(16)</sup>

Congenital hypothyroidism (CH) occurs in infants who are born without the ability to produce adequate amounts of thyroid hormone. Thyroid hormone is important for normal function of all of the body's organs and is essential for normal brain development. The incidence of congenital hypothyroidism is 1:3,000. CH is more common in Hispanic and Native American populations (1:700–2,000). There is a 2:1 female/male ratio thought to be due to an autoimmune risk factor. Infants with Down's syndrome have increased risk of CH (1:140 newborns).

### Clinical Features<sup>(17)</sup>

Deficiency of thyroid hormone in an infant may result in mental retardation and other signs of brain damage if it is not diagnosed and corrected by 2–3 weeks of life. Many infants with CH may appear clinically normal before 3 months of age, by which time some brain damage has usually occurred. Laboratory test results are the only reliable means of diagnosing CH in the newborn.

When symptoms or signs are present, they may include prolonged neonatal jaundice, constipation, lethargy and poor muscle tone, feeding problems, a large tongue, puffy face, large fontanel, distended abdomen and umbilical hernia. Approximately 10 percent of cases will have other congenital abnormalities, usually cardiac defects. Long-term neurologic damage includes mental retardation, ataxia, fine and gross motor delay, slow growth, speech disorders and hearing deficits in 20 percent. Since thyroid deficiency can occur at any age, normal tests in the newborn period do not exclude deficiency in an older infant or child.

### Causes of Congenital Hypothyroidism

The most common causes are total or partial failure of the thyroid gland to develop (aplasia or hypoplasia), or its development in an abnormal location (an ectopic gland). Less commonly, hypothyroidism is induced by medications (antithyroid drugs or excess iodine) in the mother, or maternal autoimmune thyroid disease with transfer of a maternal antibody that blocks the fetal thyroid development.

### Laboratory Tests

The initial screening test is the T4 assay. Each day 10 percent of samples with the lowest T4 results are further tested by a screening TSH assay. Different combinations of results are possible; see table below.

When the infant's physician is notified that screening results are abnormal, blood should be collected by venipuncture as soon as possible to confirm the abnormal screening results. In the case where the T4 is low and TSH is elevated, treatment can be started as soon as the serum is obtained, pending final confirmation. If the serum thyroid function tests confirm hypothyroidism, further diagnostic studies, such as a thyroid ultrasound examination or scan and X-ray to assess skeletal maturation, may be desirable to determine the type, age of onset and severity of hypothyroidism. Generally, these studies do not change management and so are optional.

### Thyroid Function in Premature Infants

In premature infants, there is a physiological reduction in blood T4 levels; TSH levels are not elevated in this situation. These cases need special observation to ensure that the low T4 levels rise into the normal range as the

# Endocrine Conditions

## Northwest Regional Newborn Screening Program

infant matures, but this may take several weeks. Serum free T4 levels (by equilibrium dialysis method) are often normal. There is no consensus that these infants benefit from thyroid supplementation during this period.

### Treatment

The American Academy of Pediatrics (AAP) recommends that infants be managed in consultation with a pediatric endocrinologist.<sup>18</sup> Treatment of CH is effective if done correctly. Thyroxine (e.g. Synthroid, levoxyl or levothyroid, or generic l-thyroxine), in pill form, is crushed, mixed with water or expressed breast milk and administered once daily. Currently, there are no FDA-approved liquid formulations. The recommended starting dose is 10–15 g/kg of body weight daily, usually 37.5 g/kg to 50 g/kg. AAP recommendations for follow-up serum T4 (or free T4) and TSH are as follows:

- Initiation of treatment and 2–4 weeks later
- Every 1–2 months in the first 6 months
- Every 3–4 months from 6 months–3 years of age
- Every 6–12 months from age 3–end of growth period

**Treatment Goals:** Maintain serum T4 or free T4 in the upper half of the normal range (10–16 µg/dL for T4 or 1.2–2.4 ng/dL for free T4), and TSH normalized (<10

µIU/mL). Clinical evaluations can occur less frequently. As infants grow, the dose of thyroxine is increased. Periodic developmental testing should be done on all patients. If treatment is started early and thyroid levels are monitored closely, development remains normal.

### Screening Practice Considerations

- Hypothyroidism is the most common disorder covered by the program.
- Ninety percent of hypothyroid infants are detected on the first specimen; in 10 percent of cases, hypothyroidism develops in the weeks after birth and is detected on a second screening test as production of thyroid hormone dwindles after birth<sup>(19,20,21)</sup>.
- Some infants (usually pre-term) will manifest a delayed rise in TSH, and so are also detected on the routine second screening test. Practitioners therefore must remain alert to clinical symptoms in older infants despite normal initial screening.
- False positive results may occur if the specimen is collected within the first few hours after birth, as the TSH rises in response to the extra-uterine environment.
- Topical iodine use on the infant or a mother who is breastfeeding may cause transient hypothyroidism.

Results	Likely Causes	Actions
T4 low/TSH elevated	* Hypothyroidism probable * False positive	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
T4 low/ TSH <100–200 µIU/mL, 0–11 hrs.	* Prematurity hypothyroidism possible * False positive	NWRNSP contacts practitioner by phone and send letters requesting further testing by mail.
T4 low/TSH normal (on two specimens unless premature)	* Thyroid binding globulin (TBG) deficiency * False positive * Pituitary gland problem with secondary hypothyroidism * Prematurity	NWRNSP contacts practitioner by mail requesting further testing.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***

## Sickle Cell Disease and other Hemoglobinopathies

### Sickle Cell Disease Essentials

- **Incidence:** 1:5,200 births; 1:400 African Americans
- **Screening Test:** Isoelectric focusing (IEF), high performance liquid chromatography (HPLC)
- **Confirmatory Tests:** IEF and/or HPLC
- **Validity:** 100% found on 1st screen (unless transfused)
- **Treatment:** Comprehensive care, prophylactic penicillin, immunizations, and empiric treatment of febrile episodes. Refer to pediatric hematologist.
- **Outcome:** Screening prevents death from sepsis in most infants. Long-term outcome depends on the severity of the hemoglobinopathy and response to treatment.

The primary goal of hemoglobinopathy screening is diagnosis of significant sickling hemoglobinopathies in the neonatal period, before symptoms occur. Newborn diagnosis of sickle cell disease, if coupled with family education and centralized comprehensive care, can markedly lower morbidity and mortality<sup>(22)</sup>.

Homozygous sickle cell disease (SCD) occurs when the recessive gene for hemoglobin S, sickle hemoglobin, is inherited from both parents. The term “clinically significant sickling syndrome” also includes conditions resulting from inheritance of one gene for hemoglobin S and certain other unusual hemoglobins, such as beta thalassemia or hemoglobin C. These doubly heterozygous conditions tend to be less severe than SCD, though all are capable of producing severe complications. The incidence of SCD in the African American population is 1:400, but also occurs at a lower frequency among all other ethnic groups. The disease incidence in a population depends on the population’s ethnic composition.

### Clinical Features

Sickle syndromes are systemic diseases and may affect any organ. They are characterized clinically by chronic hemolysis, intermittent vaso-occlusion and marked variability. Some patients experience unremitting complications, others lead a full and productive life. Early manifestations are often life-threatening and include overwhelming infection due to splenic dysfunction, splenic sequestration crisis, and aplastic crisis with profound anemia. Prior to newborn diagnosis and preventive care, mortality in the United States was 8–30 percent in the first three years of life. Some important complications include vaso-occlusive pain syndromes, osteomyelitis, acute chest syndrome, stroke, priapism, pyelonephritis, gallstones, skin ulcers, retinopathy and decreased life expectancy.

Other significant hemoglobinopathies are less common and even more variable. Their manifestations range from very mild chronic hemolysis to severe dyserythropoiesis requiring a lifetime of transfusion support. Early detection of these less common conditions, however, may prevent unnecessary diagnostic and therapeutic intervention.

### Laboratory Tests

All first filter paper samples are screened for hemoglobinopathies using isoelectric focusing (IEF). Various hemoglobin patterns occur. If an abnormality is detected, the sample is reanalyzed using high performance liquid chromatography (HPLC). If a hemoglobin abnormality is detected on the first filter paper sample, the second filter paper sample is also analyzed by IEF and HPLC. Thus, each hemoglobin abnormality is verified four times, using two different techniques on two different samples. Solubility tests (Sickle-dex, Sickle-prep, etc.) are never appropriate in infancy and should not be used to confirm screening results.

### Treatment

Infants with significant hemoglobinopathies should have a primary care provider and receive periodic evaluation in a comprehensive care setting. Therapy begins with education of care-givers and includes prophylactic

# Hemoglobin Conditions

Northwest Regional Newborn Screening Program

Results	Likely Causes	Actions
FS (absence of A)	* Sickle cell disease or Sickle beta thalassemia	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
FSC (absence of A)	* Sickle hemoglobin SC disease	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
FC (absence of A)	* Homozygous C disease	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
FE (absence of A)	* Homozygous hemoglobin E or Hemoglobin E-beta thalassemia	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
FAS	* Sickle cell carrier or * Sickle beta thalassemia or * Sickle cell disease in transfused infant	NWRNSP will report by letter regarding test results and any other recommendations.
FAC	* Hemoglobin C carrier * Homozygous C disease in a transfused infant	NWRNSP will report by letter regarding test results and any other recommendations.
FA+slow band	* Possible carrier for hemoglobin E, O, D, or G	NWRNSP will report by letter regarding test results and any other recommendations.
FA+fast band	* Possible alpha thalassemia * Bart's hemoglobin is a marker for alpha thalassemia	NWRNSP will report by letter regarding test results and any other recommendations.
F only	* Premature infant or * Beta thalassemia major	NWRNSP will report by letter regarding test results and any other recommendations.
Predominance of A	* Transfused infant or * Patient outside of neonatal age range	NWRNSP will report by letter regarding test results and any other recommendations.

penicillin, prompt evaluation and empirical treatment of any febrile illness, and immunizations including those for encapsulated bacteria. Close attention is necessary for the common problems of poor growth, recurrent pain and febrile illnesses. Organ-specific complications, sedation and general anesthesia require special attention. Other treatments, including the use of blood products and investigational therapies depend on clinical course.

## Carrier Detection Makes Hemoglobin Screening Different

This is the only newborn screening test that regularly identifies carriers (heterozygotes) as well as those affected by a given disease. In fact, many more carriers than disease states are identified for all hemoglobinopathies. If both parents are carriers of an autosomal recessive genetic trait, the risk of any infant

of that couple being homozygous is 1:4. While the best way to handle this genetic information has yet to be agreed upon, several principles are currently operative:

- 1) The family is entitled to the information and it is private.
- 2) If both parents of a SCD carrier infant are African American, they have at least a 25 percent risk of having a subsequent child with SCD, because at least one of them is now known to be a carrier. The family should be offered testing and genetic counseling. If the family declines participation, this should be documented.
- 3) The abnormal screening results may need to be confirmed using a liquid blood specimen. The medical consultant for hemoglobinopathy screening is available for assistance.

## Screening Practice Considerations

- Newborn screening for hemoglobinopathies is not done on the second specimen unless an abnormality has been identified on the first specimen. It is crucial to use the first kit for the first test; the cards are not interchangeable.
- Transfusion of red blood cells prior to drawing the newborn screening specimen will invalidate the hemoglobinopathy test. Obtain a specimen before any transfusion.
- Some hemoglobinopathies, particularly the thalassemias, are not reliably detected through newborn screening and a normal screening result does not eliminate the possibility that a patient has a hemoglobinopathy. Further testing or consultation should be sought if indicated by clinical suspicion.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***

## Amino Acid Conditions: Hypermethioninemia

### Homocystinuria (cystathionine beta-synthase deficiency) \*

#### Homocystinuria Essentials

- **Incidence:** 1:100,000
- **Screening Test:** Methionine by MS/MS
- **Confirmatory Tests:** Quantitative methionine, homocystine in blood and urine
- **Validity:** 20% 1st screen; 80% 2nd screen
- **Treatment:** Pyridoxine if responsive; if not responsive, low methionine diet with cysteine and betaine supplements
- **Outcome:** Excellent if treated early and adherence is good.

The most common form of genetic homocystinuria is cystathionine beta-synthase deficiency (CBS). CBS is required for conversion of methionine to cysteine and deficiency results in the accumulation of homocystine, methionine and cysteine-homocystine disulfides in the blood and urine. Unfortunately, methionine rises

slowly in affected infants and may not be detectable on specimens obtained in the first few days after birth. Homocystinuria is inherited as an autosomal recessive trait and occurs in approximately 1:100,000 births.

#### Clinical Features <sup>(29,30)</sup>

Untreated patients appear normal at birth, but by the first or second year mental retardation may be apparent, most will develop dislocation of the lenses and a marfanoid body habitus, osteoporosis, and ultimately thrombo-embolism may develop which can result in stroke and serious and permanent disabilities or death.

#### Methionine Adenosyltransferase (MAT) Deficiency

A number of infants in the United States, identified through newborn screening with persistently elevated methionine, have been shown to have MAT deficiency. All but two patients have been asymptomatic, with normal growth and development. Two patients have had demyelination of the brain, but it is not clear that this is a result of MAT deficiency or other causes.

#### Laboratory Test

Elevation of methionine is detected by MS/MS; normal methionine levels are <90 µM/L. Transient elevations of plasma methionine in the newborn are unusual unless the infant is premature, has liver disease and/or is receiving IV amino acid preparations. (i.e., TPN, hyperalimentation).

\* Not all forms of hypermethioninemia or even all cases of CBS deficiency will be detected by MS/MS.

Results	Likely Causes	Actions
Methionine > 120 µM/L	<ul style="list-style-type: none"> <li>* Homocystinuria/MAT deficiency possible</li> <li>* Tyrosinemia, Type I, galactosemia</li> <li>* Liver disease</li> <li>* Hyperalimentation</li> <li>* High protein diet</li> <li>* False positive</li> </ul>	NWRNSP requests repeat filter paper specimen by mail.
Methionine > 90 –120 µM/L	<ul style="list-style-type: none"> <li>* Homocystinuria/MAT deficiency probable</li> <li>* Tyrosinemia, Type I</li> <li>* Liver disease</li> <li>* Hyperalimentation</li> <li>* High protein diet</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.

## Treatment

Some patients will respond to pyridoxine in large doses (250–1,200 mg/day). For patients unresponsive or partially responsive to pyridoxine, a methionine restricted diet is supplemented with cysteine and betaine is usually effective. The outcome for treated patients is dependent on the age at diagnosis, adherence with therapy and severity of defect. For those with good compliance, outcome is normal.

## Screening Practice Considerations

- Methionine rises slowly in affected infants, so that the first screening specimen may be normal; 80 percent of the homocystinuria patients detected in the NWRNSP have been found on routine second tests.
- Methionine may be elevated secondary to liver disease, prematurity, TPN or hyperalimentation.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***



## Phenylketonuria (PKU) & Hyperphenylalaninemia

### Hyperphenylalaninemia Essentials

- **Incidence:** 1:10,000 births
- **Screening Test:** Phenylalanine >190 µM/L by MS/MS; phenylalanine/tyrosine ratio >3.0
- **Confirmatory Tests:** Quantitative amino acids; bipterins in blood and urine
- **Validity:** >99% on 1st screen
- **Treatment:** Low phenylalanine diet; bipterin supplementation
- **Outcome:** Normal if treated early and adherence is good.

Detection of elevated phenylalanine levels requires urgent follow-up. This disorder is due to a recessively inherited enzyme defect in which the body cannot use the amino acid phenylalanine properly. All other metabolic processes are intact, but phenylalanine, which comes from all dietary protein, accumulates in the blood to toxic levels. All forms of hyperphenylalaninemias from mild to severe and including bipterin defects are inherited as autosomal recessive disorders.

### Clinical Features

Infants with PKU seem to be normal for many months; however, without treatment, severe mental retardation, seizures, eczema and other problems usually develop. In older untreated patients the skin and hair may be fair, the eyes may be blue and a mousey odor of the skin or urine is common. Untreated blood phenylalanine level is often over 1,200 µM/L in infants with severe PKU. Overall, PKU occurs in about 1 in 10,000–15,000 Caucasian and Hispanic births and is less common in other races. Although severe mental deficiency is the rule in untreated cases, occasional asymptomatic adults are found with normal or near normal intelligence, despite high phenylalanine levels.

Plasma phenylalanine is not detectably elevated in cord blood. It starts rising within 24 hours after birth and often reaches 1,200 µM/L or more within a few days.

The screening test is often abnormal within 24 hours and almost uniformly abnormal within 48 hours of birth. The phenylalanine/tyrosine ratio is uniformly abnormal in true cases and can be used to differentiate false positive infants.

### Variant Forms of PKU (Hyperphenylalaninemia)

There are several intermediate forms of hyperphenylalaninemia in which the plasma phenylalanine levels are lower than in classic PKU (180–1,200 µM/L). In these cases, mental retardation is variable and, in the milder variants, is completely absent. In infancy, these patients can mimic severe PKU, and for adult women the risk of maternal PKU syndrome increases in proportion to the plasma phenylalanine.

Some forms of hyperphenylalaninemia are caused by defects of the cofactor bipterin metabolism and blood phenylalanine levels are variable. These patients have progressive neurological damage with seizures and steady deterioration that becomes noticeable sometime between 6 and 20 months of age despite early treatment with a low phenylalanine diet. Definitive tests can differentiate these variant forms of PKU. In view of the severity of this group of diseases, all infants with persistently abnormal levels of phenylalanine must have testing by special blood and urine tests for bipterin abnormalities. Information regarding this testing is provided through the metabolic consultants.

### Maternal PKU and Hyperphenylalaninemia

Women with significant hyperphenylalaninemia have an increased risk of miscarriage and their offspring (who usually do not have PKU) may have intra-uterine growth retardation that persists postnatally. More than 90 percent of infants of untreated mothers with classical PKU have microcephaly, mental retardation, and/or congenital heart defects. They have a transient elevation of phenylalanine (240–1,200 µM/L) that falls to normal within 24 hours. A screening test on the mothers of infants with transient hyperphenylalaninemia, particularly if the infant's sample was collected in the first 24 hours after birth, is recommended. A phenylalanine restricted diet begun prior to conception and during pregnancy can often prevent damage to the fetus. Most childbearing women today, if

born in the United States, would/should have been screened as infants, so the chances of undiagnosed hyperphenylalaninemia are remote but still present.

### Laboratory Tests

PKU and hyperphenylalaninemia are detected using tandem mass spectrometry (MS/MS); the normal phenylalanine level is  $< 190 \mu\text{M/L}$  and the phenylalanine/tyrosine ratio is  $< 3.0$ .

### Treatment<sup>(31,32,33,34)</sup>

With proper treatment, mental retardation is totally preventable. Treatment should be started as soon after birth as possible (preferably in the first week) in any infant recommended for treatment by the consultants and should be continued indefinitely. Frequent monitoring is required, especially in the first weeks, because variant forms of hyperphenylalaninemia may be indistinguishable from classic PKU and improper nutritional therapy can be fatal.

If treatment is not started for some weeks, the results are more variable and the IQ tends to be lower. Patients whose treatment begins after 6 months are likely to remain mentally retarded. Older patients usually show little change in IQ with treatment, but a low phenylalanine diet may help to control serious behavior problems.

### Screening Practice Considerations

- Detection may depend on the amount of protein ingested or endogenously produced by the infant, but most affected infants (90 percent) have abnormal results even in the first 24 hours of life regardless of intake. Those with milder forms of hyperphenylalaninemia require longer periods of feeding or catabolism to develop abnormal tests.
- Contamination of the filter paper with food or liquids containing NutraSweet® (Aspartame) may cause false positive results or an inadequate specimen.

Results	Likely Causes	Actions
Phenylalanine $> 190 \mu\text{M/L}$ , Phe/Tyr $> 3.0$	<ul style="list-style-type: none"> <li>* PKU possible</li> <li>* Variants forms of PKU</li> <li>* Mother has PKU</li> <li>* False positive</li> <li>* Transient hyperphenylalaninemia</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***

## Tyrosinemia, Type I, II and Transient \*

### Tyrosinemia Essentials

- **Incidence:** 1:100,000 (Types I & II) (1:1,000 transient)
- **Screening Test:** Tyrosine > 400 µM/L by MS/MS
- **Confirmatory Tests:** Succinylacetone, blood amino acids, enzyme and mutation analysis
- **Validity:** < 20% identified on either screening test
- **Treatment:** Low protein phe/tyr diet, medications and possible liver transplant in type I; low phe/tyr diet in type II; transient tyrosinemia resolves within a month or two of birth or Vitamin C supplements for a few days will shorten the time.
- **Outcome:**  
**Type I:** NTBC stops progression of disease and allows normal growth and development. The long-term risk of liver adenomas is still unknown, prompting some families to opt for liver transplant.  
**Type II and Transient:** Normal outcome

Elevated tyrosine may result from an inherited defect of tyrosine catabolism or, as in transient tyrosinemia, delayed maturation of liver enzymes or liver disease.

### Transient Tyrosinemia<sup>(35)</sup>

Transient tyrosinemia of the newborn is common (1:1,000) and more common among Inuit and Eskimo populations in Alaska. Transient tyrosinemia is thought to arise from delayed maturation of the liver enzyme, 4-hydroxyphenylpyruvic acid dehydrogenase (4HPPD), coupled with increased protein intake and/or occult ascorbic acid deficiency. Tyrosine levels may be quite high (>400 µM/L) peaking at 14 days of life and resolved by 1 month. Premature infants or those on hyper-alimentation may have prolonged hypertyrosinemia.

### Clinical Features

Transient tyrosinemia of the newborn may present with lethargy or decreased motor activity, but is usually a biochemical abnormality found in an otherwise normal newborn. Transient tyrosinemia is not associated with long-term sequelae, although this has not been systematically studied.

### Treatment

Transient tyrosinemia, while probably benign, may in some cases be treated with protein restriction to 2g/kg/day and administration of ascorbic acid (50–200 mg/day orally for 5–7 days) to infants found to have transient tyrosine (after types I & II are excluded). If the infant is breastfeeding, ascorbic acid alone may be crushed, dissolved in water and administered orally. Ascorbic acid, a co-factor for 4HPPD, helps to increase the enzyme's activity which will resolve the hypertyrosinemia more quickly if there are concerns about the infant's status.

### Hepatorenal Tyrosinemia, Type I<sup>(35)</sup>

Tyrosinemia, type I or fumarylacetoacetate hydrolase (FAH) deficiency occurs in 1:100,000 births. Hepatorenal tyrosinemia is inherited as an autosomal recessive trait.

### Clinical Features

Tyrosinemia, type I causes severe liver and renal disease and peripheral nerve damage. Presentation in infancy includes vomiting, lethargy, diarrhea and failure to thrive. Liver disease with hepatomegaly, hypoproteinemia, hyperbilirubinemia, hypoglycemia and coagulopathy may be present. In an international survey of 108 patients, 13 percent (n=14) became symptomatic in the first two weeks of life and 36 percent (n=39) in the first two months. Renal proximal tubular dysfunction results in aminoaciduria, hyperphosphaturia and hypophosphotemic rickets. Untreated, death in infancy or childhood from acute liver failure, neurological crises, or hepatocellular carcinoma is usual.

### Treatment

Therapy with oral NTBC [2-(nitro-4-trifluoromethylbenzoyl)-1-3-cyclohexanedione] blocks the formation of the toxic metabolites. NTBC is effective in preventing or halting liver and renal damage and averting acute neurological crises. Long-term ability of NTBC to prevent the development of hepatic carcinoma is yet unknown, but 4/11 patients on NTBC have developed adenomas in a recent study<sup>(36)</sup>. The ultimate treatment, liver transplantation, has been successful in many cases. Adjunct therapy with dietary restriction of phenylalanine and tyrosine as well as symptomatic treatment of clotting defects, rickets and proximal tubular losses may also be needed.

## Occulocutaneous Tyrosinemia

Tyrosinemia, type II is caused by a deficiency of the enzyme tyrosine aminotransferase (TAT) and is inherited as an autosomal recessive trait. TAT deficiency is rare, with about 100 cases described worldwide, although more infants may be identified as MS/MS screening continues to be implemented.

### Clinical Features<sup>(35,37)</sup>

TAT is manifested primarily in the eyes, the skin and the central nervous system. In the eyes, tyrosine crystals accumulate, resulting in painful corneal erosions. Equally painful hyperkeratotic plaques develop on the plantar surfaces of hands, feet and digits. Symptoms usually develop in the first year of life, but have been present on the first day of life or not occur until adulthood. A variable degree of mental retardation is present in about 50 percent of cases.

### Treatment

A diet restricting phenylalanine and tyrosine is effective in clearing and/or preventing ulcerations.

## Laboratory Tests

Tyrosinemia is detected using MS/MS; the cutoff tyrosine level is <400 µM/L. There is considerable overlap in tyrosine levels between normal infants, those with transient tyrosinemia and affected infants, making the tyrosine level itself not very specific.

Clinical correlation, blood amino acids and urine succinylacetone are necessary to differentiate these cases. More sensitive and specific tests for succinylacetone are under development and these will improve NBS for tyrosinemia, as they become available<sup>(38)</sup>.

## Screening Practice Considerations

Tyrosine may be slow to rise in affected infants, making it more likely to be found on routine second testing. Practitioners must remain alert to the possibility of tyrosinemia in any infant with liver disease, corneal or keratotic lesions.

\* Not all cases of tyrosinemia will be detected by newborn screening.

Results	Likely Causes	Actions
Tyrosine > 600 µM/L; first NBS Tyrosine ≥ 400 µM/L and >600 µM/L; first NBS Tyrosine ≥ 400 µM/L; second NBS Tyrosine ≥ 600 µM/L; second NBS	* Transient tyrosinemia * Tyrosinemia possible * Liver disease * Hyperalimentation * False Positive	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
Tyrosine ≥ 400 µM/L and ≤ 600 µM/L;	* Transient tyrosinemia * Tyrosinemia possible * Liver disease * Hyperalimentation * False positive	NWRNSP requests repeat filter paper by mail.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***

## FATTY ACID OXIDATION (FAO) CONDITIONS

### FAO Condition Essentials

- **Neonatal Emergency:** Approximately 10% of infants with FAO disorders will die in the first few days after birth, generally before screening results are known.
- **Incidence:** 1:6,000 births
- **Screening Test:** Acylcarnitines by MS/MS
- **Confirmatory Tests:** Acylcarnitine profiles, enzyme assay and/or mutation analysis
- **Validity:** 90% on the 1st screen, 10% on the 2nd screen
- **Treatment:** Avoid fasting, low fat diet
- **Outcome:** Variable depending on the FAO. MCAD patients do well if caught early and episodes are prevented. Outcome for other FAOs is still unknown.

Mitochondrial beta-oxidation of fatty acids is crucially important in the body's ability to produce energy during fasting. In infants, a "fasting" state can be produced in as little as four hours. Fatty acids must be transported into the cytoplasm and then into the mitochondria for oxidation; carnitine is required for these transport steps. Once in the mitochondria, fatty acid chains 4-18 carbons in length must be oxidized, two carbons at a time, each reaction using a chain-specific enzyme, before ketogenesis can occur. There are over 20 individual steps in beta-oxidation some with multiple enzyme complexes. An enzyme block anywhere in this process or a carnitine deficiency will result in hypoketotic hypoglycemia and tissue damage related to the toxic accumulation of unoxidized fatty acids. At least 16 separate enzyme disorders have been identified, many of which may be identified by MS/MS by measuring the accumulation of various acylcarnitines.

### Fatty acid oxidation disorders\*

- Carnitine transport defect (enzyme unknown) (CUD)
- Carnitine/acylcarnitine translocase (CT) deficiency
- Carnitine palmitoyl transferase I (CPT I) deficiency
- Carnitine palmitoyl transferase II (CPT II) deficiency
- Very long chain acyl-CoA dehydrogenase (VLCAD) deficiency
- Long chain L-3 hydroxyacyl-CoA dehydrogenase (LCHAD) deficiency
- Medium chain acyl-CoA dehydrogenase (MCAD) deficiency
- Short chain acyl-CoA dehydrogenase (SCAD) deficiency
- Multiple acyl-CoA dehydrogenase deficiency (MADD aka GA II)
- Trifunctional protein (TFP) deficiency

MCAD is the most common, approximately 1:15,000 births; LCHAD is less frequent, but not rare. The true incidence of the other disorders is unknown, but NBS has identified infants with all the FAO disorders. All are inherited as autosomal recessive traits.

### Clinical Features<sup>(44,45)</sup>

FAO disorders have overlapping symptoms and organ involvement. They fall into three major categories as described below.

**Hepatic**<sup>(48,49)</sup>: There is no typical age of presentation, which may be on the first day of life through adulthood. As of 2007, there have been three infants in our region with MCAD who presented with sudden cardio/pulmonary arrest before screening results were known; one infant survived. Precipitating factors are fasting and/or stress associated with intercurrent illness. Patients present with "Reyes-like" symptoms including vomiting, lethargy, hypoketotic hypoglycemia, mild hyperammonemia, hyperuricemia, hypocarnitinemia and abnormal liver function tests. Liver biopsy often shows steatosis. Hepatic presentation is common in MCAD, VLCAD, LCHAD, neonatal CPT I & II and mild CT deficiency. Approximately 25 percent of individuals with MCAD will die during their first episode. In survivors, about 20 percent sustain significant neurological damage, presumably due to hypoglycemia. Patients with LCHAD also develop

\* These are not all the FAO disorders, only the ones thought to be detectable with MS/MS. At this time the sensitivity and specificity of MS/MS to detect all affected infants is unknown.

retinal pigmentary changes and progressive visual loss in childhood in spite of early diagnosis and treatment.

**Cardiac:** Cardiac abnormalities include hypertrophic or dilated cardiomyopathy. Pericardial effusion or cardiac failure can lead to death in these patients. FAO disorders with cardiac involvement include carnitine transporter defects, LCHAD, TFP deficiency, neonatal CPT II and VLCAD.

**Muscular:** There is usually moderate to severe hypotonia with recurrent rhabdomyolysis. Creatinine kinase may be greatly elevated. In infants and children seizures and/or developmental delay may also be present. Rhabdomyolysis is common in the adult form of CPT II, LCHAD, TFP deficiency and VLCAD.

A mother carrying an affected LCHAD fetus is prone to developing a life-threatening acute fatty liver during pregnancy or HELLP syndrome (hemolysis, elevated liver enzymes, low platelets). The reasons for this are not yet understood, but FAO disorders should be considered in infants whose mothers have a history of these pregnancy complications<sup>(45)</sup>.

### CPT I and Alaskan Natives (AN)

The OSPHL and the Alaska State Division of Public Health have identified a high incidence of CPT I in Alaskan Native (AN) infants, approximately 1:200 births as opposed to 1:200,000 in non-Native populations in our region. The AN infants all have the same mutation (P479L), allowing easy confirmation of suspect NBS results and testing for siblings. Canadian Provinces also report a higher incidence of this condition among their native populations. The incidence of this condition in other Native American populations has not been determined.

The majority of these infants have been identified on a routine second screen obtained around 2 weeks of age or even later. Infants and young children with CPT I usually develop symptoms after the newborn period with seizures and/or coma associated with life threatening episodes of fasting hypoglycemia.

Treatment consists of avoiding fasting and intravenous glucose support during intercurrent illnesses. Long-term outcome should be favorable if hypoglycemic episodes are minimized or eliminated.

### Treatment

Even with screening, some infants with FAO disorders may die before laboratory results are available. Treatment for MCAD and some other FAOs is extraordinarily simple once the diagnosis is suspected. Avoidance of fasting, particularly as infants and young children, is the primary treatment. Carnitine supplementation (100mg/kg/day) is used to provide a pathway for removal of toxic intermediate metabolites in some FAOs. With appropriate treatment hepatic, cardiac and muscular complications can be reduced or eliminated. Patients with these disorders may require IV support for fluid and calories during intercurrent infections or illnesses. With pre-symptomatic diagnosis and appropriate therapy, outcome can be normal for infants with MCAD<sup>(46,47)</sup>. Outcomes for the other disorders are unknown.

### Screening Practice Considerations

- Neonatal forms of FAO disorders can present in the first few days of life. Collect specimens at discharge or between 24–48 hours, whichever comes first.
- Transport specimen within 4–12 hours of collection if possible and no later than 24 hours after collection.
- Practitioners must remain alert to the possibility of FAO disorders in any neonate, infant or child with hypoketotic hypoglycemia or “Reyes-like” episodes or mother’s with HELLP syndrome or fatty liver of pregnancy.
- Infants affected with an FAO who are well fed may have normal screening results, masking the presence of the disorder.
- Practitioners caring for Alaska or Canadian Native infants should ensure that infants are tested twice, once between 24–48 hours of age and the second about 2 weeks of age.



# Metabolic Conditions

## Northwest Regional Newborn Screening Program

### Laboratory Tests

Results	Likely Causes	Actions
CO (free carnitine) <15.0 µM/L, or >300 µM/L	<ul style="list-style-type: none"> <li>* Carnitine transport defects possible</li> <li>* Carnitine deficiency</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
CO/C16 + C18 >100	<ul style="list-style-type: none"> <li>* CPT I possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C14 (Tetradecanoyl) >0.80 µM/L C14:1 (Tetradecenoyl) >0.60 µM/L C16 (Palmitoyl) >9.70 µM/L C18 (Octadecanoyl) >3.10 µM/L	<ul style="list-style-type: none"> <li>* VLCAD possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C16-OH (Hydroxy Palmitoyl) >0.50 µM/L C18-OH (hydroxy octadecanoic) >0.30 µM/L C18-1OH (hydroxy oleyl) >0.30 µM/L	<ul style="list-style-type: none"> <li>* LCHAD, TFP possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C6 (Hexanoyl) >0.80 µM/L C8 (Octanoyl) >1.00 µM/L C10 (Decanoyl) >0.70 µM/L	<ul style="list-style-type: none"> <li>* MCAD possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C4(Butyryl/Isobutyryl) >2.50 µM/L	<ul style="list-style-type: none"> <li>* SCAD possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C4 >1.80 µM/L C5 >0.70 µM/L C5-DC >0.50 µM/L C6 >0.80 µM/L C8 >1.00 µM/L C10 >0.70 µM/L C16 >9.70 µM/L	<ul style="list-style-type: none"> <li>* SCAD possible</li> <li>* MADD/GA II possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***



## Organic Acid Conditions (OA)

### OA Condition Essentials

- **Neonatal Emergency:** Infants with severe forms of organic acidemias will be symptomatic within a few days of birth and may die or suffer brain damage if not diagnosed and treated promptly.
- **Incidence:** 1:20,000 births
- **Screening Test:** MS/MS detection of leucine and acylcarnitines. Approximately 15 OAs can be detected through NBS.
- **Confirmatory Tests:** Quantitative amino acids, acylcarnitines, organic acids, enzyme assay and/or mutation analysis.
- **Validity:** 100% detected on first screen
- **Treatment:** Specific amino acid dietary restrictions and medications
- **Outcome:** Variable, from poor to excellent, depending on neonatal hit, disease severity, compliance with treatment and other environmental factors.

Organic acidemias (OA) result from enzyme deficiencies involved in the catabolism of multiple amino acids and other metabolites. Maple syrup urine disease is detected by an elevation of the amino acid leucine and an abnormal leucine/alanine ratio. All the other OAs are detected through elevations in acylcarnitines. All have autosomal recessive inheritance and have a collective incidence of 1:20,000.

#### The following OAs are screened for by MS/MS:

- Beta-ketothiolase deficiency
- Glutaric acidemia, Type I (glutaryl-CoA dehydrogenase deficiency)
- Isobutyryl CoA dehydrogenase deficiency
- Isovaleric acidemia, (isovaleryl-CoA dehydrogenase deficiency)
- Malonic aciduria
- Maple syrup urine disease (branched chain alpha-ketoacid dehydrogenase deficiency)
- Methylmalonic acidemias, methylmalonyl CoA mutase deficiency and defects of B-12 metabolism
- Propionic acidemia
- 3-Hydroxy-3-methylglutaryl (HMG) CoA lyase deficiency
- 2-Methyl-3-hydroxybutyryl CoA dehydrogenase deficiency
- 2-Methylbutyryl CoA dehydrogenase deficiency (mitochondrial acetoacetyl-CoA thiolase deficiency)
- 3-Methylcrotonyl CoA carboxylase (3MCC) deficiency
- 3-Methylglutaconyl CoA hydratase deficiency (3-methyl-glutaconic aciduria, Type I)
- Multiple carboxylase deficiency

#### Clinical Features<sup>(39,40)</sup>

**Neonatal Onset:** Most of these disorders have severe forms that present in the first week of life and constitute a neonatal emergency. Infants are generally well at birth, but develop poor feeding, irritability, lethargy, vomiting, and severe metabolic ketoacidosis, with or without hypoglycemia, in the first few days of life; this progresses to coma and death in the first month if treatment is not instituted. In methylmalonic and propionic acidemias, ammonia may also be elevated. Isovaleric acidemia is associated with the odor of "sweaty socks." Maple syrup urine disease has a characteristic "burnt sugar" or "maple syrup" odor which can be noticed in the urine, sweat and ear cerumen of the affected infant as early as the fifth day of life. Isobutyryl CoA dehydrogenase deficiency is associated with a dilated cardiomyopathy. Even with prompt treatment, many infants with neonatal forms of organic acidemias sustain psychomotor damage and may have significant long-term morbidity. These infants may be ill before the results of the screening tests are known. Contact the metabolic consultants urgently if an OA is suspected.

**Late Onset:** Milder variants may present with an acute decompensation brought on by an intercurrent illness similar to those described above, or with failure to thrive, hypotonia, mental retardation or seizures and a history of bouts of vomiting, protein intolerance, acidosis and/or hypoglycemia. While these patients typically have "milder" disease, the neurological damage may be just as severe as those presenting earlier. Newborn screening may be very beneficial to these infants as the initial crisis may be prevented.

**Asymptomatic Cases:** There are numerous reports of cases of isolated 3-methylcrotonyl-CoA carboxylase deficiency who have remained asymptomatic despite biochemical and/or enzymatic confirmation of the condition. The etiology of these variant presentations is not yet understood. Mild forms of methylmalonic acidemia have been found.

**Glutaric Acidemia, Type I:** Glutaric acidemia, Type I or GA I is an organic acidemia with clinical features unlike those described above<sup>(41,42, 43)</sup>. In this disease, there is an accumulation of glutaric acid and 3-hydroxy glutaric acid, which are believed to be toxic to cells, particularly in the central nervous system. The classic presentation is macrocephaly at or shortly after birth. Infants have a period of apparently normal development but may have soft neurological signs, like jitteriness, irritability and truncal hypotonia. Generally between 6 and 18 months of age, patients will experience an acute encephalopathic episode resulting in damage to the basal ganglia and atrophy of the caudate and putamen. This occurs over the course of a few hours to a day and is irreversible and untreatable. Severe dystonia, dyskinesia and other neurological findings result, either in a static or slowly progressive form. These children are often misdiagnosed as having extra pyramidal cerebral palsy. Approximately 25 percent of GA I patients will present with motor delay, hypotonia, dystonia and dyskinesia that develop gradually during the first few years of life, without any apparent acute crisis. Intellect is relatively intact. Infants with GA I are prone to acute subdural and retinal hemorrhages, after minor head trauma. This can be misdiagnosed as child abuse. Finally, 5 percent of all Amish patients have been completely asymptomatic without any crises and normal development. Neurological crises and symptoms rarely occur after five years of age.

### Laboratory Tests

All these disorders are detected using MS/MS. Leucine can be elevated in infants receiving hyperalimentation,

usually along with other amino acid elevations. In a normal newborn, however, elevations of these compounds are unusual and require rapid follow up. There is evidence that not all affected infants will be found by NBS<sup>(43)</sup>.

### Treatment

Any infant in whom a neonatal onset organic acidemia is suspected should be treated as a neonatal emergency. Infants with these disorders should in most, if not all, cases be transferred to a major medical center with a metabolic specialist as quickly as possible. The diagnosis, investigations and management are very complicated. Death or permanent neurological deficits can occur rapidly in untreated cases. Infants who are asymptomatic at the time that abnormal screening results are reported may be handled less urgently, depending on the clinical status and individual circumstances. Treatments, which must be continued for life, consist of strict dietary amino acid restrictions and medications.

Infants with GA I, in addition to diet and medications, must have aggressive supportive care during intercurrent illness throughout the first 5–6 years of life. This generally entails hospitalization, IV fluid and calories during all febrile or flu like illnesses.

For individuals with MSUD, isovaleric acidemia and one or two other organic acidemias, prospective and early identification through newborn screening will be life saving and outcomes are expected to be good. For others, including those with GA I, the outcomes are less certain at this time.

### Screening Practice Considerations

- Affected infants must be detected early if major problems are to be prevented.
- Collect specimens before discharge and transport within 4–12 hours of collection and no later than 24 hours after collection.

Practitioners must remain alert to the possibility of these diseases in any infant with lethargy, acidosis or coma.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***

## Urea Cycle Conditions (UCD)

### Urea Cycle Essentials

- **Neonatal Emergency:** Infants with severe hyperammonemia may die in the first week to 10 days if not diagnosed and treated.
- **Incidence:** 1:60,000 births (all 3 disorders)
- **Screening Test:** Citrulline, argininosuccinic acid and arginine by MS/MS
- **Confirmatory Tests:** Quantitative amino acids, urine organic acids and enzyme assay in red blood cells or hepatocytes
- **Validity:** 100% of citrullinemia and ASA on first test. The only arginase deficient infant diagnosed in Oregon was found on the second screen.
- **Treatment:** Neonatal rescue from hyperammonemic coma is complicated and should be done under the guidance of an experienced metabolic physician. Day to day hyperammonemia is controlled with a low protein diet, medications and amino acid supplements. Complete or partial liver transplant eliminates the need for diet and may improve development.
- **Outcome:** For those with citrullinemia and ASA who survive a neonatal coma, the outcome is usually fair to poor. Brain damage is common and the risk of hyperammonemia continues throughout life. Complications from arginase deficiency should be preventable with early and continuous treatment.

The urea cycle is the metabolic pathway responsible for the detoxification of ammonia and for the synthesis of arginine and urea. There are six enzymes in the urea cycle, each of which if missing, will result in hyperammonemia and one of the six disorders of the urea cycle. Each of these enzyme deficiencies has genetic and clinical variability from mild to lethal. Only three UCDs can be detected by newborn screening:

1. Arginase deficiency
2. Argininosuccinic aciduria (ASA)
3. Citrullinemia, Type I & II

Estimated incidence of these conditions is 1:60,000. They are inherited as autosomal recessive traits.

### Arginase Deficiency<sup>(24)</sup>

#### Clinical Features

Arginase deficiency is associated with irritability, inconsolable crying, anorexia, vomiting and developmental delay in infancy. This progresses to spastic tetraplegia with lower limbs more severely affected than the upper, psychomotor retardation, hyperactivity and growth failure. Hyperammonemia may result in encephalopathy, but is often milder than that seen in other urea cycle defects. A severe neonatal form presents with cholestatic jaundice, liver failure and death.

### Citrullinemia, Type I (CTLN1) and Argininosuccinic aciduria (ASA)<sup>(26,27)</sup>

#### Clinical Features-Neonatal Onset

Infants appear normal at birth and for the first 24 hours. Usually between 24–72 hours symptoms of hyperammonemia will appear as lethargy, vomiting, hypothermia, hyperventilation progressing to coma, cerebral edema and death without intervention. Unfortunately, a misdiagnosis of sepsis is made in 50 percent of the cases, wasting precious time. In addition to ammonia, both glutamate and glutamine are usually elevated. Specific elevations in citrulline, argininosuccinic acid, arginine, and orotic acid are helpful in determining the exact type of urea cycle defect.

#### Clinical Features-Late Onset

Late onset forms of urea cycle disorders most often present as non-specific developmental delay, seizures or other neurological symptoms which are associated with a history of repeated bouts of lethargy, vomiting, irritability or headaches. Food refusal and failure to thrive are not uncommon.

#### Asymptomatic Cases

Newborn screening has detected several infants with very mild citrullinemia, who do not require any treatment when healthy, but may be at risk of decompensation under stress, infection or high protein intake.

## Citrin Deficiency (Citrullinemia, Type II & Neonatal Intrahepatic Cholestasis [NICCD]) <sup>(28)</sup>

Citrin is a mitochondrial membrane aspartate-glutamate carrier that acts to transfer cytosolic NADH into the mitochondria. There are two distinct disorders associated with citrin deficiency. It is unknown how well NBS tests will identify these patients.

### Clinical Features-Neonatal Onset

Neonatal intrahepatic cholestasis due to citrin deficiency (NICCD) has been found in over 200 Japanese and Asian infants and a handful of non-Asian infants, usually between 1–5 months of age. Liver disease may be accompanied by jaundice and fatty infiltrates. While liver failure may necessitate transplant in infancy, the liver disease generally resolves by a year of age for most patients. At least one of these infants has progressed to citrullinemia type II at the age of 16 years.

### Clinical Features-Late Onset

Patients with citrullinemia type II (CTLN2) present in childhood or adulthood (11–64 years of age). Symptoms may be acute or develop slowly. These include enuresis, delayed menarche, insomnia, night sweats and terrors, recurrent vomiting, diarrhea, tremors, confusion, lethargy, delusions and episodes of coma. Citrulline and ammonia are elevated. Within a few years of the diagnosis, episodes of pancreatitis, hyperlipidemia and death from cerebral edema generally occur. Hepatocellular carcinoma has been reported in a few cases.

## Laboratory Tests

Elevations of citrulline and arginine are detected by MS/MS. The laboratory cutoff for citrulline is <100 µM/L; for arginine, <130 µM/L; argininosuccinic acid, <6.0 µM/L. Transient elevations of plasma arginine and citrulline in the newborn are unusual unless the infant is premature and/or receiving hyperalimentation.

Infants with NICCD may or may not have citrulline elevations. Approximately half of the Japanese patients came to attention with elevated galactose, methionine and/or phenylalanine on NBS prior to the advent of MS/MS. Approximately 10 percent of NICCD patients had normal citrulline.

## Treatment (Citrullinemia, Type I & ASA)

All patients with a neonatal presentation represent medical emergencies and outcomes may be variable. Patients with neonatal onset disease will typically require aggressive treatment with hemodialysis. All patients, both late onset and those rescued from neonatal hyperammonemia, will require treatment with low protein diets and medications to prevent hyperammonemia and remove toxic compounds. The outcome for patients rescued from prolonged neonatal hyperammonemia is dismal. Brain damage is likely. Even patients treated prospectively from birth may not be normal.<sup>27</sup> Those with late onset disease fare better, and presymptomatic diagnosis and treatment may allow normal development.

Results	Likely Causes	Actions
Arginine ≥130 µM/L	<ul style="list-style-type: none"> <li>* Arginase deficiency possible</li> <li>* Transient argininemia</li> <li>* Liver disease</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
ASA ≥6.00 µM/L	<ul style="list-style-type: none"> <li>* Argininosuccinic aciduria possible</li> <li>* Liver disease</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
Citrulline ≥ 100 µM/L	<ul style="list-style-type: none"> <li>* Citrullinemia, argininosuccinic aciduria possible</li> <li>* Transient citrullinemia</li> <li>* Liver disease</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
Citrulline ≥ 200 µM/L on second specimen	<ul style="list-style-type: none"> <li>* Mild citrullinemia, argininosuccinic aciduria possible</li> <li>* Transient citrullinemia</li> <li>* Liver disease</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.

**Treatment: NICCD & CTLN2**

NICCD responds well to protein restriction in infancy for most patients. Those who do not respond or who develop progressive liver failure graduate to liver transplantation.

Patients with CTLN2 receive a liver transplant, as they will proceed to death without it. Dietary restriction of protein is ineffective. Long-term outcome is unknown.

**Screening Practice Considerations**

- Neonatal emergency.
- Infants with neonatal onset disease may be sick or die before screening results are known. Collect specimens before discharge and transport within 4–12 hours of collection and no longer than 24 hours after collection.
- Practitioners must remain alert to the possibility of these disorders in any newborn with lethargy or coma.
- Arginine may rise slowly in some cases and is more likely to be found on the second screening test.
- Citrin deficiency is more common in Asian infants.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***

## Galactosemia

### Galactosemia Essentials

- **Neonatal Emergency:** 50% will die in the first 7-10 days usually from gram-negative sepsis. Acute liver disease can produce a coagulopathy and vitreous hemorrhage.
- **Incidence:** 1:60,000
- **Screening Test:** Beutler test (qualitative enzyme assay); Second tier: Hill test (free galactose and galactose-1-phosphate) is done on every infant with abnormal Beutler test.
- **Confirmatory Tests:** Enzyme assay GALT, RBC gal-1-phosphate
- **Validity:** 100% found on 1st specimen, unless transfused
- **Treatment:** Lactose restricted diet
- **Outcome:** Somewhat diminished IQs as a group (-1SD), verbal and motor dyspraxia in 60%, ovarian failure in 80% of females and post-natal growth delay during childhood.

Dietary galactose is most commonly ingested as lactose, the principle carbohydrate of human milk and most non-soy commercial infant formulas; it is hydrolyzed to glucose and galactose in the intestine. After absorption, galactose is metabolized by several enzymes including galactokinase and galactose-1-phosphate uridyl transferase (GALT). When deficient, the latter causes galactosemia (1:60,000 births). Galactosemia is a recessively inherited condition.

### Clinical Features<sup>(24)</sup>

Detection of galactosemia requires urgent follow-up and is considered a neonatal emergency. The early clinical features of severe untreated galactosemia include neonatal hypoglycemia, liver damage, jaundice, weight loss, lethargy and sepsis. Vitreous hemorrhage from coagulopathy has been reported in some infants. Death may result from gram-negative sepsis within 1–2 weeks of birth. If the infant remains untreated and survives the neonatal period, cataracts, cirrhosis, renal Fanconi syndrome and mental retardation are usual.

There are several genetic variants with less severe reduction in the enzyme activity (e.g., the Duarte variant). The screening test is not designed to detect variant galactosemia and is not completely sensitive for this purpose. Most of these cases are asymptomatic and are detected on newborn screening because of abnormalities in GALT and/or galactose. The need for treatment of Duarte variant galactosemia is controversial and infant specific. The metabolic consultants are available for consultation.

### Laboratory Tests

Two screening tests are used to detect galactosemia in a two-tiered sequence (see diagram):

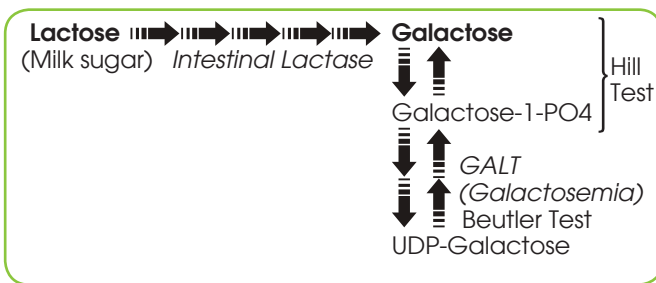
1. Galactose (Hill Test): Slight elevations (up to 20 mg/dL) can occur in normal neonates, but galactose metabolites are greatly elevated in infants with galactosemia if they are receiving a lactose-containing formula or breast milk. The Hill test is a fluorometric chemical spot test that measures galactose and galactose-1-phosphate. Liver disease may also cause an elevation of galactose

Results		Likely Causes	Actions
<b>GALT Test</b>	<b>Galactose Metabolites</b>		
Abnormal	≥20 mg/dL	<ul style="list-style-type: none"> <li>* Severe galactosemia</li> <li>* Variant galactosemia</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
Abnormal	<20 mg/dL	<ul style="list-style-type: none"> <li>* Severe galactosemia with little lactose intake</li> <li>* Variant galactosemia</li> <li>* Other enzyme defects in red blood cells</li> <li>* Improperly handled sample (heat damage or transit delay)</li> </ul>	Contact by mail if infant is ≥48 hrs old; contact by fax if <48 hrs old or if not on lactose.



metabolites. All infants with an abnormal Beutler or who have been transfused will be screened with the Hill test.

- GALT activity (Beutler Test): The enzyme test depends upon fluorescence produced by the normal galactose enzyme cascade in red blood cells. A temporarily abnormal result (diminished or absent fluorescence) is found in 1:2,000 infants. The test may be persistently abnormal if the enzyme activity is <50 percent of normal. It does not differentiate milder variants from severe defects. All infants are screened with the Beutler test.



## Treatment

Galactosemia is treated by dietary galactose restriction. This diet must be followed for life and requires close supervision. Even with early diagnosis and strict dietary restrictions children with galactosemia are at risk for speech disorders, growth and developmental delays and in females, ovarian failure.

## Screening Practice Considerations

- This disorder kills quickly. Transport all specimens 4–12 hours after collection and no later than 24 hours after collection.
- The GALT test should be abnormal in virtually all severe classic galactosemic infants even if the specimen is obtained before lactose is ingested, unless the infant has been transfused. Obtain a specimen before any transfusion.
- The GALT enzyme is prone to degradation if the sample is delayed in the mail or exposed to excess temperature or humidity. This produces a false positive GALT result.
- Galactose accumulation depends on lactose ingestion so that blood galactose metabolites may be normal in infants being fed a soy-based formula.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***



## Biotinidase Deficiency

### Biotinidase Deficiency Essentials

- **Incidence:** 1:60,000 births
- **Screening Test:** Biotinidase qualitative colorimetric enzyme assay
- **Confirmatory Tests:** Quantitative biotinidase enzyme assay
- **Validity:** 100% found on 1st screen
- **Treatment:** 5-10 mg biotin/day
- **Outcome:** Excellent if compliant with biotin therapy

This recessively inherited disorder affects the cells' ability to recycle the vitamin-cofactor biotin and this impairs the function of mitochondrial carboxylases. The incidence is 1:60,000 births. Screening is done for all infants in the region.

#### Clinical Features<sup>(2,3)</sup>

Infants with profound biotinidase deficiency are normal at birth, but develop one or more of the following symptoms after the first weeks or months of life: hypotonia, ataxia, seizures, developmental delay, alopecia, seborrheic dermatitis, hearing loss and optic nerve atrophy. Metabolic acidosis can result in coma and death.

Infants with partial deficiency (5–10 percent) have been identified through newborn screening and family studies. They may remain asymptomatic with no treatment or exhibit milder symptoms than infants with profound deficiency. A reduced dose of biotin is recommended for these infants as the consequences of complications are too great.

### Laboratory Tests

Detection of enzyme activity is by a qualitative colorimetric assay. In the presence of the enzyme a color change occurs.

Results	Likely Causes	Actions
Color change does not occur	* Biotinidase deficiency possible * False positive	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.

### Treatment

Daily biotin supplements clear the skin rash and alopecia and improve the neurological status in patients not diagnosed by screening. With early diagnosis and treatment made possible by screening, all symptoms can be prevented.

### Screening Practice Considerations

- The enzyme is prone to damage if the sample is delayed in the mail or exposed to high temperatures or excess humidity.
- Transfusion of red cells prior to drawing the newborn screening specimen will invalidate the biotinidase assay. Obtain a specimen before transfusion.

***The purpose of newborn screening is to identify infants at risk and in need of more definitive testing. As with any laboratory test, both false negative and false positive results are possible. Screening test results are insufficient information on which to base diagnosis or treatment.***

# References

1. Pass KA, Lane PA, Fernhoff PM, Hinton CF, Panny SR, Parks JS, Pelias MZ, Rhead WJ, Ross SI, Wethers DL, Elsas LJ. 2nd. US newborn screening system guidelines II: follow-up of children, diagnosis, management, and evaluation. Statement of the Council of Regional Networks for Genetic Services (CORN). *J Pediatr*. 2000 Oct;137(4 Suppl):S1-46. No abstract available.
2. American College of Medical Genetics Newborn Screening Expert Group. Newborn screening: toward a uniform screening panel and system--executive summary. *Pediatrics*. 2006 May;117(5 Pt 2):S296-307.
3. National Newborn Screening and Genetics Resource Center, Austin TX website: genes-r-us.uthscsa.edu
4. Wilson, J.M.G., and Jugner, G: Principles and Practices of Screening for Disease, Public Health Papers #34. Geneva; World Health Organization, 1968.
5. Tuerck JM, Buist NR, Skeels MR, Miyahira RS, Beach PG. Computerized surveillance of errors in newborn screening practice. *Am J Public Health*. 1987 Dec;77(12):1528-31.
6. Holtzman C, Slazyk WE, Cordero JF, Hannon WH. Descriptive epidemiology of missed cases of phenylketonuria and congenital hypothyroidism. *Pediatrics*. 1986 Oct;78(4):553-8.
7. Kosciak RL, Lai HJ, Laxova A, Zaremba KM, Kosorok MR, Douglas JA, Rock MJ, Splaingard ML, Farrell PM. Preventing early, prolonged vitamin E deficiency: an opportunity for better cognitive outcomes via early diagnosis through neonatal screening. *J Pediatr*. 2005 Sep;147(3 Suppl):S51-6.
8. Kosciak RL, Farrell PM, Kosorok MR, Zaremba KM, Laxova A, Lai HC, Douglas JA, Rock MJ, Splaingard ML. Cognitive function of children with cystic fibrosis: deleterious effect of early malnutrition. *Pediatrics*. 2004 Jun;113(6):1549-58.
9. Sontag MK, Hammond KB, Zielenski J, Wagener JS, Accurso FJ. Two-tiered immunoreactive trypsinogen-based newborn screening for cystic fibrosis in Colorado: screening efficacy and diagnostic outcomes. *J Pediatr*. 2005 Sep;147(3 Suppl):S83-8.
10. Henry RL, Boulton TJ, Roddick LG. False negative results on newborn screening for cystic fibrosis. *J Paediatr Child Health*. 1990 Jun;26(3):150-1.
11. Parad RB, Comeau AM. Diagnostic dilemmas resulting from the immunoreactive trypsinogen/DNA cystic fibrosis newborn screening algorithm. *J Pediatr*. 2005 Sep;147(3 Suppl):S78-82.
12. Frimberger D, Gearhart JP. Ambiguous genitalia and intersex. *Urol Int*. 2005;75(4):291-7. Review.
13. Mussig K, Kaltenbach S, Maser-Gluth C, Hartmann MF, Wudy SA, Horger M, Gallwitz B, Raue F, Haring HU, Schulze E. Late diagnosis of congenital adrenal hyperplasia due to 21-hydroxylase deficiency. *Exp Clin Endocrinol Diabetes*. 2006 Apr;114(4):208-14.
14. Therrell BL Jr, Berenbaum SA, Manter-Kapanke V, Simmank J, Korman K, Prentice L, Gonzalez J, Gunn S. Results of screening 1.9 million Texas newborns for 21-hydroxylase-deficient congenital adrenal hyperplasia. *Pediatrics*. 1998 Apr;101(4 Pt 1):583-90.
15. Votava F, Torok D, Kovacs J, Moslinger D, Baumgartner-Parzer SM, Solyom J, Pribilincova Z, Battelino T, Lebl J, Frisch H, Waldhauser F; Middle European Society for Paediatric Endocrinology -- Congenital Adrenal Hyperplasia (MESPE-CAH) Study Group. Estimation of the false-negative rate in newborn screening for congenital adrenal hyperplasia. *Eur J Endocrinol*. 2005 Jun;152(6):869-74.
16. Selva KA, Harper A, Downs A, Blasco PA, Lafranchi SH. Neurodevelopmental outcomes in congenital hypothyroidism: comparison of initial T4 dose and time to reach target T4 and TSH. *J Pediatr*. 2005 Dec;147(6):775-80.
17. Van Vliet G. Neonatal hypothyroidism: treatment and outcome. *Thyroid*. 1999 Jan;9(1):79-84. Review.
18. American Academy of Pediatrics AAP Section on Endocrinology and Committee on Genetics, and American Thyroid Association Committee on Public Health: Newborn screening for congenital hypothyroidism: recommended guidelines. *Pediatrics*. 1993 Jun;91(6):1203-9. No abstract available.
19. LaFranchi SH, Hanna CE, Krainz PL, Skeels MR, Miyahira RS, Sesser DE. Screening for congenital hypothyroidism with specimen collection at two time periods: results of the Northwest Regional Screening Program. *Pediatrics*. 1985 Nov;76(5):734-40.

20. Maniatis AK, Taylor L, Letson GW, Bloch CA, Kappy MS, Zeitler P. Congenital hypothyroidism and the second newborn metabolic screening in Colorado, USA. *J Pediatr Endocrinol Metab*. 2006 Jan;19(1):31-8.
21. Yunis KA, Nasr MR, Lepejian G, Najjar S, Daher R. False-negative primary neonatal thyroid screening: the need for clinical vigilance and secondary screening. *J Med Screen*. 2003;10(1):2-4.
22. Lees CM, Davies S, Dezateux C. Neonatal screening for sickle cell disease. *Cochrane Database Syst Rev*. 2000;(2):CD001913. Review.
23. Wolf B. Biotinidase Deficiency: New Directions and Practical Concerns. *Curr Treat Options Neurol*. 2003 Jul;5(4):321-328.
24. Bosch AM. Classical galactosaemia revisited. *J Inherit Metab Dis*. 2006 Aug;29(4):516-25. Epub 2006 Jul 11. Review.
25. Scaglia F, Lee B. Clinical, biochemical, and molecular spectrum of hyperargininemia due to arginase I deficiency. *Am J Med Genet C Semin Med Genet*. 2006 May 15;142(2):113-20. Review.
26. Nassogne MC, Heron B, Touati G, Rabier D, Saudubray JM. Urea cycle defects: management and outcome. *J Inherit Metab Dis*. 2005;28(3):407-14. Review.
27. Gropman AL, Batshaw ML. Cognitive outcome in urea cycle disorders. *Mol Genet Metab*. 2004 Apr;81 Suppl 1:S58-62. Review.
28. Ohura T, Kobayashi K, Tazawa Y, Abukawa D, Sakamoto O, Tsuchiya S, Saheki T. Clinical pictures of 75 patients with neonatal intrahepatic cholestasis caused by citrin deficiency (NICCD). *J Inherit Metab Dis*. 2007 Apr;30(2):139-44. Epub 2007 Feb 24.
29. Yap S, Boers GH, Wilcken B, Wilcken DE, Brenton DP, Lee PJ, Walter JH, Howard PM, Naughten ER. Vascular outcome in patients with homocystinuria due to cystathionine beta-synthase deficiency treated chronically: a multicenter observational study. *Arterioscler Thromb Vasc Biol*. 2001 Dec;21(12):2080-5.
30. Yap S, Naughten E. Homocystinuria due to cystathionine beta-synthase deficiency in Ireland: 25 years' experience of a newborn screened and treated population with reference to clinical outcome and biochemical control. *J Inherit Metab Dis*. 1998 Oct;21(7):738-47.
31. Channon S, Goodman G, Zlotowitz S, Mockler C, Lee PJ. Effects of dietary management of phenylketonuria on long-term cognitive outcome. *Arch Dis Child*. 2007 Mar;92(3):213-8. Epub 2006 Oct 26.
32. Cleary MA, Feillet F, White FJ, Vidailhet M, Macdonald A, Grimsley A, Maurin N, de Baulny HO, Rutherford PJ. Randomised controlled trial of essential fatty acid supplementation in phenylketonuria. *Eur J Clin Nutr*. 2006 Jul;60(7):915-20. Epub 2006 Mar 8.
33. Fiege B, Blau N. Assessment of tetrahydrobiopterin (BH4) responsiveness in phenylketonuria. *J Pediatr*. 2007 Jun;150(6):627-30.
34. Matalon R, Michals-Matalon K, Bhatia G, Grechanina E, Novikov P, McDonald JD, Grady J, Tyring SK, Guttler F. Large neutral amino acids in the treatment of phenylketonuria (PKU). *J Inherit Metab Dis*. 2006 Dec;29(6):732-8. Epub 2006 Sep 21.
35. Scott CR. The genetic tyrosinemias. *Am J Med Genet C Semin Med Genet*. 2006 May 15;142(2):121-6. Review.
36. Koelink CJ, van Hasselt P, van der Ploeg A, van den Heuvel-Eibrink MM, Wijburg FA, Bijleveld CM, van Spronsen FJ. Tyrosinemia type I treated by NTBC: how does AFP predict liver cancer? *Mol Genet Metab*. 2006 Dec;89(4):310-5. Epub 2006 Sep 27.
37. Tsai CP, Lin PY, Lee NC, Niu DM, Lee SM, Hsu WM. Corneal lesion as the initial manifestation of tyrosinemia type II. *J Chin Med Assoc*. 2006 Jun;69(6):286-8.
38. Magera MJ, Gunawardena ND, Hahn SH, Tortorelli S, Mitchell GA, Goodman SI, Rinaldo P, Matern D. Quantitative determination of succinylacetone in dried blood spots for newborn screening of tyrosinemia type I. *Mol Genet Metab*. 2006 May;88(1):16-21. Epub 2006 Jan 31.
39. Deodato F, Boenzi S, Santorelli FM, Dionisi-Vici C. Methylmalonic and propionic aciduria. *Am J Med Genet C Semin Med Genet*. 2006 May 15;142(2):104-12. Review.

40. Andersson HC, Marble M, Shapira E. Long-term outcome in treated combined methylmalonic acidemia and homocystinemia. *Genet Med.* 1999 May-Jun;1(4):146-50.
41. Hedlund GL, Longo N, Pasquali M. Glutaric acidemia type 1. *Am J Med Genet C Semin Med Genet.* 2006 May 15;142(2):86-94. Review.
42. Funk CB, Prasad AN, Frosk P, Sauer S, Kolker S, Greenberg CR, Del Bigio MR. Neuropathological, biochemical and molecular findings in a glutaric acidemia type 1 cohort. *Brain.* 2005 Apr;128(Pt 4):711-22. Epub 2005 Feb 2.
43. Gallagher RC, Cowan TM, Goodman SI, Enns GM. Glutaryl-CoA dehydrogenase deficiency and newborn screening: retrospective analysis of a low excretor provides further evidence that some cases may be missed. *Mol Genet Metab.* 2005 Nov;86(3):417-20. Epub 2005 Sep 23.
44. Olpin SE. Fatty acid oxidation defects as a cause of neuromyopathic disease in infants and adults. *Clin Lab.* 2005;51(5-6):289-306. Review.
45. Shekhawat PS, Matern D, Strauss AW. Fetal fatty acid oxidation disorders, their effect on maternal health and neonatal outcome: impact of expanded newborn screening on their diagnosis and management. *Pediatr Res.* 2005 May;57(5 Pt 2):78R-86R. Epub 2005 Apr 6. Review.
46. Wilcken B, Haas M, Joy P, Wiley V, Chaplin M, Black C, Fletcher J, McGill J, Boneh A. Outcome of neonatal screening for medium-chain acyl-CoA dehydrogenase deficiency in Australia: a cohort study. *Lancet.* 2007 Jan 6;369(9555):37-42.
47. Wilson CJ, Champion MP, Collins JE, Clayton PT, Leonard JV. Outcome of medium chain acyl-CoA dehydrogenase deficiency after diagnosis. *Arch Dis Child.* 1999 May;80(5):459-62.
48. Holton JB, Allen JT, Green CA, Partington S, Gilbert RE, Berry PJ. Inherited metabolic diseases in the sudden infant death syndrome. *Arch Dis Child.* 1991 Nov;66(11):1315-7.
49. Wilcox RL, Nelson CC, Stenzel P, Steiner RD. Postmortem screening for fatty acid oxidation disorders by analysis of Guthrie cards with tandem mass spectrometry in sudden unexpected death in infancy. *J Pediatr.* 2002 Dec;141(6):833-6.
50. Van Wassenae AG, Kok JH, Dekker FW, de Vijlder JJ. Thyroid function in very preterm infants: influences of gestational age and disease. *Pediatr Res.* 1997 Nov;42(5):604-9.
51. Linder N, Davidovitch N, Reichman B, Kuint J, Lubin D, Meyerovitch J, Sela BA, Dolfin Z, Sack J. Topical iodine-containing antiseptics and subclinical hypothyroidism in preterm infants. *J Pediatr.* 1997 Sep;131(3):434-9.
52. Filippi L, Cecchi A, Tronchin M, Dani C, Pezzati M, Seminara S, Gasperini S, Zammarchi E, Rubaltelli FF. Dopamine infusion and hypothyroxinaemia in very low birth weight preterm infants. *Eur J Pediatr.* 2004 Jan;163(1):7-13. Epub 2003 Nov 25.
53. Karna P. Developmental follow-up of very low birthweight premature infants with low free thyroxine. *Am J Perinatol.* 1991 Jul;8(4):288-91.

# Appendices

## Appendix I

Date: August 16, 2006  
To: Neonatal Intensive Care Units (NICU)  
From: Stephen LaFranchi, MD  
Consultant, Newborn Screening Program  
Re: Specimen Collection for Infants in Neonatal  
Intensive Care Units

Many Neonatal Intensive Care Units (NICU) obtain the first Newborn Screening specimen upon admission to the NICU. This specimen is typically obtained in the first 24 hours of life, and may be as early as one hour of age. The rationale for this early sample is to obtain the first specimen before the neonate receives any transfused blood products, which have the potential to interfere with the screening tests for some disorders (biotinidase deficiency, galactosemia and hemoglobinopathies). On the other hand, this early specimen makes screening for some disorders problematic, either because some disorders depend on analyte accumulation, which can take several days (PKU), or the screening test cutoff must be adjusted (TSH for congenital hypothyroidism). Further, preterm infants (the majority of NICU admissions) are at risk for a form of primary congenital hypothyroidism characterized by a delay in TSH elevation, which typically does not show up until after 3 weeks of age. This so-called "delayed TSH rise" occurs in approximately 1:18,000 newborns (1).

1. These issues were discussed at our last bimonthly Newborn Screening Program Meeting. The following recommendations were made for infants admitted to NICUs:
2. If the first newborn screening specimen is obtained less than 24 hours of age, then,
3. The second newborn screening specimen should be obtained between 5-7 days of age.

For infants less than 2,000 grams (or less than 34 weeks gestation), or infants who are still in the NICU at greater than 21 days of age, obtain a routine third newborn screening specimen after 21 days of age.

In rare cases, the TSH elevation was detected in newborn screening specimens obtained between 4 and 12 weeks of age. In all of these cases, the previous screen had a subnormal T4 level. Thus it is probably prudent to obtain screening results until the blood spot T4 is > 5.0 ug/dL or the serum T4 is > 7.2 ug/dL.

1. Mandel SJ, et al. Atypical hypothyroidism and the very low birth weight infant. *Thyroid* 2000; 8:693-695.

Results	Likely Causes	Actions
Leucine > 230 µM/L first NBS Leucine > 330 µM/L second NBS Leu/ala >1.30	<ul style="list-style-type: none"> <li>* MSUD possible</li> <li>* Hyeralimentation</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C3 (Propionyl) >7.50 µM/L C3/C2 >0.30 µM/L	<ul style="list-style-type: none"> <li>* Methylmalonic acidemias possible</li> <li>* Multiple carboxylase deficiency possible</li> <li>* Propionic acidemia possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C3-DC (Dicarboxyl Propionyl) >0.80 µM/L	<ul style="list-style-type: none"> <li>* Malonic aciduria possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C4 (Butyryl/Isobutyryl) >1.80 µM/L	<ul style="list-style-type: none"> <li>* Isobutyryl CoA dehydrogenase deficiency possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C5 (Isovaleryl) >0.70 µM/L	<ul style="list-style-type: none"> <li>* Isovaleric acidemia possible</li> <li>* 2-Methylbutyryl CoA dehydrogenase deficiency possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C5-DC (Glutaryl) >0.50 µM/L	<ul style="list-style-type: none"> <li>* Glutaric acidemia, Type I possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C5-OH (Methylcrotonyl) >1.50 µM/L	<ul style="list-style-type: none"> <li>* 2-Methyl-3-hydroxy butyryl CoA dehydrogenase deficiency possible</li> <li>* 3-Methylcrotonyl CoA carboxylase deficiency possible</li> <li>* 3-Methylglutaconyl CoA hydratase deficiency possible</li> <li>* Multiple carboxylase deficiency possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C5:1 (Tiglyl/3-methylcrotonyl) >0.80 µM/L	<ul style="list-style-type: none"> <li>* 2-Methyl-3-hydroxy butyryl CoA dehydrogenase deficiency possible</li> <li>* Beta ketothiolase deficiency possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.
C5-OH + Multiple elevations	<ul style="list-style-type: none"> <li>* 3-Hydroxy-3-methyl glutaric aciduria possible</li> <li>* False positive</li> </ul>	NBS nurse faxes results. Medical consultant phones practitioner with follow-up recommendations.



# Appendix II

## Northwest Regional Newborn Screening Program

### Incidence of Disorders Detected by MS/MS in Oregon

February 1, 2007

DISORDER	INCIDENCE*	ANALYTE	ANALYTE RATIO
<b>Amino acid disorders</b>	<b>1:8,600</b>		
Arginase deficiency	1:350,000	Arg	
Argininosuccinic aciduria	1:88,000	ASA	
Citrullinemia	1:71,000	Cit	
Homocystinuria	1:118,000	Met	
Hyperphenylalaninemia	1:13,600	Phe	Phe/Tyr
Maple syrup urine disease	1:350,000	Leu	Leu/Ala
Tyrosinemia	1:350,000	Tyr	
<b>Fatty acid oxidation defects</b>	<b>1:5,500</b>		
Carnitine Uptake Defect		C0↓	
Carnitine/Acylcarnitine carrier defect	1:118,000	C0↓ C16↑ C18↑	
Medium chain acyl-CoA dehydrogenase deficiency	1:11,000	C8, C10, C6, C10:1	
Short chain acyl-CoA dehydrogenase deficiency	1:350,000	C4	
Long chain 3-hydroxyacyl-CoA dehydrogenase deficiency	1:177,000	C16OH, C18:1OH, C16:1, C18OH	
Very long chain acyl-CoA dehydrogenase deficiency	1:71,000	C14:1, C16, C14, C16:1 C14:1/ C12:1↑	
Glutaric acidemia GA II	-	C5DC, C6, C8, C10, C14:1, C16:0	
(multiple acyl CoA dehydrogenase deficiency)	-	C4	
CPT I Carnitine palmitoyl transferase deficiency I	1:15,000		C0/(C16+C18)
CPT II Carnitine palmitoyl transferase deficiency II	-	C18:1, C16↑, C18↑, C0	
<b>Organic acidemias</b>	<b>1:15,000</b>		
Beta-ketothiolase deficiency	-	C5:1, C5OH	
Glutaric acidemias GA I	1:71,000	C5DC	
3-hydroxy-3-methylglutaryl-CoA lyase deficiency (HMG)	-	C6DC, C5OH	
Isobutyryl CoA dehydrogenase deficiency	-	C4	
Isovaleryl CoA dehydrogenase deficiency	1:118,000	C5	
2-methyl-3-hydroxybutyryl CoA dehydrogenase deficiency	1:350,000	C5:1, C5OH	
Methylmalonic acidemia	1:51,000	C3	C3/C2
3-methylcrotonyl CoA carboxylase deficiency	1:51,000	C5OH,	
3-methylglutaconic aciduria	-	C5OH	
Multiple carboxylase deficiency	-	C5OH, C3	
Propionyl CoA carboxylase deficiency	-	C3	C3/C2
Malonic acidemia	-	C3DC	

\*Incidence is calculated for major categories of disorders based on published reports and for individual disorders if these are well established.



N O R T H W E S T  R E G I O N A L  
N E W B O R N S C R E E N I N G P R O G R A M

*A collaborative project involving:*

Oregon Department of Human Services

Oregon Health & Science University

Alaska Department of Health and Social Services

Hawai'i Department of Health

Idaho Department of Health and Welfare

Nevada State Health Division

New Mexico Newborn Screening Program

If you need this information in an alternate format,  
please call: Oregon State Public Health Laboratory at 503-693-4100.

[www.oregon.gov/DHS/ph/nbs/index.shtm](http://www.oregon.gov/DHS/ph/nbs/index.shtm)