Occurrence of Isoxaflutole, Acetamide, and Triazine Herbicides and Their Degradation Products in 10 Iowa Rivers Draining to the Mississippi and Missouri Rivers, 2004

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Abstract

During 2004, a study to document the occurrence of herbicides and herbicide degradation products was conducted for 10 major Iowa rivers draining to the Missouri and Mississippi Rivers. Seventy-five water-quality samples were collected to measure isoxaflutole, acetamide, and triazine herbicides and their herbicide degradation products. An analytical method to measure isoxaflutole and its degradation products, diketonitrile and benzoic acid, was developed by the U.S. Geological Survey Organic Geochemistry Research Laboratory in Lawrence, Kansas, using vacuum manifold solid-phase extraction and liquid chromatography/mass spectrometry/mass spectrometry and is described in this report.

Isoxaflutole, a low application rate preemergence herbicide for control of annual broadleaf weeds, is used extensively in Iowa. Findings from the study documented in this report indicate that isoxaflutole was designed to degrade quickly to diketonitrile, which appears to be more stable, and then to benzoic acid. Of the 75 samples collected to measure isoxaflutole, there were four detections of isoxaflutole, 56 detections of diketonitrile, and 43 detections of benzoic acid. Also, results of acetamide and triazine water-quality samples correlate with past studies, which indicate that herbicide degradation products are detected more frequently and often at higher concentrations in surface water than their parent compounds.

In addition to analysis of isoxaflutole and its degradation products, samples were analyzed by the USGS National Water-Quality Laboratory schedule 2001 for about 52 pesticides and their degradation products.

Introduction

Iowa is one of the most productive agricultural areas in the United States for corn. Crops grown in the State are used to produce food, fuel, and fiber. Agricultural chemicals are needed for crops to control competing vegetation, insects, and fungus, and to supply sufficient nutrients. The use of pesticides and fertilizer has resulted in substantial crop yields, but certain climate conditions often result in agricultural chemicals being applied before rainfall that produces runoff and washing of the chemicals into streams and rivers. The presence of pesticides used in agricultural and urban areas has been documented in rivers and streams in Iowa and throughout the Midwestern United States. These contaminants can have an effect on water quality in local streams, downstream on major rivers such as the Mississippi and Missouri Rivers, and the Gulf of Mexico.

Herbicide Usage

Modern agricultural practices in the United States often involve widespread use of herbicides for production of crops. Herbicide use is one factor that affects the concentrations of herbicides measured in Midwestern streams (Battaglin and Goolsby, 1999; Coupe and Goolsby, 1999; Scribner and others, 2000a) and an important reason for monitoring the water quality in streams.

Isoxaflutole was patented by the Rhone-Poulenc Agriculture Company (Basel, Switzerland) in 1990. The Rhone-Poulenc Agriculture Company merged with AgrEvo (a company of Hoescht and Schering) in 1999 to form Aventis CropScience (Lyon, France), which was purchased by Bayer CropScience (Research Triangle Park, North Carolina) in 2001 (Luscombe and others, 1995).

Isoxaflutole, a member of the benzoyl isoxazole family, is a low-application-rate preemergence herbicide for control of annual broadleaf weeds and some grasses in field corn and sugar cane (Pallet and others, 2001). For example, in Iowa isoxaflutole was applied at 0.07 lb active ingredient per acre per year during 2003 compared to atrazine, applied at 0.91 lb active ingredient per acre per year, and metolachlor, applied at 1.9 lb active ingredient per acre per year (U.S. Department of Agriculture, 2004). The intended strategy of the manufacturer is that under field conditions, isoxaflutole, also known by



Figure 1. Herbicide use in lowa, 1990–2003 (data from U.S. Department of Agriculture, 2004).

its trade name Balance, degrades quickly to the biologically active diketonitrile (Lin and others, 2002), followed by conversion of diketonitrile to benzoic acid. Diketonitrile also is more water soluble than isoxaflutole, and therefore, its transport to surface or ground water is a concern.

Isoxaflutole was first used in 1999 in the United States, and usage has fluctuated from 213,000 lb in 1999, peaking at 439,000 lb in 2001, then decreasing to 321,000 lb in 2003 (U.S. Department of Agriculture, 2004). Isoxaflutole usage in Iowa (fig. 1) was 62,000 lb in 1999 and 2000, peaking at 229,000 lb in 2001, then decreasing to 135,000 lb in 2003.

The historical use of acetochlor, alachlor, atrazine, cyanazine, metolachlor, and S-metolachlor in Iowa is also shown in figure 1. Acetochlor was introduced in 1994 with its registration contingent on it reducing the use of other corn herbicides. As a preemergent herbicide, acetochlor rapidly replaced alachlor. As can be seen in figure 1, acetochlor usage in Iowa was about 2 million lb in 1994 increasing to 9.5 million lb in 2003 compared to alachlor usage which was at its peak of more than 9 million lb in 1992 declining to about 250,000 lb in 1998 (U.S. Department of Agriculture, 2004). Atrazine usage in Iowa stayed steady between 7 and 8.5 million lb during 1990 to 2003 regardless of the two decreases in the maximum application rate of atrazine recommended on the manufacturers' labels during the 1990s (Scribner and others, 2005). The manufacture of cyanazine was discontinued in 1999, with distribution of existing supplies discontinued on September 30, 2002 (Scribner and others, 1998). Cyanazine usage in Iowa declined to about 1 million lb in 1998 (fig. 1). The formulation of metolachlor has been refined to include primarily the active S-isomer (S-metolachlor), which reduced required application rates substantially (Benbrook, 2001). Iowa usage of metolachlor and S-metolachlor can be seen as metolachlor peaked at a rate of 12 million lb in 1997, then declined to about 600,000 lb by 2003 while S-metolachlor rapidly increased from usage of 2.7 million lb in 2001 to more than 3 million lb in 2001 (fig. 1).

Previous Studies

The effects of agricultural and urban land use on the quality of Iowa's rivers and streams have been assessed during the last 30 years by several State and Federal agencies. Monitoring networks have been established by the Iowa Department of Natural Resources (IADNR) and the U.S. Geological Survey (USGS) as described in the following sections.

Iowa Department of Natural Resources

The IADNR Ambient Water Monitoring Program conducts an ongoing assessment of the condition of Iowa's ground- and surface-water resources and reports the results to the public (Iowa Department of Natural Resources, 2001). Beginning in 1995, water samples were collected by IADNR personnel at a network of 84 sites on Iowa's rivers located upstream and downstream from Iowa's larger urban areas. Figure 2 shows IADNR ambient stream-monitoring sites in relation to USGS streamflow-gaging stations.

U.S. Geological Survey Monitoring Networks

The USGS National Stream Accounting Network (NASQAN) was begun in 1973 to provide nationally comparable information on the status of and trends in water quality (Hooper and others, 1997). Water-quality measurements were made quarterly at sites at the downstream end of most hydrologic accounting units to be consistent with the design of the national streamflow-gaging network. In 1995, the NASQAN program was redesigned to focus on monitoring water quality in four of the Nation's largest rivers, of which the Mississippi River Basin is the largest. The chemical constituents measured include 52 water-soluble pesticides. Samples are collected from 6 to 15 times per year at various NASQAN sites. Iowa NASQAN sites are shown in relation to USGS gaging stations in figure 3.



▲ U.S. Geological Survey streamflow-gaging station

Figure 2. Location of Iowa Department of Natural Resources (IADNR) ambient stream-monitoring sites in relation to U.S. Geological Survey streamflow-gaging stations (modified from Iowa Department of Natural Resources, 2001).

In 1991, the USGS began the National Water-Quality Assessment (NAWQA) Program. The long-term goals of this program are to describe the status and trends in the quality of a large, representative part of the Nation's surface- and groundwater resources and to identify the major factors that affect the quality of the resources. The NAWQA Program has identified more than 50 study units throughout the United States for which water quality was assessed (Gilliom and Hamilton, 2006; Gilliom and others, 2006). The Eastern Iowa Basins (EIWA) study unit was selected as an important hydrologic system representative of an agricultural area in the Midwest. The EIWA study unit encompasses the Wapsipinicon, the Cedar, the Iowa, and the Skunk River Basins in eastern Iowa and southern Minnesota as shown in figure 3.

Surface-water data collection activities in the EIWA study unit began in March 1996, and during the high-intensity phase of data collection (1996–98), water-quality samples were collected monthly from 12 sites on rivers and streams in the study unit. Two of these 12 sites were former NASQAN locations (Iowa River at Wapello—station 05465500; Skunk River at Augusta—station 05474000). As part of the trends network of NAWQA, 4 of the 12 EIWA sites continue to be sampled (fig. 3). Data are published annually in the Iowa annual water data report (Nalley and others, 2001). Continuous stream-discharge data are an integral part of the program, and thus streamflow gages are located at all NAWQA sites (fig. 3).

Description of Current Study

More than 26,000 mi of rivers and perennial streams drain the approximately 56,000 mi² of Iowa (Iowa Department of Natural Resources, 2001). Many of the major rivers drain directly to the Mississippi River, but a number of rivers in western Iowa drain to the Missouri River. To meet the objectives of this study, 10 sites were selected near the mouth of major rivers draining Iowa for sampling. Samples were collected monthly from March to September at each site, and high-flow samples were collected at 7 of the 10 sites



Figure 3. Location of Eastern Iowa Basin (EIWA) stream-monitoring trend sites and National Stream Accounting Network (NASQAN) sites in Iowa in relation to U.S. Geological Survey streamflow-gaging stations.

(stations 05412500, 05418600, 05422000, 06485500, 06607500, 06609500, and 06810000; table 1) and analyzed for relevant constituents. The location of sampling sites is shown in figure 4, and sampling-site drainage areas are given in table 1.

Purpose and Scope of Report

The main purpose of this report is to document the occurrence of the herbicide isoxaflutole and two of its degradation products (diketonitrile and benzoic acid) in 10 major rivers in Iowa that drain the Missouri and Mississippi Rivers (table 1) during the spring and summer of 2004. As part of quantifying isoxaflutole and its degradation products in rivers, the report describes an analytical method, using solid-phase extraction (SPE) and liquid chromatography/mass spectrometry/mass spectrometry (LC/MS/MS), developed by the USGS Organic Geochemistry Research Laboratory (OGRL), Lawrence, Kansas. The report also provides the results of analysis of waterquality samples for the occurrence of acetamide and triazine herbicides and their degradation products by two separate

Map number (fig. 4)	U.S. Geological Survey site identification number	Site name	Drainage area (square miles)
	Ν	Aississippi River Basin	
1	05412500	Turkey River at Garber	1,545
2	05418600	Maquoketa River near Spragueville	1,635
3	05422000	Wapsipinicon River near DeWitt	2,330
4	05474000	Skunk River at Augusta	4,303
5	05490500	Des Moines River at Keosauqua	14,038
		Missouri River Basin	
6	06485500	Big Sioux River at Akron	8,424
7	06607500	Little Sioux River near Turin	3,526
8	06609500	Boyer River at Logan	871
9	06810000	Nishnabotna River above Hamburg	2,806
10	06904010	Chariton River near Moulton	740

 Table 1. Sampling sites and drainage areas on 10 lowa rivers draining to the Mississippi and Missouri Rivers, 2004.

LC/MS methods at the OGRL. The USGS National Water-Quality Laboratory (NWQL), Lakewood, Colorado, analyzed for about 52 pesticides and degradation products.

Methods

Sample Collection and Processing

During 2004, samples were collected monthly from March to September using methods described in Shelton (1994) and equal-width increment sampling, which is a collection of composite samples across a section of stream with equidistant verticals and equal transit rates within each vertical that yields a representative sample of stream condition (Webb and others, 1999). The water samples were filtered through a 0.7-µm pore-size baked glass-fiber filter, using an aluminum plate filter holder and a ceramic-piston fluid-metering pump with all Teflon tubing, into precleaned 123-mL amber glass bottles. All sampling equipment was cleaned with a Liquinox solution, rinsed thoroughly with tapwater, then rinsed with organic-free or reagent water, followed by a final rinse with methanol and organic-free water to remove traces of methanol. The equipment then was air dried. The filtrate was collected in six heat-cleaned 125-L amber glass bottles. The remainder of the water in the compositing container was used for onsite measurements of specific conductance, pH, and water temperature (Wilde and others, 1998).

All samples were chilled immediately and shipped to the USGS OGRL in Lawrence, Kansas, and NWQL in Lakewood, Colorado, for analyses. Samples were refrigerated at approximately 4 °C until analysis.

Analytical Methods

Analysis of herbicides was provided by the USGS OGRL and the USGS NWQL as described in this section.

Organic Geochemistry Research Laboratory

An analytical method was developed using vacuum manifold SPE and LC/MS/MS to measure isoxaflutole [5cyclopropyl-4-(2-methylsulfonyl-4-(trifluoromethylbenzoyl) isoxazole] and its degradation products, diketonitrile [1-(2methylsulfonyl-4-trifluoromethylphenyl)-2-cyano-3-cyclopropyl propan-1, 3-dione] and benzoic acid [2-methylsulphonyl-4(trifluoromethyl) benzoic acid]. Chemical structures of these compounds are shown in figure 5. A detailed description of this method is provided in the following section. Analytical methods using SPE and LC/MS were developed to analyze water samples for acetamide and triazine herbicides (Scribner and others, 2000b). Herbicide compounds, method analysis, and reporting limits are listed in table 2.

Analysis of Isoxaflutole and Its Degradation Product

An analytical method for isoxaflutole and its degradation products was developed at the OGRL. The method includes SPE with LC/MS/MS and uses electrospray ionization (ESI) in negative-ion mode. The analytical method used to analyze isoxaflutole and its degradation products is currently (2006) undergoing USGS evaluation.

Materials

Isoxaflutole, diketonitrile, and benzoic acid were obtained from the U.S. Environmental Protection Agency (USEPA) repository (Fort Meade, Maryland). The internal



Figure 4. Location of sampling sites on 10 lowa rivers draining to the Mississippi and Missouri Rivers.

standard (ISTD), 2-4, dichlorophenoxyacetic acid- d_3 , was obtained from Cambridge Isotope Laboratory (Andover, Massachusetts). The surrogate standard, alachlor ethanesulfonic acid (ESA)-D₅, was provided by Monsanto (St. Louis, Missouri). All solvents were HPLC (high-performance liquid chromatography) grade, and the acetic acid was ACS (American Chemical Society) certified.

Preparation of Standards

Individual solutions of the ISTD, 2-4, dichlorophenoxyacetic acid-d₃, and the surrogate standard, alachlor ESA-d₅, were prepared, using volumetric procedures, at a concentration of 0.615 ng/ μ L methanol from 1-mg/mL stock solution. An intermediate standard mix containing isoxaflutole, diketonitrile, and benzoic acid was prepared by diluting individual 1-mg/mL stock solutions into acetonitrile to a concentration of 12.3 ng/ μ L. Using volumetric procedures, a working standard mix of 0.123 ng/ μ L was prepared from the intermediate standard mix by diluting it 1:100 into 9:1 formic acid (1.1 percent aqueous) in reagent water and acetonitrile.

Calibration curve solutions were prepared for extraction in new 4-oz amber bottles by adding 75 mL reagent water, 0.5 mL of 1-mg/mL aqueous humic acid solution, and 0.25 mL water pollution (WP) minerals A and B. The working standard mix then was added to each calibration standard to obtain final concentrations of 0, 0.005, 0.010, 0.050, 0.100,



soxaflutole [5-cyclopropyl-4-(2-methylsulfonyl-4-(trifluoromethylbenzoyl) isoxazole]



Diketonitrile of isoxaflutole [1-(2-methylsulfonyl-4trifluoromethylphenyl)-2-cyano-3-cyclopropyl propan-1, 3-dione]



Benzoic acid of isoxaflutole [2-methylsulfonyl-4-(trifluoromethyl) benzoic acid]

Figure 5. Chemical structures of isoxaflutole and isoxaflutole degradation products, diketonitrile, and benzoic acid.

and 0.200 μ g/L. Finally, reagent water was added to bring the final volume of the calibration solutions to 123 mL. The bottles then were capped and mixed well.

Solid-Phase Extraction

Blank samples, standards, and environmental samples were prepared by adding 100 μ L of 0.615-ng/ μ L surrogate and 100 μ L of glacial acetic acid to all bottles. One hundred microliters (100 μ L) of 0.615-ng/ μ L working standard solution was added to each matrix spike bottle. SPE cartridges were

prepared by passing 5 mL of 50/50 methanol and acetonitrile and 5 mL of reagent water slowly through each cartridge. Samples were loaded onto the cartridges within 15 to 20 minutes under vacuum. Cartridges were washed by adding 3 mL of aqueous 1-percent formic acid under vacuum. Compounds then were eluted from the cartridge with two 4-mL aliquots of 50/50 methanol/acetonitrile to each cartridge. Then 100 μ L of 0.615-ng/ μ L ISTD solution were added to each sample tube. Sample eluates were reduced to a volume of 200 μ L using a Zymark turbovap at 45 °C. Sample extracts were transferred to 200- μ L autosampler vials and stored in a freezer at -10 °C until analysis.

Liquid Chromatography/Mass Spectrometry/Mass Spectrometry

Samples were analyzed on an Agilent 1100 series liquid chromatography (LC) system (Wilmington, Delaware) and a Waters Quattro Micro API triple-stage quadrupole mass spectrometer (MS/MS) system in electrospray negative ionization mode (Milford, Massachusetts). A Keystone 250- x 4.6-mm C-18 analytical column was used to separate isoxaflutole and its degradation products (Bellefonte, Pennsylvania). The LC column was equilibrated with the mobile phase for approximately 2 hours prior to analysis. Method reporting limits are given in table 2.

Analysis of Acetamide and Triazine Herbicides and Their Degradation Products

Method number 0-2139-03 was used to analyze 6 acetamide parent herbicides and 26 acetamide degradation products. This method is identified by the USGS OGRL with the method analysis code of LCPD (Lee and Strahan, 2003). The method reporting limit for LCPD is $0.02 \mu g/L$ (table 2).

Method number 0–2138–02 was used to analyze 6 triazine parent herbicides, 12 triazine degradation products, 3 phenylurea parent compounds, and 1 phenylurea degradation product. This method is identified by the USGS laboratory with the method analysis code of LCEA (Lee and others, 2002). The method reporting limits for LCEA were 0.025 μ g/L for all triazine herbicides except deethylcyanazine, for which the reporting limit was 0.20 μ g/L. The method reporting limit for phenylurea herbicides was 0.20 μ g/L (table 2).

National Water-Quality Laboratory

The NWQL analyzed 106 samples for 52 pesticides and degradation products using laboratory schedule 2001. Analytes for this laboratory schedule are listed in table 3. Details of the analytical methods for NWQL schedule 2001 can be found in Zaugg and others (1995), Lindley and others (1996), and Madsen and others (2003). Various laboratory reporting limits are reported in table 3.

Table 2. Herbicide compounds, laboratory method analysis codes, parameter codes, method numbers, and reporting limits analyzedby the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, for water samples collected from10 lowa rivers, 2004.

[USGS, U.S. Geological Survey; µg/L, micrograms per liter; ESA, ethanesulfonic acid; OXA, oxanilic acid; SAA, sulfynil acetic acid; --, USGS method under review]

Herbicide compound	Laboratory method analysis code	USGS parameter code	USGS method number	Method reporting limit (µg/L)
	Acetamide parent	herbicides		
acetochlor	LCPD	49260X	O-2139-03	0.02
alachlor	LCPD	46342X	O-2139-03	.02
dimethenamid	LCPD	61588X	O-2139-03	.02
flufenacet	LCPD	62481X	O-2139-03	.02
metolachlor	LCPD	39415X	O-2139-03	.02
propachlor	LCPD	04024X	O-2139-03	.02
	Acetamide degrada	tion products		
acetochlor deschloro	LCPD	63778U	O-2139-03	.02
acetochlor ESA	LCPD	61029Y	O-2139-03	.02
acetochlor hydroxy	LCPD	63784U	O-2139-03	.02
acetochlor OXA	LCPD	61030Y	O-2139-03	.02
acetochlor SAA	LCPD	62847W	O-2139-03	.02
acetochlor/metolachlor ESA-2nd amide	LCPD	62850W	O-2139-03	.02
acetochlor/metolachlor-2nd amide	LCPD	63782U	O-2139-03	.02
alachlor deschloro	LCPD	63777U	O-2139-03	.02
alachlor ESA	LCPD	50009Y	O-2139-03	.02
alachlor ESA-2nd amide	LCPD	62849W	O-2139-03	.02
alachlor hydroxy	LCPD	63783U	O-2139-03	.02
alachlor OXA	LCPD	61031Y	O-2139-03	.02
alachlor SAA	LCPD	62848W	O-2139-03	.02
alachlor-2nd amide	LCPD	63781U	O-2139-03	.02
dimethenamid deschloro	LCPD	63779U	O-2139-03	.02
dimethenamid ESA	LCPD	61951W	O-2139-03	.02
dimethenamid OXA	LCPD	62482W	O-2139-03	.02
dimethenamid hydroxy	LCPD	64045U	O-2139-03	.02
flufenacet ESA	LCPD	61952W	O-2139-03	.02
flufenacet OXA	LCPD	62483W	O-2139-03	.02
metolachlor deschloro	LCPD	63780U	O-2139-03	.02
metolachlor ESA	LCPD	61043Y	O-2139-03	.02
metolachlor OXA	LCPD	61044Y	O-2139-03	.02
metolachlor hydroxy	LCPD	63785U	O-2139-03	.02
propachlor ESA	LCPD	62766W	O-2139-03	.05

Table 2. Herbicide compounds, laboratory method analysis codes, parameter codes, method numbers, and reporting limits analyzedby the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, for water samples collected from10 lowa rivers, 2004.—Continued

[USGS, U.S. Geological Survey; µg/L, micrograms per liter; ESA, ethanesulfonic acid; OXA, oxanilic acid; SAA, sulfynil acetic acid; --, USGS method under review]

Herbicide compound	Laboratory method analysis code	USGS parameter code	USGS method number	Method reporting limit (µg/L)
Ac	etamide degradation pr	oducts—Continued		
propachlor OXA	LCPD	62767W	O-2139-03	0.02
	Isoxaflutole paren	t herbicide		
isoxaflutole	LCIX	62733U		.003
	lsoxaflutole degrada	tion products		
benzoic acid	LCIX	62656U		.003
diketonitrile	LCIX	63011U		.002
	Phenylurea parent	herbicides		
diuron	LCEA	50374S	O-2138-02	0.20
fluometuron	LCEA	38811S	O-2138-02	.20
linuron	LCEA	38478S	O-2138-02	.20
	Phenylurea degrada	ation product		
demethylfluometuron (DMFM)	LCEA	61755S	O-2138-02	.20
	Triazine parent h	erbicides		
atrazine	LCEA	39632S	O-2138-02	.025
bromacil	LCEA	04029U	O-2138-02	.025
cyanazine	LCEA	04041S	O-2138-02	.025
prometon	LCEA	04037U	O-2138-02	.025
propazine	LCEA	38535S	O-2138-02	.025
simazine	LCEA	04035S	O-2138-02	.025
	Triazine degradation	on products		
deethylatrazine	LCEA	04040S	O-2138-02	.025
deethylhydroxyatrazine	LCEA	62676S	O-2138-02	.025
deethyldeisopropylatrazine/didealkylatrazine	LCEA	04039S	O-2138-02	.025
deisopropylatrazine	LCEA	04038S	O-2138-02	.025
deisopropylhydroxyatrazine	LCEA	62678S	O-2138-02	.025
hydroxyatrazine	LCEA	50355S	O-2138-02	.025
hydroxysimazine	LCEA	63154U	O-2138-02	.025
cyanazine acid	LCEA	61745S	O-2138-02	.025
cyanazine amide	LCEA	61709S	O-2138-02	.025
deethylcyanazine	LCEA	61749S	O-2138-02	.20
deethyleyanazine acid	ICEA	617508	0_2138_02	025
deethyleyanazine amide	LCEA	617518	$O_{-2138} O_{2}$.025
ucemyrcyanazine annue	LUEA	01/313	0-2136-02	.023

Quality Assurance

All water-quality measurements and water samples for this study were collected by USGS personnel in accordance with a written work plan for the study (on file with the USGS in Iowa City, Iowa). Field blank and equipment blank samples were collected to ensure cross contamination did not occur due to contaminated equipment. Replicate samples were collected to document analytical variability.

Organic Geochemistry Research Laboratory

Results of herbicide concentrations in qualityassurance samples collected during 2004 are shown in table 4. All samples were collected in duplicate or more. Analytical control was maintained by the use of carryover blanks, laboratory duplicates, laboratory spiked samples, and calibration verification standards. Types of quality assurance/quality control (QA/QC) samples collected during the study for acetamide parent and degradation products included 2 field blank, 1 field replicate, 10 laboratory duplicate, and 8 laboratory spiked samples. QA/QC samples collected for triazine parent compounds and degradation products included one field blank, seven laboratory duplicate, and five laboratory spiked samples. QA/QC samples collected for isoxaflutole and degradation products included seven laboratory duplicate and eight laboratory spiked samples. Herbicides were not detected in any of the field or laboratory blanks.

National Water-Quality Laboratory

Surrogate herbicides were added to all environmental and blank samples analyzed for pesticides by the USGS NWQL. Surrogates in samples analyzed by laboratory schedule 2001 (table 3) were diazinon- d_{10} and alpha HCH- d_6 . Surrogate herbicides were similar in chemical properties to some target analytes but were not expected to be present in the environmental samples. Surrogate herbicides are used to assess extraction efficiency and to detect sample-handling problems in the laboratory. The percentage recoveries of surrogate compounds analyzed is shown in table 5. Diazinon- d_{10} had a maximum of 131 and minimum of 65 percent recovery of the 106 water samples analyzed, whereas alpha HCH- d_6 had a maximum of 114 and a minimum of 58 percent recovery, which indicates an overall acceptable method performance.

Occurrence of Isoxaflutole, Acetamide, and Triazine Herbicides and Their Degradation Products

A summation of the number and percentage of detections of herbicides and their degradation products analyzed by the USGS OGRL are presented in table 6. Of the 75 samples analyzed, isoxaflutole was only present in four samples, and these were collected during the postplanting (May–June) season. During this season, there were 27 samples analyzed for isoxaflutole at a 15-percent detection rate. This concurs with prior findings that isoxaflutole has a very short half-life of less than 24 hours and rapidly converts to diketonitrile (Pallett and others, 2001; Lin and others, 2002). After application, isoxaflutole becomes herbicidally active through opening of the isoxazole ring, becoming the diketonitrile degradation product. Then, diketonitrile is transformed further to a herbicidally inactive benzoic acid degradation product (Viviani and others, 1998). Diketonitrile (56 detections) and benzoic acid (43 detections) were present in samples collected from all three sampling periods. However, as shown in table 6, the maximum number and percentage of detections for both compounds occurred during the postplanting season with diketonitrile detected in all 27 samples (100 percent) and benzoic acid in 23 of the 27 samples (85 percent).

The concentration results for isoxaflutole, diketonitrile, and benzoic acid are shown in figure 6 and table 7. Results show that the maximum concentration of isoxaflutole was 0.077 μ g/L, whereas diketonitrile had a maximum concentration of 0.552 μ g/L, and the highest concentration for benzoic acid was 0.166 μ g/L. Maximum concentrations for all three compounds appeared during the postplanting season.

Because isoxaflutole is applied at a low application rate and has a very short half-life, a stable analytical method proved necessary to reach acceptable method detection and reporting limits. The method developed to analyze isoxaflutole and its degradation products by LC/MS/MS was an important breakthrough and contributed to an improved understanding of the occurrence, persistence, and transport of isoxaflutole and its degradation products in the environment.

Detections for five acetamide herbicides and their degradation products are summarized by number and percentage in table 6, which shows that metolachlor was the most frequently detected acetamide parent (59 of 60 samples) during all three sampling periods, followed by acetochlor (41/60). Ethanesulfonic acid (ESA) and oxanilic acid (OXA) degradation products of acetochlor (60/60 ESA and 60/60 OXA), alachlor (56/60 and 53/60), and metolachlor (60/60 and 60/60) were detected as frequently or more frequently than their parent compounds during all three sampling periods.

Acetochlor, acetochlor ESA and OXA, metolachlor, and metolachlor ESA and OXA were detected at a 100-percent detection (29/29) frequency in water-quality samples collected during the postplanting season. Acetochlor was detected at a 100-percent detection frequency in water-quality samples collected during the postplanting season (table 6).

As shown in figure 7A and in table 8 at the back of this report, acetochlor had the highest concentration (8.6 μ g/L) of all the analyzed acetamide compounds in samples collected during the postplanting season. The maximum concentrations of acetochlor ESA and OXA collected during the postplanting

Pesticide compound	Laboratory reporting limit	Pesticide compound	Laboratory reporting limit
	(micrograms per liter)		(micrograms per liter)
2,6-diethylaniline	0.006	fipronil sulfone	.024
acetochlor	.006	fonofos	.003
alachlor	.005	lindane	.004
alpha-HCH	.005	linuron	.035
atrazine	.007	malathion	.027
azinnhas mathul	050	matalachlar	0.006
hanfluralin	.050	metribuzin	0.000
butulate	.010	melinete	.000
	.004	nanranamida	.005
	.041		.007
carboruran	.020	p,p -DDE	.005
chlorpyrifos	.005	parathion	.010
cis-permethrin	.006	parathion-methyl	.015
cyanazine	.018	pebulate	.004
dacthal (DCPA)	.003	pendimethalin	.022
deethylatrazine (DEA)	.006	phorate	.011
1 10 10 11	012		010
desulfinylfipronil	.012	prometon	.010
desulfinylfipronil amide	.029	propachlor	.025
diazinon	.005	propanil	.011
dieldrin	.009	propargite	.023
disulfoton	.021	propyzamide	.004
EPTC (s-ethyl dipropylthiocarbamate)	.004	simazine	.005
ethalfluralin	.009	tebuthiuron	.016
ethoprophos	.005	terbacil	.034
fipronil	.016	terbufos	.017
fipronil sulfide	.013	thiobencarb	.010
			007
		tri-allate	.006
		trifluralin	.009

Table 3. Pesticide compounds and laboratory reporting limits analyzed by the National Water-Quality Laboratory, Lakewood, Colorado, using laboratory schedule 2001 for water samples collected from 10 lowa rivers, 2004.

season were considerably less than their parent herbicide at 2.9 and 2.3 μ g/L, respectively (fig. 7*A*).

The highest concentration for metolachlor was 3.8 μ g/L from samples collected during the postplanting season; however, metolachlor ESA had a maximum concentration of 2.9 μ g/L, which was analyzed from a sample collected during the late summer (July–September) (fig. 7*B*). As can be seen from the analytical results in table 8, ESA and OXA

products of acetochlor, alachlor, dimethenamid, and metolachlor were detected almost as frequently or more frequently than their parent herbicides. There were no detections of alachlor deschloro, alachlor hydroxy, alachlor 2nd amide, dimethenamid hydroxy, propachlor, propachlor ESA, and propachlor OXA.

Measurable amounts of atrazine, the most frequently detected triazine parent herbicide during all three sampling periods (64 detections of 67 samples), occurred in 19 of Table 4. Concentrations of herbicides and their degradation products in quality-assurance samples collected from 10 lowa rivers as determined by the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, 2004.

[ESA, ethanesulfonic acid; FB, field blank; FR, field replicate; LD, laboratory duplicate; LS, laboratory spike; OXA, oxanilic acid; SAA, sulfynil acetic acid; <, less than; --, not detected]

				Data of		•	Concentratio	n, in microar	ams per liter	
Map number (fig. 4)	U.S. Geological Survey site identification number	Site name	Sample type	collection (month/ day/ year)	Collection - time (24-hour)	Aceto- chlor	Aceto- chlor deschloro	Aceto- chlor ESA	Ace- tochlor hydroxy	Aceto- chlor OXA
		Acetamide	e parent and	degradation p	oroducts					
1	05412500	Turkey River at Garber	LD	03/15/04	1100	0.04	1	0.17	1	0.22
			LD	04/20/04	0950	<.02	ł	.16	ł	.05
			LS	06/10/04	1000	1.1	1.3	1.3	ł	1.2
2	05418600	Maquoketa River near Spragueville	LD	05/19/04	0800	.49	.02	.43	1	44.
			LD	06/10/04	1300	.03	.03	.82	1	.42
3	05422000	Wapsipinicon River near DeWitt	FB	05/18/04	1140	<.02	1	<.02	1	<.02
			ΓS	05/18/04	1200	.56	1	.95	1	1.0
			ΓS	05/25/04	1430	1.2	1.5	.80	1	1.2
5	05490500	Des Moines River at Keosauqua	LD	07/19/04	1600	.10	.02	.90	0.08	.59
9	06485500	Big Sioux River at Akron	LD	04/13/04	0630	<.02	1	.10	1	.14
			LS	05/12/04	1030	1.2	1	.92	1	1.0
7	06607500	Little Sioux River near Turin	LS	03/10/04	0830	.88	1	1.2	1	1.1
			FB	05/11/04	1000	<.02	<.02	<.02	1	<.02
			LS	05/24/04	1100	1.3	1.5	96.	1	1.0
8	06609500	Boyer River at Logan	FR	03/12/04	0630	<.02	1	.08	1	.15
			LD	06/17/04	1000	.28	.02	.36	1	.71
6	06810000	Nishnabotna River above Hamburg	LD	03/11/04	1030	<.02	1	.07	1	60.
			LS	04/12/04	1300	1.0	1	1.0	1	1.1
			LD	05/25/04	1045	.72	<.02	.43	1	69.
10	06904010	Chariton River near Moulton	LD	05/17/04	1100	.06	.02	.22	1	.35
			TS	07/19/04	1230	1.1	66.	1.0	1.1	62.

Table 4. Concentrations of herbicides and their degradation products in quality-assurance samples collected from 10 lowa rivers as determined by the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, 2004.

[ESA, ethanesulfonic acid; FB, field blank; FR, field replicate; LD, laboratory duplicate; LS, laboratory spike; OXA, oxanilic acid; SAA, sulfynil acetic acid; </ less than; --, not detected]

				Concentrati	ion, in microgram	ıs per liter			
Map number (fig. 4)	Acetochlor SAA	Acetochlor/ metolachlor 2nd amide	Acetochlor/ metolachlor ESA - 2nd amide	Alachlor	Alachlor deschloro	Alachlor ESA	Alachlor hydroxy	Alachlor 0XA	Alachlor SAA
			Acetamide _F	parent and degra	adation products-	Continued			
1	0.08	<0.02	0.05	<0.02	1	0.26	ł	0.07	<0.02
	<.02	<.02	.03	<.02	<0.02	.40	ł	.03	<.02
	1.0	.86	.53	1.1	1.2	1.2	ł	1.1	.94
2	.13	<.02	60.	<.02	<.02	.49	ł	.02	.04
	.06	<.02	.13	<.02	<.02	.49	ł	.02	<.02
3	<.02	<.02	<.02	<.02	<.02	<.02	ł	<.02	<.02
	1.0	.93	.58	66.	1.1	1.0	ł	86.	.92
	1.4	1.1	.60	1.3	1.6	1.2	ł	1.3	.95
5	.30	<.02	.35	<.02	<.02	.32	<0.02	.02	<.02
9	.04	1	.03	<.02	1	.07	ł	.03	<.02
	LL.	1.0	.54	1.1	2.5	.92	ł	1.0	.76
L	1.2	1	.26	.93	1	.93	ł	1.1	1.6
	<.02	<.02	<.02	<.02	<.02	<.02	ł	<.02	<.02
	1.1	.80	.46	1.0	1.4	1.1	ł	1.0	96.
8	.06	1	.07	<.02	1	60.	ł	.06	<.02
	.31	<.02	.05	<.02	<.02	<.02	I	<.02	<.02
6	<.02	1	.05	<.02	ł	.08	I	.05	<.02
	1.0	<.02	.54	1.0	<.02	1.0	ł	1.1	1.0
	.34	<.02	.03	<.02	<.02	.02	ł	.02	.11
10	.06	<.02	.03	<.02	<.02	.05	ł	.02	<.02
	1.1	.61	1.2	1.1	.97	1.1	96.	.84	LL.

Table 4. Concentrations of herbicides and their degradation products in quality-assurance samples collected from 10 lowa rivers as determined by the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, 2004.

Flufenacet <.02 ESA <0.02 <.02 <.02 <.02 <.02 <.02 <.02 <.02 .95 .03 <.02 .02 <.02 .98 1.3 1.0 1.0 1.0 [ESA, ethanesulfonic acid; FB, field blank; FR, field replicate; LD, laboratory duplicate; LS, laboratory spike; OXA, oxanilic acid; SAA, sulfynil acetic acid; <, less than; --, not detected] Flufenacet <.02 <0.02 <.02 <.02 <.02 <.02 .95 <.02 .94 <.02 <.02 .10.94 .26 <.02 <.02 1.1 Ξ 5 Dimethen amid 0XA 0.02 <.02 <.02 <.02 <.02 <.02 <.02 9. <.02 <.02 .12 02 .03 1.1 1.3 1.2 0.1 5 1.1 1.2 Dimethenamid hydroxy <0.02 ł ł ł ł Ł ł ł ł ł ł ł ł H ł ł ł. ł ł Acetamide parent and degradation products—Continued **Concentration, in micrograms per liter** Dimethenamid ESA 0.05 <.02 <.02 <.02 <.02 .12 <.02 .03 .06 .02 .85 <.02 .06 .03 0 Ξ. L-∟. 2 deschloro enamid Dimeth-<.02 <.02 <.02 <.02 <.02 <.02 <.02 0.1 ł ł ł 5.1 ł ł. ł ł ł ł ł enamid Dimeth-0.02 <.02 <.02 .11 <.02 .92 <.02 <.02 <.02 <.02 <.02 .02 .03 .84 2.6 .95 42 0.1 0.1 ESA - 2nd Alachlor amide <0.02 <.02 <.02 <.02 <.02 <.02 <.02 <.02 76. <.02 <.02 <.02 <.02 <.02 .71 c. 1.2 $\tilde{\mathbf{c}}$ -Alachlor amide <0.02 <.02 <.02 <.02 .65 <.02 <.02 <.02 <.02 ć.02 <.02 2nd .79 1.01.1 1.1 1.0 ł ł ł ł number (fig. 4) Map -10 \sim 3 S 9 ∞ 6

90

0.1

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Table 4. Concentrations of herbicides and their degradation products in quality-assurance samples collected from 10 lowa rivers as determined by the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, 2004. —Continued

Map				Concentrati	on, in microgram	s per liter			
number (fig. 4)	Flufenacet 0XA	Metolachlor	Metolachlor deschloro	Metolachlor hydroxy	Metolachlor ESA	Metolachlor 0XA	Propachlor	Propachlor ESA	Propachlor 0XA
			Acetamide pa	irent and degrads	ation products—C	continued			
1	<0.02	0.07	1	1	0.39	0.12	<0.02	<0.05	<0.02
	<.02	<.02	1	1	.63	.07	<.02	<.05	<.02
	.62	1.0	1.2	1	1.3	1.1	1.1	<.05	.51
2	<.02	.19	<.02	1	1.7	.16	<.02	<.05	<.02
	<.02	.10	<.02	ł	1.9	.28	<.02	<.05	<.02
c,	<.02	<.02	<.02	1	<.02	<.02	<.02	<.05	<.02
	.52	.80	1	1	1.1	1.0	.86	<.05	.41
	.91	1.1	1.4	ł	.63	1.2	1.3	<.05	ΤΤ.
5	<.02	.27	<.02	0.08	1.6	.33	<.02	<.05	<.02
9	<.02	.08	1	1	.48	.13	<.02	<.05	<.02
	.71	1.1	1	1	.72	.91	1.0	<.05	44.
7	.30	96.	1	1	1.5	1.1	.92	<.05	.29
	<.02	<.02	<.02	1	<.02	<.02	<.02	<.02	<.02
	.54	96.	1.2	1	66.	1.0	96.	<.05	.42
8	<.02	.14	1	1	.21	.25	<.02	<.05	<.02
	.03	.48	<.02	1	.29	.23	<.02	<.05	<.02
6	<.02	.03	1	1	.14	.14	<.02	<.05	<.02
	.48	96.	ł	ł	1.1	1	89.	<.05	4.

<.02 <.02 .33

<.05 <.05 <.05

<.02 <.02 .94

.45 .28 .94

> .23 1.3

> > 2.2

54.

| |

<.02 .07 1.4

2

.08 1.4

.02 <.02

10

.86

Table 4. Concentrations of herbicides and their degradation products in quality-assurance samples collected from 10 lowa rivers as determined by the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, 2004. —Continued

[ESA, ethanesulfonic acid; FB, field blank; FR, field replicate; LD, laboratory duplicate; LS, laboratory spike; OXA, oxanilic acid; SAA, sulfynil acetic acid; <, less than; --, not detected]

	II C Pacinal			Date of			Concentratio	n, in microgra	ams per liter	
Map number (fig. 4)	u.s. Geological survey site identification number	Site name	Sample type	collection (month/ day/ year)	Collection time (24-hour)	Atrazine	Deethyl- atrazine	Deethyl- hydroxy- atrazine	Deisopro- pyl- atrazine	Deiso- propyl- hydroxy- atrazine
		Triazin	e parent and	d degradation	products					
1	05412500	Turkey River at Garber	LD	05/19/04	1200	3.1	0.35	<0.025	0.15	<0.025
2	05418600	Maquoketa River near Spragueville	LS	03/15/04	1700	.83	.94	.76	.85	1.1
			LS	03/29/04	1500	.83	80.	1.0	.87	1.0
3	05422000	Wapsinicon River near DeWitt	LS	05/18/04	1200	<.025	.70	1.1	.78	.82
			LD	06/16/04	1230	1.4	.36	<.025	.14	<.025
5	05490500	Des Moines River at Keosauqua	LS	06/14/04	1400	.70	.75	1.3	.81	.81
			LD	09/13/04	1330	.19	60.	<.025	.04	.03
9	06485500	Big Sioux River at Akron	LD	03/09/04	1200	.06	.04	<.025	<.025	<.025
7	06607500	Little Sioux near Turin	FB	05/11/04	1000	<.025	<.025	<.025	<.025	<.025
8	06609500	Boyer River at Logan	LD	05/23/04	1015	18	1.9	.53	1.2	.03
6	06810000	Nishnabotna River above Hamburg	LD	04/12/04	1300	.04	.04	<.025	<.025	<.025
			LD	05/10/04	1230	24	1.1	<.025	.42	<.025
10	06904010	Chariton River near Moulton	LS	04/22/04	1320	.81	.74	1.0	.67	96.

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Table 4. Con Survey Organ	centrations of he iic Geochemistry	rbicides and t r Research Lat	cheir degradation boratory, Lawren	products in quality- ce, Kansas, 2004. —	assurance samp -Continued	les collected fron	n 10 Iowa rivers as c	determined by the	U.S. Geological
[ESA, ethanesu	lfonic acid; FB, fiel	d blank; FR, fiel	d replicate; LD, lab	pratory duplicate; LS, li	aboratory spike; OX	A, oxanilic acid; SA	A, sulfynil acetic acid;	<, less than;, not de	stected]
Mon				Cont	centration, in mic	rograms per liter			
iviap number (fig. 4)	Didealkyl- atrazine	Hydroxy- atrazine	Bromacil	Cyanazine	Cyanazine amide	Cyanazine acid	Deethylcy- anazine	Deethyl- cyanazine acid	Deethyl- cyanazine amide
				Triazine parent and c	Jegradation produ	Icts-Continued			
1	0.36	0.11	<0.025	<0.025	<0.025	<0.025	<0.20	<0.025	<0.025
2	.72	.85	.79	1.2	1.2	1.2	1.1	1.0	1.2
	.88	1.1	.87	1.4	1.5	1.5	1.5	1.1	1.4
3	77.	1.1	.66	1.0	1.2	1.1	1.1	1.5	1.1
	.23	.14	<.025	<.025	<.025	<.025	<.20	<.025	<.025
5	66.	1.1	69.	1.1	1.4	.97	1.2	1.2	1.4
	.10	.19	<.025	<.025	<.025	<.025	<.20	<.025	<.025
9	<.025	.29	<.025	<.025	<.025	<.025	<.20	<.025	<.025
7	<.025	<.025	<.025	<.025	<.025	<.025	<.20	<.025	<.025
8	1.2	.08	<.025	.03	.24	<.025	.11	1.4	<.025
6	.26	.08	<.025	<.025	<.025	<.025	<.20	.52	<.025
	.13	.87	<.025	<.025	<.025	<.025	<.20	.32	<.025
10	.73	1.1	.59	1.0	1.3	1.1	1.1	1.5	1.1

Table 4. Concentrations of herbicides and their degradation products in quality-assurance samples collected from 10 lowa rivers as determined by the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, 2004. —Continued [ESA, ethanesulfonic acid; FB, field blank; FR, field replicate; LD, laboratory duplicate; LS, laboratory spike; OXA, oxanilic acid; SAA, sulfynil acetic acid; <, less than; --, not detected]

Map				Concentration, in I	micrograms po	er liter		
number (fig. 4)	Prometon	Propazine	Simazine	Hydroxysimazine	Diuron	Fluometuron	Demethyl- fluometuron	Linuron
			Triazine pa	ent and degradation pr	roducts-Con	tinued		
1	<0.025	<0.025	<0.025	<0.025	<0.20	<0.20	<0.20	<0.20
2	1	.86	.89	.82	.73	06.	1.0	.95
	1	.86	.79	1.1	1.0	1.1	1.1	1.1
3	.82	.70	.70	.84	.78	06.	.84	.91
	<.025	<.025	<.025	<.025	<.20	<.20	<.20	<.20
5	<.025	.67	.72	1.0	.94	.94	98.	.86
	<.025	<.025	<.025	<.025	<.20	<.20	<.20	<.20
9	1	<.025	<.025	<.025	<.20	<.20	<.20	<.20
L	<.025	<.025	<.025	<.025	<.20	<.20	<.20	<.20
8	<.025	.13	.08	.04	<.20	<.20	<.20	<.20
6	1	<.025	<.025	<.025	<.20	<.20	<.20	<.20
	<.025	.22	.08	<.025	<.20	<.20	<.20	<.20
10	1	.85	.70	.91	1.1	.92	.82	1.1

Table 4. Concentrations of herbicides and their degradation products in quality-assurance samples collected from 10 lowa rivers as determined by the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, 2004. —Continued

[]	ims per liter	Benzoic acid		0.005
nan;, not detected	ation, in microgra	Diketonitrile		0.011
cetic acid; <, less th	Concentra	lsoxaflutole		<0.003
SAA, sulfynil a	Collection	time (24-hour)		1000
e; OXA, oxanilic acid;	Date of	collection (month/day/year)	ation products	06/10/04
licate; LS, laboratory spik	Come C	type	soxaflutole and degrad	LD
R, field replicate; LD, laboratory dup		Site name		Turkey River at Garber
Ifonic acid; FB, field blank; Fl	U.S. Geological	survey site identification number		05412500
[ESA, ethanesu	Map	number (fig. 4)		1

.148 .100 .110 .110 .007 .007 .096 .111 .108 .103 .103 .014

.136 .077 .074 .074 <.002 <.017 .017

1130

07/21/04

LS

1445 1300 1500 0800 0800 1200 1230 0910 1230 1230

08/17/04

LS

03/29/04 05/19/04 05/18/04

LLD LLS LLS LLS LLS LLS LLD

Maquoketa River near Spragueville

05418600

2

Wapsipinicon River near DeWitt

05422000

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Des Moines River at Keosauqua

05490500

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09/14/04

LS

.120

.003

1030

08/16/04

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06/14/04

07/12/04

LD

.067 .012

.097 .020 .085

05/10/04

ΓD

Nishnabotna River above Hamburg

Big Sioux River at Akron

06485500

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06810000

Chariton River near Moulton

06904010

10

.003

.062

.096 .056

03/18/04

04/22/04 03/09/04

05/25/04

Man	U.S. Geological		Date of	Collection	Percentage recovery		
number (fig. 4)	survey site identifica- tion number	Site name	collection (month/day/ year)	time (24-hour)	alpha-HCH-d ₆	Diazinon-d ₁₀	
1	05412500	Turkey River at Garber	03/15/04	1100	105	113	
			04/20/04	0950	96	120	
			05/19/04	1200	93	120	
			05/24/04	1100	98	127	
			06/10/04	1000	100	114	
			07/21/04	1130	89	102	
			08/17/04	1445	92	102	
			09/14/04	1300	92	105	
2	05418600	Maquoketa River near Spragueville	03/15/04	1700	107	117	
			03/29/04	1500	101	118	
			04/20/04	1510	108	131	
			05/19/04	0800	93	117	
			05/25/04	1000	95	122	
			06/10/04	1300	101	115	
			07/21/04	0720	92	104	
			08/17/04	1025	92	91	
			09/14/04	1000	91	101	
3	05422000	Wapsipinicon River near DeWitt	03/17/04	0830	109	125	
			04/15/04	0830	99	118	
			05/18/05	1200	91	121	
			05/25/04	1400	100	128	
			06/16/04	1230	94	117	
			07/20/94	1200	93	98	
			08/17/04	0745	89	95	
			09/14/04	0730	94	103	
4	05474000	Skunk River at Augusta	03/17/04	1430	105	119	
		-	04/15/04	1345	96	112	
			05/18/04	0730	94	117	
			06/16/04	0900	95	121	
			07/20/04	0800	94	101	

Table 5. Percentage recoveries of compounds as determined by U.S. Geological Survey National Water-Quality Laboratory, Lakewood, Colorado, using laboratory schedule 2001 for water samples collected from 10 rivers in Iowa, 2004.

Man U.S. Geological			Date of	Collection	Percentage recovery		
number (fig. 4)	survey site identifica- tion number	Site name	collection (month/day/ year)	time (24-hour)	alpha-HCH-d ₆	Diazinon-d ₁₀	
4	05474000	Skunk River at Augusta	08/16/04	1615	91	105	
			09/13/04	1530	93	101	
-				1.000	100	110	
5	05490500	Des Moines River at Keosauqua	03/18/04	1230	108	118	
			04/22/04	0910	98	122	
			05/17/04	1400	94	115	
			06/14/04	1400	95	119	
			07/19/04	1600	97	104	
			08/16/04	1330	84	95	
			09/13/04	1330	95	99	
6	06485500	Big Sioux River at Akron	0309/04	1200	110	130	
			04/13/04	0930	102	122	
			05/12/04	1030	98	116	
			05/26/04	1300	102	124	
			06/02/04	1215	96	98	
			07/14/04	1030	88	97	
			08/11/04	0940	94	110	
			09/08/04	0730	93	98	
			09/17/04	1230	85	98	
7	0//07500		02/10/04	0020	114	105	
/	06607500	Little Sloux River near Turin	03/10/04	1400	114	125	
			04/13/04	1400	100	113	
			05/11/04	1020	97	114	
			05/24/04	1100	99	122	
			06/08/04	0850	95	108	
			06/17/04	1230	104	124	
			07/13/04	1000	85	102	
			08/10/04	0900	83	90	
			09/08/04	1100	90	96	
			09/20/04	1145	86	102	

Table 5. Percentage recoveries of compounds as determined by U.S. Geological Survey National Water-Quality Laboratory, Lakewood, Colorado, using laboratory schedule 2001 for water samples collected from 10 rivers in Iowa, 2004.—Continued

map survey site collection	
number identifica- Site name (month/day/ time (fig. 4) tion number (24-hour) alpha-HCH tion number year)	d ₆ Diazinon-d ₁₀
8 06609500 Boyer River at Logan 03/12/04 0930 109	117
04/14/04 0730 102	118
05/11/04 1300 98	108
05/23/04 1015 93	121
06/08/04 1130 95	105
06/17/04 1000 103	125
07/13/04 1230 58	65
08/10/04 1130 96	116
09/07/04 1530 93	96
9 06810000 Nishnabotna River above Hamburg 03/11/04 1030 106	115
03/29/04 1230 92	115
04/12/04 1300 97	114
05/10/04 1230 90	113
05/15/04 1045 95	122
06/07/04 1245 96	109
07/12/04 1230 78	107
08/09/04 1230 91	106
09/07/04 1230 93	98
10 06904010 Chariton River near Moulton 03/18/04 0900 108	120
04/22/04 1320 101	128
05/17/04 1100 96	121
06/14/04 1050 98	120
07/19/04 1230 98	115
08/16/04 1030 00	102
09/13/04 1030 88	93

 Table 5. Percentage recoveries of compounds as determined by U.S. Geological Survey National Water-Quality Laboratory,

 Lakewood, Colorado, using laboratory schedule 2001 for water samples collected from 10 rivers in Iowa, 2004.—Continued

Table 6. Summary of number and percentage of detections of herbicides and their degradation products analyzed by the U.S.

 Geological Survey Organic Research Laboratory, Lawrence, Kansas, for water samples collected from 10 rivers in Iowa, 2004.

[ESA, ethanesulfonic acid; OXA, oxanilic acid; SAA, sulfynil acetic acid; n, number of samples]

	N	umber of detecti	ons	Percentage of detections			
Herbicide compound	Pre- planting (March– April)	Post- planting (May–June)	Late summer (July–Sept.)	Pre- planting (March– April)	Post- planting (May–June)	Late summer (July–Sept.)	
	Isoxafluto	le parent and de	gradation products				
	n = 21	n = 27	n = 27	n = 21	n = 27	n = 27	
Isoxaflutole	0	4	0	0	15	0	
Diketonitrile	13	27	16	62	100	59	
Benzoic acid	7	23	13	33	85	48	
	Acetamic	le parent and deg	gradation products				
	n = 21	n = 29	n = 10	n = 21	n = 29	n = 10	
Acetochlor	9	29	3	43	100	30	
Acetochlor deschloro	0	22	2	0	76	20	
Acetochlor ESA	21	29	10	100	100	100	
Acetochlor hydroxy	0	0	4	0	0	40	
Acetochlor OXA	21	29	10	100	100	100	
Acetochlor SAA	15	27	6	71	93	60	
Acetochlor/metolachlor 2nd amide	0	8	0	0	28	0	
Acetochlor/metolachlor ESA 2nd amide	20	28	10	95	97	100	
Alachlor	0	3	0	0	10	0	
Alachlor ESA	21	26	9	100	90	90	
Alachlor OXA	21	25	7	100	86	70	
Alachlor SAA	0	12	0	0	41	0	
Alachlor ESA 2nd amide	5	6	0	24	21	0	
Dimethenamid	6	23	0	29	79	0	
Dimethenamid deschloro	0	1	0	0	3	0	
Dimethenamid FSA	14	22	7	67	76	70	
Dimethenamid OXA	6	18	5	29	62	50	
Flufenacet	1	14	0	5	48	0	
Flufenacet ESA	2	6	0	10	21	0	
Flufenacet OXA	0	5	0	0	17	0	
	-	-	-	~			
Metolachlor	20	29	10	95	100	100	
Metolachlor deschloro	0	2	1	0	7	10	
Metolachlor ESA	21	29	10	100	100	100	
Metolachlor OXA	21	29	10	100	100	100	
Metolachlor hydroxy	0	0	4	0	0	40	

Table 6. Summary of number and percentage of detections of herbicides and their degradation products analyzed by the U.S.Geological Survey Organic Research Laboratory, Lawrence, Kansas, for water samples collected from 10 rivers in Iowa, 2004.Continued

[ESA, ethanes	ulfonic acid; OXA	oxanilic acid; SAA,	sulfynil acetic	acid; n, number	of samples]
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	N	umber of detecti	ons	Percentage of detections			
Herbicide compound	Pre- planting (March– April)	Post- planting (May–June)	Late summer (July–Sept.)	Pre- planting (March– April)	Post- planting (May–June)	Late summer (July–Sept.)	
	Triazine	e parent and degr	adation products				
	n = 22	n = 29	n = 16	n = 22	n = 29	n = 16	
Atrazine	19	29	16	86	100	100	
Deethylatrazine	20	29	16	91	100	100	
Deethylhydroxyatrazine	0	2	0	0	7	0	
Deisopropylatrazine	7	29	14	32	100	88	
Deisopropylhydroxyatrazine	0	2	1	0	7	6	
Didealkylatrazine	21	29	12	95	100	75	
Hydroxyatrazine	21	29	10	95	100	63	
Cyanazine	0	1	0	0	3	0	
Deethylcyanazine acid	11	12	3	50	41	19	
Propazine	0	10	0	0	34	0	
Simazine	1	9	1	5	31	6	

22 (86 percent) preplanting (March–April) samples analyzed by the triazine herbicide method, 29 of 29 (100 percent) postplanting (May–June) samples, and 16 of 16 (100 percent) late summer (July–September) samples (table 6). Deethylatrazine (91 percent detection) occurred more frequently than atrazine (86 percent) in the preplanting-season samples, and both compounds were detected in all samples during the latesummer season. As can be seen in table 6, three other atrazine degradation products, deisopropylatrazine (32/100/88 percent), didealkylatrazine (95/100/75 percent), and hydroxyatrazine (95/100/63 percent), also occurred frequently during the three seasons in the 10 Iowa rivers sampled.

As shown in figure 8 and in table 9 at the back of this report, atrazine was not only the most frequently detected herbicide but also had the maximum concentration of $21 \mu g/L$ during the postplanting season. This maximum concentration occurred in a sample from the Nishnabotna River above Hamburg, Iowa (fig. 4). The atrazine degradation products deethylatrazine, didealkylatrazine, and hydroxyatrazine also were detected frequently during all three seasons. It is of importance to note that deethylcyanazine acid, a degradation product of cyanazine, also was detected during all three sampling seasons (table 6). The manufacture of cyanazine was discontinued on December 31, 1999, but the hydrologic system still contains the "fingerprint" of past cyanazine use.

There were no detections for the uracil herbicide, bromacil, nor the triazine degradation products, cyanazine amide, cyanazine acid, and deethylcyanazine amide. The triazine method used in this study also analyzes for three phenylurea herbicides, diuron, fluometuron, and linuron, and one degradation product, demethylfluometuron. There were no detections of the phenylurea parent compounds nor the degradation product, demethylfluometuron.

Analytical results for concentrations of selected pesticides and their degradation products determined by the USGS NWQL schedule 2001 are shown in table 10 at the back of this report. NWQL schedule 2001 (table 3) contains 52 parent pesticides and degradation products. Twenty-eight pesticide herbicides were detected above various reporting limits (table 10). Atrazine, deethylatrazine, a triazine degradation product, and metolachlor were detected in all of the 106 analyzed water samples. The highest concentration was atrazine with an estimated concentration of $3.9 \mu g/L$, followed by metolachlor with a concentration of $3.9 \mu g/L$.

This study reaffirms several previous findings of herbicides and their degradation products from the 1980s through the mid-1990s, including the fact that large concentrations of herbicides were flushed from cropland and transported in response to late spring and early summer rainfall and that





herbicides and their degradation products were detected year round in streams (Thurman and others, 1991, 1992, 1994; Scribner and others, 1993, 1998, 2000a,b, 2005; Meyer and others, 1996; Thurman and Fallon, 1996; Schnoebelen and others, 2003).

Another significant finding of this study comparable to previous studies revealed that herbicide degradation products commonly were detected as frequently or more frequently than their parent herbicide. These data agree with past studies of Iowa ground water and rivers (Kolpin and others, 1996, 1997, 1998, 2000, 2001, 2004; Kalkhoff and others, 1998, 2003; Battaglin and others, 2003, 2005; Scribner and others, 2004).

Isoxaflutole is applied in Iowa at a rate that is 8 percent of the rate of atrazine application and 4 percent of the rate of metolachlor application and is only applied to 24 percent of the number of acres to which atrazine is applied (U.S. Department of Agriculture, 2004). Thus, the frequent detection of the herbicidally active degradation product of isoxaflutole, diketonitrile, and its degradation product, benzoic acid, was unexpected.

The water-quality data resulting from this study are important to understanding effects of herbicides on the surface-water resources locally in Iowa and nationally in the Mississippi and Missouri Rivers. The data also can be used to provide information so that decisions regarding the development, management, and protection of these resources may be improved.

Table 7. Concentrations of isoxaflutole and two degradation products analyzed by the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, for water samples collected from 10 rivers in Iowa, 2004.

[<, less than]

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Map Dumboz U.S. Geological Survey site	Site nome	Date of collection	Collection	Concentration, in micrograms per liter			
(fig. 4)	identification number	Site name	(month/day/ year)	(24-hour)	lsoxa- flutole	Diketo- nitrile	Benzoic acid
1	05412500	Turkey River at Garber	03/15/04	1100	< 0.003	< 0.002	< 0.003
			04/20/04	0950	<.003	<.002	<.003
			05/19/04	1200	<.003	.035	.006
			05/24/04	1115	<.003	.092	.050
			06/10/04	1000	<.003	.010	.005
			07/21/04	1130	<.003	.004	<.003
			08/17/04	1445	<.003	<.002	<.003
			09/14/04	1300	<.003	<.002	<.003
2	05418600	Maquoketa River near	03/15/04	1700	<.003	<.005	<.003
		Spragueville	03/29/04	1500	<.003	<.002	<.003
			04/20/04	1510	<.003	<.002	<.003
			05/19/04	0800	<.003	.015	.005
			05/25/04	1015	<.003	.010	.005
			06/10/04	1300	<.003	.003	<.003
			07/21/04	0720	<.003	<.002	<.003
			08/17/04	1025	<.003	<.002	<.003
			09/14/04	1000	<.003	<.002	<.003
3	05422000	Wapsipinicon River near DeWitt	03/17/04	0830	<.003	.002	.005
			04/15/04	0830	<.003	<.002	<.003
			05/18/04	1140	<.003	.002	<.003
			05/18/04	1200	<.003	.007	.003
			05/25/04	1430	<.003	.027	.013
			06/16/04	1230	<.003	.018	.009
			07/20/04	1200	<.003	.017	.012
			08/17/04	0745	<.003	.002	<.003
			09/14/04	0730	<.003	.010	<.003
4	05474000	Skunk River at Augusta	03/17/04	1430	<.003	.002	<.003
			04/15/04	1345	<.003	<.002	<.003
			05/18/04	0730	<.003	.036	.006

Table 7. Concentrations of isoxaflutole and two degradation products analyzed by the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, for water samples collected from 10 rivers in Iowa, 2004.—Continued

Мар	U.S. Geological Survey site	a .	Date of collection	Collection	(in mi	Concentratio crograms pe	n, er liter
(fig. 4) identification number	Site name	(month/day/ year)	lime (24-hour)	lsoxa- flutole	Diketo- nitrile	Benzoic acid	
4	05474000	Skunk River at Augusta	06/16/04	0900	< 0.003	0.066	0.101
			07/20/04	0800	<.003	.008	.003
			08/16/04	1615	<.003	<.002	<.003
			09/13/04	1530	<.003	<.002	<.003
5	05490500	Des Moines River at Keosauqua	03/18/04	1230	<.003	.002	.003
			04/22/04	0910	<.003	.002	<.003
			05/17/04	1400	<.003	.015	.004
			06/14/04	1400	<.003	.100	.038
			07/19/04	1600	<.003	.039	.014
			08/16/04	1330	<.003	.002	.006
			09/13/04	1330	<.003	.010	.019
6	06485500	Big Sioux River at Akron	03/09/04	1200	<.003	.003	.016
			04/13/04	0930	<.003	<.002	<.003
			05/12/04	1030	<.003	.007	<.003
			05/26/04	1300	<.003	.042	.016
			07/14/04	1030	<.003	.006	.008
			08/11/04	0940	<.003	<.002	<.003
			09/08/04	0730	<.003	<.002	<.003
7	06607500	Little Sioux River near Turin	03/10/04	0830	<.003	.002	.008
			04/13/04	1400	<.003	.002	<.003
			05/11/04	1000	<.003	.004	<.003
			05/11/04	1020	.008	.090	.007
			05/24/04	1100	.003	.141	.051
			06/08/04	0850	<.003	.011	.003
			07/13/04	1000	<.003	.012	.005
			09/08/04	1100	<.003	<.002	<.003
8	06609500	Boyer River at Logan	03/12/04	0930	<.003	.002	.007
			04/14/04	0730	<.003	.002	<.003
			05/11/04	1300	<.003	.012	.003

Table 7. Concentrations of isoxaflutole and two degradation products analyzed by the U.S. Geological Survey Organic Geochemistry Research Laboratory, Lawrence, Kansas, for water samples collected from 10 rivers in Iowa, 2004.—Continued

[<, less than]								
Мар	U.S. Geological Survey site	al	Date of collection	Collection	(in mi	Concentration, in micrograms per liter		
number identification (fig. 4) number	identification number	Site name	(month/day/ year)	Time (24-hour)	lsoxa- flutole	Diketo- nitrile	Benzoic acid	
8	06609500	Boyer River at Logan	05/23/04	1015	0.006	0.552	0.166	
			06/08/04	1130	<.003	.013	.003	
			07/13/04	1230	<.003	.008	.004	
			09/07/04	1330	<.003	.007	.016	
9	06810000	Nishnabotna River above	03/11/04	1030	<.003	.003	.006	
		Hamburg	04/12/04	1300	<.003	.002	<.003	
			05/10/04	1230	.077	.378	.045	
			05/25/04	1045	<.003	.079	.048	
			06/07/04	1245	<.003	.010	.006	
			07/12/04	1230	<.003	.020	.012	
			09/07/04	1230	<.003	.003	<.003	
10	06904010	Chariton River near Moulton	03/18/04	0900	<.003	.005	.003	
			04/22/04	1320	<.003	.003	<.003	
			06/14/04	1050	<.003	.024	.036	
			07/19/04	1230	<.003	.011	.009	
			08/16/04	1030	<.003	<.002	.011	
			09/13/04	1030	<.003	.005	.007	



(A) Acetochlor and its degradation products







(B) Metolachlor and its degradation products



Figure 8. Concentrations of triazine herbicides and their degradation products in water samples collected from 10 rivers in Iowa, 2004.

Summary

Iowa is one of the most productive agricultural areas in the United States. Pesticides are needed for crops to control competing vegetation, insects, and fungus. The use of herbicides has produced substantial crop yields.

The use of isoxaflutole, which was registered in 1998, has increased rapidly in the Midwestern United States, but there is limited understanding of its environmental fate. However, it is known that isoxaflutole degrades to a biologically active degradation product, diketonitrile, which is more stable, and further degradation of diketonitrile produces a nonbiologically active benzoic acid degradation product.

The main purpose of this report is to document the occurrence of the herbicide isoxaflutole and its degradation products from samples collected from 10 major rivers draining Iowa to the Missouri and Mississippi Rivers and to compare their occurrence with that of more commonly measured herbicides such as acetochlor, atrazine, and metolachlor. Herbicide use is one factor that affects the concentrations of herbicides measured in Midwestern rivers and a vital reason for monitoring the water quality in these rivers.

An analytical method to quantify isoxaflutole and its two degradation products was developed by the U.S. Geological Survey (USGS) Organic Geochemistry Research Laboratory using solid-phase extraction (SPE) and liquid chromatography/mass spectrometry/mass spectrometry (LC/MS/MS). Two separate methods also were used to provide the results of analysis for the occurrence of acetamide and triazine herbicides, and their degradation products. The USGS National Water-Quality Laboratory schedule 2001 was used to analyze 52 pesticides and degradation products.

Analytical results of 75 water samples show that isoxaflutole was detected in only four of the samples collected during the postplanting (May-June) season, whereas diketonitrile was detected in 56 water samples and benzoic acid was detected in 43 water samples collected from all three sampling periods. Metolachlor was the most frequently detected acetamide parent (59 of 60 samples) during all three sampling periods, followed by acetochlor (41/60). Ethanesulfonic acid and oxanilic acid degradation products of acetochlor (60/60 ESA and 60/60 OXA), alachlor (56/60 and 53/60), and metolachlor (60/60 and 60/60) were detected as frequently or more frequently than their parent compounds. Atrazine was the most detected triazine parent compound with 64 detections of 67 samples during all three sampling periods, whereas the triazine degradation products of deethylatrazine, didealkylatrazine, hydroxyatrazine, and deisopropylatrazine were detected in 65, 62, 61, and 50 samples, respectively, which is similar to other reported conclusions of the detection frequency of the atrazine parent and its degradation.

Findings of the isoxaflutole study by the USGS include an improved understanding of the occurrence of the herbicide isoxaflutole and its degradation products in the hydrologic environment. Analytical results of the acetamide and triazine herbicides are consistent with previous studies, which show that large amounts of herbicides and their degradation products are flushed into streams with runoff. Also, the study confirmed previous findings that these herbicides occur as frequently or more frequently than their parent herbicide. The results of this study provide an important data set for future use.

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Supplemental Information