

ArrayTrack_{3,4,0} Demonstration

National Center for Toxicological Research U.S. Food and Drug Administration 3900 NCTR Road, Jefferson, AR 72079





ArrayTrack Overview

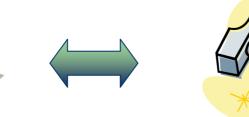
Microarray DB











An integrated environment for microarray data management, analysis and interpretation.

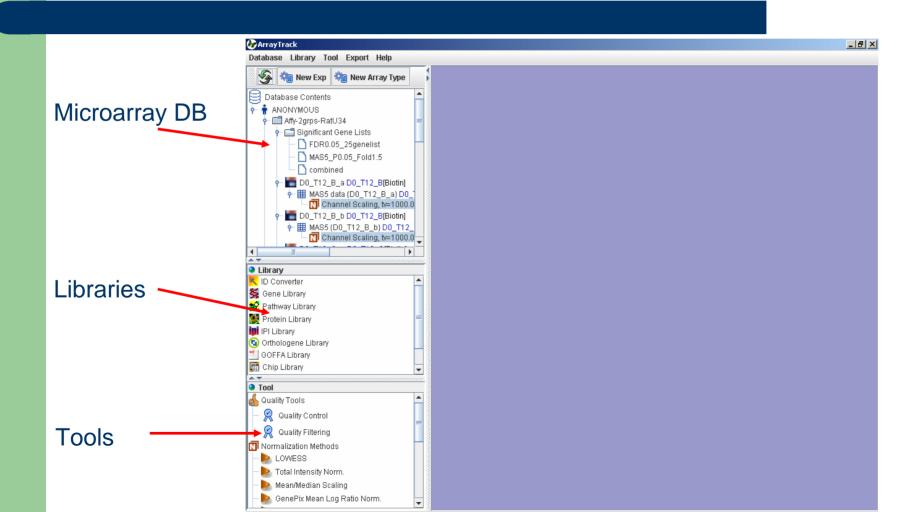
Using ArrayTrack, the user can apply analysis method from Tool to microarray DB and then get information from the linked LIB for biological interpretation





Library

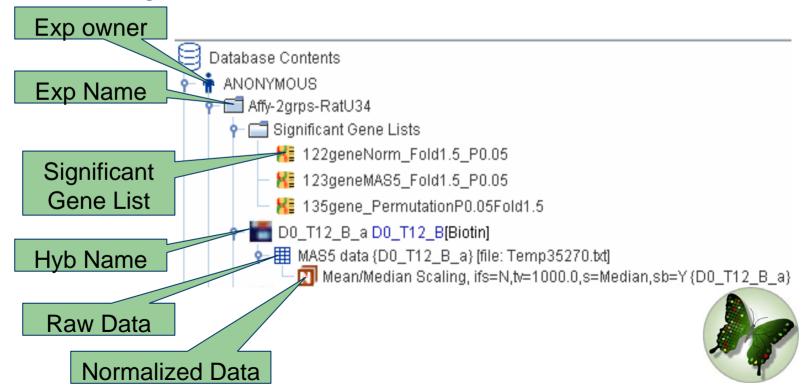
ArrayTrack Overview





Microarray DB

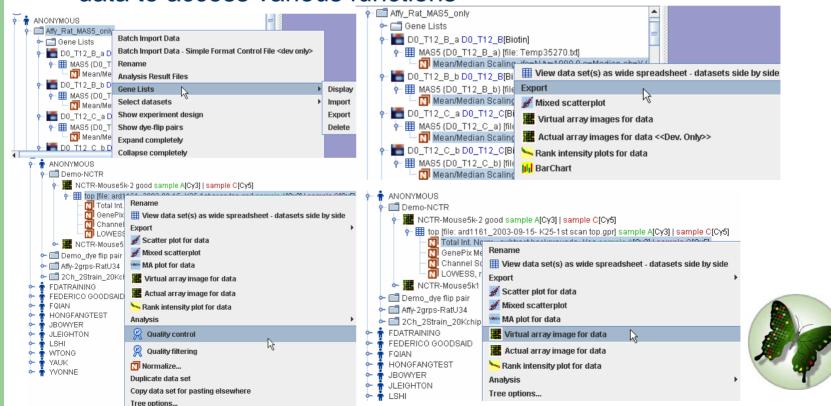
Data is organized as a hierarchical tree structure:



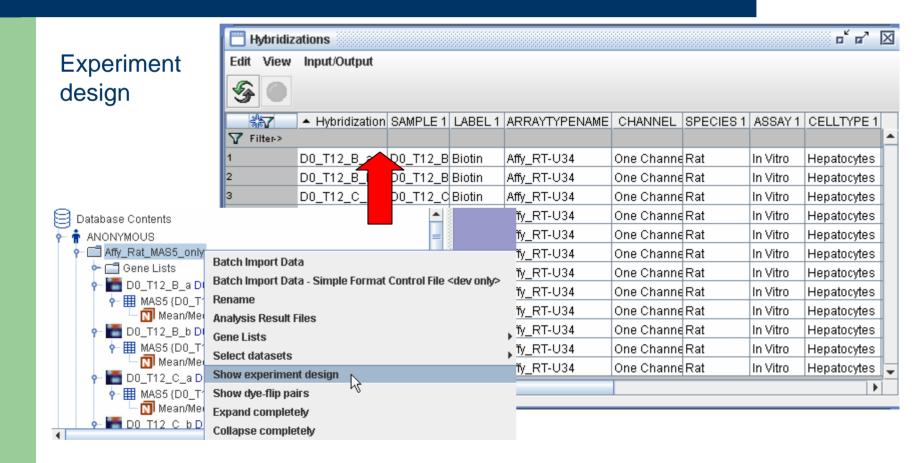


Exploring DB

 Right-click exp, hybridization, raw data and normalized data to access various functions



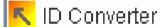
Exploring DB





Searching Libraries

Library



🥳 Gene Library

🙀 Pathway Library

🌉 Protein Library

IPI Library

📵 Orthologene Library

🐫 GOFFA Library.

👸 Chip Library

🜉 Toxicant Library

🧰 EDKB Library

There are nine libraries in ArrayTrack. All the libraries are interlinked.

The libraries integrate gene, protein, pathway and other data allowing data interrogation and mining of data across data types.

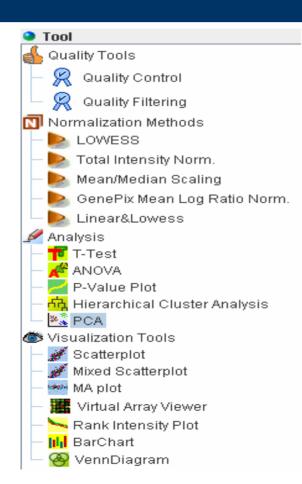
Several ways to activate these libraries

- -From library panel
- -From the Library pull-down menu
- -From the results of analysis (e.g. T-test)





Analysis Tools



Tools:

Quality Tools

Provides various visual plots and numerical parameters for measuring the quality of a hyb, and filtering the unwanted spots.

Normalization Methods

Correct systematic variations in microarray data introduced by experimental factors

Analysis

Provides different methods to evaluate the microarray data

Visualization Tools

Provides a direct view to identify abnormalities within data.





Microarray Data Processing

Importing data

Loading array data into AT

Normalization

Ensure cross-chip comparison

Gene Selection

Identify a list of significant genes

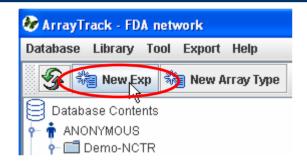
Interpretation

Interpret data using pathways and GO





Importing Data



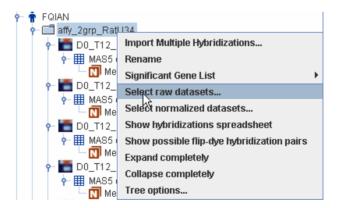
- 1. Create experiment
- 2. Create array type (if not existed in AT Chip LIB)
- 3. Use batch import wizard (see detail in Tutorial 9)

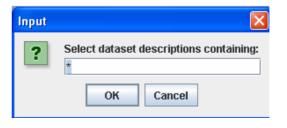
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Experiment Design		* is require	ed			
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*Exp Types:	pomperature shock; normalive :	Select	Hybridization Pr	otocol:	Import	S Export
* Key words:	rat, liver, circadian		Labeling Pro	foculer	Import	S Export
*Exp Description:	Ad lib fed rats at 6 hr. intervals; examining liver changes					
			RNA Extraction Pro	tocols:	Import	Export
* Phenotype Anchoring:	BE concernation, VITRODOSE er 2000 productione	Select	Manuscript.F	toprint:	Import	Export
Comments:			Significant Ger	se List:	Import	S Export
Comments						
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Normalization

- To remove systematic variations across chips and ensure a valid cross-chip comparison
 - 1. Right-click an experiment
 - 2. Select "raw datasets..."



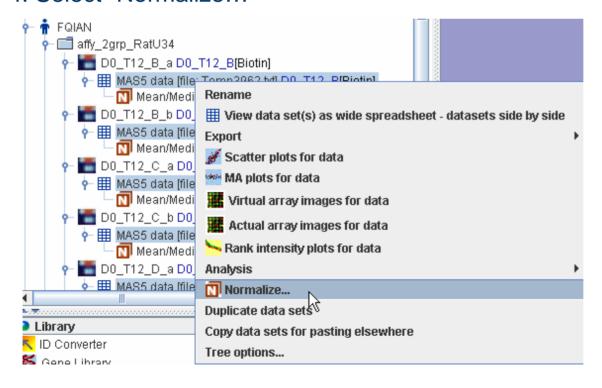






Normalization (-continued)

- 3. Right-click any highlighted raw data
- 4. Select "Normalize..."

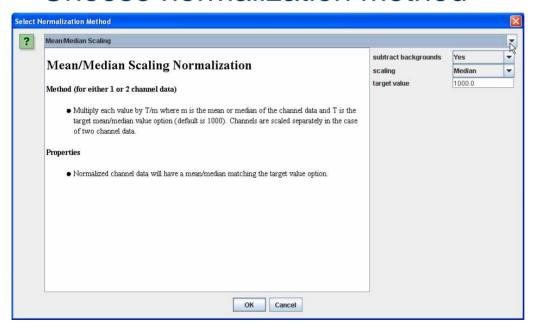






Normalization (-continued)

Choose normalization method



- •For Affy data, choose "Mean/Median Scaling"
- •For 2-channel data, the default method is "Lowess"



Gene Selections

Determining a list of genes that are differentially expressed between Strain A and Strain B

Two types of experiment: **Error rate for the exp** Single testing: 1 gene P<0.05 low error rate Multiple testing: *n* genes $P = nP_i$ If $P_i = 0.05$, high error rate e.g., If n=10 and P_i=0.05, P=0.5 for family-wise error Select a gene list based on: Bonferroni correction P value P_i/n Low sensitivity Low power False discovery rate (e.g., Benjamini & Hochberg, p-value plot)

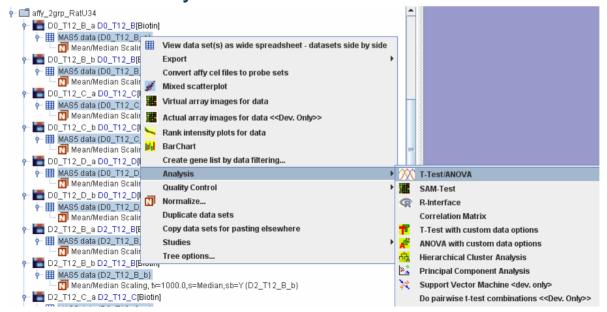
Permutation t-test (e.g., SAM)

Volcano plot (combination of p and fold change)



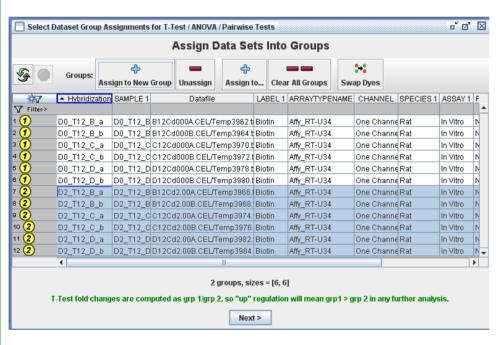
Gene Selection

- Highlight and Right-click the experiment.
- Select "Normalized datasets..." and click "OK"
- Right-click the highlighted normalized data
- Choose "Analysis->T-test"







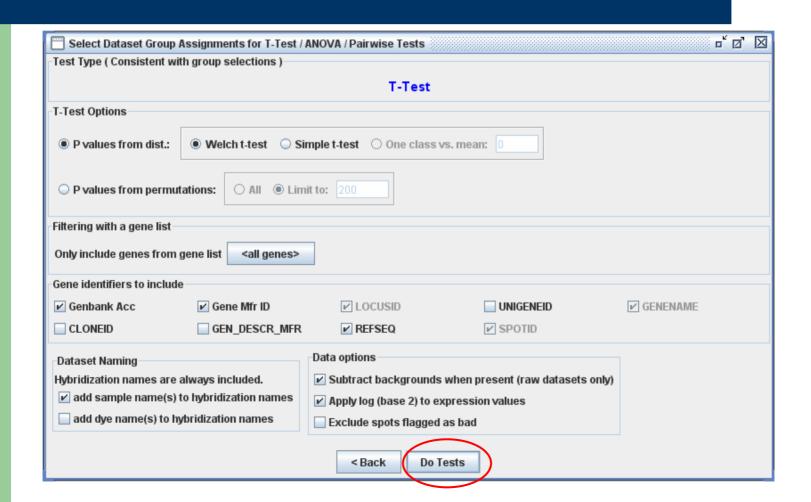


Assign the data into 2 groups

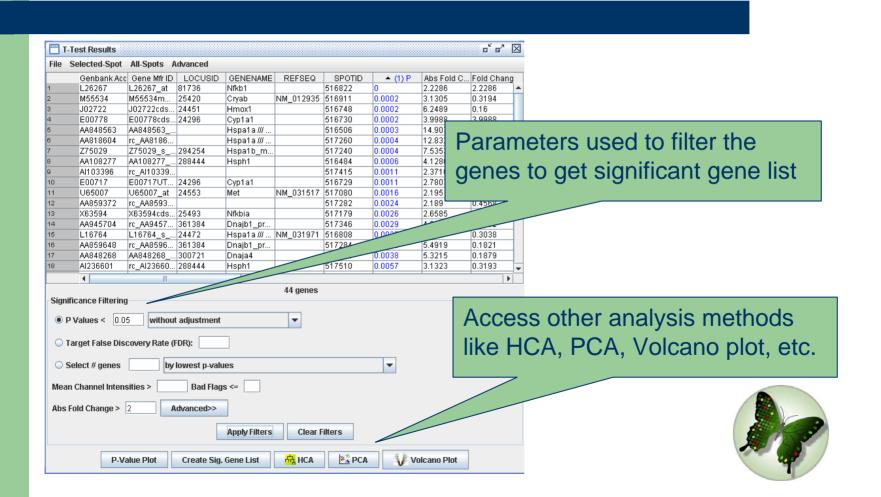
- -different dose
- -different time
- -or different animal

Note: always put control in group 2

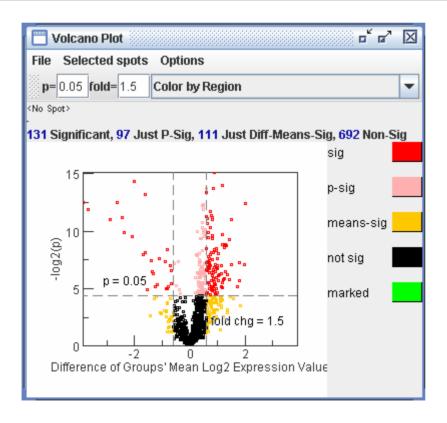












Volcano Plot



Significant Gene List

- Create
- Display
- Import
- Export
- Delete

Significant Gene List(-cont)

Create significant gene list From T-test/Anova result **6** 6 ⊠ T-Test Results File Selected-Spot All-Spots Advanced set filter criteria Create significant gene list... SPOTID Mean Grp 1 | Mean Grp 2 | Fold Change 49926 2.7783 NM 013474 6 4884 13 0868 Hierarchical Clustering Analysis 46938 3.3721 Principal Component Analysis 151257 e.g. P-value, fold change 1.3012 Library searches 🌠 Gene Library 1.5725 Select in all open viewers 🙀 Pathway Library -0.72753 2184 to get significant gene list U86105 U86105_1 18113 🔯 Protein Library 1.1372 0.2812 AK018313 AK018313 1 0.242 1.1507 10.0007 NM 008865 NM 00886... 18776 159805 0.0008 -1.748 1.8513 NM 019486 NM 01948... 56015 162758 0.0008 NM_008319 NM_00831... 15898 145874 0.0009 -0.1588 -0.4882 1.2564 NM 021447 NM 02144... 149205 0.001 0.4095 0.1626 1.1867 NM 015780 NM 01578.. Cfhl1 148342 0.0011 1.2908 3.2995 0.2485 D43759_1 156273 0.0011 5.9168 4.6198 2.4571 NM 010419 NM 01041... 15208 Hes5 157161 0.0011 0.3868 0.1695 1.1625 AF331708 AF331708 1 230103 Npr2 0.0012 0.2314 **Create significant gene list** AR041350 AR041350 1 12830 1 8511 20160 genes Significance Filtering P Values <</p> without adjustment Target False Discovery Rate (FDR): Filter criteria Select # genes with lowest p-vals: Mean Channel Intensities > Bad Flags <= Fold Change > Apply Filters P-Value Plot Create Sig. Gene List **品 HCA** E PCA Volcano Plot

Significant Gene List(-cont)

Display/Import/Export/Delete Significant Gene List

Right-click the experiment name

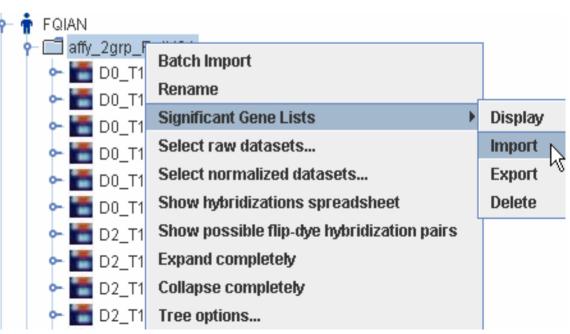
select Significant Gene Lists

select Display

Import

Export

Delete



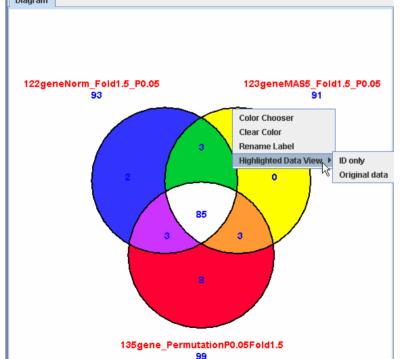
Significant Gene List(-cont)

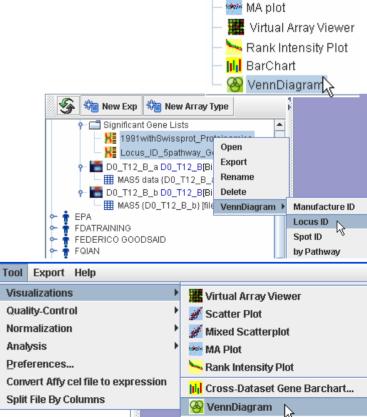
VennDiagram - get common genes from
2~3 significant gene lists

Venn Diagram on LOCUSID

Diagram

Diagram





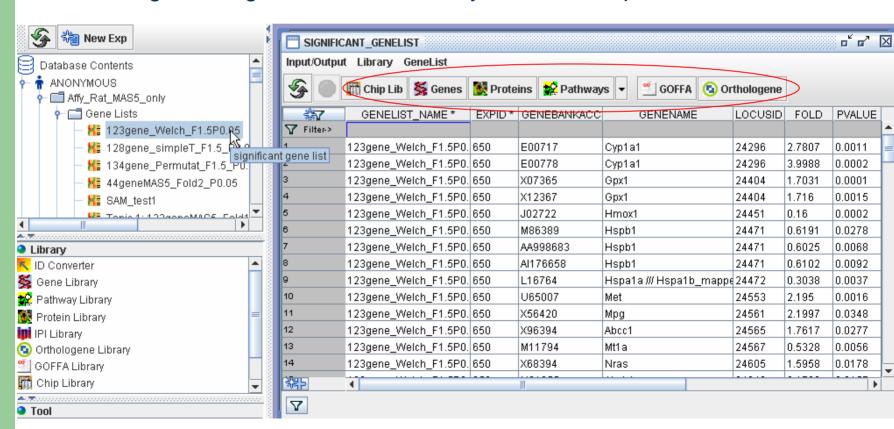
Scatterplot

🌌 Mixed Scatterplot



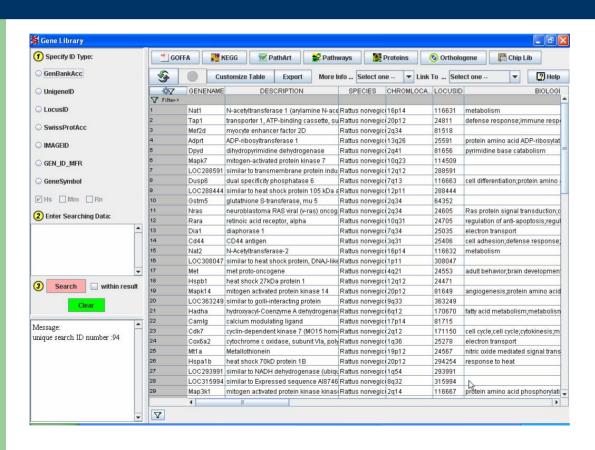
Interpretation

Link the significant genes to Gene Library for data interpretation





Interpretation (continued)



- The significant genes are listed here in Gene Library.
- Can search and sort the Gene library
- There are links to other Libraries(Kegg, Pathart)





Interpretation (continued)

- KEGG Kyoto Encyclopedia of Genes and Genomes http://www.genome.jp/kegg/
- KEGG is a suite of databases and associated software.
- KEGG Pathway database provides the information of metabolic, regulatory and disease pathways; Most of them are metabolic pathways.





Interpretation (continued)

PathArt (Jubilant) – a pathway database

- •The Pathways (over 600 mammalian disease and signaling)
- •The Pathways is a collection of manually curated information from literature and public domain databases.

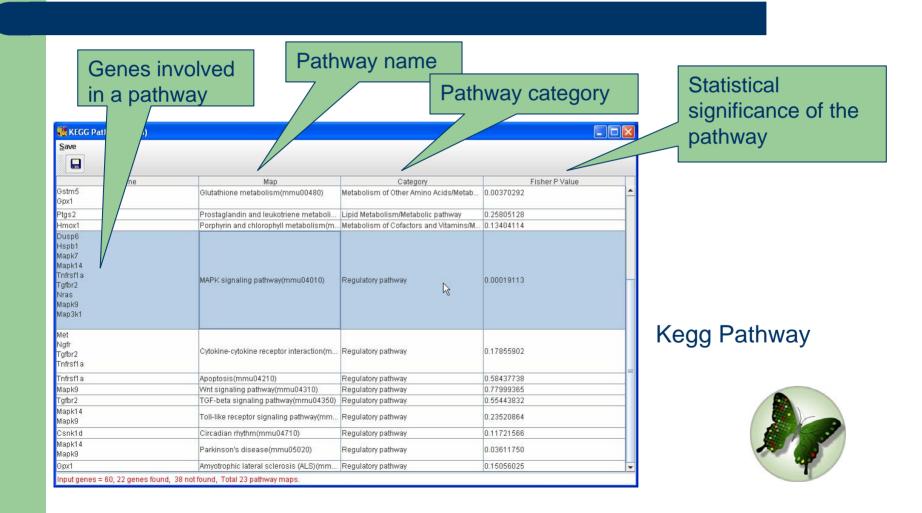
In ArrayTrack

	Human	Rat	Mouse
Kegg	200	187	193
PathArt	587	151	297





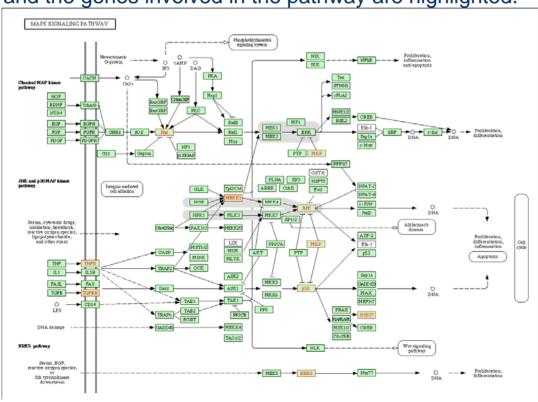
Interpretation (continued) Kegg





Interpretation (continued) Kegg

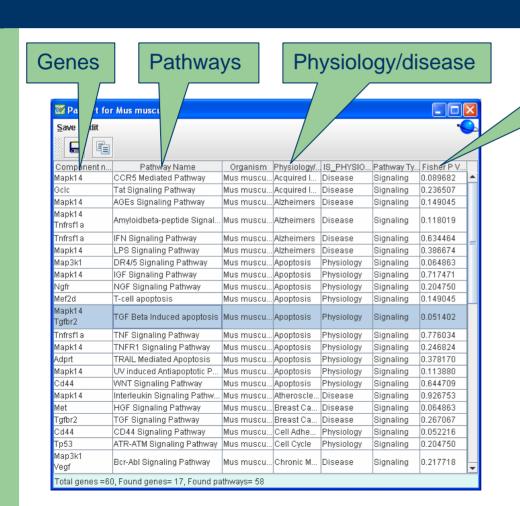
Double-click a specific pathway, the pathway map will be displayed and the genes involved in the pathway are highlighted.







Interpretation (continued) PathArt

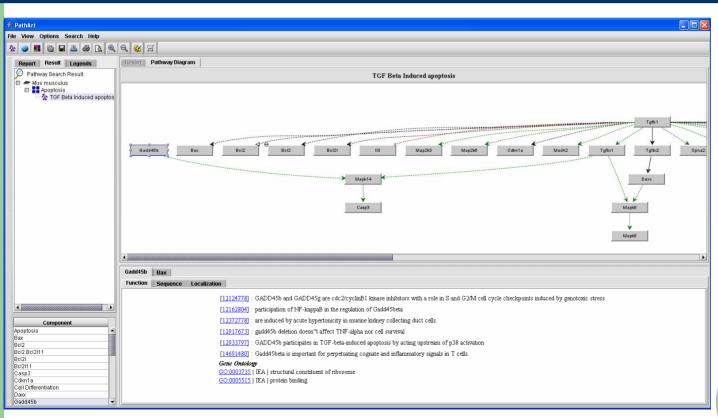


Statistical significance of the pathway





Interpretation (continued) PathArt





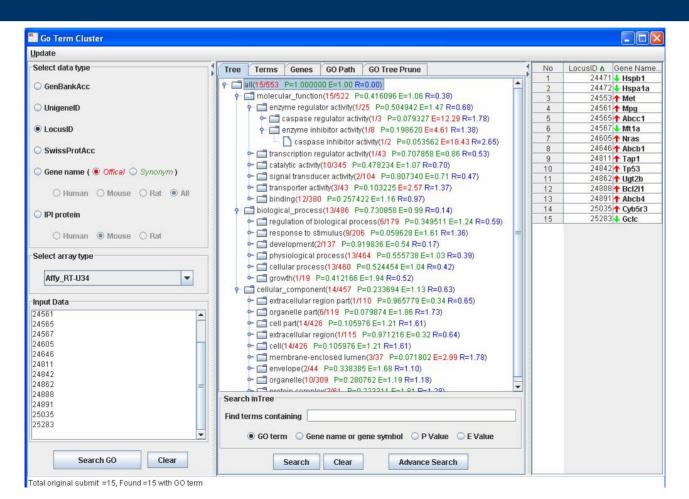


Interpretation (continued) GOFFA

- GOFFA Gene Ontology For Functional Analysis
- Developed based on Gene Ontology(GO) database
- Grouping the genes into functional classes
- GO- three ontologies
 - -Molecular function: activities performed by individual gene products at the molecular level, such as catalytic activity, transporter activity, binding.
 - -Biological process: broad biological goals accomplished by ordered assemblies of molecular functions, such as cell growth, signal transduction, metabolism.
 - -Cellular component: the place in the cell where a gene product is found, such as nucleus, ribosome, proteasome.

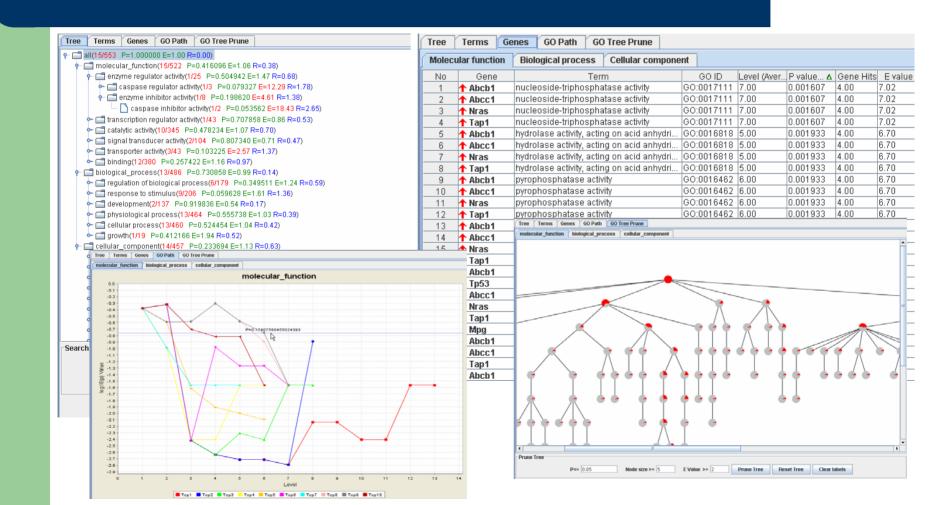


Interpretation (continued) GOFFA





GOFFA





Data Exploring

Scatter Plot

plot the fluorescence intensity data of Cy3 vs Cy5 for the same array

Mixed Scatter Plot

compare two arrays in one plot, applies to both 2-channel and 1-channel data.

Correlation Matrix

Correlation Matrix shows the correlation between column i and column j of the original matrix. It visually shows the correlation between two groups of data.

Bar Chart

displays expression data for a single gene across multiple arrays within the same experiment or across different experiment.

Principal Component Analysis (PCA)

PCA is a way of identifying the data patterns and highlighting the data's similarity and difference

P-value Plot

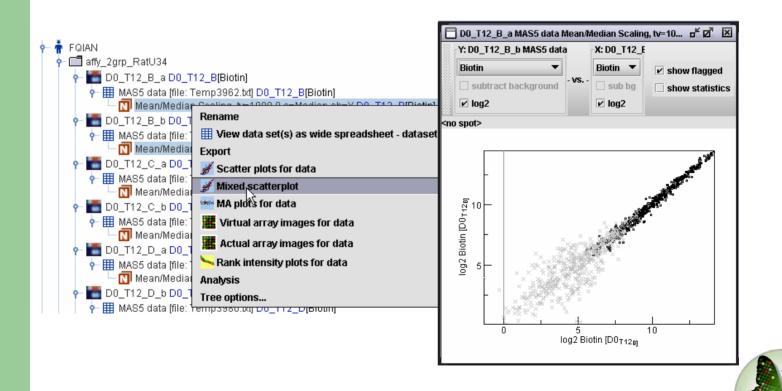
visual interpretation of P-value.

HCA

is comprised of agglomerative methods and divisive methods that finds clusters of observatic data set.

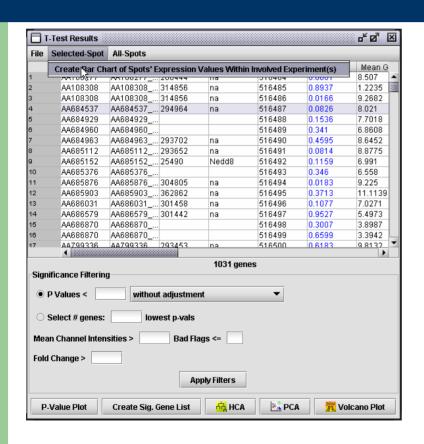


Data Exploring scatter plot





Data Exploring Bar chart



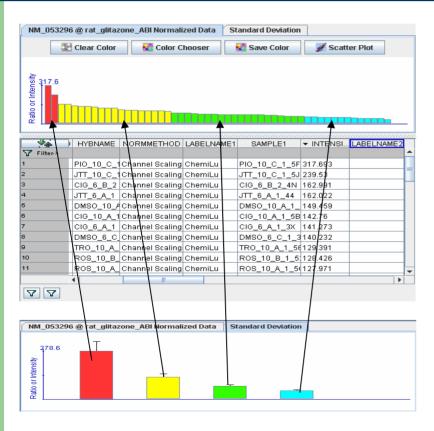
Access Bar chart from T-Test results.

Bar chart – display expression data for a single gene across multiple arrays in the same experiment or across different experiments.





Data Exploring Bar chart continued



Grouping multiple arrays marked in different colors.

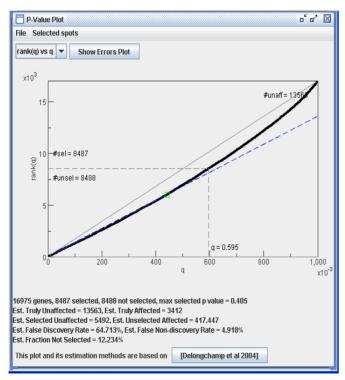
Group color could be saved.

Standard deviation bar chart for the above groups. The bar height represents the mean intensity, while the T-line above the bar stands for the value of SD. The color for each bar echoes the color of the bar chart at the top in the same order.

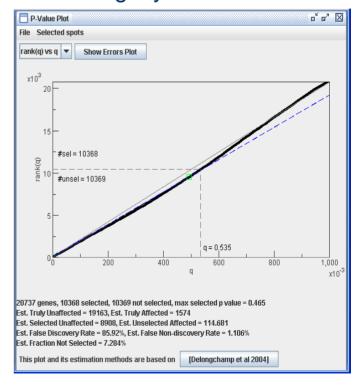
Data Exploring

P-value plot

Some effect



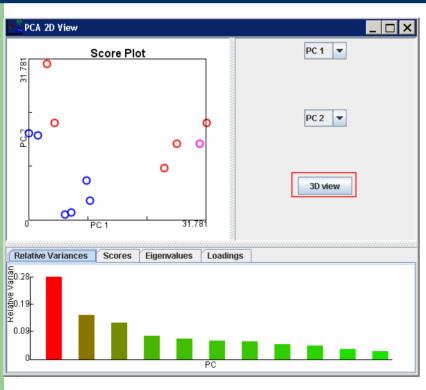
Slightly or no effect



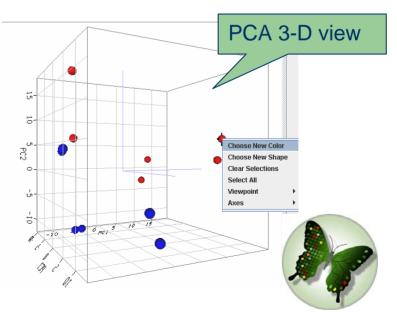
P plot curve is closer to diagonal line, the less treatment effect there is.



Data Exploring PCA

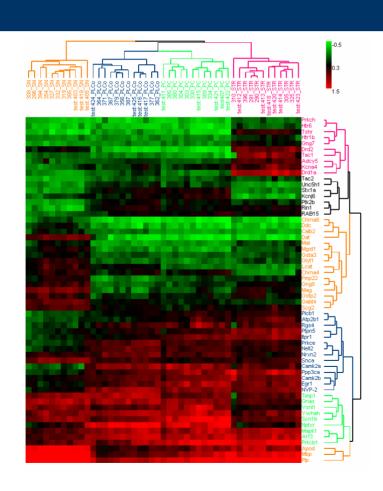


- •Commonly used before gene selection
- •To investigate the inter-sample relationship based on the gene expression profile
- •Identify the outliers in biological/technical replicates
- •View the variance of a multidimensional data





Data Exploring HCA







Accessing ArrayTrack

- FDA Internal: <u>http://weblaunch.nctr.fda.gov/jnlp/arraytrack/index.html</u>
 <u>http://weblaunch.nctr.fda.gov/jnlp/arraytrack/citrix/index.html</u>
- FDA External: http://edkb.fda.gov/webstart/arraytrack/index.html





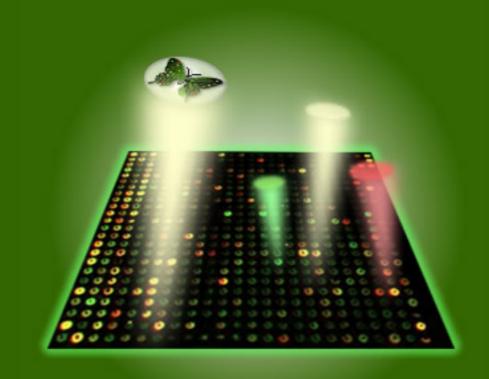
Technical Support

NCTRBioinformaticsSupport@nctr.fda.gov

ArrayTrack is developed by the U.S. Food and Drug Administration, National Center for Toxicological Research (FDA/NCTR). FDA/NCTR reserves all rights for the software.



Thank you!



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