

Glacier Bay Small Schooling Fish Project

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Project Summary

Objectives

- Sample zooplankton and fish throughout Glacier Bay to assess presence/absence of species.
- Assess relative abundance of key zooplankton and forage fish species within the bay.
- Hydroacoustically sample Glacier Bay to assess spatial patterns of pelagic fish abundance.
- Identify specific areas of particular importance associated with high abundance and/or diversity.
- Assess marine predator densities (marine birds and marine mammals).
- Link predator distributions to spatial patterns of forage fish abundance.

Key Findings

- Areas of high fish abundance can be correlated with the oceanographic or limnic features of Glacier Bay.
- Areas in close proximity to tidewater glaciers as well as in the recently non-tidewater upper-inlets support some of the highest abundances of marine life in Glacier Bay.
- The outlets from salmonid streams appear to impart a strong positive local influence on the nearshore marine ecosystem.
- Glacier Bay supports spawning stocks, and provides nursery areas for several key forage fish including sand lance, capelin, and walleye pollock.
- The areas of high euphausiid or forage fish abundance within Glacier Bay are clearly defined and encompass a relatively small part of the total bay area. These areas are critical to the biological function of the bay Upper inlets, river and stream outlets, Whidbey Passage.
- The east and west arms of Glacier Bay exhibited differing fish community structure. Capelin were much more common in the east arm than the pollock domination elsewhere in the bay and Icy Strait.
- Acoustically determined forage biomass was concentrated in relatively few areas and in shallow, nearshore waters.
- High density patches of prey were very rare, and less than 8% of the survey area contained patch densities suitable for foraging marine birds and mammals.
- Glacier Bay supports high densities of a diverse array of apex predators (marine bird and mammals) in both summer and winter.
- Marine birds and mammals in Glacier Bay were found to use predominantly coastal habitats.
- Compared to 1991 surveys, our surveys in 1999 and 2000 indicated apex predator species are generally increasing, e.g. humpback whales; but a few have declined dramatically, including harbor seals, Kittlitz's and marbled murrelets.
- Alternative survey designs were examined and could be sufficient for monitoring population trends of most of species.

<u>Abstract</u>

We studied oceanography (including primary production), secondary production, small schooling fish (SSF), and predators in Glacier Bay during 1999 and 2000. Results from these field efforts were combined with a review of current literature relating to the Glacier Bay environment. Since the conceptual model was developed by Hale and Wright (1979) 'changes and cycles' continue to be the underlying theme of the Glacier Bay ecosystem. We found marked seasonality in all parameters that we investigated and here we provide a comprehensive description of the distribution and relative abundance of a wide array of marine biota.

Glacier Bay is a tidally mixed estuary that leads into basins, which stratify in summer, with the upper arms behaving as traditional estuaries. The bay is characterized by renewal and mixing events throughout the year, and markedly higher primary production than in many neighboring southeast Alaska fjords.

Zooplankton diversity and abundance within the upper 50 meters of the water column in Glacier Bay is similar to communities seen throughout the Gulf of Alaska. Zooplankton in the lower regions of Glacier Bay peak in abundance in late May or early June, as observed at Auke Bay and in the Gulf of Alaska. The key distinction between the lower Bay and other estuaries in the Gulf of Alaska is that a second smaller peak in densities occurs in August. The upper bay behaved uniformly in temporal trends, peaking in July. Densities had begun to decline in August, but were still more than twice those observed in that region in May. The highest density of zooplankton observed was 17,870 organisms/m³ in Tarr Inlet during July. Trends in zooplankton community abundance and diversity within the lower bay were distinct from upper-Glacier Bay trends. Whereas the lower bay is strongly influenced by Gulf of Alaska processes, local processes are the strongest influence in the upper-bay.

We identified 55 species of fish during this study (1999 and 2000) from beach seines, mid-water trawls, and rod and line catches. The diversity of physical, oceanographic, and glacial chronological conditions within Glacier Bay contribute a suite of factors that influence the distribution and abundance of fish. Accordingly, we observed significant differences in the abundance and distribution of fish within the bay. Most significantly, abundance and diversity (primarily juvenile fish including walleye Pollock, eelblennies, and capelin) were greatest at the head of both the east and west arms in close proximity to tidewater glaciers and freshwater runoff.

All of Glacier Bay and Icy Strait were surveyed hydroacoustically for plankton and fish during June 1999 surveys. Acoustically determined forage biomass was concentrated in relatively few important areas such as Pt. Adolphus, Berg Bay, on the Geikie-Scidmore shelf, around the Beardslee/Marbled islands, and the upper arms of Glacier Bay. Forage biomass was concentrated in shallow, coastal waters; 50% of acoustic biomass was found at depths < 35m, 80% of biomass at depths < 80m. High density patches of prey were very rare, and less than 8% of the area surveyed in Glacier Bay contained patch densities suitable (e.g., > 0.01 fish/m³) for foraging seabirds. Less than 1% of the area contained

patches suitable (e.g., >0.1 fish/m³) for whales. High-density aggregations of 0.1-10 fish/m³ were comprised mostly of schools containing capelin, pollock, herring or euphausiids (0.1-1 kg/m³).

During predator surveys (1999-2000), we observed 63 species of birds and 7 species of marine mammals, including humpback whales, harbor seals, Kittlitz's and marbled murrelets. Seasonal distribution and abundance of these "apex" predators was highly variable by species. Glacier Bay supports high numbers of seabirds and waterfowl, although many species displayed a strong seasonality in their use of the Bay. Coastal areas had higher densities of both birds and marine mammals. Several areas, such as Pt. Adolphus, Berg Bay, on the Geikie-Scidmore shelf, around the Beardslee/Marbled islands, and the upper arms of Glacier Bay were focal points of marine bird and mammal activity. Comparisons between surveys and a prior study (1991) suggested that the assemblage of birds and marine mammals in the Bay is undergoing change. Most notable was a clear decline in *Brachyramphus spp*. murrelets and harbor seals while most other apex species are increasing.

Introduction

Reason for Work

Although this report synthesizes work from several specific projects (e.g., oceanography, plankton surveys, small-schooling-fish surveys, hydroacoustic surveys, predator surveys), the reason behind, and objectives of, any ecosystem level project in Glacier Bay can fall under the motives suggested by Catton (1995):

"In coming years, the critical test of biocentric resource management will be to assess what is happening to the park's marine environment. With greater sophistication and breadth than ever before, the NPS will attempt to differentiate between natural and anthropogenic environmental changes occurring in Glacier Bay"

Any attempt to link observed changes in Glacier Bay to either one of these causes (natural or anthropogenic) can only be speculative without a thorough understanding of the bay's ecosystem. Glacier Bay is composed of living organisms interacting both with each other and the non-living environment. These interactions should be viewed holistically, rather than as a collection of unrelated objects. This forms the foundation of the science of ecology (Hale and Wright, 1979) and is the underlying principle of this study, which quantitatively investigates the Glacier Bay ecosystem, both spatially as well as temporally. In summary, over the last 300 years as Glacier Bay has been revealed by retreating ice, a thriving marine ecosystem has developed. In this report we provide a thorough description of this ecosystem from physical processes to upper-trophic level predators as it appears in 1999 and 2000.

Glacier Bay hosts an abundant variety of marine predators during summer including significant populations of humpback whales (*Megaptera novaeangliae*) and *Brachyramphus* murrelets. The distribution and abundance of small schooling fish in Glacier Bay affects both the geographic and temporal status of many of these predators (e.g., humpback whales; Krieger and Wing, 1986). Southeast Alaska humpback whales for example, were only encountered in areas where concentrations of prey were extensive and dense (Krieger and Wing, 1986). However, with the exception of the limited work done by the NMFS Auke Bay Marine laboratory (Wing and Krieger, 1983; Krieger and Wing, 1984; Krieger and Wing, 1986), little is known about temporal and spatial patterns of forage species in the Bay (Hale and Wright, 1979). Marine predators at the trophic apex of the ecosystem, may reflect changes to other portions of the ecosystem composition and function. Understanding abundance and distribution of fish, whales, and seabirds in Glacier Bay requires knowledge about the spatial distribution and abundance of their prey. This study represents the first comprehensive survey of predators and their prey in Glacier Bay.

The distribution and health of marine predator populations, the most visible portion of marine ecosystems, depends on the abundance and distribution of their prey species. For example, declines in seabird and marine mammal populations in the Gulf of Alaska have been linked to shifts in abundance and composition of their prey species, primarily small schooling fish (SSF – see page 7 for a definition), over the past 45 years (Anderson and

Piatt, 1999). Changes in SSF populations have in turn been linked to long-term changes in the physical environment of the northern Gulf of Alaska. Small schooling fish distribution is largely influenced by local oceanography and availability of food (primarily zooplankton). Therefore, we assume that an understanding of both temporal and spatial patterns of oceanography and productivity will lead to a greater understanding of the distribution and abundance of small schooling fish and the predators that rely upon them.

Zooplankton support much of the life in the oceans as the prey of many fish, whales, and seabirds. Zooplankton abundance and composition directly influences survival and distribution of predators as well as nutrient cycling. The zooplankton community provides a nexus between primary production of the oceans and larger animals, fueling the food web of marine systems. Zooplankton includes temporary members of the planktonic community termed meroplankton (larval stages of benthic animals) and icthyoplankton (fish eggs and larvae); as well as permanent members of the community such as euphausiids, copepods, and chaetagnaths. Benthic animals such as crabs, sea urchins, sea stars, mussels, and barnacles all reproduce by releasing planktonic larval stages that disperse and as juveniles. Knowledge of zooplankton abundance and composition is key to understanding dispersal of benthic animals and the trophic web of marine systems.

Oceanography and productivity are particularly important factors to understand within the Glacier Bay ecosystem due to the complexity of sills, constrictions, large currents, and numerous glaciers. Questions, such as posed by Brattegard (1980): "Do such deep fjords have deep water faunas mainly composed of shallow water species, or do we find faunas similar to those found at corresponding depths in the ocean, or do we find elevated deep-sea faunas?" are still relevant today for fjord ecosystems like Glacier Bay. Brattegard goes on to highlight several studies that have shown deep-sea species assemblages (such as the trophically important mesopelagic myctophids; Matthews and Heimdal, 1980) living in coastal fjords. Furthermore, tidewater glaciers can significantly influence the circulation, water properties, and sedimentation of glacial fjords (Cowan, 1992), and the potential for enhanced biological activity at their faces has been known for over 60 years (Hartley and Dunbar, 1938); although Hale and Wright (1979, p83) do not make note of this in their Glacier Bay ecosystem model. Finally, in temperate fjords, such as those in Southeast Alaska, there are large diurnal and seasonal differences in freshwater and suspended sediment discharged from glaciers (Cowan, 1992). Most freshwater enters fjords during the summer from large conduits located at the base of the glacier. This turbid water, usually less dense than the more sailine ford, rises to the surface, and may result in seasonal water properties and circulation, that are unlike that of fjords where melt-water enters at sea-level (e.g., Kostaschuk, 1985).

Ecosytem-level research in Glacier Bay also provides a unique opportunity to study postglacial succession in the marine environment. Research has usually focused on vegetative colonization in Glacier Bay (e.g., Lawrence et al., 1967) and more recently the colonization of postglacial freshwater streams (e.g., Milner, 1987; Sidle and Milner, 1989). Colonization of new freshwater streams, particularly by anadromous species presumably influences the nearshore marine community. Additionally, newly exposed marine areas will also provide unique opportunities for opportunistic marine species that are able to live in highly turbid waters.

Separating natural post-glacial succession processes from natural long-term changes from anthropogenic perturbations is a challenge for both scientist and resource manager. Notable recent changes of predator distributions and abundance within Glacier Bay are humpback whales that historically made greater use of the upper bay (Hale and Wright, 1979; Wing and Krieger, 1983; Krieger and Wing, 1984; Krieger and Wing, 1986); dramatic declines in abundance of the Kittlitz's murrelet over the last decade; and the recent rapid range expansion and abundance of sea otters in the bay since first noted in 1993 (Bodkin et al. 2000). In all these cases ecosystem level research will be invaluable to separation of cause and effect, whether it be a result of anthropogenic (e.g., vessel disturbance) or natural (e.g., increased kelp in nearshore areas as a result of sea otter predation on urchins) origin.

Small Schooling Fish: A Definition

The terms "small schooling fish" (SSF) and "forage fish" have been frequently used, sometimes interchangeably in recent ecosystem studies (e.g., Exxon Valdez Trustee Council funded SEA and APEX projects). The terms are loosely defined, sometimes by species composition (e.g., Springer and Speckman, 1997) and sometimes also by size (e.g., Litzow et al., 2000). The current project was originally defined as the "small schooling fish project" and encompasses the description of forage fish used by the organizing committee for the 1996 Forage Fishes in Marine Ecosystems conference (cited in Springer and Speckman, 1997):

"Forage fishes are abundant, schooling fishes preyed upon by many species of seabirds, marine mammals, and other fish species. They provide important ecosystem functions by transferring energy from primary and secondary producers to higher trophic levels."

Springer and Speckman (1997) proposed that Gulf of Alaska SSF include Pacific sand lance, capelin, eulachon, herring, juvenile walleye pollock, lanternfishes (myctophidae), and juvenile Pacific salmon.

Marine fishing technology is still relatively primative, even though methods to find fish are highly advanced. In almost all cases, a net is put into the water that non-selectively collects fish bigger than its mesh size. Therefore, our research collects most fish in a targeted area rather than just the "small schooling fish" and our report reflects this diversity of catch. In this regard, the "Small Schooling Fish Project" has grown from the original efforts to solely characterize the prey of humpback whales. Glacier Bay is also home to numerous other predatory species that prey upon small fish - that either do not school or are demersal (solitary or schooling), and frequently left out of the 'forage' or 'SSF' definition such as Stichaeids. Furthermore, key macroinvertebrate prey species such as jellyfish, euphausiids, and cephalopods are caught in conjunction with fish catches and are all prey species for numerous predators. Although not technically SSF, we discuss all of these species, highlighting those that are of importance to specific predators within the bay.

Methods

Study Area

The rapid recession of a Neoglacial ice sheet within the last 300 years exposed Glacier Bay, a Y-shaped fjord in southeast Alaska, which is approximately 100 km long (Figure 1). Width of the bay varies from 4 to 8 km in the lower bay; widening to approximately 15 km in the middle-bay, and then narrows again in the upper-bay. The Fairweather range dominates the head of Glacier Bay, with numerous peaks over 3,000 m culminating in Mt. Fairweather at 4600 m. Numerous glaciers (12 tidewater) discharge ice and turbid water into the upper arms and inlets. Glacier Bay is connected to the Gulf of Alaska via Icy Strait (Figure 1).

Glacier Bay became a National Monument on 25 Feb 1925, and currently lies within the 11,030 km² Glacier Bay National Park and Preserve, established on 02 Dec 1980. The area was designated wilderness on 02 Dec 1980, designated a Biosphere Reserve in 1986, and a World Heritage Site in 1992.

Within Glacier Bay, numerous sills (submerged glacial moraines) separating deeper basins (up to 458 m deep) and constrictions affect water movement. Hooge and Hooge (2001) highlight the role of this bathymetry in influencing oceanography and primary productivity. Glacier Bay also experiences a very large tidal range. The tidal cycle is mixed semi-diurnal (two high and two low tides per day, of unequal heights), with a tidal range (difference between mean high and mean low tides) averaging from 3.7 m at Bartlett Cove to 4.2 m at locations approximately half-way up both the West and East arms. The tidal range further up-Bay is even greater. During the largest spring tides, the tidal range can reach 7.3 m at Bartlett Cove, and exceeds 7.8 m in the upper arms.

Glacier Bay is an important wildlife area, providing habitat and feeding opportunities for numerous marine mammals, seabirds, and commercially exploited fish species. Perhaps the best known of these wildlife species is the humpback whale. Each summer 15 to 20 humpback whales regularly feed in park waters, concentrating in the lower part of the bay. Tidewater glaciers, and wildlife species such as the humpback whale, Steller sea lion, and Kittlitz's murrelet draw numerous visitors. Despite the relativly remote setting, 442,607 tourists visited the park during 1999, most on large cruise ships.

Research Platforms

Field work for this project was conducted over two years (Table 1) and from several vessels (Figure 2). All beach seining was carried out from a 5 m aluminum hulled Naiad inflatable with 45 hp Honda 4-stroke outboard. Mid-water trawls and oceanography during 1999 were conducted from the Alaska Department of Fish and Game (ADF&G) 22 m stern trawler, the RV *Pandalus*. Oceanography and plankton samples during March and May 2000 were taken from the RV *Alaksan Gyre* (formerly RV *Tamnik*). All summer samples during 2000 (oceanography, zooplankton (after May), trawls) were from the RV *David Grey*, a 9-m Uniflight. Predator surveys were made from all the above research vessels (RV) and the *Lutris II*, an 8 m Boston Whaler.



Figure 1. Glacier Bay study area, with place names used throughout the report.

Year	Nearshore Zooplankton Hauls	Pelagic Zooplankton Hauls	Icthyoplankton Trawls	Beach Seine	Mid-Water Trawls	Predator Surveys
1999				10 Jun to 23 Jun	10 Jun to 23 Jun^H	10 Jun to 23 Jun
						5 Nov to 12 Nov
2000		17 Mar to 21 Mar				17 Mar to 22 Mar
		10 May to 17 May	24 May to 29 May			
	31 May to 9 Jun	10 Jun to 14 Jun	10 Jun to 14 Jun	31 May to 9 Jun		17 to 22 June
	12 Jul to 20 Jul	26 Jul to 31 Jul		12 Jul to 20 Jul	26 Jul to 31 Jul^{I}	
	1 Aug to 10 Aug	12 Aug to 16 Aug		1 Aug to 10 Aug	12 Aug to 17 Aug ^I	

Table 1. Time periods for full surveys of Glacier Bay by fishing and predator surveys

^H Modified Herring Trawl; ^I Isaacs Kidd Mid-Water Trawl



Figure 2. Research vessels used for the small schooling fish research project in Glacier Bay during 1999 and 2000.

Oceanography

We used three sources of oceanographic data to related physical water parameters to biological observations: 1). Previously published oceanographic information, 2). Unpublished data collected by USGS Biological Resources Division, Glacier Bay Field Station in an 9 year oceanographic investigation of Glacier Bay (Hooge and Hooge, 2001); and 3) A CTD recorder (Sea-Bird SBE 19 SEACAT Profiler profiling conductivity-temperature-depth probe (CTD; Sea-Bird Electronics, Bellevue, Washington, USA) was used to acquire oceanographic data (Figure 3). The instrument

primarily used in this study had a Sea-Bird SBE 5-01 submersible pump, a LI-COR LI-192SA photosynthetically active radiation (PAR) sensor (LI-COR, Inc., Nebraska, USA), a D&A OBS-3 turbidity sensor (D&A Intstrument Co., Port Townsend, Washington, USA) and WET Labs WetStar (WET Labs, Inc., Philomath, Oregon, USA) flurometer, in addition to the standard temperature probe, conductivity cell, and pressure port. The CTD was used in conjunction with each fishing and zooplankton tow. Casts were made to 90 % of water depth or a maximum of 240 m (Background data collected by Hooge and Hooge (2001) used a maximum depth of 335 m).

SeaSoft software modules (Sea-Bird Electronics) were used for initial processing of the raw instrument data. The data were processed through six functions; these first converted the data to engineering units, then passed conductivity and pressure through lowpass filters. The temperature and conductivity measurements were then



Figure 3. CTD apparatus ready for deployment on the *RV David Grey*.

temporally aligned to compensate for the different response times of the respective sensors. Next, all scans in which reversed pressure indicated slowdowns or failure of a minimum velocity test (< 0.25m/sec) were removed. The derived variables salinity, density, and depth in saltwater were then calculated. Finally, the data were averaged into 1-meter depth bins (for detailed processing see Hooge et al. 2000)

In this paper, depth is calculated as depth in seawater derived from pressure, temperature is presented in degrees Celsius, salinity is reported in parts per thousand (ppt) and is derived from conductivity, density is presented as sigma-t (density anomaly) in kg/m³, and fluorescence is reported in mg/m³. We did not calibrate the fluorometer for the in-situ Glacier Bay phytoplankton assemblages; therefore chlorophyll-*a* densities are most

appropriate for relative comparisons within this study and for gross comparisons with other studies. The CTD reports optical backscatter (OBS) in microvolts, which can be recalculated into mg/m 3 of sediment based on the instrument's calibration.

The OBS sensor used in this study has not yet been calibrated for the sediments found at the oceanographic stations along the length of Glacier Bay. Therefore, the voltage values do not have absolute meaning and are only presented for relative comparisons within Glacier Bay. Due to variations between casts and surveys in the time of day and amount of cloud cover, photosynthetically active radiation (PAR), which is measured in microeinsteins/ sec/ m^2 , has been standardized as a proportion of the (maximum) surface value for each cast. This standardization allows comparisons of relative light penetration between casts and surveys, but does not provide an absolute measure of PAR.

Stations established by Hooge and Hooge (2001) were used as a basis of site selection for several of the temporal collections; pelagic zooplankton stations, Tucker trawls, and Isaacs Kidd mid-water trawls. The first station is located in Icy Strait (Figure 4). Subsequent stations are spaced approximately every 9.3 km (5 nautical miles) to the head of Tarr Inlet in the West Arm and to the head of Muir Inlet in the East Arm. We chose to biologically sample every second station leading to a total of 11 regular samples sites with a separation of approximately 19 km (10 nautical miles).



Figure 4. Glacier Bay's bathymetry with pelagic study stations indicated. Locations are derived from Hooge and Hooge (2001) (exact locations are given in Appendix 1).

Hydroacoustic Survey

During 10-23 June, 1999, we conducted simultaneous hydroacoustic, predator (below), and trawl (above) surveys throughout Glacier Bay and in the eastern half of Icy Strait, from the mouth of Glacier Bay east to Lynn Canal. For these surveys we used the R/V *Pandalus*, a 22 m stern trawler operated by the Alaska Department of Fish and Game.

Hydroacoustic data were collected with a Biosonics DT4000 echosounder using a single beam 120 kHz transducer with a 6° beam angle. This transducer was capable of transmitting and receiving to depths of ca. 185m, but the quality of data collected below about 130m may be questionable. We limit most of our analyses of acoustic data to depths of less than 100m. Threshold for data collection was set at -80 dB. The transducer was attached to a hydrodynamic sled and deployed off the side of the survey vessel 1-2 m below the water surface. All data were logged directly to a computer in real time. GPS locations were obtained from a Rockwell Precision Lightweight Global-positioning Receiver (PLGR), which have a worst-case horizontal position accuracy of ± 10 m at speeds <36 kph (Anonymous 1995). At the beginning of the cruise, the hydroacoustic system (transducer, cable and sounder) was calibrated in the field using a tungsten steel sphere of known target strength.

In order to quantify forage fish biomass for GIS mapping, hydroacoustic transect data were binned into 30 second (horizontal) by 5 m (depth) bins and integrated using EchoView© (Sonar Data Pty. Ltd., Hobart, Tasmania) to determine S_A (mean backscattering per nm²) of each bin. For mapping purposes, data were further summed into 10 min bins and 25 m depth intervals. Mean backscatter (S_A) is a common currency used for spatial comparisons of hydroacoustic data (e.g., Hewitt and Demer 1993, Brodeur and Wilson 1996). The integration threshold was set at -90 dB, but because of an EchoView software error (that we later uncovered), the actual threshold for integration was 27 dB higher, i.e., -63 dB. Because we were very conservative in setting the initial threshold, the functional threshold of -63dB was still low enough to detect common forage fish such as capelin and pollock at lengths of less than 10 mm, and herring at lengths of less than 30 mm, and therefore adequate for the purpose here of describing spatial distribution of "forage fish" (typically >50mm) in Glacier Bay. As we prepare to publish these data, we plan to reanalyze the acoustic data using updated Echoview software (See Appendix 4 for detailed EchoView© protocols).

Significant schools identified on acoustic echograms were trawled to assess species composition. A total of 48 tows at 38 stations were conducted during the course of hydroacoustic surveys (see section 4.5.1). In order to assess absolute prey densities of these schools comprised of known species from trawls, relative measures of acoustic biomass (S_A) were converted to absolute estimates of fish density (fish/m²) by dividing S_A by σ (backscattering cross-sectional area of single prey) for species with the following known target strengths: Pollock TS=20Log(L_{cm})-66 (Foote and Traynor 1988); Capelin TS=20Log(L_{cm})-65 (Rose and Leggett 1988); Herring TS=26Log(L_{cm})-76 (Thomas and Kirsch 1999); Euphausiids TS= 34.8Log(L_{mm})-127.5 (Hewitt and Demer 1993); Physoclist TS=20Log(L)-65.5 (Foote 1987). Mean lengths (and ranges, in mm) of

common fish captured in trawls (previous section) were: Pollock 140.9 (12-602); capelin 65.6 (14-109); Herring 208.4 (165-235); Euphausiids \leq 30mm; All fish combined (weighted by CPUE of each species) 98.1 (10-602). Integration thresholds were set at -73 to -83 dB (corrected values) when calculating S_A for known-species schools.

<u>Zooplankton</u>

Zooplankton samples were collected using a 333µm mesh net with a 0.6-meter opening (Figure 5). Vertical hauls were taken from 50 meters, if possible, or 2 m from bottom. We sampled at even numbered physical oceanography sites (Figure 5), and immediately offshore beach seine sites (within 0.25 mile; Figure 6). In this manner our collections were synchronized with physical oceanography data collection and small schooling fish sampling at 17 nearshore and 13 mid-channel stations. Mid-channel stations were sampled for only plankton and oceanography from March to May; fish sampling and nearshore stations were added from June to August. Samples were preserved in 4% formalin.



Figure 5. Preparing the vertical-haul zooplankton net for deployment at Station 18 in upper Muir Inlet.

Samples were analyzed using a Wild dissecting scope at 30-60X magnification, according to the methods of Edmondson and Litt (1982). Samples were diluted to a known volume that varied according to abundance. Sample volume ranged from 200-1050mL (107 samples), one sample was counted entirely. Five-milliliter subsamples were pipetted out using an automated draw pipetter. We analyzed the subsamples using a modified Bogorov tray and all animals were recorded. Replicate subsamples were done with a minimum of 100 individuals of the most common taxa-lifestage grouping counted twice. The volume of water filtered was calculated for each sample and all analysis

results are quantified as number of individuals per cubic meter of water integrated over the sampling depth.

Not all species found in Glacier Bay by this study were identifiable to species. Therefore, some taxa are reported at Genus or Family taxanomic levels. Animals were identified into 44 taxa-lifestage groupings listed in Appendix 2. Copepods were not generally identified to species in samples, and species present are given in Appendix 3. Adults and copepodids were counted together, grouped by size as small (<2.5mm) or large (>2.5mm). Nauplii were counted as a separate taxa grouping. As copepod nauplii were not quantitively retained by our net, trends in nauplii abundances should be considered as relative, rather than an absolute index of abundance trends.



Figure 6. Glacier Bay nearshore zooplankton stations. Numbers and locations correspond with select beach seine stations (exact locations are given in Appendix 1).

Ichthyoplankton

Icthyoplankton samples for this study were collected from the RV *David Grey*, a 9-m glass fibre Uniflight. Horizonal plankton tows were taken at discrete depths by using a $1-m^2$ Tucker trawl, rigged with two 505-µm mesh nets (Figure 7).



Figure 7. Deploying the Tucker Trawl at Station 21 (Margerie Glacier). Note tripping mechanism holding chains connected to net frames at top of net.

Depths in the bay range to as deep as 458 m and were unattainable due to limitations of our small winch. We expected a key component of the larval fish assemblage to be walleye pollock. Based on depth distributions for this species in Prince William Sound (Müter and Norcross, 1994) we decided to obtain samples from each of the following depth strata per station: 0-25 m, 25-50 m, 50-75 m, and 75–100 m (three stations located on sills were shallower and were sampled to the bottom). Although depths were determined by wire angle and scope, precise depths were recorded using a Wildlife Computers MK7 time-depth recorder attached to the net. The nets were rigged with a double-tripper which allowed the second net to be opened and closed via a messenger from the surface. The net was towed for five minutes in the direction of tidal flow at a towing speed of 1.8 to 2.2 knots. Only daytime tows were made. Volume filtered during each tow was calculated from a General Oceanics flowmeter that was attached to the central portion of the mouth of the net. Samples used for analysis were immediately preserved in 50% isopropyl alcohol. The alcohol was renewed after 24 hours and after 2-3 days.

Littoral Zone Fish Community

We used beach seines to sample nearshore fish communities in June 1999 and in June, July, and August 2000. This fishing method effectively and non-selectively samples shallow, inshore waters with sandy or smooth bottoms (Cailliet *et al.*, 1986). Our variable-mesh net was 37 m long. The wings were tapered from 2.4 m in the middle to 0.5 m at the wing using 28 mm knotless nylon stretch mesh. The seine was equipped with a 6 mm stretch mesh bag located in the middle of the seine. Thirty meters of rope were attached to the ends for deployment. The net was set parallel to shore about 15 m from the beach as described by Cailliet *et al.* (1986; Figure 8).



Figure 8. Typical beach seine set in upper Muir Inlet.

We sampled 59 sites in Glacier Bay and Icy Strait during 1999 (Figure 9). We attempted to seine each site at both high and low tide. However, several sites could not be seined at high tide due to large rocks, or at low tide due to large rocks or mussel beds. During 2000, we streamlined the study to a more even distribution of beaches. Only those beaches that could be repeat ably sampled on both low and high tides were used (Figure 9). Four beaches from the original 1999 survey were visited to facilitate this. Several of the beaches sampled in 1999 were in close proximity to each other, particularly the case in lower regions where the bay is largely carved from a glacial outwash plain (Hale and Wright, 1979). The inlets in the upper bay are characterized by steeply sloping shores usually composed of bedrock. Few possible seine beaches exist in these areas and most were utilized in this study.

Beach seining was conducted within two-hour windows on either side of high and low tides. A single set was made as this usually provides good representation of species richness and dominant species rank (Allen *et al.*, 1992; Robards *et al.*, 2000). Fish were sorted by species, counted, and subsampled individuals were weighed and measured. In sets with large numbers of individual, subsamples were taken with the proportion of fish still in shallow water to minimize mortality.



Figure 9. Glacier Bay beach seine locations. Monitoring stations were developed as a set of sites that could be sampled repeatably on both high and low tide. Opportunistic sites were sampled at least once, but could not be sampled at both tidal states or were in close proximity to another beach seine station (exact locations are given in Appendix 1).

High-latitude fish assemblages, particularly those found in shallow water habitats, are subjected to large seasonal variations in temperature and day length. These physical factors impart a strong natural seasonality to community structure (Nash, 1988). Some fish species move from shallow water habitats to deeper waters in winter when thermal tolerances are exceeded (Allen and Horn, 1975; Allen, 1982; Bennett, 1989). Decreases in catch size between spring and fall peaks have also been observed by many investigators (e.g., Livingston, 1976; Horn, 1980; Allen, 1982; Thorman, 1986; Methven and Bajdik, 1994; Robards et al., 2000). We sampled from June until August to establish seasonality of species composition and abundance within nearshore areas of Glacier Bay during summer.

Pelagic Fish Community

Mid-water fish were caught by two means, a modified herring mid-water trawl during June, 1999; and an Isaacs Kidd mid-water trawl (IKMT) during summer, 2000.

Modified herring mid-water trawl

Mid-water trawls during summer 1999 were collected from the RV Pandalus. Small schooling fish were located with a Biosonics DT4000 digital 120kHz echosounder, and significant targets were fished. A modified herring mid-water trawl with a mouth opening of 50 m² was used to sample the bioacoustic signals in the pelagic zone. This net is larger than the Isaacs Kidd net used in 2000 and uses large steel doors to sink the net and keep it open. Mesh sizes diminished stepwise from about 50 mm in the wings to 10 mm at the cod-end, which was lined with 3 mm mesh. A plastic cod-end collecting bucket with 1 mm mesh openings was then attached to the end of the cod-end, and was detached and rinsed after each tow. A Furuno net-sounding system monitored the depth of the headrope while fishing. A temperature-depth recorder (TDR, Wildlife computers model # mk7) was mounted on the headrope to continuously record the exact depth of the net while fishing.

The target towing speed was about 2.5 knots, and the average tow duration was 20 minutes. All fishing was done during daylight hours. After each fishing tow, a CTD cast was made. If multiple fishing tows were taken at a station, only one CTD cast was taken.

All fishes were identified and measured to the nearest 1 mm fork length (FL). Stomach contents of adult walleye Pollock were identified. Capelin, Northern Smoothtongue, and Northern lampfish were frozen and weighed on land with an electronic balance to obtain length-weight relationships.

Isaacs Kidd mid-water trawl

Mid-water trawls during summer, 2000 were collected from the RV *David Grey*, a 9-m Uniflight using an IKMT (Figure 10). This net is reliable and stable, and is easily handled from a moderately sized vessel such as the *RV David Grey* in general sea conditions (Isaacs and Kidd, 1953). The net has a 2.8 m² opening and uses a depressor bar rather than weight to reach depth and can be towed at relatively high speeds (> 4

knots). Oblique tows were taken between the surface and 50 m depth. Although depths were determined by wire angle and scope, precise depths were recorded using a temperature-depth recorder (TDR, Wildlife Computers model # mk7) attached to the net. The net was towed for a total of 30 minutes in the direction of tidal flow at a towing speed of 3.5 to 4.5 knots. Only daytime tows were made. Volume filtered during each tow was calculated from a General Oceanics flowmeter that was attached to the central portion of the mouth of the net. Small schooling fish were separated from larval fish and zooplankton. Samples used for analysis were immediately preserved in



Figure 10 Deploying the Isaacs-Kidd mid-water trawl. Note the 'V' shaped depressor bar to which the net is attached.

50% isopropyl alcohol. The alcohol was renewed after 24 hours and after 2-3 days. We calculated a mean shrinkage of 6% for fish preserved in alcohol (n=100). Lengths were adjusted to fresh sizes for analysis.

During July we visited oceanographic stations 00, 02, 04, 06, 08, 10, 14, 16, 18, 20, and 21 (Figure 4), making one tow at each location. During August we visited the same stations but fished an additional 2 times at Station 21 to better quantify fish abundance at the face of a tide water glacier (Margerie). During August we also fished additional inlets of Glacier Bay to compare fish abundance between tidewater influenced and non-tidewater influenced inlets. These inlets were Johns Hopkins (Station 24), Reid (Station Z45), Rendu (Station 26), Queen (Station 27), Geike (Station 23), and Wachusetts (Station 28).

Predator Survey

Extensive predator surveys were conducted in June of 1999 and 2000. Surveys included all of the coastline areas of Glacier Bay as well as sampling pelagic areas with transects spaced approximately 2.5 km apart. In addition to Glacier Bay itself, transects were

conducted in Icy Strait and in 1999 there was also sampling in Dundas Bay following the grounding of a tourist vessel – *The Wilderness Adventurer*. Surveys were made from the research vessels (RV) *Pandalus, Alaskan Gyre, Lutris II*, and *David Grey*. Ground speed for vessels was approximately 11-15 km/h (6-8 knots). To examine seasonal patterns of use of Glacier Bay by various predatory species, we surveyed a subset (approximately 30%) of the more extensive summer surveys in November of 1999 and March of 2000.

Surveys were conducted, with some modifications, according to protocols established by the U.S. Fish and Wildlife Service for marine birds (Gould et al., 1982; Gould and Forsell, 1989; See Appendix 5 for additional resources used on surveys). Surveys from the *Pandalus* and *Alaskan Gyre* counted and identified swimming birds and mammals within 150 m on either side or 300 m forward of the boat to species. Because of their lower viewing angles we limited the area of identification to 100 m on either side of the *Lutris II*, and *David Grey*. Birds were recorded as flocks, when possible, and the following behaviors were recorded: on the water, feeding, standing on flotsam or jetsam, or swimming with a fish held in the bill.

We counted all flying birds that crossed within transects. This allowed us to make comparisons with a previous survey conducted in 1991. Birds that were flying while holding fish were noted as such. Summing the numbers of flying birds and birds on the water for a given transect yields a density estimate (birds/km²). Unusual bird or mammal sightings outside the transect boundaries were recorded as "off transect" and not used for analysis.

Bird and mammal sightings were recorded by entering them directly into a real-time computer data-entry system (DLOG; Glenn Ford, ECI) that plots sighting positions continuously using GPS coordinates. GPS locations were obtained from a Rockwell Precision Lightweight Global-positioning Receiver (PLGR). PLGR units have a worst-case horizontal position accuracy of ± 10 m at speeds < 36 kph (Anonymous, 1995). DLOG also provides the bottom depth and the distance to the nearest shore for all sightings.

At all times, 1 person entered data into a laptop computer, located in the wheelhouse, while observers surveyed from the best vantage points on each research vessel (RV). On the *Pandalus*, 1-2 observers surveyed from the bow, with a deck-height of 3.4 m above the water's surface. On the *Alaskan Gyre*, 2 observers surveyed from the wheelhouse, located 3.7 m above the water's surface. On the David Grey observers surveyed from top of the wheelhouse (2.4 m). Two observers on the *Lutris II* observed from water level (usually one of these observers was also navigating the vessel).

Observers actively scanned ahead of and alongside the survey vessel, and species identifications were confirmed using 7 - 10 power binoculars. Standard guides were used for identifications. Sightings were immediately called to the data entry person over hand-held VHF radios. If observers felt weather conditions were unsuitable for sighting small seabirds at 150-300 m, surveys were discontinued until conditions improved.
Ancillary data on weather, sea conditions, observation conditions, bird behavior or plumages, and species of fish held by birds were collected for each transect.

All bird distribution maps were plotted using spatial locations from the Dlog data file. Latitude, longitude, species, and numbers summarized in 1-minute blocks and imported into an ArcView GIS. The GIS was then used to develop maps of common species and specific species of interest.

Analysis Considerations

For gross comparisons of selected plankton and fish taxa, we divided Glacier Bay into four regions. Icy strait, the lower-bay is defined as south of a line between Rush Point and the northwestern tip of Young Island (i.e., Sitakaday Narrows); the middle bay lies between there and a line between Muir Point and the northern entrance to Geike Inlet; and the upper bay lies from there to the head of the western and eastern arms of Glacier Bay (Figure 11).

Abundance of fish caught does not represent true abundance because the efficiency of the net is unknown. Furthermore, the efficiency of this type of net differs between species. For example, in beach seines, efficiency of catch for demersal species such as flatfish may be in the order of 30 - 40 % (*Pleuronectes platessa*, cited in Gibson et al., 1996). Because of these limitations, we have made no estimates of absolute abundance in nearshore areas. However, based on the same net and deployment being used at all sites, and in both years, relative comparisons are still valid.

Data from predator surveys were summarized in a variety of ways. For completeness and basic comparisons, raw counts were provided from all surveys. For interannual comparisons, densities were calculated by averaging desities across all transects. To look at trends we used a previously unpublished survey conducted by John Piatt (USGS) and Allen Springer (UAF) in Glacier Bay during the summer of 1991 as a baseline. Then we used a subset (coastal transects) from the summer of 1999 and 2000 surveys for comparison. Finally, to compare survey intensities we used densities from varying numbers and arrangements of transects within Glacier Bay.

We have used common names throughout this report. A full list of Latin names is included in Appendices 1,6, and 7.

Results and Discussion

Oceanography

Temperature and salinity data were collected in conjunction with all fishing events connected with this project, and are now included in the Oceanography database (Hooge and Hooge 2001). This report summarizes oceanographic parameters for the bay but does not derive conclusions between specific CTD casts and fishing sets. Rather then replicating the comprehensive research of Hooge and Hooge, we have quoted directly from that report for conciseness.

Glacier Bay can be modeled as a tidally mixed estuary leading into basins that stratify in summer, with the upper arms behaving as traditional estuaries. The bay is characterized by renewal and mixing events throughout the year, and markedly higher primary production than in many neighboring southeast Alaska fjords (Hooge and Hooge, 2001).

The lower part of Glacier Bay, from station 03 to 01, extending out to Cross Sound is an area of intense mixing and upwelling due to tidally induced currents.

Detected salinities in Glacier Bay ranged from approximately 8.7 to 31.8ppt. The least saline waters were found in narrow surface lenses near tidewater glaciers, and the most saline were found at depth near and just outside the mouth of the Bay; salinity trends in general followed this same pattern, with overall salinity decreasing towards the heads of the bays. Water temperatures ranged from 1.9 to 12.2°C; they were generally coldest at the heads of the Bay's two arms near glacial input and warmer near the Bay's mouth. The density anomaly of the water varied from 5.1 to 25.0kg/m³. Values of sigma-t usually closely followed salinity patterns, and were generally least within narrow surface lenses in front of tidewater glaciers. The densest water was located at the bottom of the bay's deepest basins or at the bay's mouth. Water density increased with depth except at a few high-current stations (lower bay) during certain stages of the tide when density was virtually homogenous throughout the water column.

The lower bay is well mixed due to the constriction and shallow sill in Sitakaday narrows. Visual observations of standing waves, whirlpools, and roils; as well as images from Advanced Very High Resolution Radiometry (AVHRR; Hooge and Hooge, 2001) all attest to the CTD data. Suspended particulates that are high at the head of the bay, decrease in mid-bay (due to settling) and are resuspended in this well mixed area. The turbulence within this area was clearly demonstrated with trawls collecting particles of gravel despite fishing over 10 m from the substrate (confirmed by TDR recorder).

In sharp contrast to the lower Bay sill area the mid-bay area from Willoughby Island to the East Arm Sill and the west arm to the Gilbert Peninsula exhibits a strong pattern of stratification for much of the year (Hooge and Hooge, 2001). The upper arms of Glacier Bay are colder year round, and are characterized by a surface lens of less saline water from glacier melting. In the West arm the only sill to prevent estuarine circulation is 240 meters deep well below the depth of entrained water. In the East arm there are three sill with the shallowest sill depth at 60 meters. This depth is often below the depth of entrained water. Although, we found no evidence for strong estuarine circulation in the upper arms of the Bay the large majority of West Arm deep water was identical to the mid-bay deep water. The one notable exception, was the final basin of the upper West Arm which consistently had the greatest salinities seen in Glacier Bay indicating reduced deep-water renewal. The East Arm sills appeared to restrict some movements of water but renewal of the entire arm occurred throughout the year except for short periods (Hooge and Hooge, 2001).

Stratification is well established during the summer months at the heads of both the West and East Arms. There was no evidence for upwelling at the heads of the inlets, as indicated by localized shallowing of the isopycnals or isohalines (Hooge and Hooge, 2001). However, a clearly observable 'boil' close to the glacier face suggests that that the momentum of upwelling meltwater (from sub-surface ejection) is high resulting in a jet that intersects the water surface (Cowan, 1992). These upward currents at the face of the glacier bring deep-water nutrients to the surface. Furthermore, a "brown zone" (Hartley & Dunbar, 1938; apparently upwelling brown-colored water), and other upwelling phenomena were observed in these areas. Oceanographic results indicate that these phenomena are local and do not propagate very far from the tidewater glacier face.

In the mixed areas of the lower bay, the total amount of Chlorophyl-*a* throughout the entire water colun was less than in adjacent stratified stations. In the well-stratified upper West and East Arms, fluorescence profiles taken during summer demonstrated elevated levels of Chlorophyl-*a* throughout the water column, particularly in the sub-surface layers. Furthermore Chlorophyl-*a* levels in these periglacial areas were anomalous for both their depth and continuous distribution through the water column, as well as for the absolute magnitude of production that they represent.

Phytoplankton levels in Glacier Bay were found to be surprisingly high and sustained. The peak levels of depth-integrated chlorophyll-a were as high as the peaks of most years in Auke Bay (Ziemann et al. 1990). This was observed despite the limited number of surveys made each year, which could easily have led to missing miss much higher but short-lived peaks such as those associated with all blooms not only in Auk bay but in all Alaskan fjords (Burrell 1986). However, in sharp contrast to Auk bay and other Alaskan fjords (Burrell 1986, Sambrotto & Lorenzen 1986) phytoplankton standing crop was commonly sustained at a high level throughout the spring, summer, and fall. The early onset and peak of phytoplankton production was in March, corresponding to the onset of stratification. This was a month earlier than Auke bay (Ziemann et al. 1990) but not atypical for southeast Alaskan fjords (Burrell 1986). More surprisingly, some of the first places to experience a bloom were the central and upper arms of the Bay, which were still experiencing very cold temperatures. Subsequently, fluorescence values decreased, but not precipitously, and were seen to reach levels close to the maximum latter in mid-summer. In contrast, the typical Southeast Alaskan bloom lasts only a few weeks before nutrients are depleted and standing crop levels plummet (Burrell 1986, Sambrotto & Lorenzen 1986, Ziemann et al. 1990). Moreover, summer conditions elsewhere are characterized by general nutrient depletion and only occasional renewed phytoplankton growth driven by nutrient enhancing events (Iverson et al. 1974). In the late summer or early fall a smaller secondary bloom may occur. Heavily silted systems such as glacial fjords are thought to have suppressed fall blooms (Burrell 1986). In Glacier Bay we observed neither a precipitous drop in the standing crop following the spring bloom nor suppression of high phytoplankton levels in fall. No other fjord system in Alaska has been observed with this type of sustained productivity (Burrell 1986).

Two possible reasons for the high and sustained standing crop are lack of zooplankton predators or high nutrient availability. There are few data regarding zooplankton in Glacier Bay. However at McBride Inlet in the upper East Arm high densities of harpacticoid copepods, calenoid copopods, and other zooplankton were found even in the coldest, most brackish and most turbid environment of Glacier Bay (Simenstad & Powell 1990). In addition, preliminary results from Bay-wide plankton tows conducted synoptically to this study indicate that zooplankton populations are not depauperate in numbers (J. Anson, pers. comm.). The cold temperatures of Glacier Bay may be prevent zooplankton from responding as quickly as phytoplankton can grow, due to the decrease in efficiency of respiration as compared to photosynthesis at cold temperatures (Byron 1982). However, we believe that suppressed zooplankton respiration is not sufficient to fully explain the high and sustained phytoplankton levels found in Glacier Bay. Further research is needed into the interactions of zooplankton and phytoplankton particularly in light of the oceanographic complexity of *Glacier Bay*, which may dramatically affect temporal and spatial species distributions and metabolism. The other possible explanation for the high and sustained phytoplankton levels at Glacier Bay is nutrient enhancement. As mentioned previously Glacier Bay experiences several possible oceanographic processes that may lead to nutrient enhancement and result in high levels of phytoplankton. Foremost among these is the front created by confluence of the tidally mixed waters of the lower Bay and the Stratified waters of the central Bay. Such tidally mixed fronts are often associated not only with increased nutrient replenishment, but also with high and sustained primary production (Pingree etal. 1975, Perry et al. 1983). In addition, Glacier Bay experiences renewal more frequently than previously thought. This frequent renewal insures that deep and intermediate water layers are not depleted of nutrients; therefore when unstratified conditions occur (winter in Glacier Bay), surface waters are replenished of the nutrients lost due phytoplankton uptake and settlement. Year-round renewal of intermediate of deep water will only restore nutrients to stratified near-

surface waters if nutrients can diffuse upwards through the density gradient. The tidally induced internal waves observed once in Glacier Bay (Matthews 1981) and the hypothesized hydraulic instabilities associated with the Bay's high-current constricted entry sill provide a mechanism to increase the diffusion of nutrients upwards without disturbing stratification. Stratification is usually necessary for high phytoplankton production (Mann & Lazier 1996). Such enhancement of productivity by internal waves has been observed in several studies (e.g., Shea & Broenkow 1982, New 1988). If these nutrient enhancing events are the primary cause of high productivity then we would expect the spatial pattern of productivity to reflect these phenomena. Phytoplankton levels were most consistently high in the central bay and lower arms, which is where the strongest effects of fronts and internal waves would be expected. The high average levels of productivity in Glacier Bay can give a mistaken image of the stability of the high phytoplankton production. While there was quite a bit of temporal and spatial clustering of high standing crop, there was also significant temporal and spatial variation between individual stations. One station could have very low phytoplankton levels during one survey and extremely high levels during the next: Chlorophyll-a levels at stations only 5 km apart could vary tremendously. We believe that this represents a series of sporadically depleted conditions relieved by frequent nutrient enhancing events.

Sediment load appears to play a significant role in reducing photic depth and thus acts as a control on phytoplankton production. However, this relationship only appears at high sediment densities, which limit photic depth and are associated with low depth-integrated chlorophyll-a. At low or intermediate levels of sediment in the water column, photic depth and phytoplankton can reach their maximal levels and phytoplankton densities appear to take control of limiting photic depth. Thus, the deepest photic depths are associated with both small sediment loads and small phytoplankton levels. The shallowest photic depths were associated with high sediment loads and low phytoplankton levels. The highest phytoplankton levels were associated with intermediate photic depths. Despite the extremely high sediment loads throughout Glacier Bay and the clear role that sediments played in determining photic depth, GlacierBay nonetheless exhibited high chlorophyll-a levels; no broad-scale strong suppression of phytoplankton was seen during high sedimentation events. The interaction between sediment and phytoplankton is not limited to limitations on light penetration. During summer to early fall a high level of chlorophyll-a was seen throughout the water column at the heads of both arms. These levels, $0.5-1.5 \text{ mg/m}^3$ were as high as those seen in productive zones of the Gulf of Alaska (Sambrotto & Lorenzen 1986). Cowan (1995) has demonstrated that silt particles can interact with organic material to form flocculants that settle more rapidly than expected. We believe that the deep chloropyll-a anomaly represents extremely rapid phytoplankton settling rates that result from diatoms interacting with sediments to form larger flocculants. Rapid settlement of

phytoplankton out of the photic zone can significantly decrease production (Atlas et al. 1983, Ziemann et al. 1990). The extremely localized and brief nature of this phenomena limit its system wide effects. However, the sediments in this area may be significantly carbon and nutrient enriched (Cowan 1995).

In summary, Glacier Bay exhibits extremely high levels of phytoplankton over a very extended season. This production probably results from enhanced nutrient availability in Glacier Bay's surface waters, which in turn result from tidal mixing and frequent deep-water renewal. High phytoplankton levels were maintained despite large amounts of sediment that extended throughout the bay and often restricted light penetration. Rather than a traditional silled fjord estuary Glacier Bay should be viewed as a tidally mixed estuary in the vicinity of its sill, backed by a stratified basing. with more traditionally estuarine arms. Because the intertidal zone represents such a small percentage of the bay's waters, phytoplankton productivity must contribute the majority of carbon production to the marine environment. *Glacier Bay hosts a high density and wide variety of secondary and tertiary* consumers. It is now clear that Glacier Bay's high phytoplankton levels are commensurate with large predator populations. The seasonal variation in primary productivity and its interactions with larger-scale oceanographic events as well as with predator dynamics need to be examined. A long-term program of oceanographic monitoring is essential to understand these processes.

Hydroacoustic Survey

The most obvious feature of acoustic biomass (S_A) distribution in Glacier Bay is that it was concentrated in relatively few areas (Figures 11, 12). At the spatial scale of Glacier Bay, we found large, relatively shallow (<75m) biomass concentrations around Pt. Adolphus in Icy Strait, in Berg Bay, around and north of the Beardslee Islands, around South and North Marble Islands and near the entrance to Muir Inlet, at the entrances to Geikie Inlet and Scidmore Bay and on the shelf between them, and in upper reaches of the West Arm and Muir Inlet (including Reid, Rendu, Waschusett, and Muir inlets). Even more striking from maps of biomass distribution (Figures 11, 12) are the large areas in between concentrations of biomass in which forage fish are scarce or virtually absent.

This visual impression is corroborated by a frequency analysis of the individual integration cells used for mapping (see Methods). If we assume a target strength for swim-bladder bearing fish (physoclist) and scale S_A for the mean size of all fish combined, then we can approximate the frequencies of integration cells containing fish densities ranging from <0.00001 fish/m³ to 4.6 fish/m³ (Figure 13). Out of 6000 integrated 5x100m cells for each depth strata (5-10m, 10-15m, etc.) remarkably few cells contained mean fish densities high enough to support foraging by large predators such as seabirds and marine mammals. Indeed, less than 8% of the total area surveyed by

hydroacoustics contained cells exceeding forage fish densities of 0.01 fish/m³, and less than 1% contained cells exceeding densities of 0.1 fish/m³.

Another feature of acoustic biomass distribution in Glacier Bay evident in the maps (Figures 11, 12) is that the biomass was concentrated in shallow waters and close to shore. We can further examine depth distribution of biomass in two ways. First, when we plotted total acoustic biomass located in thick layers of water (5-25m, 25-50m, etc) versus the bottom depth of the sea floor (Figure 13), we observed that most biomass was located in the shallowest water layer (<25m) irrespective of bottom depth. Overall, acoustic biomass abundance was highest at depths of 25-50m. Secondly, we calculated the mean density of fish biomass in each depth strata (e.g., 5-10m, 10-15m, 15-20m, etc.). From this we found that, irrespective of bottom depth, mean densities of forage fish were 2-3 times higher in waters <50m in depth than in waters of greater depth (Figure 14). By plotting total cumulative biomass was found at depths <35m, more than 80% at depths <80m, and more than 90% at depths <100m.

The densest forage fish aggregations illustrated in Figs. 11 and 12 were usually composed of only a few species such as herring, capelin, pollock or euphausiids. Integration of these aggregations at much finer spatial scales (1m x 20m bins) allows us to better estimate point densities of these aggregations. For example, the highest density aggregations located at Pt. Adolphus (Figures 11 and 12) were comprised mostly of adult herring (Fig. 16) and adult capelin (Figure 17), and these schools had densities in excess of 1-10 fish/m³. Juvenile capelin and pollock aggregations at the mouth of Muir Inlet (Fig. 18) were typical of more dispersed aggregations in this area, with point densities of 0.01-0.1 fish/m³. Euphausiid aggregations— which showed up on biomass maps (Figures 11 and 12) with relatively low S_A values owing to their weaker target strength— actually contained impressive densities of euphausiids; in the order of 0.1 to 1 kg/m3 (Figure 19).



Figure 11. Distribution of fish and zooplankton Biomass (S_{A} , acoustic backscattering) in Glacer Bay during surveys in June, 1999. Upper map: Sum of $S_A/1000$ over 5-25 m depth strata. Lower map: Sum of $S_A/1000$ over 25-50 m depth strata.



Figure 12. Distribution of fish and zooplankton Biomass (S_A , acoustic backscattering) in Glacier Bay during surveys in June, 1999. Upper map: Sum of $S_A/1000$ over 50-75 m depth strata. Lower map: Sum of $S_A/1000$ over 75-100 m depth strata.



Figure 13. Frequency of high-density prey patches in Glacier Bay. Acoustic backscatter in the water column on transects was summed over moderately small (ca. 100m long by 5m deep) integration cells. Fish density was estimated in more than 6000 such cells for each 5m-depth strata. For purposes of illustration in this figure, cells from five surface strata were sorted from left to right according to fish densities in each cell. As evidenced above, cells with relatively high fish densities (>0.1 fish/m³ in a 5x100m cell) were exceedingly rare.



Figure 14. Distribution of acoustic biomass by water depth. Data from areas where bottom depths greater than 190m were combined into the 180-190 depth bin. Acoustic backscatter (SA) was summed over four depth strata (25m bins), and examined with respect to depth of the ocean floor (in 10m increments). Total biomass was prorated for the amount of area surveyed at each depth. Figure shows, for example, that most biomass is found in the upper 25m regardless of depth of the sea floor.



Figure 15. Average (bars, \pm SE) and cumulative (line) biomass distribution in the water column (irrespective of bottom depth, see previous figure). Mean acoustic biomass in shallow (<50m) strata is generally more than double that observed in deeper strata. More than 50% of total biomass was found at depths <35m, more than 80% at depths <80m, and more than 90% at depths <100m.



Figure 16. Hydroacoustic echogram (top) and integration analysis (bottom) of adult Pacific herring schools near Pt. Adolphus in June, 1999. Schools are large red-blue blobs at top of echogram. Individual herring are seen as small blue streaks in lower half of echogram. The seafloor appears at the bottom of the echogram (dark red layer). Integration graphic shows density (in fish per cubic meter) of a selected herring school in 40 m of water along a 760 m section of the transect illustrated in the echogram.



Figure 17. Hydroacoustic echogram (top) and integration analysis (bottom) of adult capelin schools near Pt. Adolphus in June, 1999. Schools are large red blobs in middle of echogram, just above the seafloor (which appears as a very dark red, narrow, wavy band across the echogram). Integration graphic shows density (in fish per cubic meter) of a selected part of this capelin school in 40 m of water along an 800 m section of the transect illustrated in the echogram.





Figure 18. Hydroacoustic echogram (top) and integration analysis (bottom) of a dispersed layer of juvenile capelin and juvenile pollock in surface waters near the entrance to Muir Inlet, in June 1999. Juvenile fish appear as tiny red dots in upper 15 m of water column. The diffuse blue band in middle is layer of euphausiids and other plankton. Large red streaks near the sea floor (which appears as a very dark red, narrow band across bottom of echogram) are adult pollock. Integration graphic shows density (in fish per cubic meter) of a selected part of the dispersed surface schools of juvenile fish in the top 40 m of water along a 780 m section of the transect illustrated in the echogram.



Figure 19. Hydroacoustic echogram (top) and integration analysis (bottom) of a dense layer of euphausiids near Hugh Miller Inlet in June 1999. Diffuse blue band in middle is layer of euphausiids and other plankton. Adult pollock appear as large red dots within the layer of euphausiids. The sea floor does not appear in the echogram. Integration graphic shows density (in kg per cubic meter) of a dense part of the euphausiid layer in the top 40 m of water along an 800 m section of the transect illustrated in the echogram.

Zooplankton

Previously, three surveys within Glacier Bay have included zooplankton sampling. Bruce L. Wing (NMFS Auke Bay Laboratory) assessed abundance at two Glacier Bay sites in August 1963. Vertical hauls were collected from the Beardsley Islands region and from close to the Lamplugh Glacier. These counts offer a snapshot of what the plankton community was like 37 years ago. Differences in sampling (primarily, we used a smaller mesh net which captures smaller animals) preclude direct community comparisons. However, despite the methodological differences, copepod abundance estimates were consistent with those of our study. We speculate that several other gross differences in community structure between Wing's and our data are real rather than functions of data collection differences. At the Beardsley Islands in 1963 arrow worms, ostracods, and decapod zoea were more abundant than we saw in 2000, when *Oikopleura* and *Asteroida* lavae were the most abundant taxa. Brachyuran zoea were seen in August, 1963, but only occurred in our samples in the Main Bay during June (Figure 20).

Bruce Wing and Kenneth Kreiger of NMFS Auke Bay Labs carried out a three-season summer survey of humpback whale prey (1981-83; Wing and Kreiger 1982, Kreiger and Wing 1984). They performed acoustic surveys and sampled using a 500-micron mesh Tucker trawl. They estimated zooplankton abundance by displacement volume, with composition listed as percent euphausiids, chaetognaths, and other. Both this and their studies indicate that euphausiids and chaetognaths, continue to be prominent parts of the Glacier Bay zooplankton community structure.

Simenstad and Powell (1990) sampled zooplankton at McBride glacier correlated with benthic sampling and tidewater sedimentation processes in 1984 and 1986. Zooplankton were collected with a 12.5cm opening 253µm mesh net. Their samples were dominated by copepods (principally *Pseudocalanus* copepodids), with barnacle nauplii and chaetognaths also present. These findings are entirely consistent with our survey in composition of upper bay inlets.

Zooplankton production is usually linked to the timing of the primary production bloom, which in turn is linked to the onset of spring stratification. The onset of stratification varies considerably between years in Glacier Bay, beginning as early as February, and as late as April (Hooge and Hooge, 2001).

Analyses of temporal and spatial zooplankton trends were done by calculating monthly means and grouping stations by region. Our samples reflect the diversity and abundance of zooplankton in the upper 50m, effectively the surface waters of the bay. The summer zooplankton community observed in Glacier Bay is similar to communities seen throughout the Gulf of Alaska. Temporally by region, the upper bay (West and East Arms) and the lower bay (Main Bay and Icy Strait) exhibited remarkably similar trends (Figure 21).



Figure 20. Relative zooplankton abundance in lower Glacier Bay in 1963 and 2000 (numerically dominant copepoda excluded to highlight less abundant taxa).

Peak zooplankton abundance of the lower bay sites in late May/early June is similar to the general pattern for the Gulf of Alaska (GOA; Mattson 1978, Cooney 1987, Paul 1991). The key distinction between the lower Bay and other estuaries in the GOA is a second smaller peak in lower-bay densities during August when zooplankton densities in other areas are usually continuing to decline.

Zooplankton in both the East and West Arms peaked in July. Although densities had begun to decline in August (our final samples), they were still more than twice the densities seen in that region during May. The highest density of zooplankton observed was 17,870 organisms/m³ seen at station Z47 (Tarr Inlet) in July. Up-bay trends were distinct from lower bay trends indicating that local processes are more strongly influencing zooplankton community dynamics in the West and East Arms compared to the Lower Bay which was more consistent with general GOA trends (Figure 21). Each region was unique in temporal trends for number of species observed (Figure 22).

Results from the different sample stations varied greatly in abundance and composition. Nearshore stations exhibited more variation in abundance and composition than midchannel stations. Mid-channel station increased in abundance and decreased in diversity with distance up the bay, never as low or as high as nearshore stations in diversity or abundance. Nearshore stations also had proportionally fewer copepods. Copepods made up an average 81.6% of nearshore samples and 87.0% of mid-channel samples.



Figure 21. Spatial and temporal patterns of numerical zooplankton abundance, grouped by region.



Figure 22. Spatial and temporal patterns of numerical zooplankton abundance, grouped by region.

Copepods at the glacier fronts had higher levels of lipids than animals in the lower bay. Copepods rely on lipid storage to survive the winter and reproduce; lipid levels also indicate the abundance and quality of food they are exposed to. This finding is correlated with the high levels of chlorophyll seen at the glacier fronts by USGS oceanography researchers (Hooge *et al.*, 2000).

Zooplankton of sessile organisms were consistently more abundant in the Main Bay and Icy Strait than further up the bay. Estuarian bivalves and crustaceans are known to cue to environmental fluxes associated with tidal change and to adjust their migrations accordingly (Young 1995). Therefore, these taxa are likely to partially control their horizontal position in the estuary by vertically migrating in response to tidal cues. Additonally, zooplankters swimming upward in the water column, on reaching the thermocline or chemocline, suspend swimming and sink down before swimming again. Considering the glacial fjord oceanography of the bay, this mechanism is certainly a possible mechanism for maintaining high densities of zooplankton in Glacier Bay; keeping these animals out of surface outflow.

Larval fish and eggs were present in plankton tows from Icy Strait to the head of the West Arm (Appendix 6). Larvae and eggs were present primarily in June, with some observations earlier in the season. Only once, in Icy Strait, were eggs seen in August, otherwise they didn't appear in our samples after June. June emergence of fish larvae correlates with the high abundances of all copepod stages.

Zooplankton Diversity Indices

We calculated the Shannon-Weiner Index for each station to examine variation in species diversity. A zero index is a population of only one species; increasing numerical indices indicate an increasingly less homogeneous community structure.

We found generally higher diversity in the lower bay than upper bay; and greater variability in diversity at nearshore stations compared to those in mid-channel. The relationship of abundance and diversity for each zooplankton station indicated a significant negative correlation (Table 2) and log(density) was the best predictor of diversity.

Glacier Bay zooplankton follow a general pattern of increasing density and decreasing diversity with distance up the bay. Physiological tolerances may have a significant contribution to this pattern, as many zooplankters are able to move 'upstream' in systems by vertically migrating with tides. The high abundance at glacier fronts is probably a result of water currents, keeping plankters within the high productivity front of mixing waters.

	Density		log(Density)		
	r	p-value	r	p-value	
Nearshore	-0.608	0.029	-0.717	0.004	
Mid-channel	-0.368	0.719	-0.319	0.935	
All Stations	-0.548	0.006	-0.605	0.002	

Table 2 Pearson correlation coefficient for Shannon-Weiner diversity index and d
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Spatial Patterns

Cluster analysis (Cooney and Coyle, 1997) helped elucidate spatial variation of zooplankton communities within the Bay, and using Cooney and Coyle's data, how Glacier Bay compares to Prince William Sound. Cooney and Coyle found that outer- and inner-Prince William Sound stations grouped together, with only a single station grouping outside its geographical region. Glacier Bay spatial patterns did not follow as straightforward a geographical trend (Figure 23; Table 3). Oceanographic features unique to Glacier Bay likely influence these patterns. Several West Arm Stations grouped together with outer-Bay stations, potentially indicating a high degree of oceanographic influence through the Bay, and supporting the apparent trend of advected animals using tidal mechanisms to remain at areas of high productivity. Species analysis of copepods would provide further insight about the spatial patterns of oceanic and estuarine species in Glacier Bay.

Table 3 Cluster analysis of dominant taxa	collected in Glacier Bay	during summer 2000.
GROUP 1		

Icy Strait		Main Bay		West Arm		East Arm	
Mid-channel	Nearshore	Mid-channel	Nearshore	Mid-channel	Nearshore	Mid-channel	Nearshore
STN00	Z55	STN02	ZO2	STN08	Z36		Z24
	Z62	STN04			Z37		
					Z40		
					Z47		
GROUP 2							
Icy Strait		Main Bay		West Arm		East Arm	
Mid-channel	Nearshore	Mid-channel	Nearshore	Mid-channel	Nearshore	Mid-channel	Nearshore
			Z06	STN06	Z45	STN14	Z25
			Z09	STN10	Z49	STN16	Z30
			Z10	STN12		STN18	
			ZI6	STN24		STN20	
			Z20				
GROUP 2 Icy S Mid-channel	trait Nearshore	Main Mid-channel	Bay Nearshore Z06 Z09 Z10 Z16 Z20	West Mid-channel STN06 STN10 STN12 STN24	Arm Nearshore Z45 Z49	East Mid-channel STN14 STN16 STN18 STN20	Arm Nearsh Z25 Z30



Figure 23. Geographic postitions showing cluster analysis by 5 top ranking Zooplankton taxa (seasonal mean).

Phylum Arthropoda

Class Copepoda

Copepods are the dominant taxa Glacier Bay, as has been found for all of the Gulf of Alaska and its estuaries (Cooney 1987). Copepods contribute most of the zooplankton biomass in the system, and trends are therefore important for predators. Copepods hatch from eggs carried externally by a female and molt through 6 naupliar and 5 copepodid stages before becoming adults. Nauplii, copepodids (juveniles), and adults are all crucial in the diets of many fish species; timing of peaks in food availability and emergence of larval fish is important for fish survival. Availability of food is not the only factor regulating survival of juvenile fish, but is accepted as primarily important (Cooney 1987).

Copepod abundance in the East and West Arms followed a similar pattern, peaking in in July and beginning to decline by the end of our sampling in mid-August (see Figures 24, 25, and 26). Copepod populations in the main bay and Icy Strait were relatively stable in abundance compared to the upper-arms. The similar patterns of copepod abundance in mid-bay and Icy Strait suggested a much closer association between these areas than the upper-bay. These results suggest that mid and lower Glacier Bay as well as Icy Strait are closely tied to the Gulf of Alaska while more local processes are affecting the upper Bay.

Large copepods that are abundant in the Gulf of Alaska were found primarily in the upper Bay. Copepods increased their lipid stores through the season, indicating that they are growing well and are storing energy for the winter and for reproduction. Lipids were primarily seen in copepods at the glacial fronts. Oceanographic data suggests that these copepods are successfully exploiting the highly productive upper-bay regions. Although the advection of copepods in and out of the bay has important ecological consequences, this question was beyond the scope of this study.



Figure 24. Monthly mean abundance for calanoid and cyclopoid copepods less than 2.5mm in length, showing lower and upper bay seasonal trends. Except for in March, small copepods dominated the zooplankton community.



Figure 25. Monthly mean abundance for large calanoids (l > 2.5mm) showing upper and lower bay trends.



Figure 26. Monthly mean abundance for copepods nauplii, showing the lower bay and upper bay seasonal trends.

Class Branchiopoda

Order Cladocera

Podon and *Evadne*, small marine cladocerans, were present in 21 % of zooplankton samples, and primarily at nearshore stations. Both species are common in the Gulf of Alaska during the summer (Cooney 1987). Abundance in Glacier Bay increased through the season, peaking in the Main Bay in August (Figure 27). Abundance increased in the upper Bay somewhat later than in lower regions.



Figure 27. Upper and lower bay densities for marine cladocerans Podon and Evadne.

<u>Class Cirripedia</u>

Order Thoracica

Balanus nauplii or cypris larvae (non-feeding settling form) were present in 81 % of our plankton samples. Barnacle nauplii and cypris larvae were present throughout our sampling (March – August), but were most abundant in March, when they dominate samples. There abundance is likely an important component for planktivores foraging in the early spring. Barnacle nauplii disperse widely (Figure 28). Oceanic individuals advected by seasonally persistent onshore Ekman transport probably supplement winter and early spring populations.



Figure 28. Upper and lower bay monthly mean densities for *Balanus* nauplii and cypis larvae.

<u>Class Malacostraca</u>

<u>Order Amphipoda</u>

Amphipods are common in aquatic systems throughout the world. Amphipods in Glacier Bay generally ranged from 2 -10 mm. Amphipods were present in 52 % of our samples, peaking in abundance in May in the Main Bay (lower- and mid-bay; Figure 29). *Hyperiid* amphipods accounted for 75 % of the amphipods collected, and *Cyphocaris chalengeri*

accounted for most of the remainder. Separate data indicates that hyperiids are an important prey item for kittiwakes, especially near glacier fronts (USGS-BRD, Elizabeth Hooge, pers comm). Numerous amphipods were also found in beach seines at the Glacial outflow from Carroll Glacier (Beach 32).



Figure 29. Monthly mean abundance for amphipods, showing upper and lower bay densities.

<u>Order Decapoda</u>

Decapods were present in 46 % of the plankton samples, peaking in abundance in May. Brachyuran zoea were markedly more abundant within the Main Bay (lower- and midbay; and Icy Strait in June; Figure 30). Decapod crustaceans reproduce synchronously with environmental cues (Forward, 1987); our sampling indicates favorable environmental cues existed for local reproduction in June. Brachyuran Zoea were only present in the lower Bay during June. However, during the 1963 sampling, they were present during August. Limits in sampling prevent understanding if this was small-scale variation or if changes in physical parameters have altered timing of larval release.

Order Euphausidacea

Euphausiids are frequently called Krill. Juvenile euphausiids were present in 58% of our plankton samples. They dominated nearshore and mid-channel lower bay samples (STN00 and STN02). Abundance was much lower in the upper bay (Figure 32). Abundance peaked in June which is later than the May peak reported for euphausiids in Resurrection Bay (Prince William Sound; Paul et al.,1991). Abundance during June in Icy Strait was over double the peak abundance reported in Resurrection Sound (during 1988). Abundance declined to zero in Resurrection Bay by June 1st, whereas juvenile euphausiids and eggs were present in Glacier Bay until August. Densities of euphausiids were much higher in nearshore samples (Figure 31 and 32). Euphausiids are important prey for numerous species including humpback whales (Wing and Krieger, 1982) and juvenile walleye pollock (Cooney 1987). Additionally, numerous euphausids were found close to the Grand Pacific (beach 47) and Reid (Beach 43) glaciers in beach seines.



Figure 30. Monthly mean abundance for decapods, showing upper and lower bay densities. Note the difference in scales.



Figure 31. Monthly mean abundance for euphausiid nauplii, metanauplii, and zoea showing upper and lower bay densities.



Figure 32. Euphausiid juveniles by station channel position.

Order Isopoda

Isopods were present in 10% of plankton samples, peaking in abundance in May in Icy Strait (Figure 33). Isopods were present throughout our study (March – August), although generally in low numbers. Parasitic isopods were grouped with parasitic copepods (see *Parasitic Zooplankton* Section). Isopods are important prey for many species of fish.



Figure 33. Monthly mean abundance for isopods, showing upper and lower bay densities.

Class Ostracoda

Ostracods were present in 34% of plankton samples, with highest abundance in the East Arm. Ostracods were present at pelagic stations from STN00 to STN08 and nearshore stations throughout the bay. Densities were relatively stable in the East Arm and had not noticeably declined by the end of the study (Figure 34). Ostracods were present in Glacier Bay in 1963.



Figure 34. Monthly mean abundance for Ostracoda showing upper and lower bay densities.

Parasitic zooplankton (Isopoda and Copepoda)

Parasitic copepods or isopods were present in 18 % of the plankton samples. These organisms were caught during their brief free-living stage before finding a host (fish). Most parasitic plankters were caught in the upper bay, peaking early in the summer and then steadily declining through the season (Figure 35). The significance and relative abundance of parasitic isopods and copepods in Glacier Bay was beyond the scope of this study.



Figure 35. Upper and lower bay monthly mean densities for parasitic copepods and isopods.

Phylum Bryozoa

Bryozoans are sessile, colonial animals that release feeding larvae, known as cyphonautes. These larvae swim freely within the plankton community for several months before settling to the substrate. Brozoan larvae were present in 28 % of our samples. Abundance was relatively consistent through the Bay in space and time (Figure 36).



Figure 36. Upper and lower bay densities for bryozoa cyphonautes larvae. Densities were relatively consistent throughout the season and region.

Phylum Chaetognatha

Sagitta elegans, the only chaetognath we caught, was present in 87 % of our plankton samples. A second subdominant species observed in low numbers by Wing and Krieger (*Eukrohnia hamata*) was not caught in this study. Peak abundance was in May for the lower Bay and in July for the upper Bay (Figure 37). Abundance remained high in the upper Bay throughout summer. Arrow worms, as they are commonly called, are carnivorous and specialize on predating copepods. They are, in turn, prey for many larger marine planktivores.



Figure 37. Upper and lower bay monthly mean densities for Sagitta elegans.

Phylum Cnidaria

Cnidaria and *Ctenophora* are carnivorous zooplankton and were present in 49 % of plankton samples. The vast majority were small medusae, ranging in size from less than 2mm to several centimeters in diameter. Highest abundance was in May in the West Arm. Abundance declines after the early summer peaks throughout the bay (Figure 38). Abundance of these species was similar to copepods – results similar for Icy Strait, Lower Bay, and Mid-Bay; but distinctly different from the East and West Arms.



Figure 38. Monthly mean abundance for *Cnidaria* and *Ctenophora* showing upper and lower bay densities.

Phylum Mollusca

Bivalve (Figure 39) and gastropod veligers (Figure 40) were present in 87 % of plankton samples, and both had similar patterns of abundance, although gastropods were always less abundant than bivalves. Both bivalve and gastropod veligers were more abundant in the upper bay. Abundance peaked in June and July and had begun declining by August.







Figure 40. Upper and lower bay monthly mean densities for gastropod veligers.

Phylum Echinodermata

Echinoderm juveniles or veligers were present in 6% of the plankton samples; *Asteroida* juveniles or veligers were seen in 48 %. Abundance in the Main Bay peaked in August

(Figure 41) with almost 60 individuals/m³. Abundance of *Asteroida* in the Main Bay was substantially higher than Icy Strait. *Holothuroidea* auricularia larvae were present in one sample, nearshore Geikie Inlet (station Z20) during July. Echinoderms were present in the lower Bay in June and July, and in the West Arm in May and July (Figure 42). They were not observed in the East Arm.



Figure 41. Upper and lower bay monthly mean densities for *asteroida* juveniles and veligers.



Figure 42. Upper and lower bay monthly mean densities for echinoderm juveniles and veligers.

Phylum Phoronida

Phoronida actinotroch larvae and juveniles were only present in five of our plankton samples. They were at low density (maximum at STN02 June, 3.7 indivuduals/m³) and only found within the Main Bay and West Arm (Figure 43).



Figure 43. Upper and lower bay monthly mean densities for *Phoronida* juveniles and actinotroch larvae.

Phylum Urochordata

Oikopleura were present in 85 % of plankton samples. Abundance was highest in Icy Strait in June, with upper-Bay numbers remaining relatively low and constant. Aside from the June peak in Icy Strait, populations grew steadily through the summer, and had not begun to decline when the study ended.

<u>Icthyoplankon</u>

We sampled the same 11 stations (Figure 4) in both May and June 2000 for a cumulative total of 76 samples (up to four depth strata per station visit). We identified a total of 9 fish taxa and 2 invertebrate taxa. Bathyagonus species caught on this project are probably *Agonsosis vulsa* and *Bathyagonis nigripinnis* (Bruce Wing NMFS Auke Bay Lab, *Pers. Comm.*). Numbers of fish per m³ were over 200 times higher in June than May. Of the total catch (adjusted for equal volume) walleye pollock was the dominant taxon, comprising 57 and 72 % of the fish caught in May and June respectively. Slender eelblenny (43 %) and capelin (20 %) were second most abundant taxa in May and June, respectively.

Walleye pollock dominance of the larval fish community mirrored that of another southeast Alaskan icthyoplankton study (Mattson and Wing, 1978) and that of Resurrection Bay in Prince William Sound (Paul et al., 1991). Mattson and Wing found capelin to be second in abundance, but Stichaeidae (e.g., slender eelblenny) only accounted for about 2 % of the catch. Their more open-channel sampling likely accounted for this difference. Both our results and those of Mattson and Wing (1978) were conspicuous in their absence of larval herring, with only 3 caught (in an Isaacs-Kidd trawl in Queen Inlet) in our pelagic samples (All herring caught in the modified herring mid-water trawl were older (minimum size 199 mm). This contrasts the Paul et al., (1991) study where herring was a significant part of the Resurrection Bay icthyoplankton community.

Distribution of juvenile pollock within the water column declined with depth (64 % at 0-25 m, 22 % at 25-50 m, 12 % at 50-75 m and 2 % at 75-100 m) as observed in Resurrection Bay, Prince William Sound (Müter and Norcross, 1994). Juvenile capelin were exclusively found in surface (0-25 m) waters and slender eelblenny were distributed as deep as 75 m.

Juvenile walleye pollock averaged 12.53 mm (sd 3.43 mm) in June. Assuming similar growth rates of juvenile pollock in Glacier Bay to Resurrection Bay (another glacially influenced fjord), we used the formula developed by Müter and Norcross (1994) to estimate mean hatch date. We estimate that the mean walleye pollock hatch-date in Glacier Bay is around April 29. This is very similar to the Müter and Norcross (1994) estimate of 22 April for Prince William Sound pollock, and within the range of observed hatch dates in other areas of the Gulf of Alaska (23 April to 2 May; cited in Müter and Norcross, 1994).

Spatially, within Glacier Bay we only found juvenile pollock at the entrance to the west arm (station 6) in May. Larval pollock may have been present elsewhere but deeper than our sampling (100 m), though none were found at the shallower sill areas where we fished the entire water column. In June, pollock were relatively widespread, occurring throughout most of the bay. Numbers increased up the bay with highest numbers found in the upper west arm; but notably not close to Margerie Glacier at Station 21. We found fewer larval pollock in the east arm and again not in the upper reaches.

Euphausiids were found in 45 of the 76 samples (59 %). Spatially they were found throughout the bay, but were exceedingly common at the face of Margerie Glacier (Station 21) in June. Numbers were generally too low to make temporal comparisons between May and June.

We examined seasonal and spatial patterns of abundance for both pelagic zooplankton and icthyoplankton in Glacier Bay. Results for May to August (Figures 44, 45, 46, 47) indicate a marked seasonal and spatial pattern to the abundance of these secondary and tertiary levels of productivity. Zooplankton abundance was highest in the upper arms and at the mid-bay station 4.



Figure 44. Spatial pattern of zooplankton (yellow) and euphausiids and forage fish (blue) in Glacier Bay during May 2000.



Figure 45. Spatial pattern of zooplankton (yellow) and euphausiids and forage fish (blue) in Glacier Bay during June 2000.



Figure 46. Spatial pattern of zooplankton (yellow) and euphausiids and forage fish (blue) in Glacier Bay during July 2000.


Figure 47. Spatial pattern of zooplankton (yellow) and euphausiids and forage fish (blue) in Glacier Bay during August 2000.

Littoral Fish Community

During 1999 and 2000 we caught a total of 50,610 fish in 271 beach seines comprising at least 38 species (not all juvenile sculpins and snailfish identified to species; Appendix 7). Small subsamples (see methods) were kept for further analysis with the proportion of fish identified, counted, and returned immediately to the ocean, thus minimizing mortality. The diversity of littoral zone species in Glacier Bay (38 species) is comparable to another southeast Alaskan study in which Orsi and Landingham (1985) found 42 species. The species composition of their study included 14 species not found in our beach seines, although, four of which were found in other Glacier Bay sampling methods. Their study was in Icy and Chatham Strait, which are more open bodies of water, possibly explaining these differences. Species found by that study and not found by this study were steelhead trout; slender cockscomb; snake prickleback; spotted snailfish; thorny, tidepool, crested, and manacled sculpins; red Irish lord; and tubenose poacher. Of these, spotted snailfish; crested and manacled sculpin; and tubenose poacher have never been reported elsewhere in the bay (see Orsi and Landingham for capture citations). Other nearshore areas in subarctic Alaska also produced comparable numbers of species; Robards et al. (1999) caught 52 species in lower Cook Inlet and Isakson et al. (1971) caught 40 species in the nearshore waters of Amchitka Island.

When broken down by area and season, as few as one species comprised 90 % of the fish caught (Table 4). In all areas and time periods the 5 most dominant species made up at least 79 % of the fish caught. As expected from these patterns of relative abundance, these fish were predominantly juveniles (Gibson *et al.*, 1996) and typically low in the trophic web (Allen, 1982). In estuarine, inshore, and bay habitats in the northeastern Pacific, our results corroborate with the tendency for five or fewer species to account for more than 75% of the individuals in local fish communities, even though the total number of species comprising these communities may be much larger (e.g., Allen and Horn, 1975; Hancock, 1975; Horn, 1980; Allen, 1982; Gordon and Leavings, 1984; Orsi and Landingham, 1985).

High-latitude fish assemblages such as those in Glacier Bay, and particularly those in shallow water, are subjected to large seasonal variations in temperature and day length. These physical factors impart a strong seasonality to community structure (Nash, 1988). In Glacier Bay species composition (relative abundance; Figure 48) and frequency at which different species are caught (Table 5) indicated that early season catches (lower and middle bay) are dominated by out-migrating salmonids and late-season catches are predominantly recruiting juveniles (middle and upper bay). Mid-summer declines in abundance may be a result of physical factors, but like here, can frequently be attributable to changes in community structure. In particular, the changes in composition and abundance are frequently related to influxes of juveniles of various species (Robards et al., 1999; Gordon and Levings, 1984). Many other investigators have also noted decreased fish abundance between spring and fall peaks (in this case July; e.g., Livingston, 1976; Horn, 1980; Allen, 1982; Thorman, 1986*b*; Methven and Bajdik, 1994).

*	June, 1999	June, 2000	July, 2000	August 2000
Icy Strait	No Data	Pink salmon (96 %)	Pacific herring (64 %) Great sculpin (18 %) Sand lance (5 %) Buffalo sculpin (5 %)	Pacific cod (35 %) Kelp greenling (25 %) Silverspotted Sculpin (20 %) Pink Salmon (10 %)
Lower Bay	Pink salmon (85 %) Unidentified sculpins (11 %)	Pink salmon (47 %) Unidentified sculpins (41%) Dolly varden (6 %)	Walleye pollock (74 %) Great sculpin (13 %) Dolly varden (9%)	Sand lance (32 %) Pink salmon (21 %) Great sculpin (18 %) Pacific herring (7 %) Dolly varden (5 %) Kelp greenling (4 %) Silverspotted sculpin (3 %)
Middle Bay	Sockeye salmon (51 %) Pacific sand lance (19 %) Pink salmon (12 %) Coho salmon (6 %) Pacific herring (3 %)	Pink salmon (77 %) Rock sole (11 %) Unidentified sculpins (8 %)	Pink salmon (46 %) Great sculpin (13 %) Pacific herring (11%) Butter sole (10 %) Slender eelblenny (5 %) Pacific sand lance (4 %) Coho salmon (3 %)	Pacific herring (87 %) Pacific sand lance (9 %)
West Arm	Unidentified sculpins (37 %) Pacific sand lance (26 %) Slender eelblenny (18 %) Walleye pollock (9 %)	Unidentified sculpins (52 %) Pink salmon (10 %) Pacific sand lance (9 %) Slender eelblenny (7 %) Butter sole (7%) capelin (5 %)	capelin (44 %) Slender eelblenny (30 %) Great sculpin (11 %) Rock sole (10 %)	capelin (73 %) Slender eelblenny (17 %) Pacific sand lance (4 %)
East Arm	Unidentified sculpins (34 %) Slender eelblenny (34 %) Pacific sand lance (15 %) Dolly varden (12 %)	Slender eelblenny (29 %) Dolly varden (22 %) Sockeye salmon (15 %) Unidentified sculpins (9 %) Butter sole (6 %) Pacific sand lance (6 %) Coho salmon (5 %)	Pacific herring (42 %) Great sculpin (19 %) Rock sole (14 %) capelin (14 %) Slender eelblenny (3 %)	Pacific sand lance (83 %) capelin (7 %)

Table 4. Species Contributing 90 % of the beach seine catch by area and season



Figure 48. Species composition of beach seine catches in Glacier Bay during 1999 and 2000. Figures represent fish caught only at monitoring locations.

Although we beach seined at numerous sites, the bulk of the fish caught were from only a few sites. Furthermore, three of the top four ranked sites for catch abundance were also in the top four for number of species present at a site; sites BS19 at Spokane Cove, BS36 at Gloomy Knob, and BS09 in South Fingers (Figure 49). The high productivity and species diversity at Spokane Cove and South Fingers are perhaps not surprising as both areas are well developed salmonid spawning areas and are therefore subject to additional nutrient inputs from established riparian zones. Perhaps more surprising was the high productivity at the Gloomy Knob site in the West Arm. Vivid Lake (outlet adjacent to the BS36 Gloomy Knob study site) is an important salmonid spawning area (Milner, 1987). However, other salmonid systems exist in the east arm that did not display this level of production (e.g., BS 25).



Figure 49. Beach seine stations in Glacier Bay (note different order of study sites in A and B) ranked by A). Mean abundance (filled circles) of fish over the four sampling periods (\Box June 1999, \circ June 2000, Δ July 2000, \Diamond August 2000); and B). Number of species caught.

Catch-per-unit-effort (per individual set) and relative taxa abundance for each month indicated a pronounced seasonality to the nearshore assemblages (Table 6; Figure 50). Additionally, marked spatial differences were observed around the bay. Salmonids dominated the June nearshore community in Icy Strait (1999 and 2000) and the middle bay (2000 only). Our sample locations in the lower bay apparently precluded catches of these salmonids that were regularly observed swarming around the dock in Bartlett Cove (Robards, pers obs.). Overall, catch was low in July throughout the bay but increased dramatically in the middle bay and upper-arms during August. A single large (18,470 individuals) catch of juvenile (first-year) herring (Figure 51) in Spokane Cove accounted for most of the increased CPUE in the middle bay for August. This catch was surprising in light of few herring being caught in Icthyoplankton samples. Juvenile (first-year) capelin (west arm) and sand lance (west and east arms) accounted for most of the increased August CPUE in the upper-arms (Figure 51).

Variability in nearshore marine communities is profound (see Robards et al., 1999). Glacier Bay's nearshore community is no exception with profound differences already discussed in seasonal assemblages. Diel variation is important for many species; however, our sampling occurred exclusively during daylight hours. Therefore, diel consideration are not considered here, except to say that for species such as cod, greater numbers may have been present at night than represented in our daytime sampling. Differences in nearshore fish catches at high and low tides is also profound. Clearly, high tide catches only represent those fish that swim from the sub-tidal up into the intertidal and a few permanent intertidal dwellers as opposed to low tide catches that encompass those and all obligate nearshore subtidal species. Our results indicated this difference in an order of magnitude difference between high and low tide catches (Figure 52). This should be considered when repeating our study or defining a new nearshore investigation.

1777 und 2000.		Jillollicu.		
	1999		2000	
	June	June	July	August
Herring	31.25	0.00	47.72	38.19
Salmonid	25.36	29.16	19.95	21.81
Dolly Varden	21.47	33.04	27.35	23.19
Smelt	18.97	13.50	53.57	43.80
Gadid	47.16	11.96	7.06	17.82
Sand Lance	41.77	20.58	28.87	42.22
Sculpin	27.11	27.24	35.17	32.52
Flatfish	16.00	13.21	28.15	23.33
Other	24.47	20.53	13.97	25.31

Table 5. Frequency of capture for key taxa in Glacier Bay beach seines during 1999 and 2000. All areas are combined.



Figure 50. Catch-per-unit-effort for beach seine sets in the different eco-regions of Glacier Bay by season.



Figure 51. Young-of-the-year (from top) Pacific sand lance, capelin, and Pacific herring caught by beach seine close to Lamplugh Glacier (Stn. 46) during August 2000.



Figure 52. Size-frequency of high and low tide beach seine catches in Glacier Bay during 1999 and 2000.

Pelagic Fish Community

Modified herring mid-water trawls in June 1999

A total of 48 tows at 34 stations were fished from June 10 to 23, 1999 (Appendix 7). A total of 5011 fish were captured, including both small schooling and larger pelagic fish. Catch numbers were standardized for catch-per-unit-effort (CPUE), standardized for 1000 m² distance towed, and determined as "good" or "bad" based on efficiency of catching the acoustic target. We fished 6 stations in Icy Strait, 7 in Lower Glacier Bay, 10 in the Middle Bay, 5 in the East Arm of Glacier Bay, and 6 in the West Arm of Glacier Bay.

Species composition was distinct between Icy Strait and the four regions of Glacier Bay (Figure 53). Pacific herring were never captured by this method in Glacier Bay although they comprised the majority of fish caught in Icy Strait (74%). Capelin were also abundant (14%) in Icy Strait, and near Point Adolphus we captured Northern lampfish at 90 m fishing depth. The most abundant fish species in Glacier Bay were walleye pollock and capelin. In the Lower Bay the remaining 35% of fish species composition were larval fish; 12% sand lance, 7% Sculpins (Cottidae), 7% Pricklebacks (Lumpenus spp.), and 6% flatfish (Pleuronectidae). In the Middle bay the 6% other fish were mostly larval sand lance and larval pricklebacks, and in the West arm the 22 % other fish were all larval pricklebacks. Within the four regions of Glacier Bay, Muir Inlet was distinct. Other fish species in Muir Inlet was predominantly Northern lampfish (1%) and Northern Smoothtongue (3%). Northern Smoothtongue were only caught by this method in front of the Muir Glacier and always in conjunction with catches of Northern lampfish (Note that the Isaacs Kidd mid-water trawl caught northern smoothtongue in front of Margerie in 2000. The Isaacs Kidd was used much closer to the glacier front which may have explained this difference).

During this study 50.6 % of all fish caught had fork length < 50 mm. Using the same equipment in Cook Inlet we caught much lower proportions of small fish during 1997 (1.01%) and July 1998 (1.5%). Mean fish length varied among the five regions. In Icy Strait 23% of fish were between 50 and 99 mm, 31% of the catch was size < 50 mm, and 35% was between 200-249 mm (this size class mostly herring). In Lower and Middle Glacier Bay the majority of the fish captured (over 90%) were larval fish with fork length < 50 mm. In the West Arm over half of the fish captured were larval species, 22% were between 50 and 99 mm, and 16% were greater than 300 mm (mostly adult walleye pollock). In Muir Inlet the majority of fish were size 50 - 99 mm (58%), with 40% larval fish size < 50 mm. We did not catch any forage size (50 - 150 mm) walleye pollock or herring in Icy Strait or Glacier Bay mid-water trawls.

Throughout Icy Strait and Glacier Bay, mean fish CPUE was relatively low compared to similar work in Cook Inlet, with Muir Inlet having the highest CPUE at 360 fish per set. Weights of euphausiids were an order of magnitude greater than jellyfish, and CPUE of both were highest in Muir Inlet.

Adult walleye pollock (fork length 231 to 602 mm) stomach contents (169 stomachs analyzed) reflect the trend of high invertebrate/low fish abundance within Glacier Bay.

The majority of pollock stomachs (65%) had only euphausiids, whereas only 15% had fish. Of the 25 stomachs with fish, 64% actually contained larval fish and euphausiids; the other 36% had capelin and northern lanternfish. Crustaceans in pollock stomachs included mysids, amphidods, isopods, and shrimp.



Figure 53. Relative composition of key taxa caught by modified herring mid-water trawl during June, 1999 in the different eco-regions of Glacier Bay.

Isaacs Kidd mid-water trawls in July and August 2000

A total of 31 tows at 18 stations were fished from July 26 to August 17, 2000 (Appendix 7). A total of 3267 fish, 64,872 euphausiids, 3 cephlapods, 132 shrimp (Pandalus, Pasiphaea, and Neomysis), and 162 jellyfish were caught.

Catches for each trawl were standardized for catch-per-unit-effort (CPUE) and adjusted for 1000 m³ of water passing through the net. Composition of the total fish caught was dominated by young-of-the-year walleye pollock (53 %) and capelin (35 %). These two species, along with slender eelblenny (6 %) and the two bathypelagic species – northern lampfish (3 %; Figure 54) and northern smoothtounge (1%) made up over 98 % of the total catch. Size composition of the fish caught included both small schooling and larger pelagic fish (Figure 55). All but two of the fish over 60 mm (n = 189; 6 % of total catch) were caught at the head of Glacier Bay adjacent to Margerie Glacier (62 %), Muir Glacier (15 %), and in Rendu Inlet (14 %), Queen Inlet (4 %), Johns Hopkins Inlet (2 %), and Reid Inlet (2 %).

Walleye pollock, capelin, and slender eelblenny larvae along with the northern lampfish and northern smoothtongue made up over 98 % of the pelagic fish larvae caught (Figure 56), similar to results described by Mattson and Wing (1978) for inland coastal waters of southeastern Alaska. These authors found 50 % of larval fish caught in June were walleye pollock, which favored channels rather than inside bays. These authors also found osmerids to be the second most common taxa and speculated that capelin was the most abundant of these osmerids (not all identified to species).

Although the Isaacs Kidd mid-water trawl was only used in the upper 50 m of the water column we saw strong and repeating patterns within Glacier Bay. These include:

-Juvenile pollock were not generally caught south of Tlingit Point during July and August by this method.

-Although waters around Margerie Glacier supported very high numbers of juvenile fish, the east arm generally supported higher numbers of (primarily juvenile) capelin than the west arm. Our fishing at Margerie Glacier may have been biased by our close proximity to the ice-front compared to for example McBride (at station 18). Zooplankton results for June indicated about twice the abundance at station 21 close to Margerie as at the closely adjacent station 12. Further research is needed to describe the extent and mechanisms driving this productivity (see recommendations for future work).

-The areas close to tidewater glaciers (upper east and west arms) appeared particularly productive for fish and euphausiids (Figure 57). Rendu Inlet also appeared remarkably productive, especially when compared to neighboring Queen Inlet. Reasons for the differences between these two sites can only be speculative based on our results. However, Queen Inlet has particularly high sedimentation rates (up to 6 cm per day). This, in conjunction with a photic depth of less than 10 m (Hooge, USGS, Glacier Bay Field Station, pers comm.) probably restricts potential productivity at this site.



Figure 54. Northern lampfish caught by the Isaacs-Kidd mid-water trawl in upper-Muir Inlet.

Figure 55. Typical catch by the Isaacs-Kidd midwater trawl at Margerie Glacier (Stn. 21). Catch is separated into (from top) large fish (pollock and hake), small schooling fish (capelin, pollock, northern smoothtongue, slender eelblenny), and euphausiids.





Figure 56. Relative composition of key taxa in Isaacs-Kidd mid-water trawls in lower (1), middle (2), west arm (3), and east arm (4) of Glacier Bay during July and August, 2000.

<u>Icthyofauna</u>

Morphometric data from the numerous fish caught in this study can be of importance to future research. Figure 57 depicts the range of length and weight data available (on accompanying data cd) for analysis.



Figure 57. Length-weight relationships for key Forage fish species within Glacier Bay. Plots are compiled from fish collected by all capture methods.

Discussion of Select Species

Pacific Herring

During June 1999, herring were only caught nearshore in small numbers (although relatively frequently) within the middle region of Glacier Bay. However, during 2000 when we sampled throughout the season, juvenile herring were found in extremely large numbers in the middle-bay, particularly at Spokane Cove. The seasonal length/frequency graph (Figure 58) clearly indicates the arrival of recruiting juvenile herring in the latter part of the summer through July and August. Herring are known to congregate in shallow embayments where they take advantage of fewer predators and relatively higher food availability for growth (Figure 59). Pelagic trawls caught adult herring in Icy Strait but not within Glacier Bay. It remains unknown whether herring spawn within Glacier Bay or the recruiting juveniles found in nearshore catches are advected into the bay from spawning that occurs outside.



Figure 58. Length-frequency distribution of Pacific herring caught in Glacier Bay during 1999 and 2000 by beach seine and pelagic trawl. Young-of-the-year fish are indicated by the dashed line.



Figure 59. Daily mean (+/- sd) size (mm) of juvenile Pacific herring caught in beach seines and pelagic trawls.

Pacific Salmon

We caught juveniles of all five species of salmon within Glacier Bay. Most salmon were out-migrating juveniles with only two adults caught (one pink and one chum). Catch rates of salmon in the nearshore were highest during early summer, in accordance with Southeast Alaska juvenile salmonid surveys conducted by the National Marine Fisheries Service Auke Bay Laboratory (Orsi et al., 1998).

Spatially, salmon species (all juvenile) ranged from common and abundant in the lower region of the bay, to infrequently caught individuals in the upper bay. Quality of streams for spawning by these anadromous fish is likely the most important factor influencing their distribution. Quality for salmon improves from the newly formed, cold, highly turbid streams in the upper reaches of the bay to the more mature streams in the lower bay (Milner, 1987). Milner (1987) clearly showed a corresponding succession from low salmonid numbers in the upper bay to an abundance in the lower bay streams. Turbid waters may result in reduction or loss of sight-feeding capabilities, reduced growth, increased stress and interference with environmental cues necessary for orientation in migration (Lloyd, 1987– cited in Milner & Bailey, 1989). Apart from turbidity and temperature, wash-out from redds during high spring and autumn discharges has been directly linked to poor salmonid production elsewhere in southeast Alaska (Tyler & Gibbons, 1983).

Pink salmon (as well as Chum) appear to be two of the most suitable initial colonizers of newly formed stream systems as their fry migrate directly into the ocean, eliminating the need for suitable rearing habitat in the highly turbid waters. However, sockeye, coho and Chinook are known to spawn in the lower reaches of turbid Alaskan rivers (Milner & Bailey, 1989). The ability of pink and chum salmon to outmigrate in their first year was reflected in their small size compared to other salmonids in this study. Pink salmon have also been reported as one of the species most likely to stray from their home streams

(Pritchard, 1939), enhancing their ability to colonize new periglacial areas. Milner (1987) did not find pink or chum salmon in Nunatak Creek (upper east arm) and suggested that although these species can avoid prolonged freshwater residence, estuarine conditions may preclude fry development in this area. We emphasize that even though we found juvenile salmon throughout the bay, our results do not establish if these fish originated in Glacier Bay or whether they were advected into the bay by currents.

Our results indicated that timing of out-migration for juvenile salmonids largely occurred in the early part of summer (May and June). Migration of juvenile salmonids out of Glacier Bay toward the open ocean is expected to take advantage of optimal water temperatures, salinities, and food availability (Sheridan, 1962). Orsi (2000) found that the timing of seaward migration of (southeast Alaska) juvenile salmon coincided with seasonal peak periods of temperature and zooplankton. With peak zooplankton abundance in the lower bay and Icy Strait in May and early June, this pattern appears to coincide with our results, at least for prey resources.

Length of pink salmon (Figures 60 and 61) for each days catch (daily mean length) indicated growth from a mean of about 30 mm in early June to about 60 mm in July. We did not continue to catch bigger and bigger pink salmon through July and August, the mean size of cohorts remaining at about 60 mm. Reasons for this apparent reduction in growth rates lies in their behavioral change from a nearshore to offshore existence (Heard, 1991). The exact size at which the shift to deeper water occurs differs between areas. Cooney et al., (1978) reported 60-70 mm for Prince William Sound. Larger individuals are the first to migrate to open waters (Heard, 1991), which may explain the lack of larger-sized juvenile pink salmon in the nearshore area in this study. Offshore sampling in Icy Strait during June by the National Marine Fishery Service (Orsi et al., 1997, 1998) reported offshore juvenile salmon with sizes ranging somewhat larger than in our nearshore study (73-136 mm in 1997 and 89-150 mm in 1998; no offshore pink salmon found in May). These results suggest that smaller pink salmon continue to reside close to shore until these minimum sizes are reached. Concurrent changes in physiology may compound this situation. LeBrasseur and Parker (1964) indicated reductions in growth of juvenile pink salmon after lengths of 60-80 mm were reached. Numerous authors (see Heard, 1991) have reported other associated changes as juvenile pinks move offshore including lower lipid levels and formation of smaller schools (500-2,000 fish).

The importance of juvenile salmonids as prey in Glacier Bay was beyond the scope of this project. However, they are a key prey in other Alaskan areas for at least walleye pollock (Prince William Sound; Willette et al., 1999), coho salmon (Wing, 1985), spiny dogfish (Beamish et al., 1992), marine mammals, and seabirds (Scheel and Hough, 1997), all of which commonly occur in this area.

Anecdotally, on July 14, 2000 one juvenile pink salmon was found with a stomach containing juvenile capelin.



Figure 60. Length-frequency distribution of pink salmon caught in Glacier Bay during 1999 and 2000 by beach seine and pelagic trawl. Young-of-the-year fish are indicated by the dashed line.



Figure 61. Daily mean (+/- sd) size of juvenile pink salmon caught in 1999 (squares) and 2000 (circles).

Table 6. Catch-per-unit-effort of Dolly Varden within Glacier Bay by sampling region during 1999 and 2000.

Area (sets)	Catch-per- unit effort			
Icy Strait (22)	3.4			
Lower Bay (31)	8.4			
Middle Bay (85)	1.5			
West Arm (91)	0.2			
East Arm (42)	1.3			

Dolly Varden

Dolly Varden were included in this discussion due to their frequency of capture and abundance within nearshore areas of Glacier Bay. Dolly Varden were the most common Char caught (only 4 cutthroat trout were caught). This species was only caught in beach seines and markedly more abundant in the lower regions of Glacier Bay than in the upper-arms.

We found few Dolly Varden in the West Arm as opposed to the East Arm despite extensive sampling (Table 7). This species is a common predator in the nearshore area of Alaskan marine bays and is typically the first salmonid to colonize new stream habitats (Milner, 1994). The lower abundance in the West Arm mirrors the colonization pattern for salmonids, which are also far more prevalent in the east arm. The geography for the lower regions of the two areas is markedly different. From Muir Point in the East Arm to Wachusetts Inlet there are wide benches of vegetated land that include several lake systems. In the West Arm the land rises quickly from the water into the coastal mountains, and only includes one lake system (Vivid Lake). This topography likely impedes colonization by salmonids and Dolly Varden in much of the West Arm, isolating these species to a few areas such as Vivid Lake. No clear class structure was present from our samples, although at least three age-classes are suggested from the length/frequency histogram (Figure 62). This species takes 3-4 years to reach average smolt size of 135 mm in fork length (Armstrong, 1970).



Figure 62. Length-frequency distribution of Dolly Varden caught in Glacier Bay during 1999 and 2000 by beach seine.

Walleye Pollock

Walleye pollock larvae and juveniles were the most abundant species in our Icthyoplankton trawls, Isaacs-Kidd mid-water trawls, and modified herring trawls.

Close to shore, in beach seines we caught juvenile pollock in all nearshore areas of the bay during June of 1999 and 2000 but not in Icy Strait. Most of these pollock were caught in the upper-arms. By July numerous juvenile pollock were present in the lower bay (Table 7). Finding juvenile pollock in the glacially influenced arms of Glacier Bay raises the question as to whether spawning occurs in this area.

Table 7. Nearshore catch-per-unit-effort for young-of-the- year walleye pollock in Glacier Bay by sampling area

Glueier Buy by Sumpling urea								
Area	June	July	August					
Icy Strait	-	-	0.8					
Lower Bay	0.1	74.5						
Middle Bay	0.1	0.1	0.2					
West Arm	2.7	0.1						
East Arm	0.5	-	-					

Length of walleye pollock (Figures 63 and 64) for each days catch (daily mean length) indicated growth from a mean of about 5 mm in early June to about 50 mm in August. We estimated a late April hatch-date for walleye pollock from the larvae we caught in Tucker trawls. This is similar to hatch dates reported for elsewhere in the northern Gulf of Alaska (e.g., Müter and Norcross, 1994) and fits with spawning potentially occurring in late winter (March to early April in Shelikof Straight; Kim and Gunderson, 1988). We have no early season pelagic trawl data to confirm this. The only demersal finfish caught in the upper bay during exploratory fishing trawls in March of 1954 (cited in Hale and Wright, 1979) was walleye pollock. Their presence in these areas at that time would be appropriate if spawning is taking place in these areas in early spring.

The only non-young-of-the-year fish in nearshore areas were captured on 6/19/99 when six juvenile pollock were caught in Reid Inlet (mean length 134 mm) and on 8/8/00 when a 403 mm adult was caught in Muir Inlet.

Anecdotally, walleye pollock were observed co-schooling with herring at the entrance to Berg Bay during underwater video work (John Brooks – 805-644 -5185 x 109).



Figure 63. Length-frequency distribution of walleye pollock caught in Glacier Bay during 1999 and 2000 by beach seine and pelagic trawl. Young-of-the-year fish are indicated by the dashed line.



Figure 64. Daily mean (+/- sd) size of juvenile walleye pollock caught in beach seines (squares) and pelagic trawls (circles); solids = 1999, open = 2000.

<u>Capelin</u>

During the spring, capelin tend to concentrate along the Alaskan coast to spawn on gravel beaches. Time of spawning varies with location and water temperature, ranging from April to August at 5-10 °C (Pahlke, 1985). Preferred beaches generally have gravel substrate of grain size from 2.5 to 25 mm (Templeman, 1948; Jangaard, 1974). Observations by coast walkers in Glacier Bay noted signs of intertidal spawning on June 30 1997 and during July 1998 (USNPS- Bill Eichenlaub, personal communication). Although speculative, based on timing (Pahlke, 1985) and maturity of the few adult capelin we caught, these events are likely the result of capelin spawning (the other intertidal spawners generally spawn earlier (herring) or later (sand lance, Robards et al., 1999).

Most of the capelin that were caught in beach seines in July and August were young-ofthe-year fish (Figures 65 and 66). These fish were a dominant component of the upperarm community in both July and August. Peak recruitment to the nearshore was clearly in August based on CPUE (Table 8). No capelin were caught in the lower bay or Icy Strait nearshore areas, although they do exist in this region based on trawl catches.

Capelin were observed being preyed on by arctic terms at the entrance to Bartlett Cove but were not caught in seines in that region.



Figure 65. A. Length-frequency distribution of capelin caught in Glacier Bay during 1999 and 2000 by beach seine and pelagic trawl. Young-of-the-year fish are indicated by the dashed line.

				Dolly						
Region	Season	Herring	Salmonid	Varden	Smelt	Gadid	Sand lance	Sculpin	Flatfish	Other
Icy Strait	2000-1		575.50					1.00		25.50
	2000-2	7.00					0.50	3.00	0.50	
	2000-3		5.00			18.00		11.00	1.50	16.50
Lower Bav	1999-1		139.50	4.00				19.38	0.63	0.13
	2000-1		29.42	3.83		0.08	0.08	25.33	0.25	1.92
	2000-2		0.30	3.60		29.80		6.40		0.30
	2000-3	1.60	5.00	1.10		0.50	7.40	5.20	0.70	1.90
Middle Bav	1999-1	1.94	25.19	0.19	0.31	0.69	34.81	2.38	2.00	0.19
	2000-1		319.79	0.42	0.05	2.21	8.68	33.95	47.58	3.68
	2000-2	8.83	39.56	1.33		0.17	3.39	11.28	9.28	4.28
	2000-3	1055.06	14.56	4.94	2.94	0.22	104.61	14.61	14.22	1.11
West Arm	1999-1			0.18	3.64	4.09	12.00	17.00	0.45	8.27
	2000-1		1.91		0.91	0.65	1.74	9.96	2.09	1.39
	2000-2	0.21	1.86	0.57	30.64	0.07	0.07	8.64	7.07	21.29
	2000-3	2.20	0.13	0.27	189.80	1.13	11.33	6.87	5.07	43.87
East Arm	1999-1		0.14	1.29	0.14		1.57	3.86		3.57
	2000-1		3.75	4.00		0.13	1.00	2.13	1.75	5.25
	2000-2	6.13	0.75		2.00		0.38	2.88	2.13	0.50
	2000-3	20.63	4.63	1.50	51.13	0.13	574.38	5.88	2.25	27.50

Table 8. Catch Per Unit-Effort (CPUE) of seine catches in Glacier Bav during 1999 and 2000.



Figure 66. Daily length and standard deviation of capelin caught in beach seines. Squares represent 1999 data, circles represent 2000 data; all fish less than 60 mm are young-of-the-year fish.

Slender Eelblenny

This species (incorrectly identified as snake prickleback in 1999 report) was widely distributed in the upper parts of Glacier Bay; few were caught in the lower and middle regions. Larval eelblennys were most common except in Reid Inlet (beaches 43-45) where large numbers of adults were caught within one-half mile of Reid Glacier. Results were similar to Chisik Island in Cook Inlet, which is also a glacial-silt dominated system. In this area, numerous snake pricklebacks were also found in otherwise depauperate silty areas. This species may be an important early colonizer of nearshore areas. Size distribution showed the newly recruiting young-of-the-year eelblenny (Figures 67 and 68).



Figure 67. Length-frequency distribution of slender eelblenny caught in Glacier Bay during 1999 and 2000 by beach seine and pelagic trawl. Young-of-the-year fish are indicated by the dashed line.



Figure 68. Daily mean (+/- sd) size of juvenile slender eelblenny caught in beach seines and pelagic trawls (squares = 1999, circles = beach seines 2000, triangles = pelagic trawls 2000).

Sand Lance

Sand lance are closely associated with clean sand substrates, in which they live for much of their lives. Areas of high water flow, such as the entrance to lagoons and outwash plains, generally provided the best substrates for sand lance. We caught sand lance in beach seines at most beaches of this type within Glacier Bay. Much of the sandy substrate in Glacier Bay is covered in mussel beds rendering it unsuitable for sand lance. Although we were unable to sample several outwash areas by seine, we established by digging in intertidally exposed substrates that sizeable sand lance populations exist at the following areas: the flats at the southwestern head of Geike inlet; the outwash from Dirt Glacier. Indications of the importance of sand lance is the large congregations of pigeon guillemots, glaucous-winged gulls, black-legged kittiwakes, cormorants, scoters, porpoises, and seals were seen feeding - some of which were observed carrying sand lance; and at beaches 33 and 34 located opposite the entrance to Adams Inlet. Bruce Wing (NMFS- Auke Bay Lab personal communication) also indicated his observations of sand lance congregating at the entrance to Berg Bay. Hooge (USGS-BRD personal communication) also observed sand lance burrowed between fingers and berg in Whidbey Passage. Size distribution of sand lance clearly showed the newly recruiting young-of-the-year and adult sand lance (Figures 69 and 70). Without otolith analysis, number of age classes of adult sand lance could not be established.

Median size of juvenile sand lance in June 1999 (33 mm) was significantly smaller (Mann-Whitney rank sum test; P<0.001) than June 2000 (41 mm) even though they were caught approximately 10 days later.



Figure 69. Length-frequency distribution of Pacific sand lance caught in Glacier Bay during 1999 and 2000 by beach seine and pelagic trawl. Young-of-the-year fish are indicated by the dashed line.



Figure 70. Daily mean (+/- sd) size of juvenile Pacific sand lance caught in beach seines and pelagic trawls (squares = 1999, circles = 2000).

Sculpins

Sculpins (mostly juveniles) were the most commonly caught taxa by beach seine in this study. Small juveniles ranging from up to about 25 mm dominated sculpin catches. Second year fish were caught regularly, but at the rate of only one or two per set. These fish were distributed throughout the bay, and unlike many of the other species sampled, did not display any preference for the different sample regions.

<u>Flatfish</u>

Flatfish were predominantly found in the lower-reaches of Glacier Bay. Rock Sole were the most common species caught within the bay. However, in the lower reaches and particularly into Icy Strait we found larger numbers of English Sole.

At least two size classes of flatfish are evident from size-frequency histograms. Newly recruiting juveniles at about 30mm for both Rock and English Sole were found, as well as second year fish ranging from 60-140 mm.

Numerous reports have described the necessity for specific substrates in flatfish nursery areas (e.g., Rogers, 1992). Fine grained or muddy, well sorted sediments in shallow water are frequently required (Rogers, 1992). Furthermore the importance of adequate supplies of benthic food organisms to the settlement of flatfish larvae at metamorphosis, and to their subsequent survival and growth has frequently been documented (e.g, Creutzberg et al., 1978).

Mesopelagic Species

Both juvenile and adult mesopelagic species (myctophids and northern smooth tongue) were found as close to the surface as 10 m (modified-herring mid-water trawl) during summer 1999 and 2000. Vertical diel migrations for these species have frequently been reported, and may be a response to light intensity or hunger (e.g., Paxton, 1967, Pearcy et al., 1977; Calilliet and Ebeling, 1990). However, our sampling was not able to establish if these species are migrating each day or are persistently present in the upper 100 m at the heads of Glacier Bay. Length-frequency distributions remained similar throughout the summer months (Figure 71)

There is tremendous silt content in the marine waters of the upper-reaches of Glacier Bay. If light intensity is the cue that triggers diel migrations, and hence their uncharacteristic daytime surface water existance, then high turbidity due to glacial silt may significantly reduce light level penetration and account for the presence of northern lampfish and northern smoothtongue.. However, if food is the primary cue for vertical migration, then these species may preferentially exist in the upper reaches of the water column, feeding on the elevated zooplankton abundance.

Myctophids have 2 – 10 times the lipid concentrations of other forage fish species, such as capelin and sand lance (Van Pelt et al., 1997), and they are the most important food source for a wide variety of larger fishes, seabirds, and marine mammals in the central North Pacific (Springer et al., 1999). Adult walleye pollock frequently consume both northern lampfish (Yang, 1996) and northern smoothtongue (Mason and Philips, 1985). Northern lampfish are also an important prey item for adult salmonids (Nagasawa et al., 1997; Shimada, 1948).

The occurrence of myctophids along the inside waters and fjords of Southeast Alaska makes them available to a suite of predators that may not usually find these characteristically offshore fish. For example, black-legged kittiwakes, which feed at night on northern lampfish at oceanic islands in the Aleutian Archipelago (Springer et al., 1996), may be able to regularly feed on myctophids during the day in the upper reaches of Glacier Bay, adjacent to tidewater glaciers. Marbled murrelets were also found almost exclusively consuming myctophids in Icy Strait (USGS-BRD, John Piatt personal communication). Given the high nutritive value of these fish, and the variety of predators that consume them, these mesopelagic species may play an important role in local food webs.



Figure 71. Length-frequency distribution of northern lamp-fish caught in Glacier Bay during 1999 and 2000 by beach seines and pelagic trawls.

Predator Survey

Species Inventory

Surveys of seabirds and marine mammals in Glacier Bay covered 1,514 km (June 1999), 440 km (November 1999), 471 km (March 2000), and 1,418 km (June 2000; Figure 72). Summer surveys covered a larger sample area including Icy Strait (1999, 2000), Dundas Bay (1999), and a denser network of pelagic transects (Figure 72). A total of 118,520 birds (64 spp.) and 1780 (7 spp.) marine mammals were counted along survey transects during the 4 surveys June 1999 - June 2000 (see Appendix 8 for a complete list).



Figure 72. Survey tracks in and around Glacier Bay National Park June 1999, November 1999, March 2000, and June 2000. Surveys were conducted aboard the *RV Padalus*, *RV Alaska Gyre*, *MV David Grey* and *MV Lutras*. Tracks represent 200 and 300m swaths (depending on vessel – see methods) within which all species were counted.

<u>Seabirds</u>

Seabirds accounted for a high of 46.8% of all birds sighted in June of 1999 to a low of 22.0% in the March 2000 survey (Table 9). Seabird numbers were dominated by black-legged kittiwakes (*Rissa tridactyla*), glaucous-winged gulls (*Larus glaucescens.*), murrelets (*Brachyramphus* spp), and pigeon guillemots (*Cepphus columba*). Together, these species accounted for 74% and 89% of all seabirds identified during surveys. Black-legged kittiwakes and glaucous-winged gulls accounted for the bulk of gull numbers. Marbled murrelets (*B. marmoratus*) greatly outnumbered Kittlitz's murrelets (*B. brevirostris*) sighted in the summer surveys; however, numbers were similar in the spring 2000 survey. Fall and spring surveys generally yielded lower densities of most seabird species, with the notable exception of Kittlitz's murrelets in the spring 2000 (similar to summer densities) may indicate an earlier arrival to Glacier Bay then the majority of migratory seabirds.

We calculated densities (birds/km²) for some of the more common species to compensate for the effects of differential survey lengths. Densities generally followed the trends of raw counts, but provide a better method for examining seasonal trends. Seabird densities were greatest during the summer months (Table 10). Many seabirds were rare or absent during the less extensive fall and spring surveys. Densities from the June 2000 survey were lower than the densities recorded in June of 1999 for all but a few species. Densities of all *Brachyramphus* murrelets (marbled, Kittlitz's, and unidentified murrelets) were the notable exceptions. Although the densities of marbled murrelets was lower in June of 2000, there were many more unidentified murrelets. When all *Brachyramphus* murrelets were combined, there was little difference between the two summer surveys. Although they did not make up a large portion of the birds sighted, four species of loon and two species of grebes were seen on transects.

<u>Waterfowl</u>

Waterfowl were the most common birds seen on surveys. Sea ducks in particular constituted from between 47-56% of all birds sighted on surveys (Table 9). Scoters (*Melanitta spp*), by themselves, accounted for 36%-40% of all birds counted on the summer surveys and 15%-17% of all birds seen on fall and spring surveys. Barrow's goldeneye (*Bucephala islandica*) accounted for less then 0.2 % of birds on the summer surveys, however, in the fall and spring surveys they accounted for 16% and 17% of all birds respectively. Common mergansers (*Mergus merganser*) accounted for 8.6% and 9.4% of birds seen in each of the summer surveys, but <0.5 % in the fall and spring surveys. Harlequin ducks (*Histrionicus histrionicus*) constituted 1.7% - 4.8% of all birds sighted. Of the remaining waterfowl, Canada geese (*Branta candensis*) were the most common, though they made up only 1% - 2% of the birds on all surveys.

Seasonal densities of waterfowl varied widely. Canada geese and harlequin ducks were seen in similar densities in all surveys (Table 10). Barrow's Goldeneye were nearly absent in summer surveys, but in the fall 1999 and spring 2000 surveys had the highest densities recorded for any bird (Table 10). Conversely, common mergansers were found at low densities in the fall and spring surveys and relatively high densities in the summer

surveys. White-winged and surf scoters were less strictly seasonal, being present in substantial densities in all surveys, white-winged scoter densities were highest in the summer surveys, and surf scoter densities highest in the fall and spring surveys (Table 9).

Shorebirds

Shorebirds were infrequently sighted compared to seabirds or waterfowl seen in Glacier Bay. While some shorebirds may have been missed due the distance of boats to the shore (100m or 150m) and the potential for decreased visibility due to surf conditions, particular care was taken to ensure that few birds were missed. Shorebirds were most common during the November Survey where they made up 3.2% of the birds seen (Table 9). Unfortunately, the majority of these birds (n=529) were unidentified. While densities for all shorebirds were relatively low, the higher densities of shorebird in the fall and spring suggested that Glacier Bay might provide wintering habitat for some shorebirds (Table 10).

Other Birds

Of the remaining birds seen on surveys, only bald eagles (*Haliaeetus leucocephalus*) and Northwestern crow (*Corus caurinus*) were seen in large numbers (Table 9). As a percentage of the surveys, "other birds", made up over 7% of November and March surveys compared to 1.8% and 2.8% of the two summer surveys. Densities suggested that Glacier Bay might provide important winter habitat for Northwestern crows, while bald eagles appear to be year-round residents (Table 10).

<u>Marine Mammals</u>

Harbor seals (*Phoca vitulina*) were the most commonly sighted marine mammals followed by sea otters (*Enhydra lutris*) and harbor porpoise (*Phocoena phocoena*; Table 9). Dall's and harbor porpoise, harbor seals and sea otters showed little seasonal variation in populations over the 4 surveys. Variations in humpback whales (*Megaptera novaeangliae*) densities by survey reflected their migratory nature, present in the summer, and absent in the winter months (Table 11). Conversely, Steller sea lions (*Eumetopius jubatus*) were sighted more frequently in the November and March surveys (Table 11). Overall, the low densities of many marine mammals may have led to a high degree of variability in the reported densities, i.e. at low densities even a small number of animals, seen or missed, can skew results. Table 9. Numbers and percent composition of bird species from marine surveys in Glacier Bay National Park during June 1999, November 1999, March 2000 and June 2000. Numbers represent totals sighted within transects, percent composition (%) was calculated both within surveys and for all surveys combined. Note that summer surveys (June) included more transects within Glacier Bay and transects in Icy Strait (see methods).

_	Jun-99		Nov-99		Mar-0	Mar-00		Jun-00		All Surveys	
	Number	%	Number	%	Number	%	Number	%	Number	%	
Aleutian Tern	0	0.0	0	0.0	0	0.0	1	0.0	1	0.0	
Arctic Tern	533	1.1	0	0.0	0	0.0	260	0.8	793	0.7	
Ancient Murrelet	0	0.0	0	0.0	0	0.0	1	0.0	1	0.0	
Black-Legged Kittiwake	3739	7.7	1109	5.8	321	1.9	1422	4.2	6591	5.6	
Bonaparte's Gull	36	0.1	0	0.0	0	0.0	4	0.0	40	0.0	
Brachyramphus Murrelet	1951	4.0	87	0.5	326	1.9	2864	8.4	5228	4.4	
Caspian Tern	4	0.0	0	0.0	0	0.0	0	0.0	4	0.0	
Common Loon	26	0.1	46	0.2	20	0.1	53	0.2	145	0.1	
Common Murre	35	0.1	5	0.0	1	0.0	18	0.1	59	0.0	
Double-crested Cormorant	0	0.0	16	0.1	0	0.0	0	0.0	16	0.0	
Fork-tailed Storm Petrel	2	0.0	7	0.0	0	0.0	1	0.0	10	0.0	
Glaucous-winged Gull	2068	4.3	808	4.2	703	4.2	1217	3.6	4796	4.0	
Herring Gull	155	0.3	35	0.2	19	0.1	110	0.3	319	0.3	
Horned Grebe	0	0.0	108	0.6	68	0.4	0	0.0	176	0.1	
Kittlitz's Murrelet	506	1.0	4	0.0	163	1.0	402	1.2	1075	0.9	
Marbled Murrelet	4049	8.4	85	0.4	177	1.0	1322	3.9	5633	4.8	
Mew Gull	798	1.7	611	3.2	123	0.7	1156	3.4	2688	2.3	
Pacific Loon	87	0.2	10	0.1	14	0.1	90	0.3	201	0.2	
Parasitic Jaeger	8	0.0	0	0.0	0	0.0	2	0.0	10	0.0	
Pelagic Cormorant	622	1.3	217	1.1	183	1.1	253	0.7	1275	1.1	
Pigeon Guillemot	1926	4.0	504	2.6	933	5.5	1702	5.0	5065	4.3	
Red-faced Cormorant	2	0.0	0	0.0	0	0.0	0	0.0	2	0.0	
Red-necked Grebe	0	0.0	33	0.2	1	0.0	1	0.0	35	0.0	
Red-throated Loon	36	0.1	0	0.0	0	0.0	0	0.0	36	0.0	
Tufted Puffin	37	0.1	0	0.0	0	0.0	14	0.0	51	0.0	
Unidentified Albatross	0	0.0	0	0.0	2	0.0	0	0.0	2	0.0	
Unidentified Alcid	0	0.0	0	0.0	0	0.0	4	0.0	4	0.0	
Unidentified Cormorant	23	0.0	13	0.1	11	0.1	13	0.0	60	0.1	
Unidentified Gull	6006	12.4	1676	8.7	761	4.5	1249	3.7	9692	8.2	
Unidentified Grebe	1	0.0	73	0.4	45	0.3	1	0.0	120	0.1	
Unidentified Large Larid	0	0.0	0	0.0	0	0.0	130	0.4	130	0.1	
Unidentified Loon	98	0.2	51	0.3	20	0.1	80	0.2	249	0.2	
Unidentified Storm Petrel	74	0.2	5	0.0	1	0.0	0	0.0	80	0.1	
Unidentified Tern	0	0.0	0	0.0	0	0.0	1	0.0	1	0.0	
Western Grebe	0	0.0	0	0.0	5	0.0	0	0.0	5	0.0	
Yellow-billed Loon	2	0.0	0	0.0	1	0.0	0	0.0	3	0.0	
Seabirds Subtotal	22824	47.3	5503	28.5	3898	23.1	12371	36.3	44596	37.6	

	Jun-99		Nov-99		Mar-0	Mar-00		Jun-00		All Surveys	
	Number	%	Number	%	Number	%	Number	%	Number	%	
American Wigeon	2	0.0	49	0.3	18	0.1	26	0.1	95	0.1	
Barrow's Goldeneye	45	0.1	3138	16.3	2870	17.0	76	0.2	6129	5.2	
Black Scoter	8	0.0	61	0.3	64	0.4	23	0.1	156	0.1	
Brant	16	0.0	0	0.0	0	0.0	0	0.0	16	0.0	
Bufflehead	2	0.0	434	2.3	522	3.1	0	0.0	958	0.8	
Canada Goose	997	2.1	177	0.9	368	2.2	824	2.4	2366	2.0	
Common Goldeneye	7	0.0	35	0.2	193	1.1	0	0.0	235	0.2	
Common Merganser	4150	8.6	47	0.2	89	0.5	3204	9.4	7490	6.3	
Gadwall	4	0.0	504	2.6	52	0.3	10	0.0	570	0.5	
Greater Scaup	0	0.0	0	0.0	0	0.0	2	0.0	2	0.0	
Green-winged Teal	3	0.0	27	0.1	10	0.1	78	0.2	118	0.1	
Harlequin Duck	1192	2.5	325	1.7	463	2.7	1645	4.8	3625	3.1	
Mallard	382	0.8	1623	8.4	1449	8.6	759	2.2	4213	3.6	
Northern Pintail	0	0.0	1	0.0	2	0.0	6	0.0	9	0.0	
Northern Shoveler	1	0.0	0	0.0	2	0.0	60	0.2	63	0.1	
Oldsquaw	11	0.0	221	1.1	24	0.1	13	0.0	269	0.2	
Red-breasted Merganser	22	0.0	158	0.8	63	0.4	65	0.2	308	0.3	
Scaup	0	0.0	19	0.1	81	0.5	169	0.5	269	0.2	
Surf Scoter	4414	9.1	1721	8.9	965	5.7	7175	21.0	14275	12.0	
Unidentified Duck	16	0.0	378	2.0	71	0.4	29	0.1	494	0.4	
Unidentified Goldeneye	39	0.1	980	5.1	1991	11.8	10	0.0	3020	2.5	
Unidentified Merganser	37	0.1	156	0.8	603	3.6	62	0.2	858	0.7	
Unidentified Scoter	8206	17.0	902	4.7	980	5.8	483	1.4	10571	8.9	
Unidentified Swan	2	0.0	0	0.0	0	0.0	0	0.0	2	0.0	
Unidentified Teal	3	0.0	0	0.0	0	0.0	0	0.0	3	0.0	
White-winged Scoter	4849	10.0	683	3.5	460	2.7	5829	17.1	11821	10.0	
Waterfowl Subtotal	24408	50.6	11639	60.4	11340	67.1	20548	60.3	67935	57.3	
Black Oystercatcher	187	0.4	74	0.4	25	0.1	214	0.6	500	0.4	
Black Turnstone	0	0.0	46	0.2	140	0.8	0	0.0	186	0.2	
Surfbird	0	0.0	0	0.0	25	0.1	0	0.0	25	0.0	
Red-necked Phalarope	1	0.0	0	0.0	0	0.0	1	0.0	2	0.0	
Spotted Sandpiper	4	0.0	0	0.0	0	0.0	0	0.0	4	0.0	
Unidentified Phalarope	5	0.0	0	0.0	0	0.0	0	0.0	5	0.0	
Unidentified Shorebird	3	0.0	529	2.7	148	0.9	4	0.0	684	0.6	
Shorebirds Subtotal	200	0.4	649	3.2	338	1.8	219	0.7	1406	1.2	
Bald Eagle	208	0.4	65	0.3	131	0.8	137	0.4	541	0.5	
Northwestern Crow	563	1.4	1391	7.2	1123	6.6	753	2.2	3830	3.2	
Other Birds	47	0.0 0	32	0.2	0 68	0.4	0 63	0.2 0	210	0.2	
Total Number of Birds	48250		19279		16898		34093		118520		

Table 9, Continued.
Table 10. Densities of common bird species from marine surveys in Glacier Bay National Park during June 1999, November 1999, March 2000 and June 2000. Densities are calculated for all transects and averaged. Note that summer surveys (June) included more transects within Glacier Bay and transects in Icy Strait (see methods).

SPECIES	Jun-99	Nov-99	Mar-00	Jun-00
Arctic Tern	1.30	0.00	0.00	0.54
Black-Legged Kittiwake	14.23	14.37	4.29	3.09
Brachyramphus Murrelet	5.74	1.05	3.13	10.03
Glaucous-winged Gull	5.59	11.05	5.38	4.22
Kittlitz's Murrelet	1.40	0.06	1.94	1.75
Barrow's Goldeneye	0.04	27.34	26.19	0.13
Marbled Murrelet	9.97	0.80	1.95	4.20
Mew Gull	1.76	5.49	0.89	2.63
Pelagic Cormorant	1.93	1.63	0.96	0.47
Pigeon Guillemot	4.91	4.32	6.10	4.04
Canada Goose	1.32	1.17	1.72	1.29
Common Merganser	7.22	0.84	0.54	6.62
Harlequin Duck	2.93	3.64	4.57	4.33
Mallard	0.64	14.24	13.31	1.29
Surf Scoter	8.80	11.32	9.64	9.15
White-winged Scoter	15.27	6.84	5.81	9.31

Table 11. Numbers and densities of all marine mammal species from marine surveys in Glacier Bay National Park during June 1999, November 1999, March 2000 and June 2000. Numbers represent totals sighted observed on all transects, densities are averages of all transects. Note that summer surveys (June) included more transects within Glacier Bay and transects in Icy Strait (see methods).

	Jun-	99	Nov-	.99	Mar	-00	Jun-00	
SPECIES	Num. Density		Num. D	ensity	Num. D	ensity	Num.	Density
Dall's Porpoise	22	0.04	0	0.00	0	0.00	12	0.03
Harbor Porpoise	67	0.22	26	0.24	57	0.69	59	0.13
Humpback Whale	13	0.06	2	0.01	1	0.02	25	0.12
Minke Whale	0	0.00	0	0.00	1	0.01	0	0.00
Sea Otter	149	0.55	137	1.21	76	0.65	167	0.55
Harbor Seal	255	0.57	161	0.85	67	0.31	331	0.78
Steller Sea Lion	14	0.03	46	0.45	76	0.68	56	0.15

Variability in Habitat Use

Comparisons between coastal and offshore transects provide useful information on the spatial distribution of species in Glacier Bay. The majority of marine bird species disproportionately used coastal areas compared to offshore areas (Figure 73, Table 12). Overall, >80% of all birds were found on coastal transects. There was also general consistency between the surveys, with species favoring either coastal or offshore in all surveys. Marbled murrelets were one of the few species whose distribution was not consistent between surveys (Table 12). This inconsistency was not related to season and may not be biologically significant. Summary densities of marine birds suggested that only Kittlitz's murrelets and unidentified (*Brachyramphus spp.*) murrelets were found predominantly in offshore transects (Table 12). However, examination of GIS maps (next section), revealed that gadwalls were in fact primarily sighted in coastal areas.



Figure 73. Distribution of seabird on coastal versus offshore transects.

Like marine birds, marine mammals were sighted more frequently on coastal as opposed to offshore transects (Table 12). The only marine mammals that were sighted more frequently on offshore transects were harbor porpoise. However, the limited numbers of marine mammals seen make generalizations tentative (when numbers sighted are low the differences made by sighting even a few animals more or less can have a disproportionate effect). There may also be a bias in the density calculations for Steller sea lion and harbor seal on and around their haul-outs surveyed in conjunction with coastal transects.

	Jun	-99	No	v-99	Ma	r-00	Jur	n-00	Ra	ıtio
SPECIES	coastal	offshore								
Arctic Tern	1.75	0.76	0.00	0.00	0.00	0.00	0.90	0.09	0.76	0.24
Barrow's Goldeneye	0.07	0.00	34.83	10.48	34.89	6.77	0.24	0.00	0.80	0.20
Black-Legged Kittiwake	20.48	6.72	7.16	30.59	5.12	2.44	4.47	1.42	0.47	0.53
Black Scoter	0.03	0.00	1.13	0.54	0.67	0.00	0.04	0.06	0.76	0.24
Brachyramphus Murrelet	4.70	6.99	0.87	1.44	2.35	4.85	4.45	16.71	0.29	0.71
Glaucous-winged Gull	7.85	2.87	15.43	1.18	6.81	2.18	6.58	1.39	0.83	0.17
Herring Gull	0.56	0.14	0.37	0.05	0.33	0.09	0.45	0.20	0.78	0.22
Kittlitz's Murrelet	0.83	2.07	0.09	0.00	1.44	3.06	0.85	2.83	0.29	0.71
Marbled Murrelet	12.01	7.53	0.50	1.46	1.72	2.44	3.04	5.59	0.50	0.50
Mew Gull	2.99	0.29	6.91	2.30	1.15	0.30	4.06	0.91	0.80	0.20
Pelagic Cormorant	3.26	0.33	1.68	1.52	1.31	0.17	0.70	0.19	0.76	0.24
Pigeon Guillemot	7.65	1.61	5.61	1.42	7.68	2.59	6.33	1.29	0.80	0.20
Bufflehead	0.01	0.00	5.18	0.33	6.17	0.00	0.00	0.00	0.97	0.03
Canada Goose	2.37	0.07	1.69	0.00	2.49	0.02	2.36	0.00	0.99	0.01
Common Goldeneye	0.01	0.00	0.54	0.00	1.72	0.37	0.00	0.00	0.86	0.14
Common Merganser	12.87	0.42	1.20	0.02	0.77	0.01	11.46	0.83	0.95	0.05
Harlequin Duck	5.11	0.30	4.83	0.97	6.46	0.34	7.35	0.70	0.91	0.09
Long-tailed Duck	0.03	0.03	2.03	0.07	0.18	0.00	0.09	0.00	0.96	0.04
Mallard	1.10	0.08	15.94	10.41	16.94	5.22	2.37	0.00	0.70	0.30
Surf Scoter	15.43	0.83	15.71	1.45	12.71	2.79	13.58	3.84	0.87	0.13
White-winged Scoter	26.63	1.61	8.91	2.17	8.37	0.09	11.95	6.15	0.85	0.15
Golden Eagle	0.03	0.00	0.00	0.00	0.00	0.00	0.00	0.00	1.00	0.00
Harbor Porpoise	0.25	0.18	0.17	0.40	0.54	1.00	0.13	0.14	0.39	0.61
Harbor Seal	0.94	0.13	1.22	0.02	0.45	0.01	1.29	0.17	0.92	0.08
Humpback Whale	0.08	0.03	0.02	0.00	0.02	0.00	0.08	0.17	0.50	0.50
Sea Otter	0.84	0.20	1.72	0.07	0.48	1.03	0.91	0.13	0.73	0.27
Steller Sea Lion	0.03	0.03	0.53	0.27	0.92	0.14	0.10	0.20	0.71	0.29

Table 12. Densities of common seabirds and marine mammals sighted on coastal and offshore surveys in and around Glacier Bay National Park June 1999 – June 2000.

The comparison of species densities on coastal and offshore transects suggests an underlying habitat association. The classification of coastal and offshore can be viewed as a proxy for bottom depth. We examined associations between bottom depth and species occurrence by looking at cumulative occurrence through 5 depth classes. We found all common species tended to use relatively shallow habitats (Figure 74). For example, >70% of all common mergansers were found in water <25m in depth, and almost all were found at <100m depth. Similarly, >50% of all birds were located in depths <50 m, and >75% of all birds were in water <100 m deep. Some species clearly favored shallow, nearshore waters (e.g., mergansers, guillemots), while others, such as murrelets (spp.) and kittiwakes, routinely foraged in deeper waters.

Marine mammals like marine birds showed distinct differences in use of habitats. A plot of cumulative occurrence by depth class indicated that harbor seals forage in extremely shallow areas whereas Dall's porpoise, an oceanic species, distinctly favored deepest waters to be found in the study area (Figure 75).



Figure 74. Distribution of common seabirds in Glacier Bay with respect to ocean bottom depth.



Figure 75. Distribution of common marine mammals in Glacier Bay with respect to ocean bottom depth.

Distribution Maps

Maps of species distributions were developed using a GIS, allowing visual comparisons between surveys, seasons, and species. While tabulated comparisons of transects categorized as "coastal" or "offshore" can assist our understanding of survey data, categories are not always exclusive and the process of categorizing tends to lose information. We mapped many of the more common species of seabirds and waterfowl, as well as a number of the marine mammals. Although only a subset of species is presented, the GIS database can be used to access any of the remaining species in the future.

Seabirds

Black-legged kittiewakes and glaucous-winged gulls were distributed throughout the Glacier Bay area (Figure 76 and Figure 77). Although sighted most commonly on coastal transects, neither species was restricted to coastal areas. Both species seasonally use Glacier Bay for nesting and reproduction. A decrease in the survey counts of June 2000 compared to the June 1999 survey was noted for both species.

Maps of marbled murrelets illustrate that they were found throughout Glacier Bay (Figure 78). Marbled murrelets were evenly distributed between coastal and offshore areas. As with the gulls, marbled murrelets were more common in Glacier Bay in June 1999 then they were in June of 2000.

Kittlitz's murrelet distribution was more clustered than that of marbled murrelets (Figure 79). There was also a difference in how these species used marine habitats. While marble murrelets were seen equally on both coastal and offshore transects, Kittlitz's murrelets were more commonly seen offshore. Although foraging distances for Kittlitz's murrelets are not well known, these patterns could also reflect the interaction between foraging and nesting habitat. If foraging distances are limited then the distribution of terrestrial nesting habitat may limit marine foraging habitat. Like marbled murrelets, Kittlitz's murrelets were clearly seasonal residents, all but disappearing during the winter months. Surprisingly, a number of Kittlitz's murrelets were sighted in Wachusett Inlet during the March 2000 survey, though they were not seen in that Inlet in large numbers during either of the summer surveys.

Pigeon Guillemots were evenly distributed throughout the coastal areas of Glacier Bay in the summer months; but few were sighted in Icy Strait. November 1999 and March 2000 survey maps suggest that pigeon guillemots use the more sheltered Beardslee Islands for over-wintering habitat (Figure 80).



Figure 76. Black-legged kittiwake sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 77. Glaucous-winged gull sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 78. Marbled murrelet sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 79. Kittlitz's murrelet sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 80. Pigeon guillemot sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.

Waterfowl

Barrow's goldeneye exhibited strong seasonal patterns of occurrence in Glacier Bay (Figure 81). Although they were one of the most common birds in the fall and spring surveys, they all but disappeared in the summer months. In terms of distribution, Barrow's goldeneye were sighted throughout the coastal areas of Glacier Bay in the fall and spring. When present, local densities were relatively high.

Common mergansers were also seasonal residents in Glacier Bay. In contrast, Barrow's goldeneye, they were seen almost exclusively in the summer months (Figure 82). Sightings of common mergansers were primarily on coastal transects. Although mergansers were seen throughout Glacier Bay, they were most numerous in the Beardslee Islands, Berg Bay, and Adams Inlet.

Mallards were seen in all surveys, though numbers clearly increased in the fall and spring surveys (Fig 83). As with other waterfowl, mallards were found only in coastal areas. In the summer months the mallards that remained in Glacier Bay were located primarily in small bays and inlets. In the fall and spring, mallard numbers rose dramatically and they expanded their range to include the Beardslee Islands, Berg Inlet, and Fingers Bay.

Harlequin ducks, though not as numerous as some other waterfowl, were found throughout Glacier Bay during the summer surveys (Fig 84). Harlequin duck use of Glacier Bay during the fall and spring appeared to shift to the south. On all survey, their distribution was coastal.

White–winged scoters were most common in the summer months, when sightings of large flocks were common (Fig 85). Scoters were sighted most frequently in the northern portion of the bay, with large numbers sighted in both Muir Inlet and the West Arm. Although most common in the summer, white-winged scoters were found in Glacier Bay year-round as evidenced from the November and March surveys.



Figure 81. Barrow's goldeneye sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 82. Common merganser sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 83. Mallard sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 84. Harlequin duck sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 85. White-winged scoter sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.

Marine Mammals

As with marine birds, marine mammal distributions varied spatially and temporally. Sea otters were seen in all seasons with no noticeable change in distribution throughout the surveys. Sea otters, once extirpated from Glacier Bay, have expanded their range to the northernmost extent of the Beardslee Islands (Figure 86; also see Bodkin et al. 2001). Concentrations of sea otters were greatest along the coast of the Sitakaday Narrows and the Beardslee Islands, with much smaller numbers located in Icy Strait.

Both harbor and Dall's porpoise were seen on surveys, however, they were segregated in their geographic distribution. Harbor porpoise were found throughout the pelagic areas of Glacier Bay (Figure 87). In contrast Dall's porpoise were found almost exclusively in the waters of Icy Strait outside of Glacier Bay. Due to a combination of low numbers and a lack of sampling in Icy Strait on the November and March surveys, little can be said about the seasonality of these two species.

Harbor seals were the most commonly sighted marine mammals. Maps illustrate the broad distribution of harbor seals within Glacier Bay proper (Figure 88). Their occurrence in the Bay in all surveys indicated year-round residency. Although they were seen throughout Glacier Bay, the Beardslee Islands were an area of particularly high harbor seal abundance in all seasons.

Humpback whales were seen throughout Glacier Bay on both June surveys (1999 and 2000). Despite their wide distribution, the majority of humpbacks on surveys were found in large concentrations at specific sites. During the 1999 survey, a large concentration of humpbacks was seen around Point Adolphus; in 2000 the largest concentration of humpbacks was seen at the mouth of Glacier Bay (Figure 89). These results were similar to patterns observed in the ongoing National Park Service whale monitoring study (Doherty and Gabriel 2002). Maps of the November 1999 and March 2000 surveys indicated that a small number of humpbacks either delayed their migration or possibly over-wintered in the Bay.

South Marble Island is the only Steller sea lion haulout in Glacier Bay, and our surveys did not pass close to it. Thus all sea lions sightings were of animals in the water. Steller sea lions were sighted on all surveys, though there appeared to be a shift to the north during the fall and spring surveys and to the south during the June surveys (Figure 90). We also noted that in the fall and spring surveys sea lions were seen in larger groups. The June 1999 survey had the fewest sightings, with no sightings in either arm.



Figure 86. Sea otter sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 87. Dall's and harbor porpoise sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 88. Harbor seal sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 89. Humpback whale sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.



Figure 90. Steller sea lion sightings in and around Glacier Bay National Park during surveys conducted between June 1999 and June 2000. Blue lines indicate survey tracks. Surveys were conducted from the *RV Padalus, RV Alaska Gyre, MV David Grey* and *MV Lutras* by U.S. Geological Survey and National Park Service personnel.

Species Trends

To look for temporal trends in species abundance, we compared our results from the June 1999 and June 2000 surveys with the results of a 1991 survey of Glacier Bay (Piatt and Springer, unpubl. data). The distances and areas sampled for all three summer surveys are listed in Table 13. Since the 1991 survey was primarily coastal, for consistency, we only used the results of the coastal portions of all three surveys, and only those transects from within Glacier Bay itself for comparison.

	19	91	19	99	2000		
	transect	area	transect	area	transect	area	
Survey Area	distance	sampled	distance	sampled	distance	sampled	
Coastal	651	130	772	204	779	181	
Pelagic	68	14	399	120	399	91	
Combination*	8	2					
TOTAL	727	145	1172	323	1178	272	

Table 13. Transect distances and areas sampled for coastal and pelagic transects in Glacier Bay 1991, 1999 & 2000. Distances are given in km. and

* Transects that sampled a combination of both pelagic and coastal areas.

Most species increased in density between 1991 and 1999-2000 (Table 14). Gulls (blacklegged kittiwakes, glaucous-winged gulls, and mew gulls) all increased. Increases were also seen in arctic tern and pelagic cormorant. Pigeon guillemot densities were essentially unchanged. The only seabird species to show evidence of decline were Kittlitz's and marbled murrelets. Comparisons with the prior survey by Piatt and Springer (1991) indicated that both Kittlitz's and marbled murrelets are declining dramatically. This appears to fit the pattern of *Brachyramphus spp*. declines state-wide (van Vliet 1983, van Vliet and McAllister 1994, Piatt and Naslund 1995). Although neither species is currently listed as threatened or endangered in Alaska, marbled murrelets are listed by the U.S. Fish and Wildlife Service as threatened in Washington, Oregon, and California. Recognition of the declines in murrelets has led to the ongoing review of Kittlitz's murrelet status in Alaska.

The general trend in marine waterfowl was one of either increasing or stable densities with time; however, only Canada geese increased dramatically. Common merganser and harlequin duck densities indicated smaller increases, while scoters appeared to be essentially unchanged.

With one exception, marine mammals were markedly increased in abundance over the 8-9 year period between the 1991 and 1999-2000 surveys. Sea otters were not seen at all on the 1991 survey, while they were quite common in the lower portion of the Bay in 1999-2000. Humpback whales were another species that was considerably more abundant in 1999-2000 than in 1991. Unlike the other marine mammals, Harbor seals appeared to decline between 1991 and 1999-2000.

Table 14. Densities of common species sighted on surveys in June of 1991, 1999,
and 2000. Density calculations include only coastal transects. Densities shaded
green indicate an increase in density, while those shaded yellow indicate a decrease
in density between 1991 and 1999-2000.

	1991		19	99	2000		
Species	$\#/km^2$	95% CI	#/km ²	95% CI	#/km ²	95% CI	
Arctic Tern	0.33	(0.38)	2.14	(2.39)	1.05	(0.92)	
Black-legged Kittiwake	4.51	(8.31)	8.38	(4.11)	5.20	(3.77)	
Canada Goose	1.45	(1.14)	2.83	(2.97)	2.74	(2.76)	
Common Merganser	10.99	(7.33)	15.40	(6.77)	13.30	(9.73)	
Glaucous-winged Gull	1.83	(0.95)	8.09	(2.87)	7.61	(4.02)	
Harlequin Duck	4.32	(2.17)	6.06	(2.32)	8.53	(4.42)	
Kittlitz's Murrelet	5.04	(1.91)	1.01	(0.55)	0.99	(0.60)	
Marbled Murrelet	31.17	(7.76)	13.01	(3.69)	3.44	(1.28)	
Mew Gull	0.98	(1.03)	2.67	(1.43)	4.63	(2.35)	
Pelagic Cormorant	0.14	(0.11)	2.76	(2.53)	0.80	(0.71)	
Pigeon Guillemot	13.63	(8.88)	8.55	(2.03)	7.29	(1.93)	
Unidentified Murrelets	19.74	(6.92)	5.54	(1.80)	5.08	(1.38)	
White-winged Scoters	28.47	(25.43)	31.30	(22.21)	13.88	(11.24)	
Humpback Whale	0.01	(0.02)	0.09	(0.07)	0.10	(0.10)	
Sea Otter	0.00	NA	0.37	(0.39)	1.04	(1.51)	
Harbor Seal	3.86	(2.45)	0.89	(0.47)	1.48	(0.93)	

Increasing No Trend Decreasing

Survey Methods

Natural resource managers seek out efficient sampling schemes to maximize their ability to monitor populations. We tested several less intensive sampling regimes against the full survey to see if they could provide accurate information about seabird and marine mammal populations. We compared 3 different sub-sets of these the June 1999 and June 2000 transects with the full surveys for each year (Figure 91). The first subset was a based on the November-March survey transects, accounting for approximately 25% of the complete set of transects. Using this set of transects would simplify surveys for all seasons with a single set of transect maps and allow direct comparisons across seasons. However, densities for the two summer surveys when compared to the densities for the reduced number of transects indicated that the reduced set of transects poorly reflected the actual densities of most species (Table 15). We also found a lack of agreement between the two years of reduced surveys. The densities of some species were overestimated in one year and underestimated in the other. Besides providing poor estimates of species densities, the lack of consistency between years further indicates that these estimates are not robust indices of density.

The second subset of transects included 35% of the full survey within Glacier Bay. This set of transects sampled the pelagic zone in approximately 5 km intervals as well as sampling approximately half the coastal area. This set of transects also failed to provide density estimates that matched those of the full set of survey transects (Table 15). As with the 25% set of transects, there was a lack of agreement in the direction of error between the two years of reduced transects, again making density estimates questionable for use as indices.

The final subset of transects included all coastal transects within Glacier Bay. These coastal transects (n=63) accounted for 63% of all transect volume sampled. Of the three subsets the coastline transects showed the greatest consistency between years, however the densities of most species were inflated (Table 15). Murrelet densities were either underestimated (Kittlitz's and all Brachyramphus), or inconsistent (marbled). Humpback whale densities were also inconsistent in comparison with the results of the complete survey.



Figure 91. Various survey designs within Glacier Bay. Subsets include 25% (27 transects), 35% (48 transects), 63% (coastal transects, n=63) of the total volume of area surveyed within Glacier Bay during June of 1999 and 2000. All transects within Glacier Bay (n=112) are displayed for comparison.

Table 15. Comparisons of species densities for the full survey and 3 reduced sets of transects. Densities are only from transects
within Glacier Bay. Percentages reflect the densities from reduced transects/all transects. Percentages shaded green are species that
were sighted more commonly then expected, while those shaded yellow were species sighted less commonly then expected. The
reduced surveys covered approximately 25%, 35%, and 46% of the full survey respectively.

	Full Survey 27			27 Tra	ansects			48 Tra	ansects			Coastal Survey			
_	Densit	ies	Densit	ies	% Of Tota	% Of Total Survey		ties	% Of Total Survey		Densi	ties	% Of Tota	Survey	
	1999	2000	1999	2000	1999	2000	1999	2000	1999	2000	1999	2000	1999	2000	
Arctic Tern	1.57	0.65	0.52	0.82	33.12	127.25	0.64	0.47	40.76	72.30	2.01	1.05	128.03	162.06	
Black-legged Kittiwake	16.43	3.41	12.29	3.22	74.80	94.55	6.38	5.01	38.83	148.08	23.10	5.20	140.60	152.49	
Brachyramphus Murrelet	6.68	9.62	7.27	9.38	108.83	97.52	4.35	13.47	65.12	141.27	5.32	5.08	79.64	52.81	
Canada Goose	1.60	1.58	3.96	3.14	247.50	198.66	0.36	0.18	22.50	11.27	2.72	2.74	170.00	173.42	
Common Merganser	8.67	8.06	16.73	16.09	192.96	199.65	5.91	9.33	68.17	116.77	14.78	13.30	170.47	165.01	
Glaucous Gull	6.41	4.94	5.43	6.46	84.71	130.80	4.16	8.28	64.90	169.16	8.68	7.61	135.41	154.05	
Harlequin Duck	3.53	5.29	5.56	8.83	157.51	166.88	2.53	4.93	71.67	94.19	5.87	8.53	166.29	161.25	
Harbor Seal	0.69	0.94	1.05	1.48	152.17	156.97	0.12	0.57	17.39	60.70	1.07	1.48	155.07	157.45	
Humpback Whale	0.07	0.11	0.02	0.13	28.57	122.49	0.01	0.11	11.15	100.79	0.09	0.10	128.57	90.83	
Kittlitz's Murrelet	1.68	2.13	1.61	1.89	95.83	<mark>88.80</mark>	1.54	2.32	91.67	110.27	0.95	0.99	56.55	46.48	
Mallard	0.77	1.58	1.41	1.38	183.12	87.19	0.23	1.19	29.87	75.86	1.27	2.75	164.94	174.05	
Marbled Murrelet	11.20	4.58	11.08	3.88	98.93	84.62	9.23	5.52	82.41	121.56	13.09	3.44	116.88	75.11	
Pigeon Guillemot	4.91	5.86	6.91	6.76	140.73	115.32	4.43	4.07	90.22	84.07	8.74	7.29	178.00	124.40	
Steller Sea Lion	0.04	0.18	0.02	0.20	50.00	111.11	0.01	0.26	25.00	47.51	0.04	0.11	100.00	61.11	
Sea Otter	0.63	0.64	0.13	0.07	20.63	10.94	0.08	0.30	12.70	149.07	0.94	1.04	149.21	162.50	
Surf Scoters	10.60	11.19	21.07	4.95	198.77	44.27	10.85	8.05	102.36	72.68	17.72	15.77	167.17	140.93	
White-winged Scoters	18.40	11.37	18.19	9.08	98.86	79.87	28.96	13.65	157.39	121.17	30.61	13.88	166.36	122.08	

Conclusions

Glacier Bay Ecosytem

Plankton and forage fish

In early spring, onshore movement of zooplankton contributes to the abundance of shelf zooplankton communities throughout the Gulf of Alaska. Cooney (1987) estimated that on the order of 10 million metric tons of zooplankton biomass are moved shoreward each year in the Gulf of Alaska. This input of biomass at the secondary trophic level probably contributes to the productivity of the bay.

Despite similarities in the zooplankton abundance and community structure between Glacier Bay and other Gulf of Alaska estuaries, seasonal patterns of production are strikingly different. In the Gulf of Alaska, zooplankton abundance peaks in late May or June, and steadily declines through the rest of the season (Cooney, 1987). However, persistent production in the West and East Arms of Glacier Bay leads to sustained seasonal zooplankton abundance within the upper-bay. Mixing fronts are often highly productive (Cooney, 1987); and the heads of both Arms are no exception. The glacially influenced oceanography provides a possible mechanism of water movement that allows plankters, largely copepods advected into the bay by Ekman transport, to congregate and remain at these fronts. The highest zooplankton abundances in this study were in Tarr Inlet – some of the coldest and most turbid waters in Glacier Bay.

At the metapopulation scale, exchange of individuals between Glacier Bay and Gulf of Alaska populations is largely the result of migration by larvae in the plankton. Local production of larvae in Glacier Bay is likely to contribute to plankton communities outside of the bay. Environmental cues regulate timing of larval release, and correlations between oceanography, release, and abundance of larvae could provide insights into population dynamics. The potential shift in timing from 1963 to 2000 may be related to changes in oceanography as glaciers within the bay continue to recede. These changes may, in turn, have a wider impact on surrounding marine areas.

Icthyoplankton surveys indicated that Glacier Bay provides nursery habitat for large numbers of juvenile pollock, capelin, and slender eelblenny. Abundances were much higher in June than in May, which is similar to results for other Alaskan fjord systems (e.g., Ressurection Bay; Müter and Norcross, 1994). Juvenile herring were conspicuous in their absence from our summer surveys, despite representing a large part of our fall littoral sampling. Juvenile herring appear to frequently avoid our nets, maybe due to their patchy distribution or use of habitat between areas sampled by our beach seines (littoral zone) and offshore pelagic trawls.

Catches of larval fish in the upper reaches of Glacier Bay were markedly reduced compared to the immediately adjacent down-bay station during June, the peak for larval abundance (for example pollock (Table16) or capelin (Table 17) at stations 21 and 10, respectively). This result suggests that a limiting physical condition exists in the upper

bays (e.g., Tarr Inlet) that restricts the presence of juvenile fish. Conversely, euphausiids were markedly more abundant at the very heads of the bay (Table 18).

Nearshore communities of fish in Glacier Bay appeared typical of those found in northern regions. Tidal, seasonal, and spatial processes all imparted strong effects on both abundance and composition of fish in the bay. Overall, the community was defined by juvenile fish that transitioned from outmigrating salmonids in the spring, to recruiting capelin, sand lance, and herring in the fall. Despite the wide coverage of our nearshore fish sampling, most fish were caught at relatively few sites – particularly those in close proximity to tidewater glaciers and those neighboring salmonid spawning streams.

Table 16. Numbers per 1000 m^3 for walleye pollock larvae in Glacier Bay Alaska. (May, June = Tucker Trawl), (July, August = Isaacs-Kidd). Zeros left out for clarity (i.e., all stations and months were fished).

	Low	er Bay	Midd	le Bay	V	Vest Arn	1			East Arm	
	00	02	04	06	08	10	21	14	16	18	20
May				3.9							
June		3.8	49.9	187.5	158.2	491.3		68.3	86.1		
July		0.1			4.1	1.0	3.6			7.9	7.9
August					0.1	1.0	35.3	0.4	0.8	2.4	4.2

Table 17. Numbers per 1000 m^3 for capelin larvae in Glacier Bay Alaska. (May, June = Tucker Trawl), (July, August = Isaacs-Kidd). Zeros left out for clarity (i.e., all stations and months were fished).

	Lowe	r Bay	Middle	e Bay	I	West Arn	n			East Arm		
	00	02	04	06	08	10	21	14	16	18	20	
May												
June		4.4	15.6		10.9	244.9			8.4			
July	1.9	3.6		2.4	0.1	0.5	24.0	0.1		8.4	6.0	
August	0.9	3.5	0.4	0.8	0.8	0.4	26.5	7.6	8.7	4.9	0.6	

Table 18. Numbers per 1000 m^3 for euphausiids in Glacier Bay Alaska. (May, June = Tucker Trawl), (July, August = Isaacs-Kidd). Zeros left out for clarity (i.e., all stations and months were fished).

	Low	er Bay	Middl	e Bay		West Ar	m	Ea			st Arm
	00	02	04	06	08	10	21	14	16	18	20
May		14.2	441.4	47.2	128.7	134.6	218.3	9.4	37.5		107.4
June	22.7		219.7	108.3		141.6	2441.5		7.5	51.9	233.2
July		11.2	67.5	59.6	4.9	66.2	622.3	67.2	70.0	941.2	268.8
August		1.1			47.9	58.4	706.2	116.1	49.0	101.6	79.7

Mid-water sampling of pelagic fish species indicated some clear patterns within Glacier Bay. First, marked spatial differences existed around the bay. For the most part, the pelagic community was dominated by juvenile walleye pollock and capelin. However, capelin dominated in Muir Inlet while walleye pollock dominated in catches in the west arm, middle bay, lower bay, and Icy Strait.

Seasonal patterns of abundance of juvenile and adult fish in Glacier Bay were also interesting. We can compare our results with similar work conducted in Lower Cook Inlet (Piatt, 2002). Most striking was the extremely high relative abundance of juveniles in Glacier Bay (particularly the upper arms) compared to Cook Inlet. The logical explanation for this pattern is that the upper reaches of Glacier Bay are important nursery areas for juvenile fish. Seasonal sampling indicated that the juveniles were present throughout the season and grew at similar rates to those published for similar species in Prince William Sound (Müter and Norcross, 1994). Abundance of these juveniles suggest beneficial conditions in Glacier Bay for early growth, despite cold and turbid waters. The few adult fish caught in the upper arms were eating juvenile fish which may represent a source of bottom-up nutrients to the Glacier Bay ecosystem.

Seasonal and spatial patterns of abundance for both pelagic zooplankton and icthyoplankton in Glacier Bay indicate a marked seasonal and spatial pattern to the abundance of these secondary and tertiary levels of productivity. Zooplankton abundance was highest in the upper arms and at the mid-bay station 4. Analysis of oceanographic data by Hooge and Hooge (2001) paralleled these results with elevated levels of primary productivity in the upper arms and mid-bay. Clearly, processes defining the productivity in the upper arms as opposed to the mid-bay are different. Perhaps the most interesting question is why are there high abundances of small fish in close proximity to Margerie Glacier but not in the mid-bay?

One of the prominent findings of our research is the relative importance of Glacier Bay as a nursery area for pelagic fish. The role of inshore habitat as an important nursery area for juveniles of many marine fish species is well known (e.g., Poxton *et al.*, 1983; Orsi and Landingham, 1985; Bennett, 1989; Blaber *et al.*, 1995; Dalley and Anderson, 1997). However, our results suggest that pelagic fish in Glacier Bay are also generally smaller than those in Cook Inlet, and more juveniles are present (Piatt, 2002). Several key forage species (e.g., capelin, sand lance, and juvenile walleye pollock and herring) probably spawn within the bay.

Hooge and Hooge (2001) indicated that outside Glacier Bay, no other fjord system in Alaska has been observed with the sustained seasonal productivity. They gave two possible reasons for these observations in Glacier Bay: 1) lack of zooplankton predators; or 2) nutrient enhancement. The high zooplankton densities observed in the upper-bays during this study suggest that the former reason is unlikely. However, this poses as many questions as it answers; establishing sources and flows of biomass and energy will be a demanding question requiring further research.

Forage fish predators

Predator surveys provided us with extensive information about species occurrence and distribution in and around Glacier Bay. Although the Bay had been surveyed in the past, our efforts were the first comprehensive surveys covering the greater Glacier Bay ecosystem and will serve as a baseline for future inventories.

The heterogeneous marine environment provides a basis for explanation of the diverse and productive assemblage of species found in the Bay. Glacier Bay provides foraging, nesting, and wintering habitat for a wide variety of species (n=71 sighted in surveys). Although overall marine bird densities were somewhat less then that recorded in some of the most productive areas in the northern Gulf of Alaska, such as the Barren Islands (about 100 bird/km², Piatt 1994), the summer densities were nonetheless quite high (about 80 birds/km²). Fall and spring surveys had not been carried out previously to our surveys. Seasonal trends in abundance and diversity of seabird assemblages indicated that they dominated the summer surveys, whereas waterfowl were dominant in the fall and spring.

Summer surveys indicated that while marine birds and mammals use all portions of the bay, several areas were focal points for many species. These areas included the Beardslee Islands, Sitakaday Narrows, Hugh Miller Inlet, Berg Bay, and Adams Inlet. An integration of colony site, hydroacoustics, and fishing data may provide additional information on whether these sites are primarily associated with proximity to breeding areas, food resources, or a combination of the two. The decreased sampling area during the Fall and spring surveys limited our ability to comprehensively identify marine bird and mammal distributions during this period. However, we were able to confirm that areas that were high-use during the June surveys (Beardslee Islands, Sitakaday Narrows, Hugh Miller Inlet, Berg Bay, and Adams Inlet) were also high-use areas during the fall and spring surveys. However, a primary difference was the shift from use by seabirds in the summer to use by waterfowl in the winter. This use during the colder months suggested that these sites might also provide some measure of shelter to marine birds.

Distribution of marine birds in Glacier Bay was primarily coastal. Only Kittlitz's murrelets were more common on the offshore transects. This distribution likely reflects the coastal nature of foraging resources. Surveys also illustrated the seasonal attendance by most marine bird species. Summer residents, including black- legged kittiwake, common mergansers, murrelets and scoters use the bay as reproductive habitat. The vast majority of these species migrate from Glacier Bay in the fall; whereas other species including Barrow's goldeneye, mallard and northwestern crow overwinter in the Bay. Generally, the two summer surveys showed a consistent pattern of habitat use for most species. Of the more abundant species, only Kittlitz and marbled murrelets were not present in the same portions of the Bay in 2000 that they were in 1999. The explanation for this lack of similarity in distribution may be related to foraging resources or differing levels of disturbance by vessels; however, shifts in distribution between the 2 surveys were not recorded for other species.

Harbor porpoise were only found in the Bay itself while Dall's porpoise were only seen in Icy Strait. Humpback whale distributions were clumped near Point Adolphus in June of 1999 and Point Carolus in June of 2000. This localized abundance is attributed to their use of known foraging areas. The changes in whale distribution between June 1999 and June 2000 may suggest changes in the distribution of forage species during the period of the surveys. Humpbacks migrate to Hawaii during winter; therefore, no sightings of these whales were made in the November and March surveys. Harbor seals were the most common marine mammals in all surveys and found in all parts of Glacier Bay during all surveys. Steller sea lions were sighted on all surveys indicating year-round residency in the Bay. We did note that an apparent shift in their distribution, to the north during the fall and spring surveys (possibly in association with the ice-edge) and to the south during the June surveys. One question that we could not address was whether these larger groups lead to greater probability of detection.

Comparison of our surveys to those conducted in 1991 indicated that Glacier Bay is a system undergoing change. For predators, the trend was toward increasing densities of marine birds and mammals. Perhaps that is to be expected given the increased protection given to some species, e.g. humpback whales, but suggests, in general, an increase in fish abundance within the bay. There may also be some connection with the continuing changes in the physical environment of Glacier Bay. For example, continued recession of glaciers and changes in the terrestrial environment that either directly, thorough terrestrial habitat quality (e.g., nesting habitat) or indirectly through the effects of these changes on inputs to the marine system (e.g., runoff from rapidly developing riparian areas).

Of the marine birds, densities of black-legged kittiwakes and all gull species increased between 1991 and the 1999-2000 surveys. In fact densities of most marine birds were higher in the 1999-2000 surveys than recorded previously in 1991 (Piatt and Springer unpubl. data). Among marine mammals, humpback whale and sea otters, numbers increased dramatically between 1991 and our 1999-2000 surveys. These increases reflected the generally acknowledged increases in populations of both (previously harvested) species throughout their range.

A few species declined between 1991 and 1999-2000. Average declines were on the order of 60% for Kittlitz's murrelets and 75% for marbled murrelets. We cannot discount the possibility that the distribution of murrelets was more offshore in 1999-2000 than in 1991. Even so, the scale of the declines are so great that we are confident that they reflect a significant changes in these populations. Given that Glacier Bay represents the primary concentration of Kittlitz's murrelets in Southeast Alaska (Kendall and Agler 1998), population declines in this area are of particular concern. Although declines in both marbled and Kittlitz's murrelets appear to be a northern Gulf of Alaska phenomena, the dramatic declines in an area thought to be a core for their populations was not anticipated.

Our surveys indicated that Harbor seals had also declined. While this is not a new finding, it corroborates the findings of Mathews and Pendleton (2000). This earlier study

only used counts at specific haulout sites, leaving some doubt as to whether seal populations were actually declining because seals were spending more time foraging (i.e, in the water), or because they had shifted to using unmonitored haulout sites. Agreement of our pelagic surveys with those of Mathews and Pendleton (2000) suggest that the lower number of seals at haulouts (resting) is not a result of seals spending more time in the water (foraging); and therefore likely represents real declines for this species.

Our efforts to identify a lower intensity-sampling regime for monitoring predators were only marginally successful. The first two sets of reduced transects (25% and 35%) did a poor job of sampling the true densities of many species. That might not be a fatal flaw if only an index was required; but the variability of this index between years also appeared to be high. In large part, this appears to be a product of variable species distributions and the low abundance of some species. When we looked at using all coastal transects (63%) we found that while most species densities were overestimated, they were consistent for all species. This is not surprising given that the majority of species were sighted predominantly on coastal transects. Unfortunately, we found that indices for two species of concern to the park, humpback whales and marbled murrelets, were inconsistent. Other species such as Kittlitz's and unidentified *Brachyramphus* murrelets were underestimated in the coastal surveys as their densities declined, leading to greater variance in results. Some combination of coastal surveys and species-specific surveys might be sufficient to develop population indices and we will continue to examine this issue.

Critical Habitat Areas

Fjords have traditionally provided fish in good quantity to the local populations in Norway, Scotland, British Columbia, and Greenland. This is perhaps not surprising based on the known propensity for fjord systems to be very productive (Matthews and Heimdal, 1980). Our results over the two years of study have highlighted distinct areas within Glacier Bay -- Whidbey Passage, upper Inlets, mouths of Inlets, and outflows from salmon streams -- that are highly productive. Furthermore, our results strongly suggest spawning of key forage species (walleye pollock, capelin, slender eelblenny, and sand lance) within the bay. These areas of concentration and spawning are likely to be vital to the function of the Glacier Bay system. Current research into the value of Glacier Bay as a Marine Reserve (USGS-BRD – Spencer Taggart) should elucidate the relative importance of these spawning and nursery areas.

Glacier Bay's local oceanographic patterns result in a patchy distribution of optimal marine pelagic fish habitat. In contrast, anadromous species usage of habitat is more easily defined. Salmon spawning streams are clearly important areas both to the terrestrial system as well as the nearshore marine system.

Recommendations

State of Current Knowledge

Hale and Wright (1979) summarized the Glacier Bay ecosystem as it was known at the time as:

"Glacier Bay is not a homogeneous body of water, either in time or space. Successional processes following the most recent rapid deglaciation have resulted in the development of a gradient of biological communities of the lower bay where the ice has been gone the longest to the less diverse communities found in the upper bay. These biological differences are teamed with physical differences such as greater water clarity and more constant salinity in the lower bay. The result is a gradual transition from a complex to a more simple ecosystem structure as one moves up the bay."

Since that time, our knowledge has grown. In this report, we have highlighted some anomalies to Hale and Wright's basic ideas, most notable of which is the extraordinary level of biomass found in close proximity to tidewater glaciers. The 3-dimensionsal spatial extent of this biomass, the driving forces (e.g., source of carbon), and the downstream dispersion of organisms and nutrients are important questions to resolve, both scientifically as well as from a resource management perspective. In conjunction with this question, it is hard to conceptually separate the locations of the black-legged kittiwake colonies at Margerie and Riggs glaciers from locally elevated productivity. However, at least the Margerie colony has suffered poor reproductive success over the last decade (Elizabeth Hooge, *Pers. Comm.*). What conditions may have favored kittiwake foraging close to their colony as opposed to the southern end of the bay (if this was the case) is a compelling question that could dovetail with an analysis of the Tarr Inlet ecosystem.

We have provided a detailed description of some seasonal and spatial processes within the bay from production, small schooling fish, and predator perspective. We are now able to point to 'productive' areas of the bay and those areas that are routinely used (important) to predators. However, the goal is to tie predator locations to food. We would like to see work focused on several key predators and their prey. Humpback whales and Kittlitz's murrelets would be obvious choices due to their listed status.

Recommendations for Further Research

Euphausiids are an important food resource for whales and fish. Documenting the distribution of adults, zoea, meganauplii, and nauplii will improve the understanding of euphausiid ecology in the bay.

Describing larval development of benthic animals that are presently undescribed would be valuable, and with the incorporation of source-sink philosophy in marine resource management would provide basic data for decisions about marine reserves.

Conducting a survey targeting crab veligers in coordination with USGS-BRD Jim Taggart's Marine Reserve program on adult movement patterns would be of wide interest, and useful for identifying the spatial range of crab dispersal.

Continued zooplankton surveys using a reduced number of stations and discrete depth sampling, with the additional analysis of copepod species. This would allow for more detailed analysis of correlations between zooplankton and physical parameters. We would also recommend biomass analysis, which would provide a clear summary of the variations in secondary productivity around the bay, and allow comparison to many other systems. We would recommend extending the sampling season.

Further work should be initiated to describe the spatial extent and processes that drive the extraordinary levels of productivity at tidewater glacier fronts.

Further work should be initiated to establish connections between nutrient input from adult salmon to nearshore streams and the marine interface. The value of salmon derived nutrients to nearshore marine systems in development of peri-glacial marine communities is of great interest. Furthermore, this research would provide a valuable nexus between salmon studies being conducted by Sandy Milner and Chad Soiseth in freshwater systems and State of Alaska research in the marine system.

The rapid declines, as well as the potentially related increases, of specific Apex predators are of concern. Given the rapid decline in murrelets and the steady decline of harbor seals, a better understanding of the factors influencing these species will be necessary before any measures can be taken to mitigate these changes.

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Literature Cited

Abookire, A. A., J. F. Piatt, & M. D. Robards. 2000. Nearshore fish distributions in an Alaskan estuary in relation to stratification, temperature, and salinity. *Estuarine, coastal, and shelf science* **51**: 45-59.

Allen, D. M., S. K. Service, and M. V. Ogburn-Matthews. 1992. Factors influencing the collection efficiency of estuarine fishes. *Transactions of the American Fisheries Society* **121**: 234-244.

Allen, L. 1982. Seasonal abundance, composition, and productivity of the littoral fish assemblage in upper Newport Bay, California. *Fishery Bulletin US* **80**: 769-790.

Allen, L. G. and M. H. Horn. 1975. Abundance, diversity, and seasonality of fishes in Colorado Lagoon, Alamitos Bay, California. *Estuarine, Coastal and Marine Science* **3**: 371-380.

Anderson, P. J. and J. F. Piatt. 1999. Community reorganization in the Gulf of Alaska following ocean climate regime shift. *Marine Ecology Progress Series*. **189**: 117-123.

Anonymous. 1995. Technical manual: operations and maintenance manual, satellite signals navigation sets, AN/PSN-11 and AN/PSN-11(V)1. Collins Avionics & Communications Division, Rockwell International Corporation, Cedar Rapids, Iowa, 231p.

Armstrong, R. H. 1970. Age, food, and migration of Dolly Varden smolts in southeastern Alaska. *Journal of the Fisheries Research Board of Canada* **27**: 991-1004.

Beamish, R. J., B. L. Thomson, and G. A. McFarlane. 1992. Spiny dogfish predation on chinook and coho salmon and the potential effects on hatchery-produced salmon. *Transactions of the American Fisheries Society* **121**: 444-455.

Brattegard, T. 1980. Why biologists are interested in fjords. Pages 53-56 in *Fjord Oceanogaphy*. Freeland, H. J., Farmer, D. M., and Levings, C. D. editors. Plenum Press, New York.

Bodkin, J. L., K.A. Kloecker, G.G. Esslinger, and D. H. Monson. Sea otter studies in Glacier Bay National Park and Preserve: Aerial surveys, foraging observations, and intertidal clam sampling. 2000 Annual Report from USGS Biological Resources Division to Glacier Bay National Park, Resources Management.

Cailliet, G. M. and A. W. Ebeling. 1990. The vertical distribution and feeding habits of two common midwater fishes (*Leuroglossus stilbius* and *Stenobrachius leucpsarus*) off Santa Barbara. CalCOFI Rep. **31**: 106-123.

Cailliet, G. M., M. S. Love, and A. W. Ebeling. 1986. Fishes: A field and laboratory manual on their structure, identification, and natural history. Wadsworth Publishing Company, Belmont, California: 132-134.

Catton, T. 1995. Land Reborn: A history of administration and visitor use in Glacier Bay National Park and Preserve. National Park Service, Anchorage, Alaska 99503.

Chilton, L., P. N. Hooge, and S. J. Taggart. 1995. Prey preferences of Pacific halibut (*Hippoglossus stenolepis*) in Glacier Bay National Park. In: D. R. Engstrom (ed.). Proceedings of the Third Glacier Bay Science Symposium. National Park Service, Anchorage, Alaska 1993.

Cooney, R. T. 1987. Zooplankton. The Gulf of Alaska: physical environment and biological resources. D. W. Hood and S. T. Zimmerman. Washington D.C., Alaska Office, Ocean Assessments Division, NOAA: 285-303.

Cooney, R. T. and K. O. Coyle. 1997. The role of zooplankton in the Prince William Sound ecosystem. Fairbanks, University of Alaska Fairbanks: 30.

Cowan, E. A. Meltwater and tidal currents: Controls on Circulation in a small glacial fjord. Estuarine, *Coastal, and Shelf Science* **34**: 381-392.

Creutzberg, F, A.T.G.W. Eltink, & Van Noort 1978. The migration of plaice larvae Pleuronectes platessa into the western Dutch Wadden Sea. Pages 243-252 *in: Physiology and behavior of marine organisms*. D.S. McLusky and A. J. Berry (eds.) Proceedings of the 12th. European Marine Biology Symposium, Stirling, Scotland. Pergamon Press: 243-251.

Doherty, J. L. and C. M. Gabriele. Population characteristics of humpback whales in Glacier Bay and adjacent waters: 2001. Glacier Bay National Park Service research report 28 pp.

Edmondson, W. and A. Litt 1982. Daphnia in Lake Washington. Limnology and Oceanography **27**: 272-293.

Epifanio, C. E. 1987. The role of tidal fronts in maintaining patches of brachyuran zoeae in estuarine waters. Journal of Crustacean Biology **7**: 513-517.

Forward, R. B., Jr. 1987. Larval release rhythms of decapod curstaceans: and overview. *Bulletin of Marine Science* **41**: 165-176.

Gordon, D. K. and C. D. Leavings. 1984. Seasonal changes of inshore fish populations on Sturgeon and Roberts Bank, Fraser River estuary, British Columbia. *Canadian Technical Report of Fisheries and Aquatic Science* **1240**: 1-12.

Gould, P. J., and D. J. Forsell. 1989. Techniques for shipboard surveys of marine birds. U.S. Department of the Interior, Fish and Wildlife Service, Washington, DC. Fish and Wildlife Technical Report **25**. 22pp.

Gould, P. J., D. J. Forsell, and C. J. Lensink. 1982. Pelagic distribution and abundance of seabirds in the Gulf of Alaska and eastern Bering Sea. U.S. Department of the Interior, Fish and Wildlife Service, Biological Services Program, OBS 82/48. 294pp.

Greisman, P. 1979. On upwelling driven by the melt of ice shelves and tidewater glaciers. *Deep Sea Research* **26A**: 1051-1065.

Hale, L. Z., & R. G. Wright. 1979. The Glacier Bay marine ecosystem: A conceptual ecological model. U.S. National Park Service, Anchorage Area Office.

Hancock, M. J. 1975. A survey of the fish fauna in the shallow marine waters of clam lagoon, Adak, Alaska. Florida Atlantic University, M.S. Thesis, 49pp.

Hartley, C. H. and M. J. Dunbar. 1938. On the hydrographic mechanism of the so-called brown zones associated with tidal glaciers. *Journal of Marine Research* **1**: 305-311.

Hooge, P. N. and E. R. Hooge (2001). Fjord oceanographic processes in Glacier Bay, Alaska. Gustavus, USGS Glacier Bay Field Station: 134.

Horn, M. H. 1980. Diel and seasonal variation in abundance and diversity of shallowwater fish populations in Morro Bay, California. *Fishery Bulletin US* **78**: 759-770.

Isakson, J. S., C. A. Simenstad, and R. L. Burgner. 1971. Fish communities and food chains in the Amchitka area. *Bioscience* **21**: 666-670.

Isaacs, J. D., and L. W. Kidd. 1953. Isaacs-Kidd midwater trawl. Final Report. Scripps Institution of Oceanography, University of California 53-3.

Jangaard, P. M. 1974. The capelin (*Mallotus villosus*). Department of the Environment, Fisheries, and Marine Service. Ottawa. *Fisheries Research Board of Canada Bulletin* **186**.

Kim, S., and D.R. Gunderson. 1988. Walleye Pollock, *Theragra chalcogramma*, in the Gulf of Alaska. Pages 70-82 *in*: Species synopses, life histories of selected fish and shellfish of the Pacific and Bering Sea. Wilmovsky, N. J., Incze, L. S., Westrheim, S. J. (eds.). Washington Sea Grant Program.

Krieger, K. J. and B. L. Wing. 1984. Hydroacoustic surveys and identification of humpback whale forage in Glacier Bay, Stephens Passage, and Frederick Sound, Southeastern Alaska, Summer 1983. U. S. Dep. Commer., NOAA Technical Memorandum. NMFS/NWC-66, 60p.

Krieger, K. J. and B. L. Wing. 1986. Hydroacoustic monitoring of prey to determine humpback whale movements. U.S. Dep. commer., NOAA Technical Memorandum. NMFS/NWC-98, 62p.

Lawrence, D. B., R. C. Schonike, Q. Quispel, and G. Bond. 1967. The role of *Dryas drummondii* in vegetation development following ice recession at Glacier Bay, Alaska, with special reference to its nitrogen fixation by root nodules. *Journal of Ecology* **55**: 793-813.

Litzow, M. A., J. F. Piatt, A. A. Abookire, A. K. Prichard, and M. D. Robards. 2000. Monitoring temporal and spatial variability in Pacific sand lance abundance with pigeon guillemot diets. *ICES Journal of Marine Science* **57**: 976-986.

Livingston, R. J. 1976. Diurnal and seasonal fluctuations or organisms in a north Florida estuary. *Estuarine Coastal and Marine Science* **4**: 373-400.

Lloyd, D. S. 1987. Turbidity as a water quality standard for salmonid habitats in Alaska. *North American Journal of Fisheries Management* **7**, 34-45.

Mann, K. H. 1993. Physical oceanography, food chains, and fish stocks: a review. *ICES Journal of Marine Science* **50**: 105-119.

Mason, J. C. and A. C. Phillips. 1985. Biology of the bathylagid fish, *Leuroglossus schmidti*, in the Strait of Georgia, British Columbia, Canada. *Canadian Journal of Fisheries and Aquatic Sciences* **42**: 1144-1153.

Matthews, J. B., and A. V. Quinlan. 1975. Seasonal characteristics of water masses in Muir Inlet, a fjord with tidewater glaciers. *Journal of the Fisheries Research Board of Canada* **32**: 1693-1703.

Matthews, J. B., and B. R. Heimdal. 1980. Pelagic productivity and food chains in fjord systems. Pages 377-398 *in Fjord Oceanogaphy*. Freeland, H. J., D. M. Farmer, and C. D. Levings (eds.). Plenum Press, New York.

Mattson, C. R. W., and L. Bruce. 1978. Ichthyoplankton Composition and Plankton Volumes From Inland Coastal Waters of Southeaster Alaska, April-November 1972. Auke Bay, NOAA: 11.

Milner, A. M. 1987. Colonization and ecological development of new streams in Glacier Bay National Park, Alaska. *Freshwater Biology* **18**: 53-70.

Milner, A. M. 1994. Colonization and succession of invertebrate communities in a new stream in Glacier Bay National Park, Alaska. *Freshwater Biology* **32**: 387-400.

Muench, R. D. and D. T. Heggie. 1978. *Deep water exchange in Alaskan subarctic fjords*. Pages 239-267 *in*: Estuarine Transport and Processes (Kjerfie, B. ed.). University of South Carolina Press, Columbia.

Müter, F. J. and B. L. Norcross. 1994. Distribution, abundance, and growth of larval walleye pollock, *Theragra chalcogramma*, in an Alaskan fjord. *Fishery Bulletin* **92**: 579-590.

Nagasawa, K., A. Nishimura, T. Asanuma, and T. Marubayashi. 1997. Myctophids in the Bering Sea: Distribution, abundance, and significance as food for salmonids. Pages 337-350 *in* Forage Fishes in Marine Ecosystems. Proceedings of the International Symposium on the Role of Forage Fishes in Marine Ecosystems. (C. W. Mechlenburg, ed.). 13-16 Nov. 1996. Anchorage, Alaska. Alaska Sea Grant College Program Report No. 97-01. University of Alaska, Fairbanks, AK.

Orsi, J. A. and J. H. Landingham. 1985. Numbers, species, and maturity stages of fish captured with beach seines during the spring 1981 and 1982 in some nearshore marine waters of southeastern Alaska. U.S. Dept. Comm., NOAA Tech. Mem. NMFS F/NWC-86, 34pp.

Orsi, J. A., J. M. Murphy, and A. L. J. Brase. 1997. Surveys of juvenile salmon in the marine waters of southeastern Alaska, May-August 1997. (NPAFC Doc. 277) 27p. Auke Bay Laboratory, Alaska Fisheries Science Center, National Marine Fisheries Service, NOAA, 11305 Glacier Highway, Juneau, AK 99801-8626, USA.

Orsi, J. A., J. M. Murphy, and D. G. Mortensen. 1998. Surveys of juvenile salmon in the marine waters of southeastern Alaska, May-August 1998. (NPAFC Doc. 346) 26p. Auke Bay Laboratory, Alaska Fisheries Science Center, National Marine Fisheries Service, NOAA, 11305 Glacier Highway, Juneau, AK 99801-8626, USA.

Orsi, J. A., M. V. Sturdevant, J. M. Murphy, D. G. Mortensen, and B. L. Wing. 2000. Seasonal habitat use and early marine ecology of juvenile Pacific salmon in southeastern Alaska. 29p. Auke Bay Laboratory, Alaska Fisheries Science Center, National Marine Fisheries Service, NOAA, 11305 Glacier Highway, Juneau, AK 99801-8626, USA.

Paul, A. J., J. M. Paul, K. Coyle, and R. Smith 1991. Phytoplankton, zooplankton, and ichthyoplankton in Resurrection Bay, Northern Gulf of Alaska in 1988. Fairbanks, Alaska Sea Grant, University of Alaska Fairbanks: 24pp.

Paxton, J. R. 1967. A distributional analysis for the lanternfishes (family Myctophidae) of the San Pedro Basin, California. *Copeia* **1967(2)**: 422-440.

Pearcy, W. G. 1964. Some distributional features of mesopelagic fishes off Oregon. Journal of Marine Research **22(1)**: 83-102.

Pritchard, A. L. 1939. Homing tendancy and age at maturity of pink salmon (*Oncorhynchuys gorbuscha*) in British Columbia. *Journal of the fisheries research board of Canada* **4**: 233-251.

Robards, M. D., J. F. Piatt, A. B. Kettle, and A. A. Abookir. 2000. Temporal and Geographic Variation in Fish Communities of Lower Cook Inlet, Alaska. *Fishery Bulletin* **97**: 962-977.

Rogers, B. J., M. E. Wangerin, K. J. Garrison, and D. E. Rogers 1981. Epipelagic meroplankton, juvenile fish, and forage fish: distribution and relative abundance in coastal waters near Yakutat. Seattle, Outer Continental Shelf Environmental Assessment Program (OCSEAP) **12**: 603-708.

Rogers, S. I. 1992. Environmental factors affecting the distribution of sole (*Solea solea* [L]) within a nursery area. *Netherlands Journal of Sea Research* **29**: 153-161.

Scheel, D., and K. R. Hough. 1997. Salmon fry predation by seabirds near an Alaskan hatchery. *Marine Ecology Progress Series* **150**: 35-48.

Shimada, B. M. 1948. Records of lanternfish in Puget Sound. Copeida 1948(3): 227.

Sidle, R. C., and A. M. Milner. 1989. Stream development in Glacier Bay National Park, Alaska, USA. *Arctic and Alpine Research* **21**: 350-363.

Simenstad, C. A. and R. D. Powell. 1990. Benthic, epibenthic, and planktonic invertebrates in ice-proximal glacimarine environs: Life in the turbidity lane. Pages 120-126 *in* Proceedings of the Second Glacier Bay Science Symposium. Milner, A. M., and Wood, J. D. Jr., (Eds.). September 19-22, 1988. Glacier Bay National Park and Preserve, Gustavus, Alaska.

Springer, A. M., and S. G. Speckman. 1997. A forage fish is what? Summary of the Symposium. Pages 773-794 *in*: Proceedings of the Symposium on the Role of Forage Fish in the Marine Ecosystem. Baxter, B. R. (Ed.). Alaska Sea Grant College Program AK-SG-97-01.

Springer, A. M. J. F. Piatt, V. P. Shunton, G. B. Van Vliet, V. L. Vladimirov, A. E. Kuzin, and A. S. Perlov. 1999. Marine birds and mammals of the Pacific subarctic gyres. Progress in Oceanography **3(2-4)**: 443-487.

Templeman, W. 1948. The life history of capelin (*Mallotus villosus*) in Newfoundland waters. *Bulletin of the Newfoundland Government Laboratory* **17** (research). 151pp.

Thorman, S. 1986. Seasonal colonization and effects of salinity and temperature on species richness and abundance of fish of some brackish and estuarine shallow waters in Sweden. *Holarctic Ecology* **9**: 126-132.

Tyler, R. W., and D. R. Gibbons. 1973. Observations of the effects of logging on salmon producing tributaries on the Staney Creek watershed and the Thorne River watershed and of logging in the Sitka district. Fisheries Research Institute Report No 7307. University of Washington, Seattle, Washington.

Van Pelt, T. I., J. F. Piatt, B. K. Lance, and D. D. Roby. 1997. Proximate composition and energy density of some North Pacific forage fishes. *Comparative Biochemistry and Physiology* **118A(2)**: 1393-1398.

Willette, T. M., R. T. Cooney, and K. Kyer. 1999. Predator foraging mode shifts affecting mortality of juvenile fishes during the subarctic spring bloom. *Canadian Journal of Fisheries and Aquatic Sciences* **56**, 364-376.

Wing, B. L. 1985. Salmon stomach contents from the Alaska troll logbook program 1977-84. U.S. Dep. Commer., NOAA Tech. Memo. NMFS-F/NWC-91.

Wing, B. L. and K. Krieger. 1983. Humpback whale prey studies in southeastern Alaska, summer 1982. Unpubl. rep., 60p. Auke Bay Laboratory, Northwest and Alaska Fisheries Center, National Marine Fisheries Service, NOAA, P.O. Box 210155, Auke Bay, AK 99821.

Yang, M-S. 1996. Diets of the important groundfishes in the Aleutian Islands in summer 1991. U.S. Dep. Commer., NOAA Tech. Memo. NMFS-AFSC-60, 105p.

Young, C. M. 1995. Behavior and locomotion during the dispersal phase of larval life. Ecology of Marine invertebrate larvae. L. McEdward. Boca Raton, CRC Press: 249-277.

Appendices

Sample Type	Station	Location	Longitude	Latitude	Region
Beach Seine	BS01	North Side Bartlett Cove	-135.90	58.47	Lower Bay
Beach Seine	BS02	Southwestern beach on Young Island	-135.99	58.47	Lower Bay
Beach Seine	BS03	First Beach North of BS02	-135.99	58.47	Lower Bay
Beach Seine	BS04	Second Beach North of BS02	-136.00	58.47	Lower Bay
Beach Seine	BS05	Northwestern Beach on Young Island	-136.00	58.48	Lower Bay
Beach Seine	BS06	Ripple Cove	-136.09	58.45	Lower Bay
Beach Seine	BS07	South Side of Berg Bay	-136.18	58.51	Lower Bay
Beach Seine	BS08	Northern Entrance to South Fingers Bay	-136.20	58.58	Middle Bay
Beach Seine	BS09	Northwestern Corner of South Fingers Bay	-136.21	58.58	Middle Bay
Beach Seine	BS10	West of Southern Tip of Drake at base of Marble Mountain	-136.25	58.63	Middle Bay
Beach Seine	BS11	Southern Beardslee Islands	-135.90	58.50	Lower Bay
Beach Seine	BS12	Eider Island in Beardslee Islands	-135.93	58.51	Lower Bay
Beach Seine	BS13	East side of Link Island in Beardslee Islands	-135.89	58.56	Lower Bay
Beach Seine	BS14	Northern Beardslee Islands	-135.92	58.57	Lower Bay
Beach Seine	BS15	Flapjack Island (South side)	-135.98	58.59	Lower Bay
Beach Seine	BS16	West of Island on North Shore of Bear Track Cove	-135.87	58.60	Middle Bay
Beach Seine	BS17	Beach on south side of broad promentory north of Bear Track	-135.91	58.63	Middle Bay
Beach Seine	BS18	York Creek	-135.92	58.64	Middle Bay
Beach Seine	BS19	North side of Spokane Cove	-135.97	58.70	Middle Bay
Beach Seine	BS20	Mount Wright drop-off	-136.05	58.79	Middle Bay
Beach Seine	BS21	Mid-channel island in Geike Inlet	-136.38	58.64	Middle Bay
Beach Seine	BS22	Southeastern corner of Geike Inlet	-136.48	58.59	Middle Bay
Beach Seine	BS23	Southwest beach of Geike	-136.51	58.59	Middle Bay

Appendix 1. Sample site locations throughout Glacier Bay.

Station Sample Type Location Longitude Latitude Region Beach Seine **BS24** Northwestern beach at head of -136.5258.61 Middle Bay Geike Inlet Beach Seine BS25 Immediately south of Sealers -136.1058.95 East Arm Island Beach Seine BS26 Immediately north of Sealers -136.12 58.97 East Arm Island **BS27** South of McBride entrance Beach Seine -136.12 59.02 East Arm Beach Seine BS28 Just north of entrance to -136.15 59.04 East Arm **McBride** Beach Seine BS29 Camping beach north of -136.16 59.04 East Arm McBride entrance **Beach Seine** BS30 Northeastern corner of Muir -136.19 59.07 East Arm Inlet Mid-way into Wachusett Inlet Beach Seine BS31 -136.35 58.95 East Arm on North Side Beach Seine BS32 Outwash flats of Wachusett -136.42 58.99 East Arm Glacier Beach Seine BS33 Directly opposite Maquinna -136.11 58.86 East Arm Cove on west of Muir Inlet Beach Seine BS34 On outwash of Morse Creek -136.1258.84 East Arm Beach Seine BS35 Muir Point -136.0958.82 Middle Bay Beach nestled into south side of Beach Seine BS36 -136.4658.83 Middle Bay Gloomy Knob Beach Seine BS37 Northwestern corner of Queen -136.56 58.96 West Arm Inlet **Beach Seine** BS38 Western side of Queen Inlet -136.58 58.94 West Arm **Beach Seine** BS39 Head of Rendu Inlet -136.72 59.01 West Arm Beach Seine BS40 Northwestern Rendu. Beach to -136.70 58.99 West Arm north of largest aluvial fan Old Queen drop-off site east of Beach Seine BS41 -136.51 58.88 West Arm Composite Island **Beach Seine** Gloomy knob by Vivid Lake BS42 -136.50 58.85 West Arm outlet **Beach Seine** Northeastern corner of Reid Inlet BS43 -136.81 58.87 West Arm **Beach Seine BS44** Mid-eastern side of Reid Inlet -136.8058.86 West Arm Beach Seine BS45 Southeastern Corner of Reid 58.85 West Arm -136.81

Inlet

Appendix 1. Continued.

Sample Type	Station	Location	Longitude	Latitude	Region
Beach Seine	BS46	Large beach east of Lamplugh Glacier	-136.89	58.89	West Arm
Beach Seine	BS47	Northeastern corner of Tarr by outlet (difficult seine)	-137.04	59.06	West Arm
Beach Seine	BS48	Large outwash fan on east side of Tarr Inlet	-136.94	59.00	West Arm
Beach Seine	BS49	Southeastern corner of Tarr before entering Russel Passage	-136.86	58.96	West Arm
Beach Seine	BS50	Southeastern Scidmore Bay	-136.56	58.77	Middle Bay
Beach Seine	BS51	Northern point in Scidmore Bay	-136.65	58.82	Middle Bay
Beach Seine	BS52	Southwestern point of Strawberry Island	-136.02	58.51	Lower Bay
Beach Seine	BS53	Just south of Ripple Cove	-136.07	58.43	Lower Bay
Beach Seine	BS54	Inside lagoon at Point Carolus	-136.04	58.38	Lower Bay
Beach Seine	BS55	West of broad headland between Carolus and Pt. Dundas	-136.14	58.34	Lower Bay
Beach Seine	BS56	Point Gustavus	-135.92	58.38	Lower Bay
Beach Seine	BS57	On outside of Point Corolus (seineable at low tide only)	-136.04	58.37	Lower Bay
Beach Seine	BS58	North side of Lemesurier Island	-136.10	58.30	Lower Bay
Beach Seine	BS59	Head of Mud Bay	-135.98	58.19	Lower Bay
Beach Seine	BS61	Point Adolphus	-135.83	58.27	Lower Bay
Beach Seine	BS62	West side of Pleasant Island	-135.71	58.36	Icy Strait
Beach Seine	BS63	Southwestern Reid Inlet	-136.82	58.85	West Arm
Beach Seine	BS64	Northwestern Reid Inlet	-136.82	58.87	West Arm
Nearshore Zooplankton	Z02	Young Island	-136.00	58.47	Lower Bay
Nearshore Zooplankton	Z06	Ripple Cove	-136.07	58.45	Lower Bay
Nearshore Zooplankton	Z09	Fingers Bay	-136.20	58.58	Middle Bay
Nearshore Zooplankton	Z10	Whidbey Passage	-136.25	58.63	Middle Bay
Nearshore Zooplankton	Z16	Beartrack Cove	-135.88	58.60	Middle Bay
Nearshore Zooplankton	Z20	Geikie Inlet	-136.05	58.78	Middle Bay
Nearshore Zooplankton	Z24	Mt. Wright	-136.51	58.60	Middle Bay
Nearshore Zooplankton	Z25	Sealers Island	-136.10	58.95	East Arm
Nearshore Zooplankton	Z30	Riggs Glacier	-136.19	59.07	East Arm
Nearshore Zooplankton	Z36	Gloomy Knob	-136.45	58.83	Middle Bay

Appendix 1. Continued

Appendix 1. Continued

Sample Type	Station	Location	Longitude	Latitude	Region
Nearshore Zooplankton	Z37	Queen Inlet	-136.55	58.96	West Arm
Nearshore Zooplankton	Z40	Rendu Inlet	-136.70	58.99	West Arm
Nearshore Zooplankton	Z45	Reid Inlet	-136.81	58.85	West Arm
Nearshore Zooplankton	Z47	Tarr Inlet	-136.99	59.03	West Arm
Nearshore Zooplankton	Z49	N. of Russel Island	-136.87	58.96	West Arm
Nearshore Zooplankton	Z55	North Passage	-136.13	58.34	Lower Bay
Nearshore Zooplankton	Z62	Pleasant Island	-135.73	58.36	Icy Strait
Non-Routine Sample	Random	Various Locations			Random
Pelagic	STN00	Icy Strait, Mouth of Glacier Bay	-135.87	58.33	Lower Bay
Pelagic	STN01	Mouth Glacier Bay	-135.99	58.41	Lower Bay
Pelagic	STN02	Sitakaday	-136.05	58.49	Lower Bay
Pelagic	STN03	SE of Willoughby Island	-136.06	58.57	Middle Bay
Pelagic	STN04	N of Drake I and N of Marble I.	-136.11	58.65	Middle Bay
Pelagic	STN05	Between N Drake and SW Tlingit PT	-136.23	58.70	Middle Bay
Pelagic	STN06	E of Hugh Miller Inlet	-136.35	58.76	Middle Bay
Pelagic	STN07	N of Blue Mouse, W of Tidal Inlet	-136.47	58.81	Middle Bay
Pelagic	STN08	S of Rendu Inlet	-136.59	58.87	West Arm
Pelagic	STN09	S of Russell Island	-136.73	58.90	West Arm
Pelagic	STN10	E of Russell Island	-136.84	58.90	West Arm
Pelagic	STN11	Tarr Inlet	-136.91	58.97	West Arm
Pelagic	STN12	Head of Tarr Inlet	-137.02	59.03	West Arm
Pelagic	STN13	SE of Tlingit PT, NW of Sturgess	-136.11	58.73	Middle Bay
Pelagic	STN14	Muir sill	-136.11	58.79	Middle Bay
Pelagic	STN15	W of Muir PT	-136.10	58.82	Middle Bay
Pelagic	STN16	E of Hunter Cove	-136.09	58.90	East Arm
Pelagic	STN17	E of Westdahl Pt	-136.13	58.98	East Arm
Pelagic	STN18	S of Riggs, NW of McBride	-136.18	59.05	East Arm
Pelagic	STN19	Muir Inlet	-136.33	59.07	East Arm
Pelagic	STN20	Head of Muir Inlet	-136.37	59.09	East Arm
Pelagic	STN21	Marjorie/Grand Pacific	-137.06	59.05	West Arm
Pelagic	STN22	Entrance to Geikie	-136.36	58.66	Middle Bay
Pelagic	STN23	Head of Geikie	-136.50	58.60	Middle Bay
Pelagic	STN24	Johns Hopkins	-137.06	58.88	West Arm

Appendix 1. Continued

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Sample Type	Station	Location	Longitude	Latitude	Region
Pelagic	STN25	Tidal Inlet	-136.32	58.82	Middle Bay
Pelagic	STN26	Rendu Inlet	-136.68	58.98	West Arm
Pelagic	STN27	Queen Inlet	-136.55	58.93	West Arm
Pelagic	STN28	Wachusetts Inlet	-136.39	58.98	West Arm

Appendix 2: Phylogeny of zooplankton taxa-lifestage groupings used.

Phylum	<i>Arthropoda</i> Crustacian (nauplii) Unidentified (nauplii)			Phylum Phylum	<i>Annelida</i> Annelid (Annelid Bryozoa	A <i>nnelida</i> Annelid (larvae) Annelid Bryozoa		
Class		Branchiopoda Order Cladesere			Bryozoar	(cyphona	utes larvae)	
		Evadne Podon	Claudera	Phylum	<i>Chaetogr</i> Class	natha Sagittoide Order	ea Aphragmophora	
	Class	Cirripedia					Sagitta elegans	
		Balanus Balanus	(cypris larvae) (nauplii)	Phylum	Cnidaria Cnidariar Medusae	ı (juvenile)		
	Class	Copepoo	la Colonaid and Cyclonaid		Class	Lludrozog		
		Orders	Calanoid/cyclopoid (small) Calanoid (large) Parasitic copepod		Class	Order	<i>Siphonophora</i> Siphonophore	
	Class	Harpaction Harpaction	Harpacticoida Harpacticoid		um Ctenophora Ctenophora			
		Order	Siphonostomatid Siphonostomatid	Phylum	Echinode Echinod Echinod	ermata lerm (juver lerm (larva	nile) e)	
	Class	<i>Malacosi</i> Order	rraca Amphipoda Amphipod Cyphocaris chalengeri Hyperiid Hyperiid (larvae)		Class	Asteroide Asteroida Asteroida Holothurc	ea (larvae) (juvenile) bidea	
					01000	Holothuro	nidea (auricularia larvae)	
		Order	Decapoda Brachyuran (larvae) Decapod (larvae) Decapod (megalop)	Phylum	<i>Mollusca</i> Class	<i>Bivalvia</i> Bivalve (v	veliger)	
		Order	Euphausiacea Euphausiid (metanauplii) Euphausiid (nauplii)		Class	<i>Gastropo</i> Gastropo	<i>da</i> d (veliger)	
			Euphausiid (zoea)	Phylum	Nematoda Nematoda			
		Order	Isopoda Cryptoniscid (larvae) Isopod Parasitic Isopod	Phylum	Phoronid Phoronid Phoronid	a a (juvenile a (actinotro) och larvae)	
	Class	<i>Ostracoc</i> Order	la Halocyprida Concoecia elegans	Phylum	<i>Urochoro</i> Class	lata Larvacea Oikopleui	ra spp.	

Appendix 3: Calanoid copepod species.

Centropages abdominalis	Neocalanus cristatus
Acartia clausi	Neocalanus plumchris
Acartia longiremis	Euchaeta elongata
Tortanus discaudatus	Epilabidocera longipedata
Eurytemora affinis	Metridia pacifica
Lucicutia flavicornis	Metridia ohkotensis
Calanus marshallae	Metridia lucens
Calanus pacificus	Pseudocalanus spp.

Appendix 4: Protocol for Echointegration of .dt4 files.

1) You must ensure that all files are named in such a way that, when listed in alphanumeric order, they will also be ordered in time. This is critical.

2) To begin working in Echoview, you must make a new .ev file. To make a new .ev file: go to **File**, **New**

open Transceiver 1 as the template go to **Data**, **Add Files**

Go to the location of the files you want to analyze, select them using the shift/control keys, and hit **Open**.

3) To see the echogram, go to View, Echogram.

4) To customize analysis options, go to **View**, **Options** or click on the icon of a hand pointing to a sheet of paper.

 5) Choose the Echogram tab, and set the following parameters: Change Lower Display Depth to the deepest depth of the entire .ev file (200 m) Set the minimum Sv (-90 dB for Glacier Bay) Set the grid limits. For Glacier Bay:

For time/distance, choose Ping Number and set it to 60 For depth, choose Relative to Surface and set it to 5 meters

6) To set the constants under the **Constants** tab, you must first use another menu. Close and save the Properties settings by clicking **Okay**. Then go to **Help**, **Sonar Calculator**.

Make sure the seawater box is checked

Enter the Temperature in °C (10° C for Glacier Bay)

Enter the Salinity in ppt (32 ppt for Glacier Bay)

Enter the frequency of the system (120 kHz)

Hit **Calculate**, and then copy the speed of sound in water (MacKenzie) and the absorption coefficient. Close the **Sonar Calculator** and return to the **View Options** menu, and choose the **Constants** tab.

Enter the speed of sound (1478.64 m/sec) Enter the absorption coefficient (0.0336230 dB/m) Enter the frequency as 120 kHz (book-keeping)

7) Still in the Properties menu system, choose the Integration tab. Check Exclude below bottom, and keep the 0 offset Check Apply minimum Sv threshold Check Apply maximum Sv threshold Appendix 4. Continued.

8) Still in the Properties menu system, choose the Export tab.

Select the spreadsheet format

Select all the following:

Sv mean Sa mean Sv max Sv min Samples LDepth U LDepth L Ping S Dist S Date S Time S Lat S Lon S Depth int Program version Num layers Num intervals Min depth

Max_depth Processing_date EV_filename Alpha Gain_constant Noise_power Minimum_Sv_threshold_applied Minimum_integration_threshold Maximum_Sv_threshold_applied Maximum_integration_threshold Surface_excluded Surface_excluded Surface_excluded Below_bottom_excluded Bottom_offset

9) To bottom track, first select the entire .ev file using the horizontal band tool. Right mouse click on the echogram, and a menu will pop up.

Select Pick Bottom

Wait until it finishes

Right mouse click again on the echogram, and select Move Bottom Line

Move the line –3 meters, which will move it 3 meters above the bottom. It should be above the visible bottom at all times. Check carefully where the bottom is steep, or has a thick sediment layer. You will need to painstakingly scroll along the entire .ev file, and should zoom in to a smaller area (such as 300 pings by 30 meters) to see well. To edit the bottom line, use the line draw tool to draw the bottom where you like by clicking along the line you want to draw. Then right mouse click and choose **Define Bottom**. All bottom drawn by hand will automatically be labeled "good" and have a green color. Be careful in places where fish are schooled along the bottom.

After the entire bottom is edited, use the horizontal band tool to select the entire .ev file, and right mouse click. Choose **Set Bottom**, **Good**. The entire bottom tracking line should be green.

10) In order to get an output that labels each transect by name, you must select each transect and define it as a region. Each transect can have more than one region, as long as each is named the same, i.e., uses the same transect name (for example, 45 or 87). Each region must begin at the line that forms the left-hand side (earliest side) of an interval. This is because we are using start time and position to define each interval.

Appendix 4. Continued.

Transects are broken into smaller parts according to the classification of the bottom. We defined 3 categories:

A) good bottom B) unknown bottom C) bottom >160 m

Define a region using the vertical band tool, starting from the left and scrolling to the right using the right arrow key and holding the mouse button down until you reach the end of the region.

Right mouse click on the region, and select Define Region

Erase the label; type in the transect number

Type will remain integration

For classification, choose **Edit**, **Add**, and then add the types you wish to classify (good, unknown, >160m). Make sure you leave Bad Data as an option.

Then choose one of the 3 types from the classification menu

Scroll through the entire .ev file, naming regions and classifying them. The left side must begin where an interval begins, but the right side can end in the middle of an interval. Be sure to check both sides of a region by scrolling to look, and zooming in if necessary—the lines tend to wander. Each transect may contain multiple sub-regions, all labeled with the same transect name (number) but with different bottom classifications.

11) To define Bad Data Regions, circle or enclose the affected area. Bad data regions can be of any shape or size. A variety of tools exists that draw in different ways. Once a bad data region is outlined, right mouse click, choose **Define Region**, and select the classification of Bad Data. The label does not matter and using the default will just result in regions numbered consecutively.

12) Finally, after the parameters are defined, the bottom is edited, the transects are labeled and the regions classified, you are ready to echointegrate. go to Data, Export, Integrate Regions by Cells name the output and choose the destination choose All Classifications (the default) and hit okay after 20-30 minutes or less, your output(s) will show up in the directory you indicated above

Appendix 5: Materials Used to Design and Conduct Predator Surveys

Protocols

Gould, P. J., and D. J. Forsell. 1989. Techniques for shipboard surveys of marine birds. U.S. Department of the Interior, Fish and Wildlife Service, Washington, DC. Fish and Wildlife Technical Report 25. 22pp.

Gould, P. J., D. J. Forsell, and C. J. Lensink. 1982. Pelagic distribution and abundance of seabirds in the Gulf of Alaska and eastern Bering Sea. U.S. Department of the Interior, Fish and Wildlife Service, Biological Services Program, OBS 82/48. 294pp.

Tips for Bird Surveys/Bird Identifications/Observsation Conditions/Sea Conditions.

DLOG Data entry and real-time mapping program overview (Need Ref information)

Marine Biodiversity Monitoring, protocol for monitoring seabirds. Diamond. http://www.cciw.ca/eman-temp/research/protocols/seabirds/intro.html

Species Identifications

Field Identification of Kittlitz's Murrelet, John Piatt. http://www.absc.usgs.gov/research/seabird&foragefish/index.html Field guide to the birds of north america. nat'l geographic society Peter Harrison (1983) Seabirds: An Identification Guide Peter Harrison (1997) Seabirds of the World: A Photographic Guide.

Appendix 6: Fish Larvae and Eggs Observed in Zooplankton Sampling.

Fish eggs STN00 March STN06 May West Arm EOIN* June Z9 June Z10 June Z20 June Z24 June Z36 June Z37 June Z45 June Z49 June Z55 August

.

Unidentified fish larvae STN00 March May STN16 March STN06 May West Arm EOIN* June Z02 June Z24 June Z49 June

Capelin larvae Z45 June

Unidentified fish alevin Z02 June

*EOI- Edge of sheet ice, N-neashore, M-mid-channel

Appendix 7: Fish Species List.

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Common Name	Latin Name
Pacific Herring	Clupea pallasi
Pink Salmon	Oncorhynchus gorbuscha
Chum Salmon	Oncorhynchus keta
Chinook Salmon	Oncorhynchus tshawytscha
Coho Salmon	Oncorhynchus kisutch
Sockeye Salmon	Oncorhynchus nerka
Dolly Varden	Salvelinus malma
Cutthroat Trout	Salmo clarki
Pacific Hake	Merluccius productus
Walleye Pollock	Theragra chalcogramma
Pacific Cod	Gadus macrocephalus
Capelin	Mallotus villosus
Northern Lampfish	Stenobrachius leucopsarus
Northern Smoothtongue	Leuroglossus schmidii
Unidentified Ronquil	Bathymaster Spp.
High Cockscomb	Anoplarchus purpurescens
Crescent Gunnel	Pholis laeta
Penpoint Gunnel	Apodichthys flavidus
Unidentified Gunnel	Pholidae Spp.
Arctic Shanny	Stichaeus punctatus
Slender Eelblenny	Lumpenus fabricii
Longsnout Prickleback	Lumpenella longirostris
Unidentified Prickleback	Stichaeidae Spp.
Wolf-Eel	Anarrhichthys ocellatus
Pacific Sandfish	Trichodon trichodon
Pacific Sand Lance	Ammodytes hexapterus
Rockfish Spp.	Sebastes Spp.
Lingcod	Ophiodon elongatus
Kelp Greenling	Hexagrammos decagrammus
Rock Greenling	Hexagrammos lagocephalus
Masked Greenling	Hexagrammos octogrammus
Whitespotted Greenling	Hexagrammos stelleri
Unidentified Greenling	Hexagrammidae Spp.
Unidentified Snailfish	Cyclopteridae
Unidentified Liparid (snailfish)	Liparis Spp.
Slipskin Snailfish	Liparis fucensis
Ribbon Snailfish	Liparis cyclopus

Appendix 7. Continued

Common Name	Latin Name
Pacific Spiny Lumpsucker	Eumicrotremus orbis
Smooth Lumpsucker	Aptocyclus ventricosus
3-spine stickleback	Gasterosteus aculeatus
Northern Sculpin	Icelinus borealis
Great Sculpin	Myoxocephalus polyacanthocephalus
Warty Sculpin	Myoxocephalus verrucosus
Buffalo Sculpin	Enophrys bison
Silverspotted Sculpin	Blepsias cirrhosus
Tadpole Sculpin	Psychrolutes paradoxus
Armorhead Sculpin	Gymnocanthus galeatus
Pacific Staghorn Sculpin	Leptocottus armatus
Brown Irish Lord	Hemilepidotus spinosus
Shaggy Sea Raven	Hemitripterus villosus
Sailfin Sculpin	Nautichthys oculofasciatus
Blackfin Sculpin	Malacocottus kincaidi
Unidentified Sculpin	Cottidae Spp.
Sturgeon Poacher	Podothecus acipenserinus
Bathyagonus Spp.	Bathyagonus Spp.
Unidentified Poacher	Agonidae Spp.
Rock Sole	Pleuronectes bilineatus
Butter Sole	Pleuronectes isolepis
English Sole	Pleuronectes vetulus
Curlfin Sole	Pleuronichthys decurrens
Pacific Halibut	Hippoglossus stenolepis
Starry Flounder	Platichthys stellatus
Unidentified Righteye Flounde	r Pleuronectidae Spp.
Unidentified Juvenile Flatfish	Pleuronectes Spp.
Squid	
Octopus	
Pandalus eos (Shrimp)	Pandalus eos
Pasiphaea pacifica (Shrimp)	Pasiphaea pacifica
Neomysis rayii (Shrimp)	Neomysis rayii
Pandalid Shrimp	Pandalus Spp.
Euphausiid	
Jellyfish	

SPECIES	LATIN NAME	GROUP	FUNC. GROUP	Species Code
Aleutian Tern	Sterna aleutica	Tern	Fish-feeder	ALTE
American Robin	Turdus migratorius	Terrestrial Bird	Other	AMRO
American Wigeon	Anas americana	Duck	Vegfeeder	AMWI
Ancient Murrelet	Synthliboramphus antiquus	Alcid	Plankton-feeder	ANMU
Arctic Tern	Sterna paradisaea	Tern	Fish-feeder	ARTE
Bald Eagle	Haliaeetus leucocephalus	Raptor	Other	BAEA
Barrow's Goldeneye	Bucephala islandica	Duck	Invert-feeder	BAGO
Barn Swallow	Hirundo rustica	Terrestrial Bird	Insect-Feeder	BASW
Black-billed Magpie	Pica pica	Terrestrial Bird	Other	BBMA
Belted Kingfisher	Ceryle alcyon	Terrestrial Bird	Fish-feeder	BEKI
Black Bear	Ursus americanus	Terrestrial Mammal	Other	BLBE
Black-Legged Kittiwake	Rissa tridactyla	Gull	Invert-feeder	BLKI
Black Oystercatcher	Haematopus bachmani	Shorebird	T idal Feeder	BLOY
Black Scoter	Melanitta nigra	Duck	Invert-feeder	BLSC
Black Turnstone	Arenaria melanocephala	Shorebird	T idal Feeder	BLTU
Bonaparte's Gull	Larus philadelphia	Gull	Invert. Feeder	BOGU
Brant	Branta bernicla	Goose	Vegfeeder	BRAN
Brown Bear	Ursus arctos	Terrestrial Mammal	Other	BRBE
Brachyramphus Murrelet	Brachyramphus spp.	Alcid	Fish-feeder	BRMU
Bufflehead	Bucephala albeola	Duck	Invert-feeder	BUFF
Canada Goose	, Branta candensis	Goose	Vegfeeder	CAGO
Canada Jay (Gray Jay)	Perisoreus canadensis	Terrestrial Bird	Other	CAJA
Caspian Tern	Sterna caspia	Tern	Fish-feeder	CATE
Cliff Swallow	Hirundo pyrrhonota	Terrestrial Bird	Insect-Feeder	CLSW
Common Goldeneye	Bucephala clangula	Duck	Invert-feeder	COGO
Common Loon	Gavia immer	Loon	Fish-feeder	COLO
Common Merganser	Mergus merganser	Duck	Fish-feeder	COME
Common Murre	Uria aalge	Alcid	Fish-feeder	COMU
Common Raven	Corvus corax	Terrestrial Bird	Other	CORA
Dall's Porpoise	Phocoenoides dalli	Marine Mammal	Fish-feeder	DAPO
Double-crested Cormorant	Phalacrocorax auritus	Cormorant	Fish-feeder	DCCO
Fork-tailed Storm Petrel	Oceanodroma furcata	Tubenose	Plankton-feeder	FTSP
Gadwall	Anas strepera	Duck	Veafeeder	GADW
Great Blue Heron	Ardea herodias	Shorebird	T idal Feeder	GBHE
Golden Eagle	Aquila chrvsaetos	Raptor	Other	GOEA
Grev Wolf	Canis lupus	Terrestrial Mammal	Other	GRWO
Greater Scaup	Avthva marila	Duck	Invert-feeder	GRSC
Glaucous-winged Gull	Larus glaucescens	Gull	Fish-feeder	GWGU
Green-winged Teal	Anas crecca	Duck	Veafeeder	GWTE
Harlequin Duck	Histrionicus histrionicus	Duck	Invert-feeder	HADU
Harbor Porpoise	Phocoena phocoena	Marine Mammal	Fish-feeder	HAPO
Harbor Seal	Phoca vitulina	Marine Mammal	Fish-feeder	HASE
Herring Gull	Larus argentatus	Gull	Fish-feeder	HEGU
Horned Grebe	Podiceps auritus	Grebe	Fish-feeder	HOGR
Humpback Whale	Megaptera novaeangliae	Marine Mammal	Fish-feeder	HUWH
Kittlitz's Murrelet	Brachvramphus brevirostris	Alcid	Fish-feeder	KIMU
Killer Whale	Orcinus orca	Marine Mammal	Fish-feeder	KIWH
Marbled Murrelet	Brachvramphus marmoratus	Alcid	Fish-feeder	MAMU
Mew Gull	Laurs canus	Gull	Fish-feeder	MEGU

Appendix 8: Predator Survey Species List.

.

Appendix 8. Continued

SPECIES	LATIN NAME	GROUP	FUNC. GROUP	Species Code
American Mink	Mustela vison	Terrestrial Mammal	Other	MINK
Mallard	Anas platyrhynchos	Duck	Vegfeeder	MALL
Minke Whale	Balaenoptera acutorostrata	Marine Mammal	Fish-feeder	MIWH
Mountain Goat	Oreamnos americanus	Terrestrial Mammal	Other	MOGO
Moose	Alces alces	Terrestrial Mammal	Other	MOOS
Northwestern Crow	Corus caurinus	Terrestrial Bird	Other	NOCR
Northern Pintail	Anas acuta	Duck	Vegfeeder	NOPI
Northern Shoveler	Anas clypeata	Duck	Vegfeeder	NOSH
Long-tailed Duck	Clangula hyemalis	Duck	Invert-feeder	OLDS
Parasitic Jaeger	Stercorarius parasiticus	Jaeger	Fish-feeder	PAJA
Pacific Loon	Gavia pacifica	Loon	Fish-feeder	PALO
Pelagic Cormorant	Phalacrocorax pelagicus	Cormorant	Fish-feeder	PECO
Pigeon Guillemot	Cepphus columba	Alcid	Fish-feeder	PIGU
Red-breasted Merganser	Mergus serrator	Duck	Fish-feeder	RBME
Red-faced Cormorant	Phalacrocorax urile	Cormorant	Fish-feeder	RFCO
River Otter	Lutra canadensis	Terrestrial Mammal	Other	RIOT
Red-necked Grebe	Podiceps grisegena	Grebe	Fish-feeder	RNGR
Red-necked Phalarope	Phalaropus lobatus	Phalarope	Plankton-feeder	RNPH
Red-throated Loon	Gavia stellata	Loon	Fish-feeder	RTLO
Scaup	Aythya spp.	Duck	Invert-feeder	SCAU
Sea Otter	Enhydra lutris	Marine Mammal	Invert-feeder	SEOT
Spotted Sandpiper	Actitis macularia	Shorebird	T idal Feeder	SPSA
Stellar's Jay	Cyanocitta stelleri	Terrestrial Bird	Other	STJA
Stellar's Sea Lion	Eumetopias jubatus	Marine Mammal	Fish-feeder	STSL
Surfbird	Aphriza virgata	Shorebird	T idal Feeder	SURF
Surf Scoter	Melanitta perspicillata	Duck	Invert-feeder	SUSC
Tufted Puffin	Fratercula cirrhata	Alcid	Fish-feeder	TUPU
Unidentified Alcid	Alcidae spp.	Alcid	Unknown	UNAC
Unidentified Albatross	Phoebastria spp.	Tubenose	Squid-feeder	UNAL
Unidentified Cormorant	Phalacrocorax spp.	Cormorant	Fish-feeder	UNCO
Unidentified Duck	Anas spp.	Duck	Unknown	UNDU
Unidentified Eagle	Haliaeetus spp.	Raptor	Other	UNEA
Unidentified Goldeneye	Bucephala spp.	Duck	Invert-feeder	UNGO
Unidentified Grebe	Podiceps spp.	Grebe	Fish-feeder	UNGR
Unidentified Gull	Larus spp.	Gull	Fish-feeder	UNGU
Unidentified Jaeger	Stercorarius spp.	Jaeger	Fish-feeder	UNJA
Unidentified Large Larid	Larid spp.	Gull	Fish-feeder	UNLL
Unidentified Loon	Gavia spp.	Loon	Fish-feeder	UNLO
Unidentified Merganser	Mergus spp.	Duck	Fish-feeder	UNME
Unidentified Phalarope	Phalaropus spp.	Phalarope	Plankton-feeder	UNPH
Unidentified Raptor	Accipitridae spp.	Raptor	Other	UNRA
Unidentified Shorebird	shorebird spp.	Shorebird	Tidal Feeder	UNSB
Unidentified Scoter	Melanitta spp.	Duck	Invert-feeder	UNSC
Unidentified Swan	Cygnus spp.	Goose	Vegfeeder	UNSN
Unidentified Storm Petrel	Oceanodroma spp.	Tubenose	Plankton-feeder	UNSP
Unidentified Swallow	Hirundinidae spp.	Terrestrial Bird	Insect-Feeder	UNSW
Unidentified Tern	Sterna spp.	Tern	Fish-feeder	UNTE
Unidentified Teal	Teal spp.	Duck	Vegfeeder	UNTL
Unidentified Whale	Baleen Whale spp.	Marine Mammal	Unknown	UNWH
Western Grebe	Aechmophorus occidentalis	Grebe	Fish-feeder	WEGR
Western Screech Owl	Otus kennicottii	I errestrial Bird	Other	WSOW
White-winged Scoter	Melanitta fusca	Duck	Invert-feeder	WWSC
Yellow-billed Loon	Gavia adamsii	Loon	⊢ish-feeder	YBLO