

Thermodynamics of Enzyme-Catalyzed Reactions:

Part 4. Lyases

Robert N. Goldberg and Yadu B. Tewari

Biotechnology Division, National Institute of Standards and Technology, Gaithersburg, Maryland 20899-0001

Received January 27, 1995

Equilibrium constants and enthalpy changes for reactions catalyzed by the lyase class of enzymes have been compiled. For each reaction the following information is given: the reference for the data; the reaction studied; the name of the enzyme used and its Enzyme Commission number; the method of measurement; the conditions of measurement (temperature, pH, ionic strength, and the buffer(s) and cofactor(s) used); the data and an evaluation of it; and, sometimes, commentary on the data and on any corrections which have been applied to it or any calculations for which the data have been used. The data from 106 references have been examined and evaluated. Chemical Abstract Service registry numbers are given for the substances involved in these various reactions. There is a cross reference between the substances and the Enzyme Commission numbers of the enzymes used to catalyze the reactions in which the substances participate. © 1995 American Institute of Physics and American Chemical Society.

Key words: apparent equilibrium constants; enthalpies of reaction; enzyme-catalyzed reactions; evaluated data; lyases; transformed thermodynamic properties.

Contents

1. Introduction.....	1670
2. Some Aspects and Uses of Thermodynamic Data on Biochemical Reactions	1671
3. Acknowledgments.....	1671
4. References for the Introductory Discussion	1672
5. Table of Equilibrium Constants and Enthalpies of Reaction	1672
5.1 Enzyme: phosphoenolpyruvate carboxykinase (GTP) (EC 4.1.1.32).....	1672
5.2. Enzyme: phosphoenolpyruvate carboxykinase (diphosphate) (EC 4.1.1.38)...	1672
5.3. Enzyme: ribulose-biphosphate carboxylase (EC 4.1.1.39).....	1672
5.4. Enzyme: ketotetraose-phosphate aldolase (EC 4.1.2.2).....	1673
5.5. Enzyme: deoxyribose-phosphate aldolase (EC 4.1.2.4).....	1673
5.6. Enzyme: fructose-biphosphate aldolase (EC 4.1.2.13).....	1673
5.7. Enzyme: 2-dehydro-3-deoxyphosphogluconate aldolase (EC 4.1.2.14).....	1675
5.8. Enzyme: L-fuculose-phosphate aldolase (EC 4.1.2.17).....	1675
5.9. Enzyme: 2-dehydro-3-deoxy-L-pentonate aldolase (EC 4.1.2.18).....	1676
5.10. Enzyme: rhamnulose-1-phosphate aldolase (EC 4.1.2.19).....	1676
5.11. Enzyme: 2-dehydro-3-deoxy-6-phosphogalactonate aldolase (EC 4.1.2.21). .	1676
5.12. Enzyme: D-arabino-3-hexulose phosphate formaldehyde lyase (EC 4.1.2.a).....	1676
5.13. Enzyme: isocitrate lyase (EC 4.1.3.1).....	1676
5.14. Enzyme: malate synthase (EC 4.1.3.2)....	1677
5.15. Enzyme: N-acetylneuraminate lyase (EC 4.1.3.3).....	1677
5.16. Enzyme: citrate (<i>pro</i> -3 <i>S</i>)-lyase (EC 4.1.3.6).....	1677
5.17. Enzyme: citrate (<i>si</i>)-synthase (EC 4.1.3.7)..	1678
5.18. Enzyme: ATP citrate (<i>pro</i> -3 <i>S</i>)-lyase (EC 4.1.3.8).....	1678
5.19. Enzyme: 4-hydroxy-2-oxoglutarate aldolase (EC 4.1.3.16).....	1678
5.20. Enzyme: citramalate lyase (EC 4.1.3.22)...	1679
5.21. Enzyme: malyl-CoA lyase (EC 4.1.3.24)...	1679
5.22. Enzyme: 2,3-dimethylmalate lyase (EC 4.1.3.32).....	1679
5.23. Enzyme: tryptophanase (EC 4.1.99.1).....	1679
5.24. Enzyme: fumarate hydratase (EC 4.2.1.2)..	1680
5.25. Enzyme: aconitate hydratase (EC 4.2.1.3)..	1682
5.26. Enzyme: 3-dehydroquinate dehydratase (EC 4.2.1.10).....	1684
5.27. Enzyme: phosphopyruvate hydratase (EC 4.2.1.11).....	1685
5.28. Enzyme: enoyl-CoA hydratase (EC 4.2.1.17).....	1686

©1995 by the U. S. Secretary of Commerce on behalf of the United States. This copyright is assigned to the American Institute of Physics and the American Chemical Society.
Reprints available from ACS; see Reprints List at back of issue.

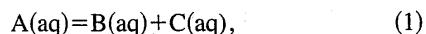
5.29. Enzyme: tryptophan synthase (EC 4.2.1.20).....	1686
5.30. Enzyme: maleate hydratase (EC 4.2.1.31).....	1687
5.31. Enzyme: (<i>S</i>)-2-methylmalate dehydratase (EC 4.2.1.34).....	1688
5.32. Enzyme: (<i>R</i>)-2-methylmalate dehydratase (EC 4.2.1.35).....	1688
5.33. Enzyme: D-glutamate cyclase (EC 4.2.1.48).....	1688
5.34. Enzyme: urocanate hydratase (EC 4.2.1.49).....	1689
5.35. Enzyme: crotonoyl-[acyl-carrier-protein] hydratase (EC 4.2.1.58).....	1689
5.36. Enzyme: dimethylmaleate hydratase (EC 4.2.1.85).....	1689
5.37. Enzyme: 3-hydroxybutyryl-CoA dehydratase (EC 4.2.1.a).....	1689
5.38. Enzyme: aspartate ammonia-lyase (EC 4.3.1.1).....	1689
5.39. Enzyme: methylaspartate ammonia-lyase (EC 4.3.1.2).....	1691
5.40. Enzyme: histidine ammonia-lyase (EC 4.3.1.3).....	1691
5.41. Enzyme: phenylalanine ammonia-lyase (EC 4.3.1.5).....	1692
5.42. Enzyme: β -alanyl-CoA ammonia lyase (EC 4.3.1.6).....	1692
5.43. Enzyme: arginosuccinate lyase (EC 4.3.2.1).....	1692
5.44. Enzyme: adenylosuccinate lyase (EC 4.3.2.2).....	1692
5.45. Enzyme: ureidoglycolate lyase (EC 4.3.2.3).....	1693
5.46. Enzyme: lactoylglutathione lyase (EC 4.4.1.5).....	1693
5.47. Enzyme: adenylate cyclase (EC 4.6.1.1)....	1693
6. List of Substances with Chemical Abstract Service (CAS) Registry Numbers with Cross References to Enzyme Commission Numbers	1694
7. Abbreviations.....	1696
8. Glossary of Symbols.....	1696
9. Reference Codes and References in the Table	1697

1. Introduction

This article is a continuation of the first three reviews in this series¹⁻³ that dealt, respectively, with the thermodynamics of the reactions catalyzed by the oxidoreductases, transferases, and hydrolases. These are the first three classes of enzymes classified by the Nomenclature Committee of the International Union of Biochemistry.⁴ In the current review a critical compilation of thermodynamic data is provided for the reactions catalyzed by the fourth class of enzymes—the lyases. These reactions play significant roles in metabolic processes such as glycolysis, the glyoxylic pathway, and the Krebs' or citric acid cycle. There is also interest in many of these reactions due to their importance in the utilization of biomass; an example here is the production of succinic acid from glucose. The data presented herein is limited to equi-

librium and calorimetric measurements performed on these reactions under *in vitro* conditions. The thermodynamic quantities that are generally given are apparent equilibrium constants K' and calorimetrically determined enthalpies of reaction $\Delta_r H(\text{cal})$. Apparent equilibrium constants calculated from kinetic data are also tabulated. If the change in binding of hydrogen ion $\Delta_r N(H^+)$ in a biochemical reaction is known, the standard transformed enthalpy of reaction $\Delta_r H^\circ$ can be calculated from the calorimetrically determined enthalpy of reaction.⁵ Equilibrium constants K and standard molar enthalpies of reaction $\Delta_r H^\circ$ for chemical reference reactions are also given if they have been reported in the literature. The standard transformed enthalpy of reaction $\Delta_r H^\circ$ can be used to calculate the temperature dependence of apparent equilibrium constants K' in the same way that the standard enthalpy of reaction $\Delta_r H^\circ$ is used to calculate the temperature dependence of the equilibrium constant K .

The data are presented in the same format as in Parts 1 to 3.¹⁻³ Thus, the following information is given for each entry in this review: the reference for the data; the biochemical reaction studied; the recommended name⁴ of the enzyme used and its Enzyme Commission number; the method of measurement; the conditions of measurement (temperature, pH, ionic strength, and the buffer(s) and cofactor(s) used); the data and its evaluation; and, sometimes, commentary on the data and on any corrections that have been applied or any calculations for which the data have been used. The absence of a piece of information indicates that it was not found in the cited paper. The arrangement of the data, its evaluation, and the thermodynamic conventions have been discussed previously.¹ One should note that equilibrium constants should be expressed as dimensionless quantities. However, the numerical value obtained for the equilibrium constant of an unsymmetrical reaction will depend upon the measure of composition and standard concentration selected for the reactants and products. Thus, for the chemical reaction



$K_c = c(B)c(C)/\{c(A)c^\circ\}$, $K_m = m(B)m(C)/\{m(A)m^\circ\}$, and $K_x = x(B)x(C)/x(A)$. Here, c , m , and x are, respectively, concentration, molality, and mole fraction, $c^\circ = 1 \text{ mol dm}^{-3}$, and $m^\circ = 1 \text{ mol kg}^{-1}$. The equilibrium constant expressed in terms of mole fractions is automatically dimensionless. Similar definitions and considerations apply to the apparent equilibrium constant K' . The symbols used in this review are given in the Glossary (see Section 8).

The subjective evaluation of the data in this review consisted of the assignment of a rating: A (high quality), B (good), C (average), or D (low quality). In making these assignments we considered the various experimental details which were provided in the study. These details include the method of measurement, the number of data points determined, and the extent to which the effects of varying temperature, pH, and ionic strength were investigated. A low rating was generally given when few details of the investigation were reported. For example, in many of the papers cited, the major aim of the study was the isolation and purification of the enzyme of interest. Thus, the equilibrium data

were obtained as only a small part of an investigation to characterize many of the properties of that enzyme and the reaction it catalyzes.

This effort has been given additional impetus by the recent completion of the IUBMB-IUPAC document "Recommendations for Nomenclature and Tables in Biochemical Thermodynamics".⁶ The work described in this review paper has also been accepted by the Thermodynamics Commission (1.2) and by the Steering Committee on Biophysical Chemistry of IUPAC as a project of particular timeliness and importance. The project has therefore been conducted under the auspices of these bodies, has been endorsed by them, and has been written to be consistent with recommended IUPAC nomenclature.

2. Some Aspects and Uses of Thermodynamic Data on Biochemical Reactions

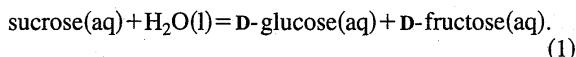
While a full discussion of the applications of thermodynamic data would be beyond the scope of this review, it seems useful to indicate briefly the utility of the information presented. A primary motivation for performing thermodynamic studies on biochemical reactions is to determine the position of equilibrium of the reaction(s) studied as well as to establish definitely the substrates that participate in the biochemical reaction(s). This information is concisely summarized in terms of the apparent equilibrium constants given herein. These apparent equilibrium constants can be conveniently used to calculate the extent of reaction under the stated set of conditions and therefore are very useful to engineers concerned with the optimization of product yield in bioreactors. The enthalpy changes accompanying these reactions are also required and determine how much heating or cooling is required to maintain a bioreactor at its proper temperature. To perform this calculation one must know both the standard transformed enthalpy of reaction $\Delta_r H^\circ$, the change in binding of the hydrogen ion $\Delta_r N(H^+)$, and the enthalpy of protonation of the buffer(s) in the bioreactor. In general, both $\Delta_r H^\circ$ and $\Delta_r N(H^+)$ are functions of temperature, pH, ionic strength, and the concentration of free metal ion(s).

Apparent equilibrium constants obtained from studies of *in vitro* systems can also be used to calculate the position of equilibrium in metabolic processes involving several reactions. Glycolysis is probably the best example of an enzymatic process where data are available and for which such calculations have been performed.^{7,8} The results of these calculations can then be compared with information on the concentrations of the various substrates obtained from the analysis of *in vivo* systems. This comparison can provide valuable insight into the chemical machinery of living systems.

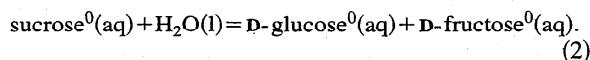
For many biochemical reactions, the apparent equilibrium constant is a function of temperature, pH, pX, and ionic strength. Therefore, when performing Hess' Law and thermochemical cycle calculations, it is necessary that the data for all of the reactions in such a calculation refer to the same set of conditions. The dependencies of apparent equilibrium constants and standard transformed enthalpies of reaction on the conditions of reaction can be very complex. Also, the

reduction of such results to a common standard state generally requires auxiliary information on the binding of protons and metal ions to the various reactants as well as information on or assumptions about the activity coefficients of the species in solution. These types of calculations have been performed by Kuby and Noltmann,⁹ Alberty,^{10,11} Guynn, Gelberg, and Veech,¹² Langer *et al.*,¹³ Goldberg and Tewari,¹⁴ and others.

For some reactions, none of the reactants or products in a given biochemical reaction have ionizable groups or bind metal ions under the conditions of the study. In such a case, and when there are no hydrogen or metal ions as reactants in the chemical reference reaction, the thermodynamic quantities for the chemical reference reaction correspond directly to the transformed thermodynamic quantities for the overall biochemical reaction. For example, the biochemical reaction for the hydrolysis of sucrose to D-glucose and D-fructose is:



A chemical reference reaction involving specific species is:



Here, the charges of the reference species have been specified to distinguish these species from the biochemical reactants which, in principle, can consist of a mixture of pseudoisomer species.⁸ Since the extent of ionization of these sugars for $\text{pH} < 10$ is negligible, sucrose⁰, D-glucose⁰, and D-fructose⁰ are the predominant species and for $\text{pH} < 10$, $K'(1) = K(2)$, $\Delta_r H^\circ(1) = \Delta_r H^\circ(2)$, and $\Delta_r G^\circ(1) = \Delta_r G^\circ(2)$. Also, since $\Delta_r N(H^+) = 0$ for reaction (2), $\Delta_r H(\text{cal}) = \Delta_r H^\circ(2)$.

Tables of standard formation properties¹⁵ have proven to be a useful way of generalizing upon and presenting thermodynamic data for many chemical substances. However, tables of this type have been prepared for only limited classes of biochemical substances.¹⁶⁻¹⁸ It has recently been shown¹⁹ how it is possible to prepare tables of standard transformed formation properties for biochemical reactants (i.e., sums of species) as distinct from standard formation properties for individual biochemical species. The adenosine 5'-triphosphate series was used as a prototype for this purpose. Thus, it appears likely and desirable that several different types of thermodynamic tables will eventually appear in the literature. Clearly, the larger the scope of such tables, the more useful they are for calculating thermodynamic quantities for reactions which have not been the subject of a direct investigation.

3. Acknowledgments

We thank Dr. Ellen Anderson and Dr. Edgar Etz for their assistance with the papers written in German and Dr. Mikhail V. Rekharsky and Dr. Vytaus Reipa for their help with the papers in Russian. Continuing discussions with Dr. Robert A. Alberty on various aspects of biochemical thermodynamics have been very helpful. Ms. Kari Fazio and Donna Bell provided valuable assistance in the early collection of the refer-

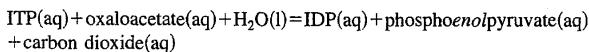
ences containing the data and in the preliminary abstracting of information. Support given to this project by the Offices of Industrial Technologies and Transportation Technologies in the U.S. Department of Energy is gratefully acknowledged.

4. References for the Introductory Discussion

- ¹R. N. Goldberg, Y. B. Tewari, D. Bell, K. Fazio, and E. Anderson, *J. Phys. Chem. Ref. Data* **22**, 515 (1993).
- ²R. N. Goldberg and Y. B. Tewari, *J. Phys. Chem. Ref. Data* **23**, 547 (1994).
- ³R. N. Goldberg and Y. B. Tewari, *J. Phys. Chem. Ref. Data* **23**, 1035 (1994).
- ⁴E. C. Webb, *Enzyme Nomenclature 1992* (Academic Press, San Diego, 1992).
- ⁵R. A. Alberty and R. N. Goldberg, *Biophys. Chem.* **47**, 213 (1993).
- ⁶R. A. Alberty, "Recommendations for Nomenclature and Tables in Biochemical Thermodynamics," *Pure Appl. Chem.* **66**, 1641 (1994).
- ⁷S. Minakami and H. Yoshikawa, *J. Biochem. (Tokyo)* **59**, 139 (1966).
- ⁸R. A. Alberty, *Biophys. Chem.* **42**, 117 (1992).
- ⁹S. A. Kuby and E. A. Noltmann, in *The Enzymes*, Volume 6, edited by P. D. Boyer, H. Lardy, and K. Myrbäck (Academic Press, New York, 1962), pp. 515–603.
- ¹⁰R. A. Alberty, *J. Biol. Chem.* **244**, 3290 (1969).
- ¹¹R. A. Albert, *Proc. Natl. Acad. Sci. U.S.A.* **88**, 3268 (1991).
- ¹²R. W. Guynn, H. J. Gelberg, and R. L. Veech, *J. Biol. Chem.* **248**, 695 (1973).
- ¹³R. S. Langer, C. R. Gardner, B. K. Hamilton, and C. K. Colton, *AIChE J.* **23**, 1 (1977).
- ¹⁴R. N. Goldberg and Y. B. Tewari, *Biophys. Chem.* **40**, 241 (1991).
- ¹⁵D. D. Wagman, W. H. Evans, V. B. Parker, R. H. Schumm, I. Halow, S. M. Bailey, K. L. Churney, and R. L. Nuttall, "The NBS Tables of Chemical Thermodynamic Properties," *J. Phys. Chem. Ref. Data* **11**, Supplement No. 2 (1982).
- ¹⁶R. C. Wilhoit, in *Biochemical Microcalorimetry*, edited by H. D. Brown (Academic Press, New York, 1969), pp. 33–81, 305–317.
- ¹⁷R. N. Goldberg and Y. B. Tewari, *J. Phys. Chem. Ref. Data* **18**, 809 (1989).
- ¹⁸S. L. Miller and D. Smith-Magowan, *J. Phys. Chem. Ref. Data* **19**, 1049 (1990).
- ¹⁹R. A. Alberty and R. N. Goldberg, *Biochemistry* **31**, 10610 (1992).

5. Table of Equilibrium Constants and Enthalpies of Reaction

5.1 Enzyme: phosphoenolpyruvate carboxykinase (GTP) (EC 4.1.1.32)



T K	pH	K' _c
303.15	7.6	≈12

Reference: 54UTT/KUR

Method: chromatography and enzymatic assay

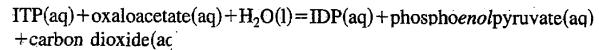
Buffer: acetate (0.050 mol dm⁻³)

pH: 7.6

Cofactor(s): MnCl₂ (0.002 mol dm⁻³)

Evaluation: C

The apparent equilibrium constant given here was calculated from the data given in Utter and Kurahashi's Table VIII.



T K	pH	c(MnCl ₂) mol dm ⁻³	I' _c mol dm ⁻³	K' _c
298.15	7.80	0.0020	0.1	1.7
298.15	7.06	0.0025	0.1	0.50
298.15	7.03	0.0025	0.1	0.71

Reference: 66WO0/DAV

Method: spectrophotometry and enzymatic assay

Buffer: Tris (0.010 mol dm⁻³) + HCl

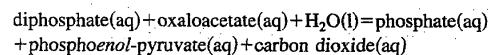
pH: 7.03–7.80

Cofactor(s): MnCl₂

Evaluation: A

Wood *et al.* also calculated $K_c = 4.5 \times 10^{-8}$ at $T = 298.15$ K and $I = 0.1$ mol dm⁻³ for the chemical reference reaction: $\text{ITP}^{4-}(\text{aq}) + \text{oxaloacetate}^{2-}(\text{aq}) = \text{IDP}^{3-}(\text{aq}) + \text{phosphoenolpyruvate}^{3-}(\text{aq}) + \text{HCO}_3^-(\text{aq}) + \text{H}^+(\text{aq})$.

5.2 Enzyme: phosphoenolpyruvate carboxykinase (diphosphate) (EC 4.1.1.38)



T K	pH	I' _c mol dm ⁻³	K' _c
298.15	6.5	0.1	0.013

Reference: 66WO0/DAV

Method: spectrophotometry and enzymatic assay

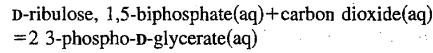
Buffer: carbonate + bicarbonate

pH: 6.5

Cofactor(s): MgCl₂ (0.0024 mol dm⁻³)

Evaluation: A

5.3 Enzyme: ribulose-biphosphate carboxylase (EC 4.1.1.39)



T K	pH	Buffer	ΔH(cal) kJ mol ⁻¹
298.15	8.0	Tris+HCl	-61.1
298.15	8.0	triethanolamine	-43.1

Reference: 56KIT/HOR

Method: microcalorimetry

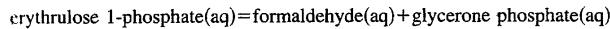
Buffer: Tris (0.1 mol dm⁻³) + HCl and triethanolamine

pH: 8.0

Evaluation: C

The temperature is assumed to be 298.15 K. After correction for the enthalpies of buffer protonation, Kitzinger *et al.* calculated $\Delta_f H^\circ (T = 298.15 \text{ K}, \text{pH}=8.0) = -20.1 \text{ kJ mol}^{-1}$.

5.4. Enzyme: ketotetraose-phosphate aldolase (EC 4.1.2.2)



$\frac{T}{K}$	pH	K'_c
301.15	7.4	4.3E-4

Reference: 54CHA

Method: radioactivity

Buffer: NH_4OH

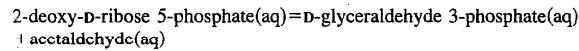
pH: 7.4

Cofactor(s): MgCl_2 (0.075 mol dm⁻³)

Evaluation: B

The apparent equilibrium constant given here was calculated from the data given in Charalampous' Table III.

5.5. Enzyme: deoxyribose-phosphate aldolase (EC 4.1.2.4)



$\frac{T}{K}$	pH	K'_c
310.15	6.3	2.35E-4

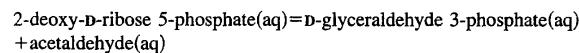
Reference: 60PRI/HOR

Method: enzymatic assay, chemical analysis, and spectrophotometry

Buffer: malate (0.025 mol dm⁻³)

pH: 6.3

Evaluation: B



$\frac{T}{K}$	pH	K'_c
295.15	7.5	2.5E-4

Reference: 67GRO

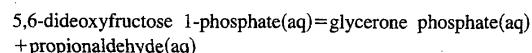
Method: spectrophotometry

Buffer: Tris (0.005 mol dm⁻³)

pH: 7.5

Evaluation: B

5.6. Enzyme: fructose-biphosphate aldolase (EC 4.1.2.13)

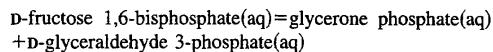


$\frac{T}{K}$	pH	K'_c
310.15	7.0	6.9E-4

Reference: 55LEH/SIC

pH: 7.0

Evaluation: C



$\frac{T}{K}$	pH	K'_c
301.15	7.2	6.0E-5
311.15	7.2	1.19E-4
311.15	7.33	1.22E-4
321.15	7.2	2.34E-4

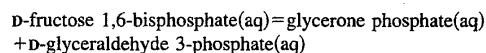
Reference: 40HER/GOR

Buffer: borate (0.8 mol dm⁻³)

pH: 7.2–7.33

Evaluation: C

Herbert *et al.* also calculated $\Delta_f H^\circ$ ($\bar{T}=311$ K, pH=7.2)=55 kJ mol⁻¹.



$\frac{T}{K}$	pH	K'_c
303.15	7	8.0E-5
311.15	7	1.5E-4
313.15	7	1.1E-4
333.15	7	4.5E-4

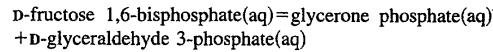
Reference: 43MEY/JUN

Method: chemical analysis and polarimetry

pH: 7

Evaluation: B

These measurements were performed in the absence of a buffer but near pH=7. From the temperature dependence of the apparent equilibrium constant we calculate $\Delta_f H^\circ$ ($\bar{T}=318$ K, pH≈7)≈48 kJ mol⁻¹.

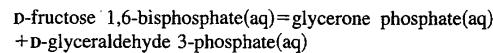


$\frac{T}{K}$	pH	K'_c
310.15	7.0	1.18E-4

Reference: 55LEH/SIC

pH: 7.0

Evaluation: C



$\frac{T}{K}$	pH	K'_c
311.15	7.1	9.3E-5

Reference: 64LOW/PAS

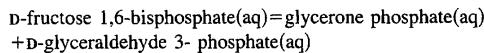
Method: fluorimetry

Buffer: imidazole (0.020 mol dm⁻³)

pH: 7.1

Cofactor(s): MgCl_2 (0.005 mol dm⁻³)

Evaluation: A



$\frac{T}{K}$	pH	$c(\text{MgCl}_2)$ mol dm ⁻³	K'_c
311.15	≈6.7	0.005	9.9E-5
311.15	≈6.7	0.050	5.6E-5

Reference: 69VEE/RAI

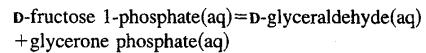
Method: spectrophotometry

Buffer: sodium phosphate (0.010 mol dm⁻³)

pH: 6.35–7.12

Cofactor(s): MgCl₂

Evaluation: A



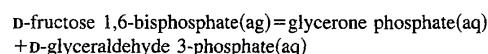
$\frac{T}{K}$	pH	K'_c
282.15	8.8	2.1E-6
296.15	8.8	4.1E-6
303.15	8.8	6.2E-6
313.15	8.8	2.8E-6

Reference: 75KUR/KON

Buffer: Tris + acetate

pH: 8.8

Evaluation: C



$\frac{T}{K}$	pH	K'_c
282.15	8.8	6.4E-5
296.15	8.8	3.90E-4
303.15	8.8	6.10E-4
313.15	8.8	5.20E-4

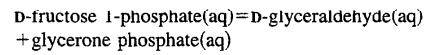
Reference: 75KUR/KON

Buffer: Tris + acetate

pH: 8.8

Evaluation: C

From the temperature dependence of the apparent equilibrium constant we calculate $\Delta_r H^\circ$ ($\bar{T}=298$ K, pH=8.8)≈53 kJ mol⁻¹.



$\frac{T}{K}$	$\frac{\Delta_r H(\text{cal})}{\text{kJ mol}^{-1}}$
298.15	-60.2

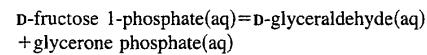
Reference: 36MEY/SCH

Method: calorimetry

Buffer: phosphate

Evaluation: C

Meyerhof and Schulz also performed measurements using the racemic mixture of D- and L-glyceraldehyde. The result was $\Delta_r H(\text{cal})=-63.2$ kJ mol⁻¹. The pH was not reported.

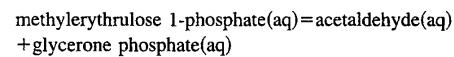


$\frac{T}{K}$	pH	K'_c
310.15	7.0	2.8E-6

Reference: 55LEH/SIC

pH: 7.0

Evaluation: C



$\frac{T}{K}$	K'_c
273.15	0.0021
293.15	0.0029
313.15	0.0067

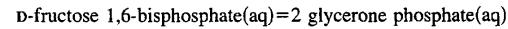
Reference: 36MEY/LOH

Method: chemical analysis

Buffer: phosphate

Evaluation: C

The approximate values of the apparent equilibrium constants given here were calculated from the results given in Meyerhof *et al.*'s Tables I and II. The pH was not reported.



$\frac{T}{K}$	pH	K'_c
266.15	7	0.00018
273.15	7	0.00030
293.15	7	0.0015
333.15	7	0.013
343.15	7	0.022

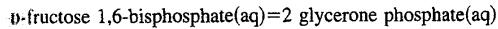
Reference: 34MEY/LOH

Method: spectrophotometry

pH: ≈7

Evaluation: C

From the temperature dependence of K'_c we calculate $\Delta_r H^\circ$ ($\bar{T}=255$ K, pH=7)=47 kJ mol⁻¹.



$\frac{T}{K}$	pH	Salt	$c(\text{salt})$ mol dm ⁻³	K'_c
273.15	7	none	...	0.00032
293.15	7	none	...	0.0015
313.15	7	none	...	0.0064
333.15	7	none	...	0.019
293.15	7	NaCl	0.043	0.0011
293.15	7	NaCl	0.21	0.00075
293.15	7	NaCl	0.90	0.00047
293.15	7	Na ₂ SO ₄	0.2	0.00055
293.15	7	MgCl ₂	0.06	0.00035
293.15	7	MgCl ₂	0.12	0.00035
293.15	7	MgCl ₂	0.24	0.00035
273.15	7	MgCl ₂	0.12	0.000071
293.15	7	MgCl ₂	0.12	0.00029
333.15	7	MgCl ₂	0.12	0.0012
273.15	7	MgCl ₂	0.006	0.00014
293.15	7	MgCl ₂	0.06	0.00037
293.15	7	MgCl ₂	0.031	0.00031
293.15	7	MgCl ₂	0.012	0.00051
293.15	7	MgCl ₂	0.006	0.00064
293.15	7	MgCl ₂	0.0025	0.0011
293.15	7	MgCl ₂	0.0012	0.0014
313.15	7	MgCl ₂	0.031	0.0014
313.15	7	MgCl ₂	0.012	0.0020
313.15	7	MgCl ₂	0.006	0.0025
313.15	7	MgCl ₂	0.0012	0.0056
333.15	7	MgCl ₂₂	0.031	0.0046
333.15	7	MgCl ₂	0.012	0.0056
333.15	7	MgCl ₂	0.006	0.0082
333.15	7	MgCl ₂	0.0012	0.015

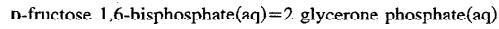
Reference: 35MEY

Method: spectrophotometry

pH: ≈7

Evaluation: C

Triose-phosphate isomerase (EC 5.3.1.1) was also present. The pH was not well controlled in this study. From the temperature dependence of the apparent equilibrium constant we calculate $\Delta_r H^\circ$ ($\bar{T}=303$ K, pH≈7)=52 kJ mol⁻¹.



$\frac{T}{K}$		$\Delta_r H(\text{cal})$ kJ mol ⁻¹
293.15		58
313.15		64

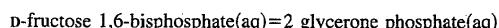
Reference: 35MEY/LOH

Method: calorimetry

Buffer: phosphate

Evaluation: C

Triose-phosphate isomerase (EC 5.3.1.1) was also present.



$\frac{T}{K}$	pH	K'_c
278.15	9.0	4.35E-4
298.15	9.0	1.82E-3
313.15	9.0	6.37E-3

Reference: 41UTT/WER

Method: chemical analysis

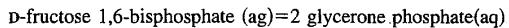
Buffer: glycine+NaOH

pH: 9.0

Cofactor(s): MgCl₂

Evaluation: B

Triose-phosphate isomerase (EC 5.3.1.1) was also present. From the temperature dependence of the apparent equilibrium constant we calculate $\Delta_r H^\circ$ ($\bar{T}=296$ K, pH=9.0)≈55 kJ mol⁻¹.



$\frac{T}{K}$	pH	K'_c
303.15	7	0.0020
311.15	7	0.0024
313.15	7	0.0030
333.15	7	0.011

Reference: 43MEY/JUN

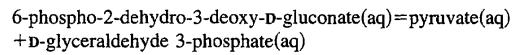
Method: chemical analysis and polarimetry

pH: 7

Evaluation: B

Triose-phosphate isomerase (EC 5.3.1.1) was also present. These measurements were performed in the absence of a buffer but near pH=7. From the temperature dependence of the apparent equilibrium constant we calculate $\Delta_r H^\circ$ ($\bar{T}=318$ K, pH≈7)=50 kJ mol⁻¹

5.7. Enzyme: 2-dehydro-3-deoxyphosphogluconate aldolase (EC 4.1.2.14)



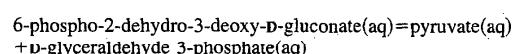
$\frac{T}{K}$	pH	K'_c
298.15	6.8	1.6E-3

Reference: 62DOU/SHU

pH: 6.8

Evaluation: C

Few details were given in this Proceedings abstract.



$\frac{T}{K}$	pH	K'_c
298.15	8.0	≈0.0012

Reference: 64MEL/WOO

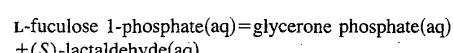
Method: spectrophotometry and radioactivity

Buffer: imidazole

pH: 8.0

Evaluation: C

5.8. Enzyme: L-fuculose-phosphate aldolase (EC 4.1.2.17)



$\frac{T}{K}$	pH	K'_c
310.15	7.2	4.6E-4

Reference: 62GHA/HEA

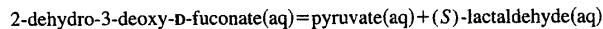
Method: spectrophotometry

Buffer: Tris (0.05 mol dm⁻³)

pH: 7.2

Evaluation: C

5.9. Enzyme: 2-dehydro-3-deoxy-L-pentonate aldolase (EC 4.1.2.18)



$\frac{T}{K}$	pH	K'_c
301.15	7.5	1.2E-4

Reference: 72DAH/AND

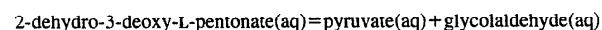
Method: enzymatic assay and spectrophotometry

Buffer: Hepes ($0.012 \text{ mol dm}^{-3}$)

pH: 7.5

Cofactor(s): MnCl_2 ($0.0038 \text{ mol dm}^{-3}$)

Evaluation: B



$\frac{T}{K}$	pH	K'_c
301.15	7.4	3.7E-4

Reference: 69DAH/AND

Method: enzymatic assay and spectrophotometry

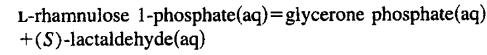
Buffer: Hepes ($0.015 \text{ mol dm}^{-3}$)

pH: 7.4

Cofactor(s): MnCl_2 ($0.005 \text{ mol dm}^{-3}$)

Evaluation: B

5.10. Enzyme: rhamnulose-1-phosphate aldolase (EC 4.1.2.19)



$\frac{T}{K}$	pH	K'_c
310.15	7.5	0.083

Reference: 65CHI/FEI

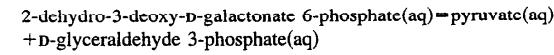
Method: spectrophotometry

Buffer: glycylglycine

pH: 7.5

Evaluation: C

5.11. Enzyme: 2-dehydro-3-deoxy-6-phosphogalactonate aldolase (EC 4.1.2.21)



$\frac{T}{K}$	pH	K'_c
298.15	6.8	3.7E-3

Reference: 62DOU/SHU

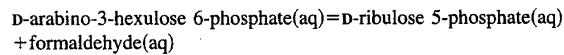
Method: spectrophotometry

pH: 6.8

Evaluation: C

Few details were given in this Proceedings abstract. Also see [66SHU].

5.12. Enzymes: D-arabino-3-hexulose phosphate formaldehyde lyase (EC 4.1.2.0)



$\frac{T}{K}$	pH	K'_c
303.15	7.0	4.0E-5

Reference: 74FER/STR

Method: enzymatic assay and spectrophotometry

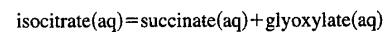
Buffer: phosphate (0.05 mol dm^{-3})

pH: 7.0

Cofactor(s): MgCl_2 ($0.005 \text{ mol dm}^{-3}$)

Evaluation: B

5.13. Enzyme: isocitrate lyase (EC 4.1.3.1)



$\frac{T}{K}$	pH	K'_c
300.15	7.6	0.029

Reference: 56SMI/STA

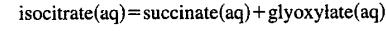
Method: chemical analysis and enzymatic assay

Buffer: Tris (0.05 mol dm^{-3})

pH: 7.6

Cofactor(s): MgCl_2 ($0.002 \text{ mol dm}^{-3}$)

Evaluation: B



$\frac{T}{K}$	pH	K'_c
300.15	7.6	0.029

Reference: 57SMI/GUN

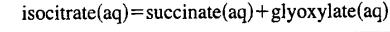
Method: spectrophotometry

Buffer: Tris ($0.067 \text{ mol dm}^{-3}$)

pH: 7.6

Cofactor(s): MgCl_2 ($0.002 \text{ mol dm}^{-3}$)

Evaluation: B



$\frac{T}{K}$	pH	K'_c
303.15	7.7	0.0023

Reference: 71WIL/ROC

Method: spectrophotometry

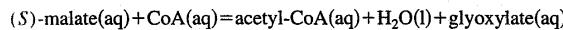
Buffer: Mops (0.10 mol dm^{-3}) + NaOH

pH: 7.7

Cofactor(s): MgCl_2 ($0.003 \text{ mol dm}^{-3}$)

Evaluation: B

Williams *et al.* also obtained K' ($T=303.15 \text{ K}$, pH=7.7) ≈ 0.0016 from kinetic data.

5.14 Enzyme: malate synthase (EC 4.1.3.2)

$\frac{T}{K}$	pH	K'_c
298.15	8.5	29

Reference: 62GOL/WAG

Method: spectrophotometry

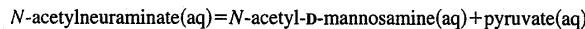
Buffer: Tris (0.04 mol dm^{-3})

pH: 8.5

Cofactor(s): MgCl_2 ($0.006 \text{ mol dm}^{-3}$)

Evaluation: C

Goldman *et al.* report $\{K'_c(\text{H}_2\text{O})\}^{-1}=0.00063$. The apparent equilibrium constant given here was calculated from this result.

5.15 Enzyme: N-acetylneuraminate lyase (EC 4.1.3.3)

$\frac{T}{K}$	pH	K'_c
310.15	7.1	0.064

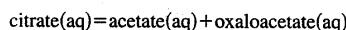
Reference: 60COM/ROS

Method: spectrophotometry

Buffer: phosphate (0.1 mol dm^{-3})

pH: 7.1

Evaluation: C

5.16. Enzyme: citrate (pro-3S)-lyase (EC 4.1.3.6)

$\frac{T}{K}$	pH	K'_c
300.15	7.6	0.64

Reference: 56SMI/STA

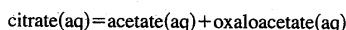
Method: chemical analysis

Buffer: Tris (0.05 mol dm^{-3})

pH: 7.6

Cofactor(s): MnSO_4 ($0.002 \text{ mol dm}^{-3}$)

Evaluation: B



$\frac{T}{K}$	pH	K'_c
303.15	7.0	0.0637

Reference: 63HAR/COL

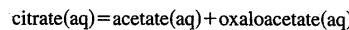
Method: spectrophotometry

Buffer: phosphate (0.13 mol dm^{-3})

pH: 7.0

Cofactor(s): Mg^{2+}

Evaluation: C



$\frac{T}{K}$	pH	$c(\text{Mg}^{2+})$ mol dm^{-2}	I_c mol dm^{-3}	K'_c
298.15	8.4	0.00031	0.1	0.118
298.15	8.4	0.00032	0.1	0.138
298.15	8.4	0.00061	0.1	0.0592
298.15	8.4	0.00062	0.1	0.123
298.15	8.4	0.00063	0.1	0.0986
298.15	8.4	0.00063	0.1	0.0739
298.15	8.4	0.00069	0.1	0.0640
298.15	8.4	0.00101	0.1	0.0667
298.15	8.4	0.00136	0.1	0.0430
298.15	8.4	0.00165	0.1	0.0457
298.15	8.4	0.00378	0.1	0.0293
298.15	8.4	0.00609	0.1	0.0218
298.15	8.4	0.00855	0.1	0.0159
298.15	8.4	0.00858	0.1	0.0178
298.15	8.4	0.00892	0.1	0.0176
298.15	8.4	0.00932	0.1	0.0243
298.15	8.4	0.00966	0.1	0.0251
298.15	8.4	0.01097	0.1	0.0184

Reference: 65STAT/DAT

Method: chemical analysis and spectrophotometry

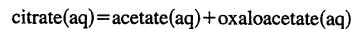
Buffer: triethanolamine + HCl

pH: 8.4

Cofactor(s): MgCl_2

Evaluation: A

Tate and Datta also calculated $K_c=0.325$ at $T=298.15 \text{ K}$, $I_c=0.1 \text{ mol dm}^{-3}$ for the chemical reference reaction: $\text{citrate}^{3-}(\text{aq}) = \text{acetate}^-(\text{aq}) + \text{oxaloacetate}^{2-}(\text{aq})$.



$\frac{T}{K}$	pH	$c(\text{MgSO}_4)$ mol dm^{-3}	I_c mol dm^{-3}	K'_c
311.15	7.0	0.010	0.25	0.168
311.15	7.0	0.00508	0.25	0.314
311.15	7.0	0.00201	0.25	0.592
311.15	7.0	0.00102	0.25	0.909
311.15	7.0	0.000116	0.25	0.877

Reference: 73GUY/GEL

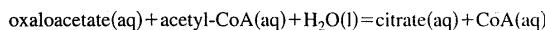
Method: spectrophotometry

Buffer: potassium phosphate ($0.025 \text{ mol dm}^{-3}$)

pH: 7.0

Cofactor(s): MgSO_4

Evaluation: A

5.17. Enzyme: citrate (*si*)-synthase (EC 4.1.3.7)


$\frac{T}{K}$	pH	$c(\text{MgSO}_4)$ mol dm ⁻³	I_c mol dm ⁻³	K'
311.15	6.39	2.77E-3	0.25	0.290E6
311.15	6.42	2.77E-3	0.25	0.418E6
311.15	6.52	2.77E-3	0.25	0.409E6
311.15	6.75	2.77E-3	0.25	0.697E6
311.15	6.79	2.77E-3	0.25	0.595E6
311.15	6.82	2.77E-3	0.25	0.773E6
311.15	6.91	2.77E-3	0.25	0.733E6
311.15	7.05	2.77E-3	0.25	1.14E6
311.15	7.20	2.77E-3	0.25	1.23E6
311.15	7.22	2.77E-3	0.25	1.80E6
311.15	7.28	2.77E-3	0.25	1.73E6
311.15	7.10	0.01E-3	0.25	1.03E6
311.15	7.10	0.10E-3	0.25	1.09E6
311.15	7.09	1.00E-3	0.25	1.29E6
311.15	7.06	10.0E-3	0.25	3.11E6
311.15	7.00	20.0E-3	0.25	5.40E6
311.15	6.94	30.0E-3	0.25	6.75E6
311.15	6.81	2.77E-3	0.25	1.31E6
311.15	7.11	2.77E-3	0.25	2.26E6
311.15	7.08	2.77E-3	0.25	1.53E6
311.15	7.35	2.77E-3	0.25	2.42E6
311.15	6.36	2.77E-3	0.25	0.992E6
311.15	6.82	2.77E-3	0.25	1.34E6

Reference: 73GUY/GEL

Method: spectrophotometry

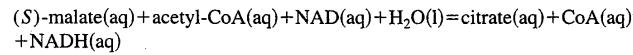
Buffer: potassium phosphate (0.025 mol dm⁻³)

pH: 6.39–7.28

Cofactor(s): MgSO₄

Evaluation: A

Guynn *et al.* also calculated K' ($T=311.15$ K, pH=7.0, $c(\text{Mg}^{2+})=0$, $I=0.25$ mol dm⁻³)=(2.24±0.11)E6.



$\frac{T}{K}$	pH	K'
295.15	6.25	0.480
295.15	7.20	10.8
295.15	7.60	101
295.15	8.20	599
295.15	8.70	5610

Reference: 52STE/OCH

Method: spectrophotometry

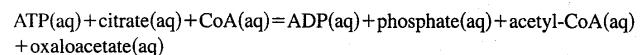
Buffer: potassium phosphate (0.025 mol dm⁻³)

pH: 6.25–8.70

Cofactor(s): MgCl₂ (0.003 mol dm⁻³)

Evaluation: B

Malate dehydrogenase (EC 1.1.1.37) was also present. The results given by Stern *et al.* in their Table III and Fig. 3 included the concentration of water as 55.5 mol dm⁻³. The apparent equilibrium constants given here have been adjusted so that the activity of water is unity.

5.18. Enzyme: ATP citrate(*pro-3S*)-lyase (EC 4.1.3.8)


$\frac{T}{K}$	pH	K'_c
298.15	8.1	≈1.2

Reference: 67PLO/CLE

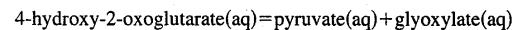
Method: radioactivity and paper chromatography

Buffer: Tris+HCl

pH: 8.1

Cofactor(s): MgCl₂

Evaluation: C

5.19. Enzyme: 4-hydroxy-2-oxoglutarate aldolase (EC 4.1.3.16)


$\frac{T}{K}$	pH	K'_c
310.15	7.5	≈0.0037

Reference: 63KUR/FUK

Method: spectrophotometry

Buffer: potassium phosphate (0.075 mol dm⁻³)

pH: 7.5

Evaluation: C

$\frac{T}{K}$	pH	K'_c
310.15	8.4	0.012

Reference: 64MAI/DEK

Method: spectrophotometry

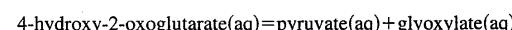
Buffer: Tris (0.1 mol dm⁻³)+HCl

pH: 8.4

Cofactor(s): MgCl₂ (0.005 mol dm⁻³)

Evaluation: C

The apparent equilibrium constant given here is based on the percentages of the various substrates reported by Maitra and Dekker. Note that the apparent equilibrium constant calculated by Maitra and Dekker is in error by a factor of 1000.



$\frac{T}{K}$	pH	K'_c
310.15	8.4	0.010

Reference: 67ROS/ADA

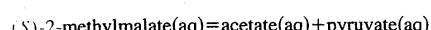
Method: enzymatic assay, spectrophotometry, and chemical analysis

Buffer: Tris (0.013 mol dm⁻³)

pH: 8.4

Evaluation: B

The apparent equilibrium constant given here was calculated from the data given in Rosso and Adams' Table III on the assumption that the total volume of solution in an experiment was 1.0 cm³.

5.20. Enzyme: citramalate lyase (EC 4.1.3.22)

$\frac{T}{K}$	pH	K'_c
298.15	7.4	8.3

Reference: 67BAR

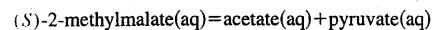
Method: spectrophotometry

Buffer: potassium phosphate (0.05 mol dm^{-3})

pH: 7.4

Cofactor(s): MgCl_2 ($0.050 \text{ mol dm}^{-3}$)

Evaluation: C



$\frac{T}{K}$	pH	I_c mol dm^{-3}	K'_c
298.15	7.4	0.845	0.151
298.15	7.4	0.648	0.212
298.15	7.4	0.399	0.188
298.15	7.4	0.306	0.207
298.15	7.4	0.212	0.211

Reference: 87BUC/MIL

Method: enzymatic assay and spectrophotometry

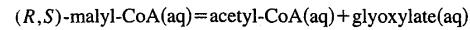
Buffer: phosphate ($0.050 \text{ mol dm}^{-3}$)

pH: 7.4

Cofactor(s): MgCl_2

Evaluation: A

Buckel and Miller also calculated $K_c = 0.32$ at $T = 298.15 \text{ K}$ and $I = 0$ for the chemical reference reaction: $(S)\text{-2-methylmalate}^{2-}(\text{aq}) = \text{acetate}^-(\text{aq}) + \text{pyruvate}^-(\text{aq})$.

5.21. Enzyme: mallyl-CoA lyase (EC 4.1.3.24)

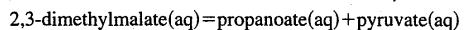
$\frac{T}{K}$	pH	K'_c
303.15	7.4	0.0029

Reference: 73HER

Buffer: Tris (0.05 mol dm^{-3}) + HCl

pH: 7.4

Evaluation: B

5.22. Enzyme: 2,3-dimethylmalate lyase (EC 4.1.3.32)

$\frac{T}{K}$	pH	K'_c
298.15	8.0	0.50

Reference: 79PIR/LIL

Method: gas chromatography and spectrophotometry

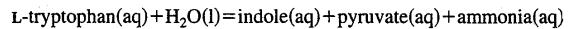
Buffer: Tris (0.10 mol dm^{-3})

pH: 8.0

Cofactor(s): MgCl_2 ($0.002 \text{ mol dm}^{-3}$)

Evaluation: B

It is not clear which optical isomer of 2,3-dimethylmalate is the active one, although the (*2R,3S*) form seems likely.

5.23. Enzyme: tryptophanase (EC 4.1.99.1)

$\frac{T}{K}$	pH	I_m mol kg^{-1}	K'_m
287.45	7.98	0.727	0.83E-4
287.45	7.98	0.726	0.89E-4
292.15	7.90	0.726	1.37E-4
292.15	7.91	0.725	1.54E-4
298.15	7.76	0.723	2.35E-4
298.15	7.77	0.722	2.28E-4
298.15	7.84	1.47	2.40E-4
298.15	7.85	1.47	2.77E-4
298.15	7.89	0.494	2.12E-4
298.15	7.97	0.494	2.22E-4
298.15	7.93	0.933	2.65E-4
298.15	7.93	0.934	2.61E-4
298.15	8.85	0.491	2.59E-4
298.15	8.85	0.491	2.23E-4
304.25	7.71	0.721	4.24E-4
304.25	7.71	0.721	3.59E-4
310.15	7.81	0.720	5.98E-4
310.15	7.81	0.720	5.71E-4
316.15	7.94	0.697	9.59E-4
316.15	7.92	0.703	10.1E-4

Reference: 94TEW/GOL

Method: HPLC

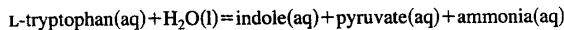
Buffer: potassium phosphate (0.10 mol kg^{-1})

pH: 7.76–8.85

Cofactor(s): pyridoxal 5-phosphate

Evaluation: A

Tewari and Goldberg also calculated $K_c = (1.05 \pm 0.13) \times 10^{-4}$, $\Delta_r G^\circ = (22.71 \pm 0.33) \text{ kJ mol}^{-1}$, $\Delta_r H^\circ = (62.0 \pm 2.3) \text{ kJ mol}^{-1}$, and $\Delta_r S^\circ = (132 \pm 8) \text{ J K}^{-1} \text{ mol}^{-1}$ at $T = 298.15 \text{ K}$ and $I = 0$ for the chemical reference reaction: L-tryptophan(aq) + $\text{H}_2\text{O(l)}$ = indole(aq) + pyruvate⁻(aq) + NH_4^+ (aq).



$\frac{T}{\text{K}}$	pH	I_m mol kg ⁻¹	$\Delta_r H(\text{cal})$ kJ mol ⁻¹
298.15	7.82	0.30	63.2

Reference: 94TEW/GOL

Method: calorimetry

Buffer: potassium phosphate (0.10 mol kg⁻¹)

pH: 7.82

Evaluation: A

Tewari and Goldberg also calculated $\Delta_r H^\circ = (62.0 \pm 2.3)$ kJ mol⁻¹ at $T = 298.15$ K and $I = 0$ for the chemical reference reaction:
 $\text{L-tryptophan(aq)} + \text{H}_2\text{O(l)} = \text{indole(aq)} + \text{pyruvate}^-(\text{aq}) + \text{NH}_4^+(\text{aq})$.

5.24. Enzyme: fumurate hydratase (EC 4.2.1.2)



$\frac{T}{\text{K}}$	pH	K'
298.15	6.81	3.1
298.15	7.12	3.3

Reference: 31BOR/SCH

Method: electrochemistry

pH: 6.81–7.12

Evaluation: C



$\frac{T}{\text{K}}$	pH	K'
278.15	6.8	8.4
288.15	6.8	6.5
298.15	6.8	5.1
311.15	6.8	3.8
327.15	6.8	2.1

Reference: 34JAC

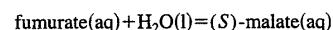
Method: polarimetry

Buffer: barbital

pH: 6.8

Evaluation: C

From the temperature dependence of the apparent equilibrium constant we calculate $\Delta_r H^\circ (T = 303 \text{ K}, \text{pH} = 6.8) = -21 \text{ kJ mol}^{-1}$.



$\frac{T}{\text{K}}$	pH	K'
293.15	7.4	4.57
303.15	7.4	3.54
313.15	7.4	3.17
323.15	7.4	2.65

Reference: 40KRE/SMY

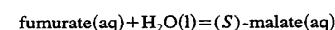
Method: manometry

Buffer: phosphate (0.1 mol dm⁻³)

pH: 7.4

Evaluation: B

From the temperature dependence of the apparent equilibrium constant we calculate $\Delta_r H^\circ (\bar{T} = 308 \text{ K}, \text{pH} = 7.4) = -14 \text{ kJ mol}^{-1}$.



$\frac{T}{\text{K}}$	pH	$\Delta_r H^\circ$ kJ mol ⁻¹
297.45	6.8	-16.0

Reference: 45OHL

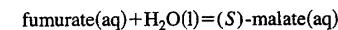
Method: calorimetry

Buffer: barbital

pH: 6.8

Evaluation: B

This same result was reported later by Ohlmeyer [46OHL] in a separate publication.



$\frac{T}{\text{K}}$	pH	K'
288.3	7.29	4.86
292.8	7.29	4.48
298.5	7.29	4.07
303.0	7.29	3.64
308.1	7.29	3.31
313.4	7.29	3.13
318.3	7.29	2.79
323.3	7.29	2.46

Reference: 48SCO/POW

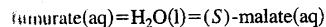
Method: chemical analysis

Buffer: phosphate

pH: 7.29

Evaluation: B

The apparent equilibrium constants given here were taken from Scott and Powell's Fig. 2. Scott and Powell calculated $\Delta_r H^\circ (\bar{T} = 305 \text{ K}, \text{pH} = 7.29) = -14.9 \text{ kJ mol}^{-1}$ from the temperature dependence of the apparent equilibrium constant.



$\frac{T}{K}$	pH	Buffer	I_c mol dm ⁻³	K'
298.15	7.3	phosphate	0.0011	4.28
298.15	7.3	phosphate	0.010	4.37
298.15	7.3	phosphate	0.049	4.44
298.15	7.3	phosphate	0.100	4.59
298.15	7.3	phosphate	0.25	4.87
298.15	4.06	?	0.05	9.49
298.15	5.00	?	0.05	5.66
298.15	5.05	?	0.05	4.66
298.15	6.00	?	0.05	4.55
298.15	6.01	?	0.05	4.42
298.15	6.06	?	0.05	4.47
298.15	7.99	?	0.05	4.31
298.15	8.00	?	0.05	4.35
298.15	9.01	?	0.05	4.43
313.0	7.30	phosphate (0.05 mol dm ⁻³)	0.10	3.11
313.0	7.30	phosphate (0.05 mol dm ⁻³)	0.10	3.13
308.3	7.30	phosphate (0.05 mol dm ⁻³)	0.10	3.81
308.3	7.30	phosphate (0.05 mol dm ⁻³)	0.10	3.72
307.7	7.30	phosphate (0.05 mol dm ⁻³)	0.10	4.01
307.6	7.30	phosphate (0.05 mol dm ⁻³)	0.10	3.97
307.5	7.30	phosphate (0.05 mol dm ⁻³)	0.10	4.00
307.5	7.30	phosphate (0.05 mol dm ⁻³)	0.10	3.93
307.3	7.30	phosphate (0.05 mol dm ⁻³)	0.10	3.97
304.1	7.30	phosphate (0.05 mol dm ⁻³)	0.10	4.68
304.1	7.30	phosphate (0.05 mol dm ⁻³)	0.10	4.74
303.6	7.30	phosphate (0.05 mol dm ⁻³)	0.10	4.69
303.6	7.30	phosphate (0.05 mol dm ⁻³)	0.10	4.85
294.1	7.30	phosphate (0.05 mol dm ⁻³)	0.10	4.88
294.1	7.30	phosphate (0.05 mol dm ⁻³)	0.10	4.82
293.4	7.30	phosphate (0.05 mol dm ⁻³)	0.10	4.99
293.3	7.30	phosphate (0.05 mol dm ⁻³)	0.10	5.04
291.2	7.30	phosphate (0.05 mol dm ⁻³)	0.10	5.70
291.2	7.30	phosphate (0.05 mol dm ⁻³)	0.10	5.54
290.8	7.30	phosphate (0.05 mol dm ⁻³)	0.10	5.68
290.8	7.30	phosphate (0.05 mol dm ⁻³)	0.10	5.61
277.8	7.30	phosphate (0.05 mol dm ⁻³)	0.10	7.22
277.8	7.30	phosphate (0.05 mol dm ⁻³)	0.10	7.37
307.5	4.91	acetate (0.20 mol dm ⁻³)	0.10	5.94
295.2	4.91	acetate (0.20 mol dm ⁻³)	0.10	6.34
292.6	4.91	acetate (0.20 mol dm ⁻³)	0.10	6.69
292.6	4.91	acetate (0.20 mol dm ⁻³)	0.10	6.88
290.8	4.91	acetate (0.20 mol dm ⁻³)	0.10	7.01
290.8	4.91	acetate (0.20 mol dm ⁻³)	0.10	7.08
290.8	4.91	acetate (0.20 mol dm ⁻³)	0.10	7.20
290.6	4.91	acetate (0.20 mol dm ⁻¹)	0.10	7.36
290.6	4.91	acetate (0.20 mol dm ⁻³)	0.10	7.51
277.9	4.91	acetate (0.20 mol dm ⁻³)	0.10	8.14

Reference: 53BOC/ALB

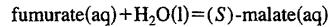
Method: spectrophotometry

Buffer: phosphate and acetate

pH: 4.06–9.01

Evaluation: A

The results given here were taken from Bock and Alberty's Figs. 1 to 3. From the temperature dependence of the apparent equilibrium constant, Bock and Alberty calculated $\Delta_f H^\circ$ ($T=294$ K, pH=7.3, $I_c=0.1$ mol dm⁻³)=-(16.6±0.4) kJ mol⁻¹ and $\Delta_f H^\circ$ ($T=293$ K, pH=4.91, $I_c=0.1$ mol dm⁻³)=-(10.0±1.7) kJ mol⁻¹.



$\frac{T}{K}$	pH	K'
298.15	7.4	4.43

Reference: 53MAS

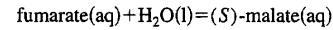
Method: spectrophotometry

Buffer: phosphate (0.067 mol dm⁻³)

pH: 6.35–7.30

Evaluation: A

The apparent equilibrium constants given here were taken from Massey's Fig. 1. Massey also calculated $\Delta_f H^\circ$ ($T=294$ K, pH≈6.6)= -15.1 kJ mol⁻¹ from the temperature dependence of the apparent equilibrium constant.



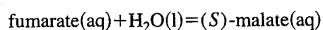
$\frac{T}{K}$	K'
298.15	4.45

Reference: 69BEN

Method: calorimetry

Evaluation: C

This result is based on unpublished data of Kitzinger and Hems. There is very little information given about the measurement.



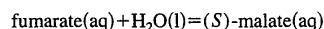
$\frac{T}{\text{K}}$	$\Delta_r H(\text{cal})$ kJ mol^{-1}
298.15	-15.5

Reference: 69BEN

Method: calorimetry

Evaluation: C

This result is based on unpublished data of Kitzinger and Hems. There is very little information given about the measurement.



$\frac{T}{\text{K}}$	pH	K'
298.15	8.2	6.2

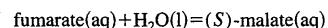
Reference: 69WAN/BAR

Method: spectrophotometry

Buffer: Tris ($0.050 \text{ mol dm}^{-3}$) + HCl

pH: 8.2

Evaluation: C



$\frac{T}{\text{K}}$	pH	K'
298.15	8.0	4.25

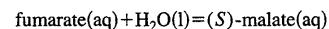
Reference: 80COO/BLA

Method: spectrophotometry

Buffer: Tris (0.1 mol dm^{-3}) + HCl

pH: 8.0

Evaluation: B



$\frac{T}{\text{K}}$	pH	I_m mol kg^{-1}	K'
298.15	7.30	0.00048	4.25
298.15	7.63	0.0035	4.16
298.15	7.99	0.0055	4.14
309.41	7.99	0.0055	3.28
320.05	7.99	0.0055	2.66

Reference: 85GAJ/GOL

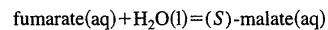
Method: gas chromatography

Buffer: Tris + HCl

pH: 7.30–7.99

Evaluation: A

Gajewski *et al.* also calculated $K=4.20$, $\Delta_r G^\circ=-3.56 \text{ kJ mol}^{-1}$, $\Delta_r H^\circ=-15.7 \text{ kJ mol}^{-1}$, and $\Delta_r C_p^\circ=-36 \text{ J K}^{-1} \text{ mol}^{-1}$ at $T=298.15 \text{ K}$ and $I=0$ for the chemical reference reaction: fumarate²⁻(aq) + H₂O(l) = (S)- malate²⁻(aq).



$\frac{T}{\text{K}}$	pH	I_m mol kg^{-1}	$\Delta_r H(\text{cal})$ kJ mol^{-1}
298.15	7.92	0.33	-16.12
298.15	8.24	0.41	-16.27
298.15	7.80	0.44	-16.26
298.15	8.04	0.56	-16.40
298.15	8.11	0.78	-16.82
313.15	7.98	0.32	-16.48
313.15	7.99	0.51	-16.84
313.15	8.03	0.76	-17.12
320.15	8.06	0.42	-16.94
320.15	8.11	0.60	-17.27
320.15	8.14	0.76	-17.44

Reference: 85GAJ/GOL

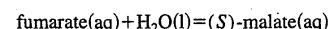
Method: calorimetry

Buffer: Tris + HCl

pH: 7.80–8.24

Evaluation: A

The calorimetric enthalpies of reaction given here were calculated from the results given in Gajewski *et al.*'s Tables 3 and 4.



$\frac{T}{\text{K}}$	pH	K'
298.15	7.6	4.3

Reference: 92KER/KER

Method: spectrophotometry

Buffer: potassium phosphate (0.05 mol dm^{-3})

pH: 7.2

Evaluation: C

5.25. Enzyme: aconitate hydratase (EC 4.2.1.3)



$\frac{T}{\text{K}}$	pH	K'
298.15	6.8	0.048

Reference: 43KRE/EGG

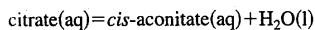
Method: polarimetry and manometry

Buffer: phosphate

pH: 6.8

Evaluation: B

The apparent equilibrium constant given here was calculated from the percentages of the substrates in solution.



$\frac{T}{K}$	pH	K'
298.15	7.4	0.032

Reference: 53KRE

Method: polarimetry and manometry

Buffer: carbonate + bicarbonate

pH: 7.4

Evaluation: A

The apparent equilibrium constant given here was calculated from the percentages of the substrates in solution.



$\frac{T}{K}$	pH	Solvent	K'
298.15	7.4	H ₂ O	0.046
298.15	7.4	D ₂ O	0.042

Reference: 66THO/NAN

Method: spectrophotometry

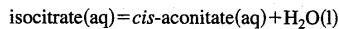
Buffer: Tris (0.033 mol dm⁻³) + HCl

pH: 7.4

Cofactor(s): MgSO₄ (0.003 mol dm⁻³)

Evaluation: C

The apparent equilibrium constant given here was calculated from the percentages of the various substrates given in Thomson *et al.*'s Table II.



$\frac{T}{K}$	pH	K'
311.15	1.4	0.69

Reference: 43KRE/EGG

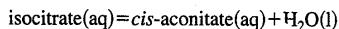
Method: polarimetry

Buffer: phosphate

pH: 6.8

Evaluation: B

The apparent equilibrium constant given here was calculated from the percentages of the substrates in solution.



$\frac{T}{K}$	pH	K'
298.15	7.4	0.47

Reference: 53KRE

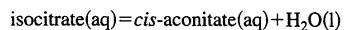
Method: polarimetry and manometry

Buffer: carbonate + bicarbonate

pH: 7.4

Evaluation: A

The apparent equilibrium constant given here was calculated from the percentages of the substrates in solution.



$\frac{T}{K}$	pH	Solvent	K'
298.15	7.4	H ₂ O	0.55
298.15	7.4	D ₂ O	0.44

Reference: 66THO/NAN

Method: spectrophotometry

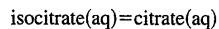
Buffer: Tris (0.033 mol dm⁻³) + HCl

pH: 7.4

Cofactor(s): MgSO₄ (0.003 mol dm⁻³)

Evaluation: C

The apparent equilibrium constant given here was calculated from the percentages of the various substrates given in Thomson *et al.*'s Table II.



$\frac{T}{K}$	pH	K'
298.15	6.8	14.4

Reference: 43KRE/EGG

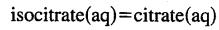
Method: polarimetry and manometry

Buffer: phosphate

pH: 6.8

Evaluation: B

The apparent equilibrium constant given here was calculated from the percentages of the substrates in solution.



$\frac{T}{K}$	pH	K'
298.15	7.4	14.7

Reference: 53KRE

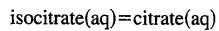
Method: polarimetry and manometry

Buffer: carbonate + bicarbonate

pH: 7.4

Evaluation: A

The apparent equilibrium constant given here was calculated from the percentages of the substrates in solution.



$\frac{T}{K}$	pH	Solvent	K'
298.15	7.4	H ₂ O	11.8
298.15	7.4	D ₂ O	10.3

Reference: 66THO/NAN

Method: spectrophotometry

Buffer: Tris (0.033 mol dm⁻³) + HCl

pH: 7.4

Cofactor(s): MgSO₄ (0.003 mol dm⁻³)

Evaluation: C

The apparent equilibrium constant given here was calculated from the percentages of the various substrates given in Thomson *et al.*'s Table II.

isocitrate(aq)=citrate(aq)

$\frac{T}{K}$	pH	$c(\text{MgSO}_4)$ mol dm ⁻³	$c(\text{CaCl}_2)$ mol dm ⁻³	K'
310.15	7.3	0	0	7.5
310.15	7.3	0.00065	0	14
310.15	7.3	0.0011	0	18
310.15	7.3	0.0016	0	25
310.15	7.3	0.0021	0	35
310.15	7.3	0.0031	0	35
310.15	7.3	0.0050	0	33
310.15	7.3	0.050	0	28
310.15	7.3	0	0.00065	11
310.15	7.3	0	0.0011	15
310.15	7.3	0	0.0016	17
310.15	7.3	0	0.0021	19
310.15	7.3	0	0.031	25
310.15	7.3	0	0.0050	26
310.15	7.3	0	0.050	30

Reference: 67ENG/DEN

Method: spectrophotometry

Buffer: triethanolamine (0.1 mol dm⁻³) + HCl

pH: 7.3

Cofactor(s): MgSO₄ and CaCl₂

Evaluation: B

The apparent equilibrium constants given here were taken from England *et al.*'s Fig. 1.

isocitrate(aq)=citrate(aq)

$\frac{T}{K}$	pH	$c(\text{Na}^+)$ mol dm ⁻³	$c(\text{MgCl}_2)$ mol dm ⁻³	I_c mol dm ⁻³	K'
298.15	7.5	0	0	0.10	9.31
298.15	7.5	0.080	0	0.10	12.2
310.15	7.5	0.080	0	0.10	10.8
298.15	7.5	0.022	0	0.044	11.4
298.15	7.5	0.048	0	0.070	11.6
298.15	7.5	0.130	0	0.15	12.7
298.15	7.5	0	0.001	0.10	21.3
298.15	7.5	0	0.002	0.10	36.1
298.15	7.5	0	0.003	0.10	46.1
298.15	7.5	0	0.004	0.10	52.5
298.15	7.5	0	0.007	0.10	64.8
298.15	7.5	0	0.010	0.10	69.0
298.15	7.5	?	0.001	0.044	27.6
298.15	7.5	?	0.002	0.044	48.3
298.15	7.5	?	0.004	0.044	66.8
298.15	7.5	?	0.008	0.044	77.7
298.15	7.5	?	0.0005	0.070	16.4
298.15	7.5	?	0.001	0.070	23.3
298.15	7.5	?	0.002	0.070	37.9
298.15	7.5	?	0.004	0.070	54.5
298.15	7.5	?	0.006	0.070	63.8
298.15	7.5	?	0.008	0.070	67.3
298.15	7.5	?	0.001	0.15	21.5
298.15	7.5	?	0.002	0.15	31.2
298.15	7.5	?	0.004	0.15	46.8
298.15	7.5	?	0.008	0.15	59.7

Reference: 69BLA

Method: spectrophotometry

Buffer: triethanolamine

pH: 7.5

Cofactor(s): MgCl₂

Evaluation: A

The results given here were taken from Blair's Tables 1 and 2 and Fig. 2.

isocitrate(aq)=citrate(aq)

$\frac{T}{K}$	pH	$c(\text{MgCl}_2)$ mol dm ⁻³	K'
311.15	7.07	0	9.89
311.15	7.07	0.009	9.92
311.15	7.07	0.047	10.0
311.15	7.07	0.094	11.0
311.15	7.07	0.230	12.2
311.15	7.07	0.470	13.9
311.15	7.07	0.700	16.1
311.15	7.07	0.930	18.0
311.15	7.07	2.30	28.8
311.15	7.07	3.60	37.2
311.15	7.07	4.50	43.6
311.15	7.07	6.90	47.0
311.15	7.07	8.50	46.0
311.15	7.07	10.0	52.3
311.15	7.07	12.3	57.9
311.15	7.07	0.000	9.83
311.15	7.07	0.047	10.6
311.15	7.07	0.700	17.1
311.15	7.07	2.30	28.8
311.15	7.07	6.90	47.5
311.15	6.64	0.000	9.51
311.15	6.64	0.047	10.3
311.15	6.64	0.700	15.8
311.15	6.64	2.30	29.5
311.15	6.64	6.90	46.0
311.15	7.45	0.000	9.48
311.15	7.45	0.047	9.95
311.15	7.45	0.700	14.8
311.15	7.45	2.30	27.6
311.15	7.45	6.90	46.6

Reference: 73VEL/GUY

Method: enzymatic assay and fluorometry

Buffer: phosphate

pH: 6.64–7.45

Cofactor(s): MgCl₂

Evaluation: A

5.26. Enzyme: 3-dehydroquinate dehydratase (EC 4.2.1.10)

3-dehydroquinate(aq)=3-dehydroshikimate(aq)+H₂O(l)

$\frac{T}{K}$	pH	K'
302.15	7.4	15

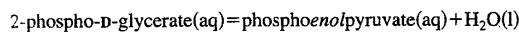
Reference: 54MIT/DAV

Method: spectrophotometry

Buffer: potassium phosphate (0.033 mol dm⁻³)

pH: 7.4

Evaluation: C

**5.27. Enzyme: phosphopyruvate hydratase
(EC 4.2.1.11)**


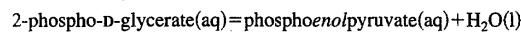
$\frac{T}{K}$	K'
273.15	0.33
293.15	0.39
313.15	0.54
333.15	0.70

Reference: 34LOH/MEY

Method: chemical analysis

Evaluation: C

The pH was not reported.

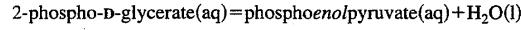


$\frac{T}{K}$	K'
301.15	2.3

Reference: 35AKA

Evaluation: C

The pH was not reported.



$\frac{T}{K}$	pH	K'
293.15	7.34	1.43

Reference: 41WAR/CHR

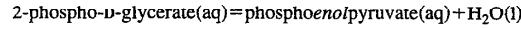
Method: spectrophotometry

Buffer: bicarbonate and phosphate

pH: 7.34

Cofactor(s): MgSO₄

Evaluation: C



$\frac{T}{K}$	K'
297.15	2.9

Reference: 49MEY/OES

Method: chemical analysis and radioactivity

Buffer: bicarbonate

Cofactor(s): Mg²⁺

Evaluation: C

The pH was not reported.



T K	pH	c(MgSO ₄) mol dm ⁻³	c(MnSO ₄) mol dm ⁻³	c(KCl) mol dm ⁻³	K'
288.0	7.5	0.008	0	0.4	3.54
299.5	7.5	0.008	0	0.4	4.55
307.5	7.5	0.008	0	0.4	5.28
298.15	6.0	0.001	0	0	2.50
298.15	6.5	0.001	0	0	3.50
298.15	7.0	0.001	0	0	4.29
298.15	7.5	0.001	0	0	4.68
298.15	8.0	0.001	0	0	5.00
298.15	8.5	0.001	0	0	5.15
298.15	6.0	0.01	0	0	2.65
298.15	6.5	0.01	0	0	3.40
298.15	7.0	0.01	0	0	3.92
298.15	7.5	0.01	0	0	4.29
298.15	8.0	0.01	0	0	4.41
298.15	8.5	0.01	0	0	4.52
298.15	6.00	0	0.001	0	2.35
298.15	6.50	0	0.001	0	3.19
298.15	6.88	0	0.001	0	3.48
298.15	7.28	0	0.001	0	3.89
298.15	8.08	0	0.001	0	4.20
298.15	5.92	0	0.002	0	2.12
298.15	6.50	0	0.002	0	2.91
298.15	6.90	0	0.002	0	3.51
298.15	7.28	0	0.002	0	3.48
298.15	8.08	0	0.002	0	3.80
298.15	5.92	0	0.005	0	2.32
298.15	6.50	0	0.005	0	2.84
298.15	6.90	0	0.005	0	3.16
298.15	7.28	0	0.005	0	3.19
298.15	8.08	0	0.005	0	3.27
298.15	6.03	0.008	0	0.4	2.63
298.15	6.04	0.008	0	0.4	2.83
298.15	6.08	0.008	0	0.4	2.45
298.15	6.08	0.008	0	0.4	2.76
298.15	6.13	0.008	0	0.4	2.76
298.15	6.13	0.008	0	0.4	2.55
298.15	6.23	0.008	0	0.4	3.21
298.15	6.23	0.008	0	0.4	3.27
298.15	6.23	0.008	0	0.4	3.32
298.15	6.29	0.008	0	0.4	3.17
298.15	6.29	0.008	0	0.4	3.21
298.15	6.34	0.008	0	0.4	3.10
298.15	6.34	0.008	0	0.4	3.21
298.15	6.34	0.008	0	0.4	3.36
298.15	6.43	0.008	0	0.4	3.78
298.15	6.49	0.008	0	0.4	3.63
298.15	6.49	0.008	0	0.4	3.68
298.15	6.55	0.008	0	0.4	3.61
298.15	6.55	0.008	0	0.4	3.84
298.15	6.74	0.008	0	0.4	4.09
298.15	6.74	0.008	0	0.4	4.37
298.15	6.78	0.008	0	0.4	4.26
298.15	6.84	0.008	0	0.4	4.00
298.15	6.84	0.008	0	0.4	4.24
298.15	6.84	0.008	0	0.4	4.26
298.15	7.00	0.008	0	0.4	4.38
298.15	7.00	0.008	0	0.4	4.43
298.15	7.00	0.008	0	0.4	4.52
298.15	7.00	0.008	0	0.4	4.68
298.15	7.12	0.008	0	0.4	4.40
298.15	7.12	0.008	0	0.4	4.80
298.15	7.16	0.008	0	0.4	4.80
298.15	7.16	0.008	0	0.4	4.97
298.15	7.38	0.008	0	0.4	4.56
298.15	7.38	0.008	0	0.4	4.64
298.15	7.38	0.008	0	0.4	4.70



$\frac{T}{K}$	pH	$c(\text{MgSO}_4)$ mol dm^{-3}	$c(\text{MnSO}_4)$ mol dm^{-3}	$c(\text{KCl})$ mol dm^{-3}	K'
298.15	7.38	0.008	0	0.4	5.00
298.15	7.46	0.008	0	0.4	4.68
298.15	7.46	0.008	0	0.4	4.82
298.15	7.46	0.008	0	0.4	4.86
298.15	7.85	0.008	0	0.4	4.71
298.15	7.85	0.008	0	0.4	5.25
298.15	7.88	0.008	0	0.4	5.06
298.15	7.88	0.008	0	0.4	5.10
298.15	7.88	0.008	0	0.4	5.25
298.15	7.88	0.008	0	0.4	5.92
298.15	7.94	0.008	0	0.4	5.15
298.15	7.94	0.008	0	0.4	5.25
298.15	8.25	0.008	0	0.4	4.97
298.15	8.25	0.008	0	0.4	5.05
298.15	8.25	0.008	0	0.4	5.08
298.15	8.25	0.008	0	0.4	5.25

Reference: 57WOL/BAL

Method: spectrophotometry

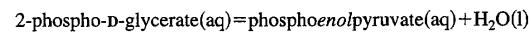
Buffer: imidazole (0.05 mol dm^{-3})

pH: 5.9 to 8.5

Cofactor(s): MgSO_4 and MnSO_4

Evaluation: A

The apparent equilibrium constants given here were taken from Wold and Ballou's Figs. 4-6 and Table IV. Wold and Ballou also calculated $K=6.3$ at $T=298.15 \text{ K}$ for the chemical reference reaction: 2-phospho-D-glycerate⁻(aq) = phosphoenolpyruvate⁻(aq) + $\text{H}_2\text{O(l)}$. From the temperature dependence of the apparent equilibrium constant we calculate $\Delta_r H^\circ (\bar{T}=298 \text{ K}, \text{pH}=7.5)=15 \text{ kJ mol}^{-1}$.



$\frac{T}{K}$	pH	K'
311.15	7.1	4.6

Reference: 64LOW/PAS

Method: fluorimetry

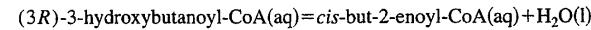
Buffer: imidazole ($0.020 \text{ mol dm}^{-3}$)

pH: 7.1

Cofactor(s): MgCl_2 ($0.005 \text{ mol dm}^{-3}$)

Evaluation: A

5.28. Enzyme: enoyl-CoA hydratase (EC 4.2.1.17)



$\frac{T}{K}$	pH	K'
298.15	7.5	0.18

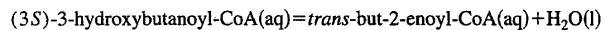
Reference: 56STE/DEL

Method: spectrophotometry

Buffer: Tris

pH: 7.5

Evaluation: B



$\frac{T}{K}$	pH	K'
298.15	7.5	0.29

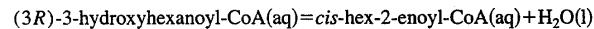
Reference: 56STE/DEL

Method: spectrophotometry

Buffer: Tris

pH: 7.5

Evaluation: C



$\frac{T}{K}$	pH	K'
298.15	7.5	0.20

Reference: 56STE/DEL

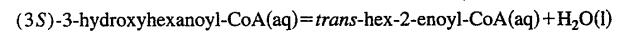
Method: spectrophotometry

Buffer: Tris

pH: 7.5

Evaluation: B

Stern and del Campillo reported $\{K' \cdot c(\text{H}_2\text{O})\}^{-1}=0.0919$. The apparent equilibrium constant given here was calculated from this result.



$\frac{T}{K}$	pH	K'
298.15	7.5	0.66

Reference: 56STE/DEL

Method: spectrophotometry

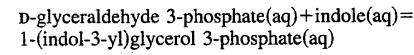
Buffer: Tris

pH: 7.5

Evaluation: B

Stern and del Campillo reported $\{K' \cdot c(\text{H}_2\text{O})\}^{-1}=0.0274$. The apparent equilibrium constant given here was calculated from this result.

5.29. Enzyme: tryptophan synthase (EC 4.2.1.20)



$\frac{T}{K}$	pH	K'_c
298.15	7.8	≈ 2300

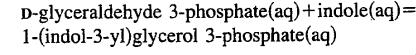
Reference: 76WEI/KIR

Method: spectrophotometry

Buffer: Tris (0.05 mol dm^{-3})

pH: 7.8

Evaluation: C



$\frac{T}{K}$	pH	$\Delta_r H^\circ(\text{cal})$ kJ mol^{-1}
298.15	7.5	-54.0

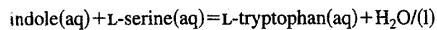
Reference: 85WIE/HIN

Method: calorimetry and spectrophotometry

Buffer: sodium diphosphate (0.1 mol dm^{-3})

pH: 7.5

Evaluation: A



T K	pH	Buffer	$\Delta_r H(\text{cal})$ kJ mol ⁻¹
283.15	7.5	sodium diphosphate (0.1 mol dm ⁻³)	78.7
298.15	7.5	sodium diphosphate (0.1 mol dm ⁻³)	80.3
298.15	7.5	Tris (0.2 mol dm ⁻³)	81.2
308.15	7.5	sodium diphosphate (0.1 mol dm ⁻³)	80.6

Reference: 85WIE/HIN

Method: calorimetry and spectrophotometry

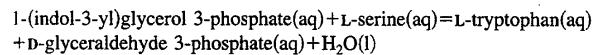
Buffer: sodium diphosphate (0.1 mol dm⁻³) and Tris (0.2 mol dm⁻³)

pH: 7.5

Cofactor(s): pyridoxal 5-phosphate

Evaluation: A

This reaction is catalyzed by the β_2 subunit of tryptophan synthase. Since the calorimetrically determined reaction enthalpies $\Delta_r H(\text{cal})$ were the same (within experimental error) in both diphosphate and Tris buffers at pH=7.5, the change in binding of hydrogen ion $\Delta_r N(\text{H}^+)$ in this biochemical reaction is zero and $\Delta_r H(\text{cal})=\Delta_r H^\circ$. From the temperature dependence of $\Delta_r H^\circ$, we calculate $\Delta_r C_p^\circ$ ($\bar{T}=296$ K, pH=7.5) ≈ 76 J K⁻¹ mol⁻¹.



T K	pH	$\Delta_r H(\text{cal})$ kJ mol ⁻¹
298.15	7.5	-13.4

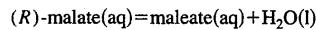
Reference: 85WIE/HIN

Method: calorimetry and spectrophotometry

Buffer: sodium diphosphate (0.1 mol dm⁻³)

pH: 7.5

Evaluation: A



T K	pH	Buffer	I_c mol dm ⁻³	K'
298.15	6.50	potassium phosphate (0.10 mol dm ⁻³) + Hepes (0.10 mol dm ⁻³) + NaOH	0.10	7.41E-4
298.15	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	4.88E-4
298.15	7.50	potassium phosphate (0.10 mol dm ⁻³) + Hepes (0.10 mol dm ⁻³) + NaOH	0.10	4.46E-4
298.15	8.00	potassium phosphate (0.10 mol dm ⁻³) + Hepes (0.10 mol dm ⁻³) + NaOH	0.10	4.28E-4
298.15	8.50	potassium phosphate (0.10 mol dm ⁻³) + Hepes (0.10 mol dm ⁻³) + NaOH	0.10	4.17E-4
288.3	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	4.39E-4
292.9	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	5.00E-4
297.8	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	5.58E-4
302.9	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	6.24E-4
307.9	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	7.20E-4
313.2	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	8.11E-4

Reference: 93WER/TWE2

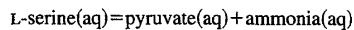
Method: HPLC

Buffer: {potassium phosphate (0.20 mol dm⁻³) + NaOH} and {potassium phosphate (0.10 mol dm⁻³) + Hepes (0.10 mol dm⁻³) + NaOH}

pH: 7.0

Evaluation: B

The apparent equilibrium constants given here were taken from van der Werf *et al.*'s Figs. 2 and 3. From the temperature dependence of K' van der Werf *et al.* calculated $\Delta_r H^\circ$ ($\bar{T}=301$ K, pH=7.0) = 18.1 kJ mol⁻¹. They also calculated $K=0.000419$ at $T=298.15$ K for the chemical reference reaction: $(R)\text{-malate}^{2-}\text{(aq)} \rightleftharpoons \text{maleate}^{2-}\text{(aq)} + \text{H}_2\text{O(l)}$.



T K	pH	$\Delta_r H(\text{cal})$ kJ mol ⁻¹
308.15	7.5	-7.3

Reference: 85WIE/HIN

Method: calorimetry

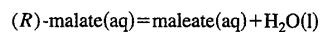
Buffer: sodium diphosphate (0.1 mol dm⁻³)

pH: 7.5

Evaluation: A

This reaction is catalyzed by the β_2 subunit of tryptophan synthase.

5.30. Enzyme: maleate hydratase (EC 4.2.1.31)



T K	pH	I_c mol dm ⁻³	K'
298.15	7.0	0.1	4.88E-4

Reference: 93WER/TWE

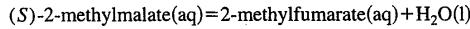
Method: HPLC

Buffer: potassium phosphate (0.025 mol dm⁻³)

pH: 7.0

Evaluation: B

**5.31. Enzyme: (S)-2-methylmalate dehydratase
(EC 4.2.1.34)**



$\frac{T}{K}$	pH	K'
298.15	8.2	0.17

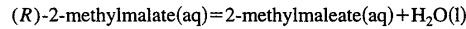
Reference: 69WAN/BAR

Method: spectrophotometry

Buffer: Tris (0.050 mol dm⁻³) + HCl

pH: 8.2

Evaluation: C



$\frac{T}{K}$	pH	Buffer	$\frac{I_c}{\text{mol dm}^{-3}}$	K'
298.15	6.00	potassium phosphate (0.10 mol dm ⁻³) + Hepes (0.10 mol dm ⁻³) + NaOH	0.10	0.0153
298.15	6.50	potassium phosphate (0.10 mol dm ⁻³) + Hepes (0.10 mol dm ⁻³) + NaOH	0.10	0.0105
298.15	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	0.00962
298.15	7.50	potassium phosphate (0.10 mol dm ⁻³) + Hepes (0.10 mol dm ⁻³) + NaOH	0.10	0.00945
298.15	8.00	potassium phosphate (0.10 mol dm ⁻³) + Hepes (0.10 mol dm ⁻³) + NaOH	0.10	0.00929
298.15	8.50	potassium phosphate (0.10 mol dm ⁻³) + Hepes (0.10 mol dm ⁻³) + NaOH	0.10	0.00917
283.5	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	0.00553
288.1	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	0.00675
292.5	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	0.00758
298.2	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	0.00864
302.8	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	0.0103
307.9	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	0.0119
313.2	7.00	potassium phosphate (0.20 mol dm ⁻³) + NaOH	0.10	0.0144

Reference: 93WER/TWE2

Method: HPLC

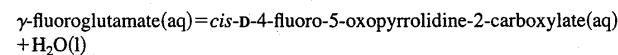
Buffer: {potassium phosphate (0.20 mol dm⁻³) + NaOH} and {potassium phosphate (0.10 mol dm⁻³) + Hepes (0.10 mol dm⁻³) + NaOH}

pH: 7.0

Evaluation: B

The apparent equilibrium constants given here were taken from van der Werf *et al.*'s Figs. 2 and 3. From the temperature dependence of K' van der Werf *et al.* calculated $\Delta_f H^\circ (\bar{T}=298 \text{ K}, \text{pH}=7.0)=22.6 \text{ kJ mol}^{-1}$. They also calculated $K=0.00909$ at $T=298.15 \text{ K}$ for the chemical reference reaction: $(R)\text{-2-methylmalate}^{2-}\text{(aq)} = 2\text{-methylmaleate}^{2-}\text{(aq)} + \text{H}_2\text{O(l)}$.

5.33. Enzyme: D-glutamate cyclase (EC 4.2.1.48)



$\frac{T}{K}$	pH	K'
299.4	7.9	0.58
303.2	7.9	0.44
305.9	7.9	0.50
313.5	7.9	0.43

Reference: 71UNK/GOL

Method: spectrophotometry

Buffer: 2-methylimidazole (0.15 mol dm⁻³) + HCl

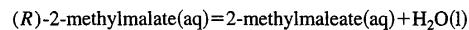
pH: 7.9

Cofactor(s): MgSO₄ (0.033 mol dm⁻³)

Evaluation: B

The apparent equilibrium constants given here were taken from Unkeless and Goldman's Table II and Fig. 4. Unkeless and Goldman also calculated $\Delta_f H^\circ (\bar{T}=307 \text{ K}, \text{pH}=7.9)=15.7 \text{ kJ mol}^{-1}$ from the temperature dependence of the apparent equilibrium constant.

**5.32. Enzyme: (R)-2-methylmalate dehydratase
(EC 4.2.1.35)**



$\frac{T}{K}$	pH	$\frac{I_c}{\text{mol dm}^{-3}}$	K'
298.15	7.0	0.1	0.0962

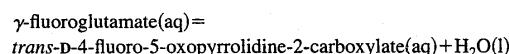
Reference: 93WER/TWE

Method: HPLC

Buffer: potassium phosphate (0.025 mol dm⁻³)

pH: 7.0

Evaluation: B



$\frac{T}{K}$	pH	K'
303.15	7.9	2

Reference: 71UNK/GOL

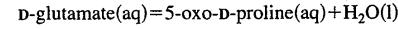
Method: spectrophotometry

Buffer: 2-methylimidazole (0.15 mol dm⁻³) + HCl

pH: 7.9

Cofactor(s): MgSO₄ (0.033 mol dm⁻³)

Evaluation: B



$\frac{T}{K}$	pH	K'
310.15	8.3	16

Reference: 63MEI/BUK

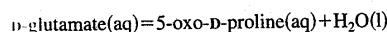
Method: chromatography and radioactivity

Buffer: Tris (0.17 mol dm⁻³) + HCl

pH: 8.3

Cofactor(s): MgCl₂ (0.033 mol dm⁻³)

Evaluation: B



$\frac{T}{K}$	pH	K'
293.4	7.9	24.3
302.3	7.9	27.0
303.2	7.9	26.8
310.9	7.9	30.3

Reference: 71UNK/GOL

Method: spectrophotometry

Buffer: 2-methylimidazole (0.15 mol dm^{-3}) + HCl

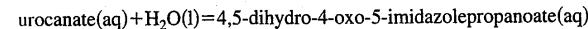
pH: 7.9

Cofactor(s): MgSO_4 ($0.033 \text{ mol dm}^{-3}$)

Evaluation: B

The apparent equilibrium constants given here were taken from Unkeless and Goldman's Table II and Fig. 4. Unkeless and Goldman also calculated $\Delta_c H^\circ$ ($\bar{T}=320 \text{ K}$, pH=7.9)=9.6 kJ mol $^{-1}$ from the temperature dependence of the apparent equilibrium constant.

5.34. Enzyme: urocanate hydratase (EC 4.2.1.49)



$\frac{T}{K}$	pH	K'
277.15	7.5	35.5
288.15	7.5	49.1
293.15	7.5	58.2
298.15	7.5	69.8
310.15	7.5	91.2

Reference: 75COH/LYN

Method: spectrophotometry

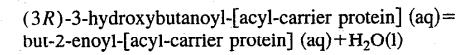
Buffer: potassium phosphate ($0.025 \text{ mol dm}^{-3}$)

pH: 7.5

Evaluation: A

Cohn *et al.* also calculated $\Delta_c H^\circ$ ($\bar{T}=294 \text{ K}$, pH=7.5)=21.8 kJ mol $^{-1}$ from the temperature dependence of K' .

5.35. Enzyme: crotonyl-[acyl-carrier-protein] hydratase (EC 4.2.1.58)



$\frac{T}{K}$	pH	K'
298.15	8.5	≈ 0.32

Reference: 68MIZ/WEE

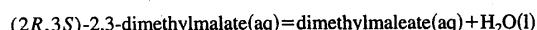
Method: enzymatic assay and spectrophotometry

Buffer: Tris (0.11 mol dm^{-3}) + HCl

pH: 8.5

Evaluation: C

5.36. Enzyme: dimethylmaleate hydratase (EC 1.4.2.1.85)



$\frac{T}{K}$	pH	K'
308.15	7.0	0.43

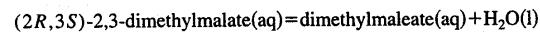
Reference: 84KOL/EGG

Method: spectrophotometry

Buffer: phosphate (0.10 mol dm^{-3})

pH: 7.0

Evaluation: B



$\frac{T}{K}$	pH	I_c mol dm^{-3}	K'
298.15	7.0	0.1	0.089

Reference: 93WER/TWE

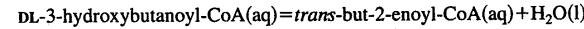
Method: HPLC

Buffer: potassium phosphate ($0.025 \text{ mol dm}^{-3}$)

pH: 7.0

Evaluation: B

5.37. Enzyme: 3-hydroxybutyryl-CoA dehydratase (EC 4.2.1.a)



$\frac{T}{K}$	pH	K'
308.15	7.4	0.28

Reference: 85COO/PRA

Method: spectrophotometry and HPLC

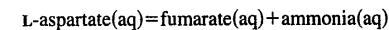
Buffer: Tris (0.10 mol dm^{-3}) + HCl

pH: 7.4

Evaluation: C

Cook *et al.* reported $\{K' \cdot c(\text{H}_2\text{O})\}^{-1} = 0.0638$. The apparent equilibrium constant given here was calculated from this result. Also see data given under EC 4.2.1.17.

5.38. Enzyme: aspartate ammonia-lyase (EC 4.3.1.1)



$\frac{T}{K}$	pH	K'_c
310.15	7.4	≈ 0.04

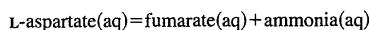
Reference: 26QUA/WOO

Method: chemical analysis

Buffer: phosphate

pH: 7.4

Evaluation: C



$\frac{T}{\text{K}}$	pH	K'_c
310.15	7.4	0.01

Reference: 29WOO

Method: chemical analysis and polarimetry

Buffer: phosphate

pH: 7.4

Evaluation: B



$\frac{T}{\text{K}}$	pH	K'_c
278.15	7.0	0.0024
310.15	7.0	0.0076

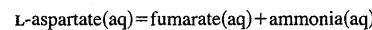
Reference: 35JAC/TAP

Method: polarimetry and chemical analysis

Buffer: barbital

pH: 7.0

Evaluation: C



$\frac{T}{\text{K}}$	pH	K'_c
310.15	6.8	0.023

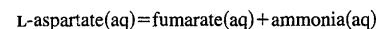
Reference: 55WIL/MCI

Method: chemical analysis

Buffer: phosphate (0.05 mol dm^{-3})

pH: 6.8

Evaluation: C



$\frac{T}{\text{K}}$	pH	K'_c
302.15	7.0	0.010
312.15	7.0	0.022

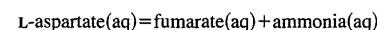
Reference: 61WIL/WIL

Method: chemical analysis

Buffer: phosphate (0.10 mol dm^{-3})

pH: 7.0

Evaluation: B

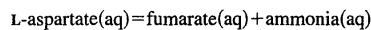


$\frac{T}{\text{K}}$	K'_c
310.15	0.020

Reference: 65SEK/SUN

Evaluation: C

The value of the apparent equilibrium constant (forward direction of the reaction) we calculate from the data given in Sekijo *et al.*'s Table 10 differs from the value calculated by Sekijo *et al.*



$\frac{T}{\text{K}}$	pH	$I_m \text{ mol dm}^{-3}$	K'_m
278.05	7.0	0.1	0.00215
288.45	7.0	0.1	0.00307
300.55	7.0	0.1	0.00511
309.45	7.0	0.1	0.00673
309.45	5.7	0.1	0.0071
309.45	7.0	0.1	0.0067
309.45	7.7	0.1	0.0064
309.45	7.0	0.10	0.0067
309.45	7.0	0.30	0.0091
309.45	7.0	0.52	0.0098
309.45	7.0	0.73	0.0098

Reference: 68BAD/MIL

Method: amino acid analysis, Conway diffusion, and UV absorption

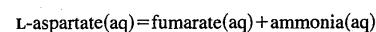
Buffer: Hepes, Mes, Tris, and phosphate

pH: 5.7–7.0

Cofactor(s): MgSO_4

Evaluation: A

Several of the apparent equilibrium constants given here were taken from Bada and Miller's Figs. 1 and 3. Bada and Miller also calculated $K_m = 0.00230$, $\Delta_f H^\circ = 26.9 \text{ kJ mol}^{-1}$, $\Delta_f S^\circ = 39.7 \text{ J K}^{-1} \text{ mol}^{-1}$, and $\Delta_f C_p^\circ = -117 \text{ J K}^{-1} \text{ mol}^{-1}$ at $T = 298.15 \text{ K}$ and $I_m = 0.1 \text{ mol kg}^{-1}$ for the chemical reference reaction: $\text{L-aspartate}^-(\text{aq}) \rightleftharpoons \text{fumarate}^{2-}(\text{aq}) + \text{NH}_4^+(\text{aq})$.



$\frac{T}{\text{K}}$	pH	K'_m
286.15	7.43	0.00331
292.15	7.39	0.00374
298.25	7.37	0.00471
304.25	7.33	0.00559
310.15	7.30	0.00507
310.25	7.29	0.00669
310.15	7.24	0.00854
316.25	7.35	0.00763

Reference: 86GOL/GAJ

Method: HPLC

Buffer: phosphate

pH: 7.25–7.43

Evaluation: A

Goldberg *et al.* also calculated $K_m = (0.00148 \pm 0.00001)$, $\Delta_f H^\circ = (24.5 \pm 1.0) \text{ kJ mol}^{-1}$, and $\Delta_f C_p^\circ = -(147 \pm 100) \text{ J K}^{-1} \text{ mol}^{-1}$ at $T = 298.15 \text{ K}$ and $I = 0$ for the chemical reference reaction: $\text{L-aspartate}^-(\text{aq}) \rightleftharpoons \text{fumarate}^{2-}(\text{aq}) + \text{NH}_4^+(\text{aq})$.



T K	pH	I_m mol kg^{-1}	$\Delta_r H'^\circ$ kJ mol^{-1}
298.15	7.27	0.133	24.8
304.15	7.28	0.133	24.7
310.15	7.30	0.066	24.2
310.15	7.32	0.134	23.7
310.15	7.28	0.182	23.8
310.15	7.27	0.334	23.1

Reference: 86GOL/GAJ

Method: calorimetry

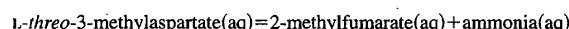
Buffer: phosphate

pH: 7.27–7.32

Evaluation: A

Goldberg *et al.* applied corrections for protonation of the buffer and for the side reaction of fumarate to malate to obtain the values of $\Delta_r H'^\circ$ given here.

5.39. Enzyme: methylaspartate ammonia-lyase (EC 4.3.1.2)



T K	pH	Buffer	K'_c
298.15	7.9	Tris (0.05 mol dm ⁻³) + HCl	0.238
298.15	9.7	ethanolamine (0.05 mol dm ⁻³)	0.306

Reference: 59BAR/SMY

Method: spectrophotometry

Buffer: {Tris (0.05 mol dm⁻³) + HCl} and ethanolamine (0.05 mol dm⁻³)

pH: 7.9–9.7

Cofactor(s): MgCl₂ (0.001 mol dm⁻³)

Evaluation: B

5.40. Enzyme: histidine ammonia-lyase (EC 4.3.1.3)



T K	pH	K'_c
298.15	8.0	3

Reference: 67WIL/HIR

Method: spectrophotometry

Buffer: Tris (0.033 mol dm⁻³) + acetate

pH: 8.0

Cofactor(s): MgSO₄ (0.001 mol dm⁻³)

Evaluation: C



T K	pH	K'_c
303.2	7.2	0.016
305.3	7.2	0.025
308.2	7.2	0.028
311.3	7.2	0.040
316.4	7.2	0.10

Reference: 73MCC/KOL

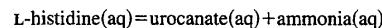
Method: spectrophotometry and thin layer chromatography

Buffer: phosphate (0.040 mol dm⁻³)

pH: 7.2

Evaluation: C

The apparent equilibrium constants given here were taken from McClard and Kolenbrander's Fig. 4. McClard and Kolenbrander also calculated $\Delta_r H'^\circ$ (pH=7.2)=108 kJ mol⁻¹ for this reaction from the temperature dependence of the apparent equilibrium constant. Larson *et al.* [93LAR/TEW] have pointed out that the results of McClard and Kolenbrander may be in error because equilibrium was not reached in their experiments.



T K	pH	I_m mol kg^{-1}	K'_m
298.25	8.41	0.167	3.01
298.25	8.70	0.145	2.35

Reference: 93LAR/TEW

Method: HPLC

Buffer: Tris (0.101 mol kg⁻¹) + HCl

pH: 8.41–8.70

Evaluation: A

Larson *et al.* also calculated $K_m = (2.7 \pm 0.7)$, $\Delta_r G^\circ = -(2.5 \pm 0.7)$ kJ mol⁻¹, $\Delta_r H^\circ = (7.6 \pm 0.8)$ kJ mol⁻¹, and $\Delta_r S^\circ = (34 \pm 4)$ J K⁻¹ mol⁻¹ at $T = 298.15$ K and $J = 0$ for the chemical reference reaction: L-histidine(aq) \rightleftharpoons urocanate⁻(aq) + NH₄⁺(aq).



T K	pH	I_m mol kg^{-1}	$\Delta_r H(\text{cal})$ kJ mol^{-1}
298.15	8.15	0.30	9.24
310.15	8.15	0.29	9.44

Reference: 93LAR/TEW

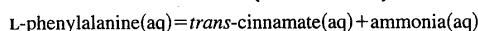
Method: calorimetry

Buffer: phosphate

pH: 8.15

Evaluation: A

**5.41. Enzyme: phenylalanine ammonia-lyase
(EC 4.3.1.5)**



$\frac{T}{K}$	pH	Buffer	K'_c
303.15	6.8	potassium phosphate (0.13 mol dm^{-3})	6.0
303.15	6.8	potassium phosphate (0.08 mol dm^{-3})	4.7
303.15	6.8	potassium phosphate (0.08 mol dm^{-3})	5.7
303.15	6.8	potassium phosphate (0.12 mol dm^{-3})	5.2
303.15	6.8	potassium phosphate (0.12 mol dm^{-3})	6.1
303.15	6.8	potassium phosphate (0.08 mol dm^{-3})	5.9
303.15	6.8	potassium phosphate (0.08 mol dm^{-3})	7.4
303.15	6.8	potassium phosphate (0.19 mol dm^{-3})	7.9
303.15	6.8	potassium phosphate (0.19 mol dm^{-3})	7.9
303.15	6.8	potassium phosphate (0.12 mol dm^{-3})	8.0
303.15	6.8	potassium phosphate (0.08 mol dm^{-3})	7.4
303.15	8.5	sodium diphosphate (0.08 mol dm^{-3})	3.9
303.15	8.5	sodium diphosphate (0.08 mol dm^{-3})	5.4
303.15	8.5	sodium diphosphate ($0.069 \text{ mol dm}^{-3}$)	6.2
303.15	8.5	sodium diphosphate ($0.084 \text{ mol dm}^{-3}$)	5.4
303.15	8.5	sodium diphosphate (0.08 mol dm^{-3})	5.5
303.15	8.5	sodium diphosphate (0.08 mol dm^{-3})	6.6
303.15	8.5	sodium diphosphate ($0.069 \text{ mol dm}^{-3}$)	6.3
303.15	8.5	sodium diphosphate ($0.084 \text{ mol dm}^{-3}$)	6.2
303.15	8.5	sodium diphosphate (0.08 mol dm^{-3})	6.7

Reference: 68HAV/HAN

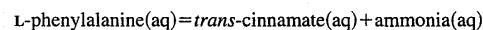
Method: chromatography

Buffer: sodium diphosphate and potassium phosphate

pH: 6.8–8.5

Evaluation: B

The apparent equilibrium constants given here were taken from Havar and Hansen's Fig. 2.



$\frac{T}{K}$	pH	K'_m
285.65	7.68	1.54
292.15	7.62	2.09
298.05	7.69	2.47
304.45	7.62	3.11
309.95	6.98	3.61
309.95	7.25	3.74
309.95	7.71	3.46
309.95	7.39	3.70
309.95	7.28	3.58
316.45	7.23	4.48

Reference: 87TEW/GAJ

Method: HPLC

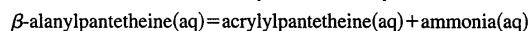
Buffer: Tris (0.1 mol dm^{-3}) + HCl

pH: 7.0–7.7

Evaluation: A

Tewari *et al.* also calculated $K_m = (1.16 \pm 0.3)$ and $\Delta_r H^\circ = (24.8 \pm 2.0) \text{ kJ mol}^{-1}$ at $T = 298.15 \text{ K}$ and $I = 0$ for the chemical reference reaction: $\text{L-phenylalanine}^+(aq) = \text{transcinnamate}^-(aq) + \text{NH}_4^+(aq)$.

**5.42. Enzyme: β -alanyl-CoA ammonia lyase
(EC 4.3.1.6)**



$\frac{T}{K}$	pH	K'_c
298.15	7.5	1.22E-6

Reference: 59VAG/EAR

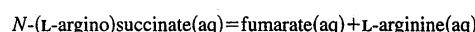
Method: spectrophotometry

Buffer: triethanolamine (0.05 mol dm^{-3}) + HCl

pH: 7.5

Evaluation: B

5.43. Enzyme: arginosuccinate lyase (EC 4.3.2.1)



$\frac{T}{K}$	pH	K'_c
311.15	7.5	0.0114

Reference: 53RAT/ANS

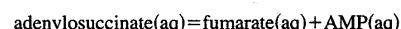
Method: chemical analysis

Buffer: potassium phosphate (0.05 mol dm^{-3})

pH: 7.5

Evaluation: B

5.44. Enzyme: adenylosuccinate lyase (EC 4.3.2.2)



$\frac{T}{K}$	pH	K'_c
298.15	7.0	0.012

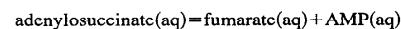
Reference: 55CAR/COH

Method: spectrophotometry

pH: 7.0

Evaluation: C

The temperature was assumed to be 298.15 K. This was a preliminary result.



$\frac{T}{K}$	pH	K'_c
308.15	6.0	0.0063
308.15	6.75	0.0074
308.15	7.0	0.0068

Reference: 56CAR/COH

Method: spectrophotometry

Buffer: sodium phosphate (0.05 mol dm^{-3})

pH: 6.0–7.0

Evaluation: B

N-(5-amino-1- β -D-ribosylimidazol-4-yl-carbonyl)-L-aspartic acid
 $5'$ -phosphate(aq)=fumarate(aq)+5-amino-1- β -D-ribosylimidazole-
 4-carboxamide $5'$ -phosphate(aq)

$\frac{T}{K}$	pH	K'_c
310.15	7.2	0.0023

Reference: 59MIL/LUK

Method: spectrophotometry, chromatography, and radioactivity

Buffer: phosphate ($0.023 \text{ mol dm}^{-3}$)

pH: 7.2

Evaluation: C

5.45. Enzyme: ureidoglycolate lyase (EC 4.3.2.3)

(-)ureidoglycolate(aq)=glyoxylate(aq)+urea(aq)

$\frac{T}{K}$	pH	K'_c
303.15	8.4	0.15
303.15	8.8	0.14

Reference: 65GAU/WOL

Method: chemical analysis

Buffer: sodium barbital ($0.040 \text{ mol dm}^{-3}$)

pH: 8.4–8.8

Evaluation: C

(-)ureidoglycolate(aq)=glyoxylate(aq)+urea(aq)

$\frac{T}{K}$	pH	K'_c
303.15	7.5	0.14

Reference: 67TRI/VOG

Method: polarimetry

Buffer: triethanolamine (0.07 mol dm^{-3})+HCl

pH: 7.5

Cofactor(s): $ZnSO_4$ ($0.010 \text{ mol dm}^{-3}$)

Evaluation: B

5.46. Enzyme: lactoylglutathione lyase (EC 4.4.1.5)

(*R*)-*S*-lactoylglutathione(aq)=glutathione (reduced)(aq)+methylglyoxal(aq)

$\frac{T}{K}$	pH	K'_c
303.15	7.0	9.0E-5

Reference: 83SEL/MAN

Method: spectrophotometry

Buffer: sodium phosphate (0.1 mol dm^{-3})

pH: 7.0

Evaluation: C

The approximate value of the apparent equilibrium constant given here was obtained from kinetic data.

5.47. Enzyme: adenylate cyclase (EC 4.6.1.1)

ATP(aq)=adenosine 3',5'-(cyclic)phosphate(aq)+diphosphate(aq)

$\frac{T}{K}$	pH	K'_c
298.15	7.3	0.065

Reference: 71HAY/GRE

Method: enzymatic assay

Buffer: Tris ($0.0590 \text{ mol dm}^{-3}$)+HCl

pH: 7.3

Cofactor(s): $MgSO_4$ ($0.010 \text{ mol dm}^{-3}$)

Evaluation: B

ATP(aq)=adenosine 3',5'-(cyclic)phosphate(aq)+diphosphate(aq)

$\frac{T}{K}$	pH	K'_c
298.15	7.3	0.255

Reference: 71TAK/KUR

Method: chromatography, fluorometry, and radioactivity

Buffer: Tris ($0.050 \text{ mol dm}^{-3}$)+HCl

pH: 7.3

Cofactor(s): $MgSO_4$ ($0.005 \text{ mol dm}^{-3}$)

Evaluation: C

ATP(aq)=adenosine 3',5'-(cyclic)phosphate(aq)+diphosphate(aq)

$\frac{T}{K}$	pH	K'_c
298.15	6.2	0.029
298.15	6.5	0.036
298.15	6.8	0.049
298.15	7.0	0.065
298.15	7.3	0.116
298.15	7.7	0.270

Reference: 74KUR/TAK

Method: fluorometry, spectrophotometry, and radioactivity

Buffer: Hepes ($0.060 \text{ mol dm}^{-3}$)+KOH

pH: 6.2–7.7

Cofactor(s): $MgSO_4$ ($0.005 \text{ mol dm}^{-3}$)

Evaluation: A

6. List of Substances with Chemical Abstract Service (CAS) Registry Numbers With Cross References to Enzyme Commission Numbers

Substance	CAS Registry Number ^a	Enzyme Commission Numbers
acetaldehyde	75-07-0	4.1.2.4, 4.1.2.13
acetate	64-19-7	4.1.3.6, 4.1.3.22
acetyl-coenzyme A	102029-73-2	4.1.3.2, 4.1.3.7, 4.1.3.8, 4.1.3.24
<i>N</i> -acetyl-D-mannosamine	3615-17-6	4.1.3.3
<i>N</i> -acetylneuraminate	131-48-6	4.1.3.3
cis-aconitate	585-84-2	4.2.1.3
acrylylpantetheine	4515-49-5	4.3.1.6
adenosine 3',5'-(cyclic)phosphate	60-92-4	4.6.1.1
adenosine 5'-diphosphate	58-64-0	4.1.3.8
adenosine 5'-monophosphate	18422-05-4	4.3.2.2
adenosine 5'-triphosphate	56-65-5	4.1.3.8, 4.6.1.1
adenylosuccinate	19046-78-7	4.3.2.2
β -alanyl pantetheine	110180-69-3	4.3.1.6
5-amino-1- β -D-ribosylimidazole-4-carboxamide 5'-phosphate	3031-94-5	4.3.2.2
<i>N</i> -(5-amino-1- β -D-ribosylimidazol-4-yl-carbonyl)-L-aspartic acid 5'-phosphate	3031-95-6	4.3.2.2
ammonia	1336-21-6	4.1.99.1, 4.2.1.20, 4.3.1.1, 4.3.1.2, 4.3.1.3, 4.3.1.5, 4.3.1.6
D-arabino-3-hexulose 6-phosphate	53010-97-2	4.1.2.a
L-arginine	74-79-3	4.3.2.1
<i>N</i> -(L-argino)succinate	2387-71-5	4.3.2.1
L-aspartate	56-84-8	4.3.1.1
but-2-enoyl-[acyl-carrier protein]	b	4.2.1.58
cis-but-2-enoyl-coenzyme A	6244-90-2	4.2.1.17
trans-but-2-enoyl-coenzyme A	102680-35-3	4.2.1.17, 4.2.1.a
carbon dioxide	124-38-9	4.1.1.32, 4.1.1.38, 4.1.1.39
trans-cinnamate	140-10-3	4.3.1.5
citrate	77-92-9	4.1.3.6, 4.1.3.7, 4.1.3.8, 4.2.1.3
coenzyme A	85-61-0	4.1.3.2, 4.1.3.7, 4.1.3.8
2-dehydro-3-deoxy-D-fuconate	35902-09-1	4.1.2.18
2-dehydro-3-deoxy-D-galactonate 6-phosphate	32120-43-7	4.1.2.21
2-dehydro-3-deoxy-L-pentonate	3495-27-0	4.1.2.18
3-dehydroquinate	10534-44-8	4.2.1.10
3-dehydroshikimate	2922-42-1	4.2.1.10
2-deoxy-D-ribose 5-phosphate	102916-66-5	4.1.2.4
5,6-dideoxyfructose 1-phosphate	29024-88-2	4.1.2.13
4,5-dihydro-4-oxo-5-imidazolepropanoate	17340-16-8	4.2.1.49
2,3-dimethylmalate	31519-20-7	4.1.3.32
(2 <i>R</i> ,3 <i>S</i>)-2,3-dimethylmalate	73522-92-6	4.2.1.85
dimethylmaleate	624-48-6	4.2.1.85
diphosphate	2466-09-3	4.1.1.38, 4.6.1.1
erythrulose 1-phosphate	2547-10-6	4.1.2.2
γ -fluoroglutamate	2708-77-2	4.2.1.48
<i>cis</i> -D-4-fluoro-5-oxopyrrolidine-2-carboxylate	160705-72-6	4.2.1.48
<i>trans</i> -D-4-fluoro-5-oxopyrrolidine-2-carboxylate	160705-73-7	4.2.1.48
formaldehyde	50-00-0	4.1.2.2, 4.1.2.a
D-fructose 1,6-bisphosphate	488-69-7	4.1.2.13
D-fructose 1-phosphate	103213-46-3	4.1.2.13
L-fuculose 1-phosphate	92418-41-2	4.1.2.17
fumurate	110-17-8	4.2.1.2, 4.3.1.1, 4.3.2.1, 4.3.2.2
D-glutamate	6893-26-1	4.2.1.48
glutathione (reduced)	70-18-8	4.4.1.5
D-glyceraldehyde	453-17-8	4.1.2.13
D-glyceraldehyde 3-phosphate	142-10-9	4.1.2.4, 4.1.2.13, 4.1.2.14, 4.1.2.21, 4.2.1.20
glycerone phosphate	102783-56-2	4.1.2.2, 4.1.2.13, 4.1.2.17, 4.1.2.19
glycolaldehyde	23147-58-2	4.1.2.18
glyoxylate	298-12-4	4.1.3.1, 4.1.3.2, 4.1.3.16, 4.1.3.24, 4.3.2.3

6. List of Substances with Chemical Abstract Service (CAS) Registry Numbers
With Cross References to Enzyme Commission Numbers—Continued

Substance	CAS Registry Number ^a	Enzyme Commission Numbers
H ₂ O	7732-18-5	4.1.1.32, 4.1.1.38, 4.1.3.2, 4.1.3.7, 4.1.99.1, 4.2.1.2, 4.2.1.3, 4.2.1.10, 4.2.1.11, 4.2.1.17, 4.2.1.20, 4.2.1.31, 4.2.1.34, 4.2.1.35, 4.2.1.48, 4.2.1.49, 4.2.1.58, 4.2.1.85, 4.2.1.a
<i>cis</i> -hex-2-enoyl-coenzyme A	160705-74-8	4.2.1.17
<i>trans</i> -hex-2-enoyl-coenzyme A	10018-93-6	4.2.1.17
L-histidine	71-00-1	4.3.1.3
(3 <i>R</i>)-3-hydroxybutanoyl-[acyl-carrier protein]	b	4.2.1.58
D,L-3-hydroxybutanoyl-coenzyme A	103404-51-9	4.2.1.a
(3 <i>R</i>)-3-hydroxybutanoyl-coenzyme A	21804-29-5	4.2.1.17
(3 <i>S</i>)-3-hydroxybutanoyl-coenzyme A	22138-45-0	4.2.1.17
(3 <i>R</i>)-3-hydroxyhexanoyl-coenzyme A	117249-47-5	4.2.1.17
(3 <i>S</i>)-3-hydroxyhexanoyl-coenzyme A	79171-47-4	4.2.1.17
4-hydroxy-2-oxoglutarate	1187-99-1	4.1.3.16
indole	120-72-9	4.1.99.1, 4.2.1.20
1-(indol-3-yl)glycerol 3-phosphate	4220-97-7	4.2.1.20
inosine 5'-diphosphate	81012-88-6	4.1.1.32
inosine 5'-triphosphate	35908-31-7	4.1.1.32
isocitrate	320-77-4	4.1.3.1, 4.2.1.3
(<i>S</i>)-lactaldehyde	3913-64-2	4.1.2.17, 4.1.2.18, 4.1.2.19
(<i>R</i>)-S-lactoylglutathione	41656-56-8	4.4.1.5
(<i>R</i>)-malate	636-61-3	4.2.1.31
(<i>S</i>)-malate	97-67-6	4.1.3.2, 4.1.3.7, 4.2.1.2
maleate	110-16-7	4.2.1.31
(<i>R,S</i>)-mallyl-coenzyme A	2043-93-8	4.1.3.24
L-threo-3-methylaspartate	31571-69-4	4.3.1.2
methylerthrulose 1-phosphate	160705-75-9	4.1.2.13
2-methylfumarate	498-24-8	4.2.1.34, 4.3.1.2
methylglyoxal	78-98-8	4.4.1.5
(<i>R</i>)-2-methylmalate	6236-10-8	4.2.1.35
(<i>S</i>)-2-methylmalate	6236-09-5	4.1.3.22, 4.2.1.34
2-methylmaleate	498-23-7	4.2.1.35
β-nicotinamide-adenine dinucleotide (oxidized)	53-84-9	4.1.3.7
β-nicotinamide-adenine dinucleotide (reduced)	606-68-8	4.1.3.7
oxaloacetate	328-42-7	4.1.1.32, 4.1.1.38, 4.1.3.6, 4.1.3.7, 4.1.3.8
5-oxo-D-proline	4042-36-8	4.2.1.48
L-phenylalanine	63-91-2	4.3.1.5
phosphate	10049-21-5	4.1.1.38, 4.1.3.8
6-phospho-2-dehydro-3-deoxy-D-gluconate	27244-54-8	4.1.2.14
2-phospho-D-glycerate	70195-25-4	4.2.1.11
3-phospho-D-glycerate	80731-10-8	4.1.1.39
phosphoenolpyruvate	4265-07-0	4.1.1.32, 4.1.1.38, 4.2.1.11
propanoate	79-09-04	4.1.3.32
propionaldehyde	123-38-6	4.1.2.13
pyruvate	127-17-3	4.1.2.14, 4.1.2.18, 4.1.2.21, 4.1.3.3, 4.1.3.16, 4.1.3.22, 4.1.3.32, 4.1.99.1, 4.2.1.20
L-rhamnulose 1-phosphate	444-09-7	4.1.2.19
D-ribulose 1,5-biphosphate	108321-97-7	4.1.1.39
D-ribulose 5-phosphate	108321-99-9	4.1.2.a
L-serine	56-45-1	4.2.1.20
succinate	110-15-6	4.1.3.1
L-tryptophan	73-22-3	4.1.99.1, 4.2.1.20
urea	57-13-6	4.3.2.3
(-)ureidoglycolate	103192-53-6	4.3.2.3
urocanate	104-98-3	4.2.1.49, 4.3.1.3

^aIn some cases the CAS registry number refers to a salt of the substance.

^bIn the absence of an amino acid sequence, no CAS registry number is assigned to this substance.

7. Abbreviations

ADP	adenosine 5'-diphosphate	IDP	inosine 5'-diphosphate
AMP	adenosine 5'-monophosphate	ITP	inosine 5'-triphosphate
ATP	adenosine 5'-triphosphate	Mes	2-(<i>N</i> -morpholino)ethanesulfonic acid
CoA	coenzyme A	Mops	3-(<i>N</i> -morpholino)propanesulfonic acid
Hepes	<i>N</i> -(2-hydroxyethyl)piperazine- <i>N'</i> -2-ethanesulfonic acid	NAD	β -nicotinamide-adenine dinucleotide (oxidized)
		NADH	β -nicotinamide-adenine dinucleotide (reduced)
		Tris	tris(hydroxymethyl)aminomethane

8. Glossary of Symbols

Symbol	Name	Unit
<i>c</i>	concentration	mol dm^{-3}
<i>c</i> [°]	standard concentration ($c^{\circ} = 1 \text{ mol dm}^{-3}$)	mol dm^{-3}
$\Delta_f C_p^{\circ}$	standard heat capacity of reaction at constant pressure	$\text{J K}^{-1} \text{ mol}^{-1}$
$\Delta_f G^{\circ}$	standard Gibbs energy of reaction	kJ mol^{-1}
$\Delta_f G'^{\circ}$	standard transformed Gibbs energy of reaction	kJ mol^{-1}
$\Delta_f H^{\circ}$	standard enthalpy of reaction	kJ mol^{-1}
$\Delta_f H'^{\circ}$	standard transformed enthalpy of reaction	kJ mol^{-1}
$\Delta_i H(\text{cal})$	calorimetrically determined enthalpy of reaction	kJ mol^{-1}
<i>I_c</i>	ionic strength, concentration basis	mol dm^{-3}
<i>I_m</i>	ionic strength, molality basis	mol kg^{-1}
<i>K</i>	equilibrium constant ^a	dimensionless
<i>K'</i>	apparent equilibrium constant ^a	dimensionless
<i>m</i>	molality	mol kg^{-1}
<i>m</i> [°]	standard molality ($m^{\circ} = 1 \text{ mol kg}^{-1}$)	mol kg^{-1}
$\Delta_f N(\text{H}^+)$	change in binding of hydrogen ion in a biochemical reaction	dimensionless
pH	$-\log_{10}[c(\text{H}^+)/c^{\circ}]^b$	dimensionless
pX	$-\log_{10}[c(X)/c^{\circ}]$	dimensionless
$\Delta_f S^{\circ}$	standard entropy of reaction	$\text{J K}^{-1} \text{ mol}^{-1}$
<i>T</i>	thermodynamic temperature	K
<i>x</i>	mole fraction	dimensionless

^aWhen needed, a subscript *c*, *m*, or *x* is added to these quantities to designate a concentration, molality, or mole fraction basis.

^bThis is an approximate definition. The IUPAC Green Book (I. Mills, T. Cvitaš, K. Homann, N. Kallay, and K. Kuchitsu, "Quantities, Units and Symbols in Physical Chemistry," Blackwell Scientific Publications, Oxford, 1993) contains a discussion of the operational definition of pH.

9. Reference Codes and References in the Table

26QUA/WOO	Quastel, J. H.; Woolf, B.; Biochem. J.; 20 , 545 (1926).	60COM/ROS	Comb, D. G.; Roseman, S.; J. Biol. Chem.; 235 , 2529 (1960).
29WOO	Woolf, B.; Biochem. J.; 23 , 472 (1929).	60PRI/HOR	Pricer, W. E.; Horecker, B. L.; J. Biol. Chem.; 235 , 1292 (1960).
31BOR/SCH	Borsook, H.; Schott, H. F.; J. Biol. Chem.; 92 , 559 (1931).	61WIL/WIL	Wilkinson, J. S.; Williams, V. R.; Arch. Biochem. Biophys.; 93 , 80 (1961).
34JAC	Jacobsohn, K. P.; Biochem. Z.; 274 , 167 (1934).	62DOU/SHU	Doudoroff, M.; Shuster, C. W.; Bacteriol. Proc. p. 44 (1962).
34LOH/MEY	Lohman, K.; Meyerhof, O.; Biochem. Z.; 273 , 60 (1934).	62GHA/HEA	Ghalambor, M. A.; Heath, E. C.; J. Biol. Chem.; 237 , 2427 (1962).
34MEY/LOH	Meyerhof, O.; Lohmann, K.; Biochem. Z.; 271 , 89 (1934).	62GOL/WAG	Goldman, D. S.; Wagner, M. J.; Biochim. Biophys. Acta; 65 , 297 (1962).
35AKA	Akano, R.; Biochem. Z.; 280 , 110 (1935).	63HAR/COL	Harvey, R. J.; Collins, E. B.; J. Biol. Chem.; 238 , 2648 (1963).
35JAC/TAP	Jacobsohn, K. P.; Tapadinhas, J.; Biochem. Z.; 282 , 374 (1935).	63KUR/FUK	Kuratomi, K.; Fukunaga, K.; Biochim. Biophys. Acta; 78 , 617 (1963).
35MEY	Meyerhof, O.; Biochem. Z.; 277 , 77 (1935).	63MEI/BUK	Meister, A.; Bukenberger, M. W.; Strassburger, M.; Biochem. Z.; 338 , 217 (1963).
35MEY/LOH	Meyerhof, O.; Lohman, K.; Biochem. Z.; 275 , 430 (1935).	64LOW/PAS	Lowry, O. H.; Passonneau, J. V.; J. Biol. Chem.; 239 , 31 (1964).
36MEY/SCH	Meyerhof, O.; Schulz, W.; Biochem. Z.; 289 , 87 (1936).	64MAI/DEK	Maitra, U.; Dekker, E. E.; J. Biol. Chem.; 239 , 1485 (1964).
36MEY/LOH	Meyerhof, O.; Lohmann, K.; Schuster, Ph.; Biochem. Z.; 286 , 301 (1936).	64MEL/WOO	Meloche, H. P.; Wood, W. A.; J. Biol. Chem.; 239 , 3511 (1964).
40HER/GOR	Herbert, D.; Gordon, H.; Subrahmanyam, V.; Green, D. E.; Biochem. J.; 34 , 1108 (1940).	65CHI/FEI	Chiu, T. H.; Feingold, D. S.; Biochem. Biophys. Res. Commun.; 19 , 511 (1965).
40KRE/SMY	Krebs, H. A.; Smyth, D. H.; Evans, E. A., Jr.; Biochem. J.; 34 , 1041 (1940).	65GAU/WOL	Gaudy, E. T.; Wolfe, R. S.; J. Bacteriol.; 90 , 1531 (1965).
41UTT/WER	Utter, M. F.; Werkman, C. H.; J. Bacteriol.; 42 , 665 (1941).	65SEK/SUN	Sekijo, C.; Sunhara, N.; Iwakumo, S.; Hakko To Taisha; 2 , 139 (1965).
41WAR/CHR	Warburg, O.; Christian, W.; Biochem. Z.; 310 , 384 (1941).	65TAT/DAT	Tate, S. S.; Datta, S. P.; Biochem. J.; 94 , 470 (1965).
43KRE/EGG	Krebs, H. A.; Eggleston, L. V.; Biochim. J.; 37 , 334 (1943).	66SHU	Shuster, C. W.; Methods Enzymol.; 9 , 524 (1966).
43MEY/JUN	Meyerhof, O.; Junowicz Kocholaty, R.; J. Biol. Chem.; 149 , 71 (1943).	66THO/NAN	Thomson, J. F.; Nance, S. L.; Bush, K. J.; Szczepanik, P. A.; Arch. Biochem. Biophys.; 117 , 65 (1966).
45OHL	Ohlmeyer, P.; Hoppe-Seyler's Z. Physiol. Chem.; 282 , 37 (1945).	66WOO/DAV	Wood, H. G.; Davis, J. J.; Lochmüller, H.; J. Biol. Chem.; 241 , 5692 (1966).
46OHL	Ohlmeyer, P.; Z. Naturforsch.; 1 , 30 (1946).	67BAR	Barker, H. A.; Arch. Mikrobiol.; 59 , 4 (1967).
48SCO/POW	Scott, E. M.; Powell, R.; J. Am. Chem. Soc.; 70 , 1104 (1948).	67ENG/DEN	England, P. J.; Denton, R. M.; Randle, P. J.; Biochem. J.; 105 , 32c (1967).
49MEY/OES	Meyerhof, O.; Oesper, P.; J. Biol. Chem.; 179 , 1371 (1949).	67GRO	Groth, D. P.; J. Biol. Chem.; 242 , 155 (1967).
52STE/OCH	Stern, J. R.; Ochoa, S.; Lynen, F.; J. Biol. Chem.; 198 , 313 (1952).	67PLO/CLE	Plowman, K. M.; Cleland, W. W.; J. Biol. Chem.; 242 , 4239 (1967).
53BOC/ALB	Bock, R. M.; Alberty, R. A.; J. Am. Chem. Soc.; 75 , 1921 (1953).	67ROS/ADA	Rosso, R. G.; Adams, E.; J. Biol. Chem.; 242 , 5524 (1967).
53KRE	Krebs, H. A.; Biochem. J.; 54 , 78 (1953).	67TRI/VOG	Trijbels, F.; Vogels, G. D.; Biochim. Biophys. Acta; 132 , 115 (1967).
53MAS	Massey, V.; Biochem. J.; 53 , 72 (1953).	67WIL/HIR	Williams, V. R.; Hiroms, J. M.; Biochim. Biophys. Acta; 139 , 214 (1967).
53RAT/ANS	Ratner, S.; Anslow, W. P.; Jr.; Petrack, B.; J. Biol. Chem.; 204 , 115 (1953).	68BAD/MIL	Bada, J. L.; Miller, S. L.; Biochemistry; 7 , 3403 (1968).
54CHA	Charlampous, F. C.; J. Biol. Chem.; 211 , 249 (1954).	68HAV/HAN	Haviv, E. A.; Hanson, K. R.; Biochemistry; 7 , 1904 (1968).
54MIT/DAV	Mitsuhashi, S.; Davis, B. D.; Biochim. Biophys. Acta; 15 , 54 (1954).	68MIZ/WEE	Mizugaki, M.; Weeks, G.; Toomey, R. E.; Wakil, S. J.; J. Biol. Chem.; 243 , 3661 (1968).
54UTT/KUR	Utter, M. F.; Kurahashi, K.; J. Biol. Chem.; 207 , 821 (1954).	69BEN	Benzinger, T. H.; in "A Laboratory Manual of Analytical Methods of Protein Chemistry," P. Alexander and H. P. Lundgren, eds.; Pergamon Press, New York (1969), pp. 93-149.
55CAR/COH	Carter, C. E.; Cohen, L. H.; J. Biol. Chem.; 77 , 499 (1955).	69BLA	Blair, J. McD.; Eur. J. Biochem.; 8 , 287 (1969).
55WIL/MCI	Williams, V. R.; McIntyre, R. T.; J. Biol. Chem.; 217 , 467 (1955).	69DAH/AND	Dahms, A. S.; Anderson, R. L.; Biochem. Biophys. Res. Comm.; 36 , 809 (1969).
56CAR/COH	Carter, C. E.; Cohen, L. H.; J. Biol. Chem.; 222 , 17 (1956).	69VEE/RAI	Veech, R. L.; Rajzman, L.; Dalziel, K.; Krebs, H. A.; Biochem. J.; 115 , 837 (1969).
56KIT/HOR	Kitzinger, C.; Horecker, B. L.; Weisbach, A.; Abstr. 20th Int. Physiol. Congress, Brussels (1956).	69WAN/BAR	Wang, C. C.; Barkers, H. S.; J. Biol. Chem.; 244 , 2516 (1969).
56SMI/STA	Smith, R. A.; Stamer, J. R.; Gunslaus, I. C.; Biochim. Biophys. Acta; 19 , 567 (1956).	71HAY/GRE	Hayaishi, O.; Greengard, P.; Colowick, S. P.; J. Biol. Chem.; 246 , 5840 (1971).
56STE/DEL	Stern, J. R.; del Campillo, A.; J. Biol. Chem.; 218 , 985 (1956).	71TAK/KUR	Takai, K.; Kurashina, Y.; Suzuki, C.; Okamoto, H.; Ueki, A.; Hayaishi, O.; J. Biol. Chem.; 246 , 5843 (1971).
57WOL/BAL	Wold, F.; Ballou, C. E.; J. Biol. Chem.; 227 , 301 (1957).	71UNK/GOL	Unkeless, J. C.; Goldman, P.; J. Biol. Chem.; 246 , 2354 (1971).
57SMI/GUN	Smith, R. A.; Gunsalus, I. C.; J. Biol. Chem.; 229 , 305 (1957).	71WIL/ROC	Williams, J. O.; Roche, T. E.; McFadden, B. A.; Biochemistry; 10 , 1384 (1971).
59BAR/SMY	Barker, H. A.; Smyth, R. D.; Wilson, R. M.; Weissbach, H.; J. Biol. Chem.; 234 , 320 (1959).	72DAH/AND	Dahms, A. S.; Anderson, R. L.; J. Biol. Chem.; 247 , 2238 (1972).
59MIL/LUK	Miller, R. W.; Lukens, L. N.; Buchanan, J. M.; J. Biol. Chem.; 234 , 1806 (1959).		
59VAG/EAR	Vagelos, P. R.; Earl, J. M.; Stadtman, E. R.; J. Biol. Chem.; 234 , 490 (1959).		

- 73GUY/GEL Guynn, R. W.; Gelberg, H. J.; Veech, R. L.; *J. Biol. Chem.*; **248**, 6957 (1973).
- 73HER Hersh, L. B.; *J. Biol. Chem.*; **248**, 7295 (1973).
- 73MCC/KOL McClard, R. W.; Kolenbrander, H. M.; *Can. J. Biochem.*; **51**, 556 (1973).
- 73VEL/GUY Veloso, D.; Guynn, R. W.; Oskarsson, M.; Veech, R. L.; *J. Biol. Chem.*; **248**, 4811 (1973).
- 74FER/STR Ferenci, T.; Ström, T.; Quayle, J. R.; *Biochem. J.*; **144**, 477 (1974).
- 74KUR/TAK Kurashina, Y.; Takai, K.; Suzuki-Hori, C.; Okamoto, H.; Hayaishi, O.; *J. Biol. Chem.*; **249**, 4824 (1974).
- 75COH/LYN Cohn, M. S.; Lynch, M. C.; Phillips, A. T.; *Biochim. Biophys. Acta*; **337**, 444 (1975).
- 75KUR/KON Kurski, M. D.; Kondratyook, T. P.; Litvinenko, L. T.; Kosterin, S. O.; Dopov. Akad. Nauk. Ukr. RSR Ser. B; Geol. Geofiz. Khiml. Biol.; **3**, 256 (1975).
- 76WEI/KIR Weischet, W. O.; Kirschner, K.; *Eur. J. Biochem.*; **65**, 365 (1976).
- 79PIR/I.II. Pirzer, P.; Lill, U.; Eggerer, H.; *Hoppe-Seyler's Z. Physiol. Chem.*; **360**, 1693 (1979).
- 80COO/BLA Cook, P. F.; Blanchard, J. S.; Cleland, W. W.; *Biochemistry*; **19**, 4853 (1980).
- 82DEM de Meis, L.; *Ann. N. Y. Acad. Sci.*; **402**, 535 (1982).
- 83SEL/MAN Sellin, S.; Mannervik, B.; *J. Biol. Chem.*; **258**, 8872 (1983).
- 84KOL/EGG Kollmann-Koch, A.; Eggerer, H.; *Hoppe-Seyler's Z. Physiol. Chem.*; **365**, 847 (1984).
- 85COO/PRA Cook, L.; Prasad, M. R.; Vieth, R.; Cinti, D. L.; *Arch. Biochem. Biophys.*; **236**, 26 (1985).
- 85GAJ/GOL Gajewski, E.; Goldberg, R. N.; Steckler, D. K.; *Biophys. Chem.*; **22**, 187 (1985).
- 85WIE/HIN Wiesinger, H.; Hinz, H. J.; *Arch. Biochem. Biophys.*; **242**, 440 (1985).
- 86GOL/GAJ Goldberg, R. N.; Gajewski, E.; Steckler, D. K.; Tewari, Y. B.; *Biophys. Chem.*; **24**, 13 (1986).
- 87BUC/MIL Buckel, W.; Miller, S.; *Eur. J. Biochem.*; **164**, 565 (1987).
- 87TEW/GAJ Tewari, Y. B.; Gajewski, E.; Goldberg, R. N.; *J. Phys. Chem.*; **91**, 904 (1987).
- 92KER/KER Keruchenko, J. S.; Keruchenko, I. D.; Gladilin, K. L.; Zaitsev, V. N.; Chirgadze, N. Y.; *Biochim. Biophys. Acta*; **1122**, 85 (1992).
- 93LAR/TEW Larson, J. W.; Tewari, Y. B.; Goldberg, R. N.; *J. Chem. Thermodyn.*, **25**, 73 (1993).
- 93WER/TWE van der Werf, M. J.; van den Tweel, W. J. J.; Hartmans, S.; *Appl. Environ. Microbiol.*; **59**, 2823 (1993).
- 93WER/TWE2 van der Werf, M. J.; van den Tweel, W. J. J.; Hartmans, S.; *Eur. J. Biochem.*; **217**, 1011 (1993).
- 94TEW/GOL Tewari, Y. B.; Goldberg, R. N.; *J. Solution Chem.*, **23**, 167 (1994).