

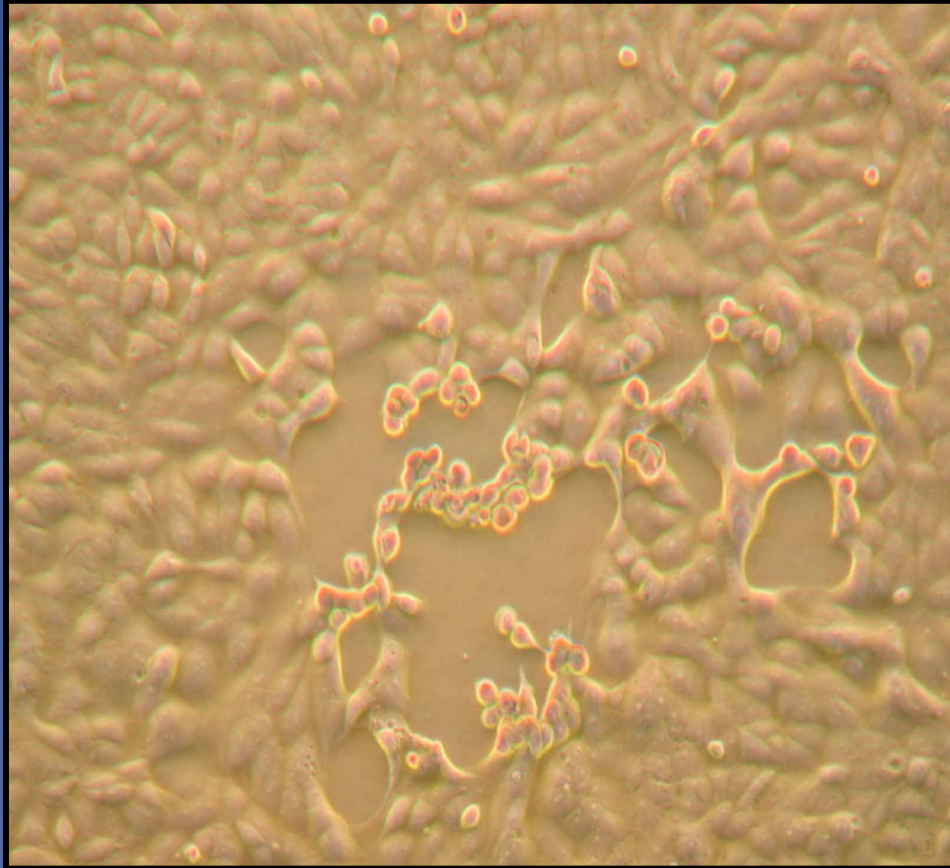
SARS Diagnostics

Preparedness

- **Performance of current diagnostic tests**
 - **Real-time RT-PCR**
 - **Serology**
- **New diagnostic tools**
- **Optimal specimen types and timing**
- **Quality assessment**
- **Other respiratory pathogens – “rule-out testing”**

SARS Diagnostics

Cell culture



P Rollin, Special Pathogens Branch

BSL-3 Activity

Restricted culture range

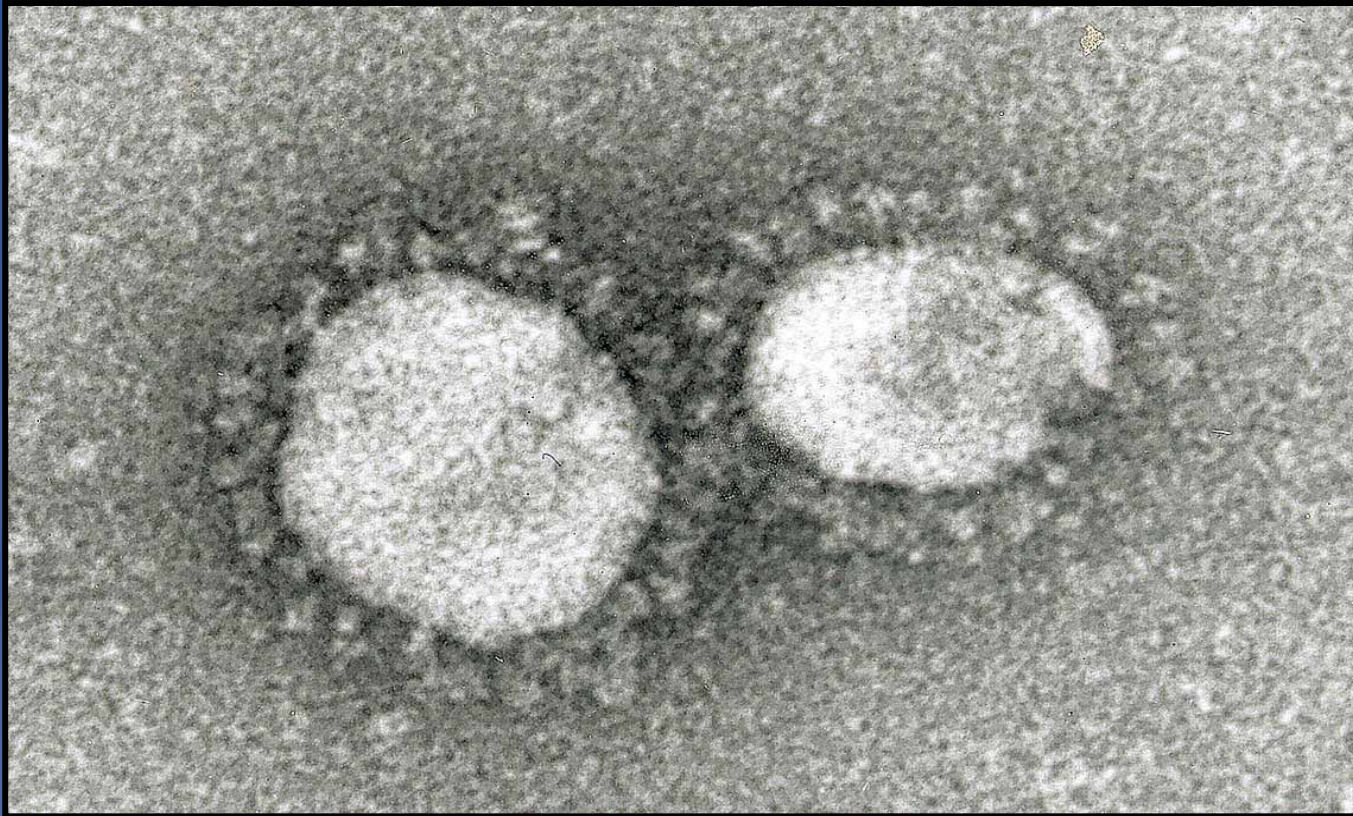
Vero E6 cells

CPE:

- focal
- cell rounding
- retractile appearance

***SARS* Diagnostics**

Electron Microscopy



C Humphrey, Pathology Activity Program

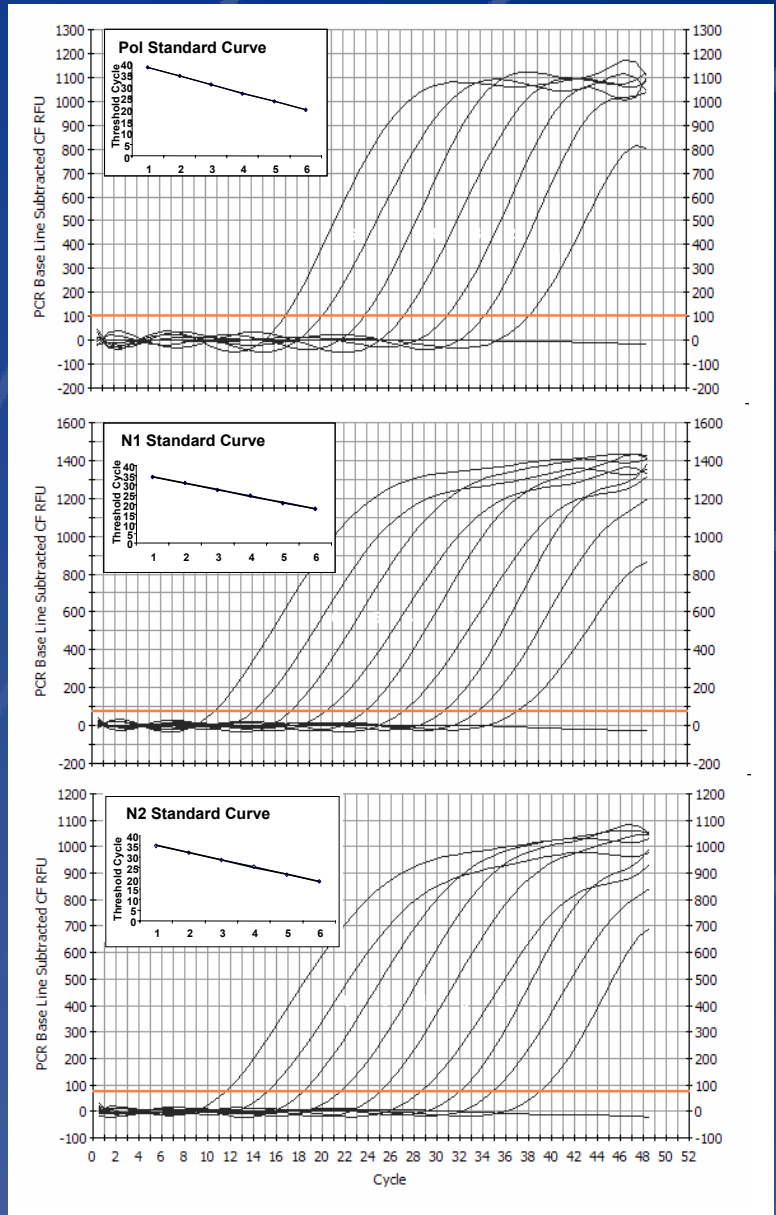
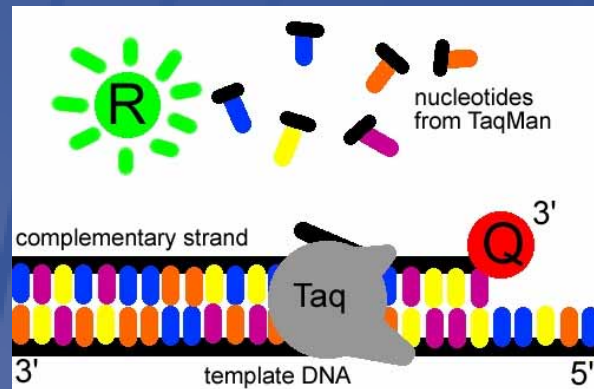
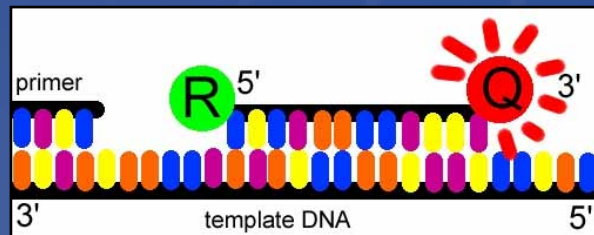
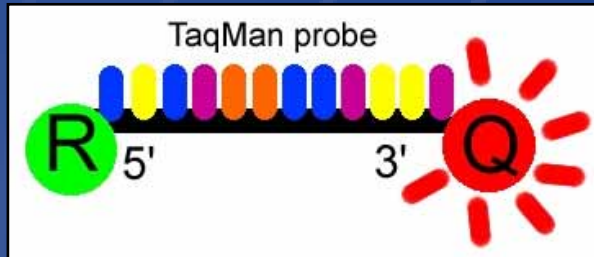
SARS Diagnostics

Real-time RT-PCR

- **Conventional vs Real-time RT-PCR (TaqMan™)**
 - increased sensitivity (1-10 transcript copies)
 - increased speed/throughput
 - quantitative
 - reduced risk of amplicon contamination
- **Multiple genetic targets**
 - nucleocapsid and polymerase genes
 - amplification of 2 of the 3 targets required for a positive test

SARS Diagnostics

Real-time RT-PCR



SARS Diagnostics

RT-PCR – Interpretation of Test Results

- **Potential for false negative results**
 - **low titer virus in respiratory secretions in first few days after onset of illness**
- **Potential for false positive results**
 - **contamination from previously amplified DNA**
 - **cross-contamination between specimens**
- **A positive test result should be considered provisional until confirmed by independent testing**
- **A negative test result does not rule out SARS and should not affect patient management decisions**

SARS Diagnostics

RT-PCR – Interpretation of Test Results

- **Confirmation of a positive SARS RT-PCR test (specimen)**
 - repeat the RT-PCR from new aliquot of the original sample
 - if positive, have the sample tested in a second laboratory
- **Positive SARS diagnostic test finding (patient)**
 - at least 2 different clinical specimens
 - the same specimen type collected on 2 or more days

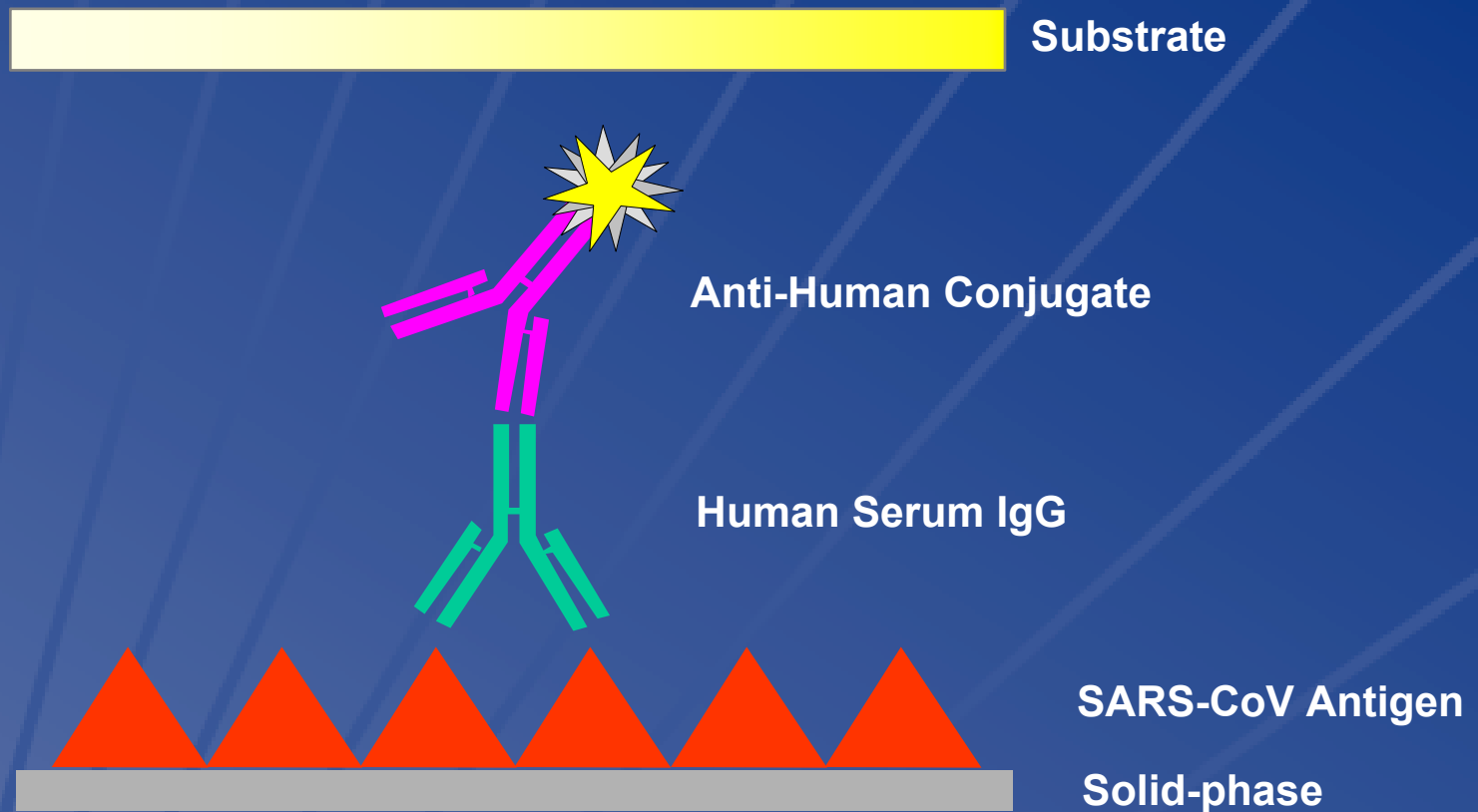
SARS Diagnostics

Serology – Current EIA

- **Serology appears to be highly specific**
 - **no reactions with other documented CoV infections (OC43 and 229E)**
 - **no reactions with “normal” blood donors (U.S. and Hong Kong populations)**
- **Serology can be positive in as few as 8 to 10 days after onset of symptoms**
- **Serology cannot be considered negative until >28 days after onset of symptoms**

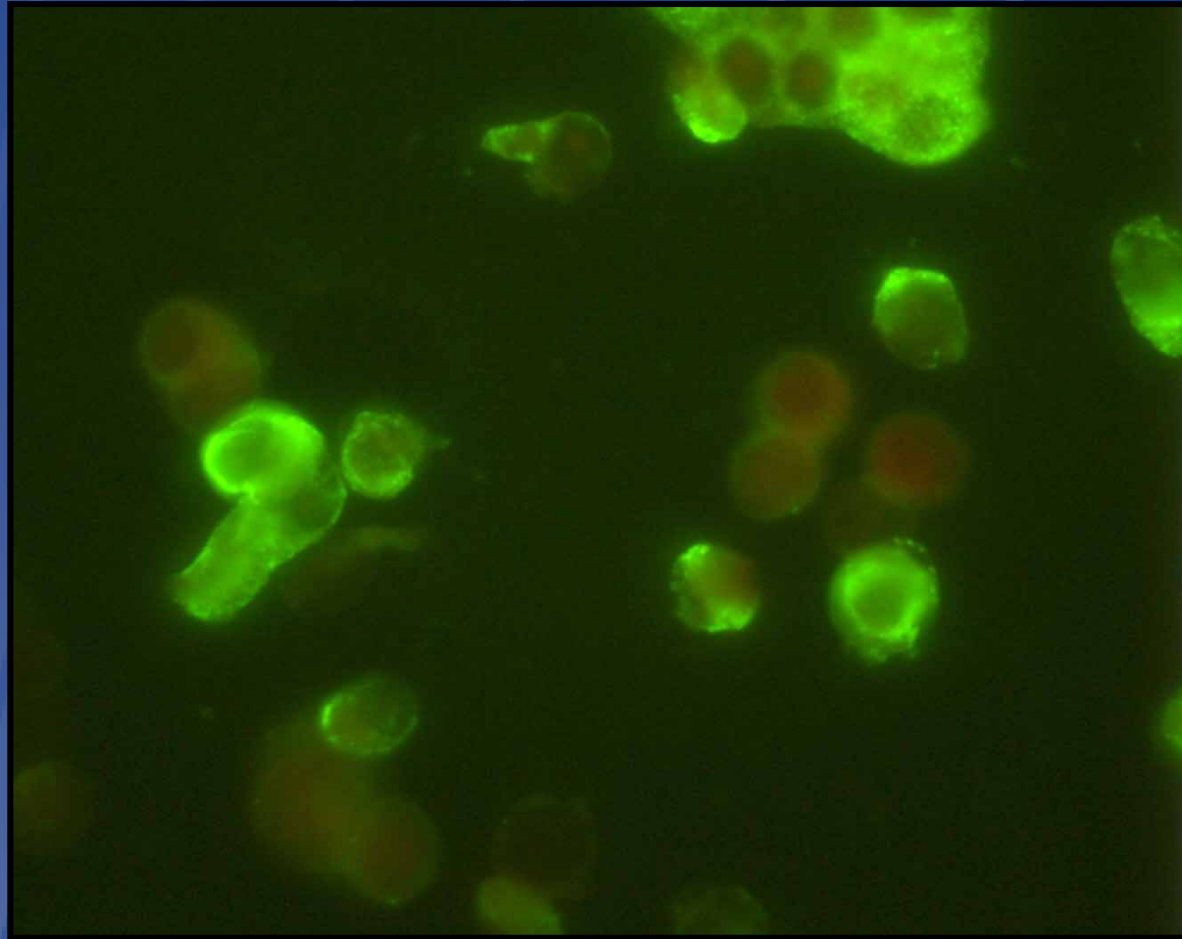
SARS Diagnostics

Antibody tests: Enzyme immunoassay



SARS Diagnostics

Antibody tests: Immunofluorescence Assay



SARS Diagnostics

Serology - New assays

- **Native virus vs recombinant antigens**
 - **nucleocapsid, spike, and membrane proteins**
 - **safety, standardization, and sensitivity**
 - **need to rule out cross-reactions with other human coronaviruses**
- **IgM assays**
 - **IgM antibodies may be detectable earlier in the course of infection**
 - **Transient response**
- **Neutralization and other immunological markers**

SARS Diagnostics

Specimen Selection and Timing

- **Respiratory tract specimens**
 - **LRT > URT**
 - **Sputum > NP aspirates > NP/OP swabs**
 - **More sample**
 - **Multiple samples**
- **Others specimens**
 - **Blood plasma**
 - **Stool**
- **Timing of specimen collection**

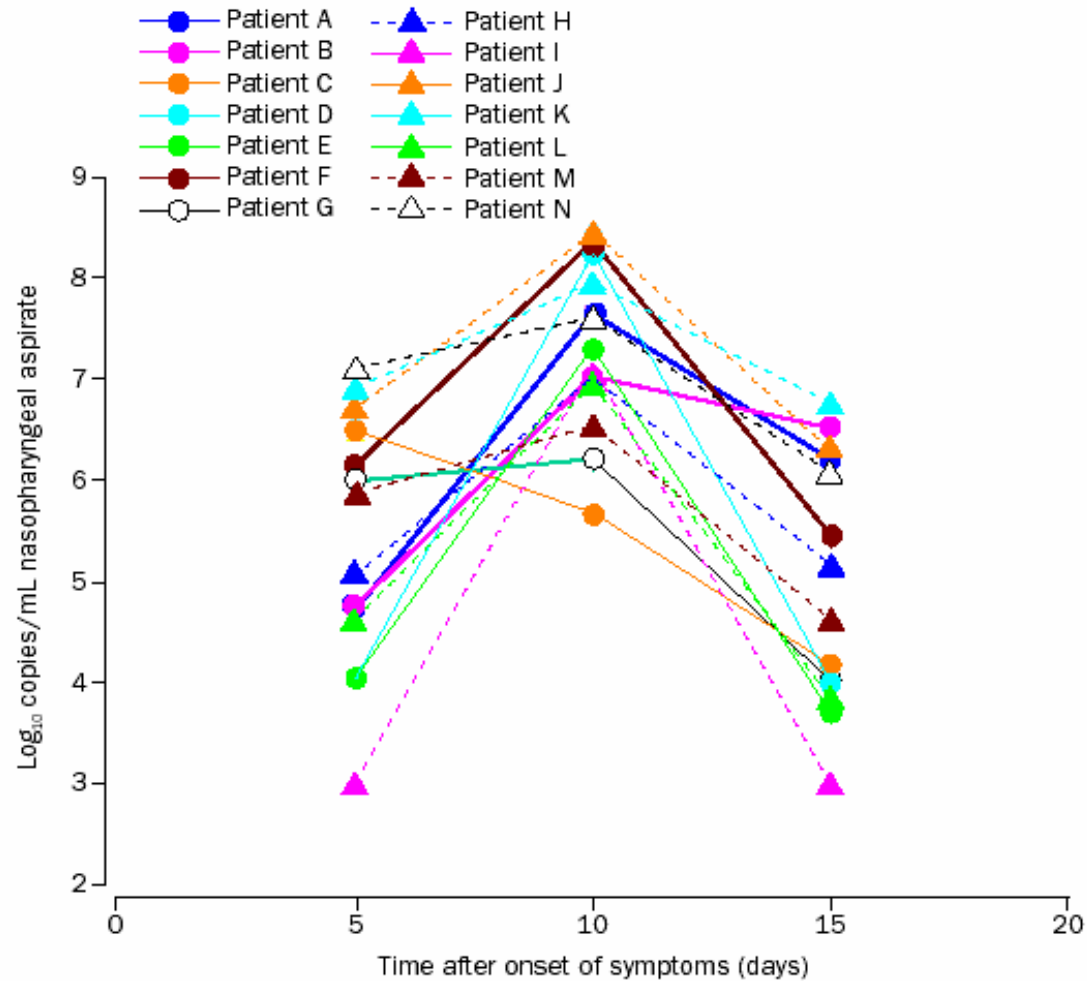
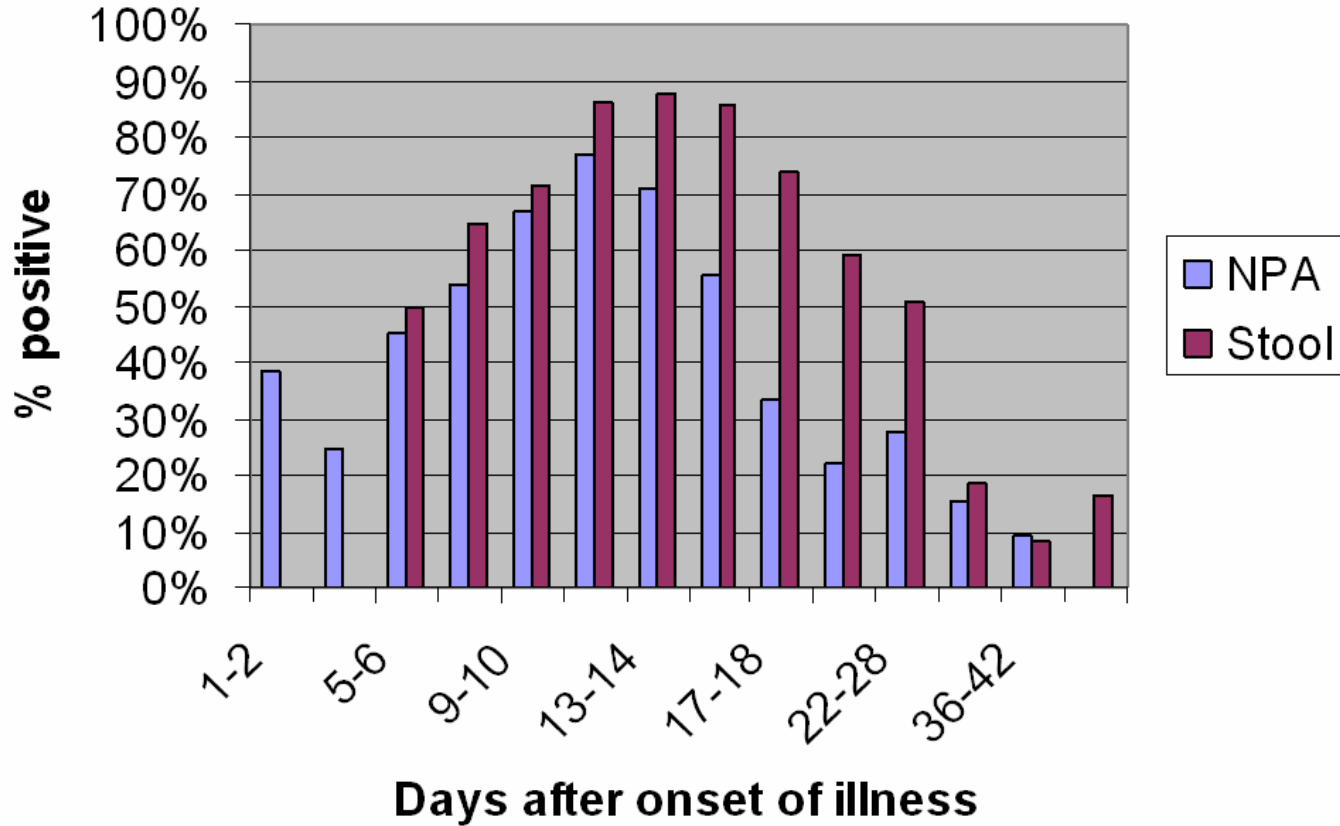


Figure 4: **Sequential quantitative RT-PCR for SARS-associated coronavirus in nasopharyngeal aspirates of 14 SARS patients**

Peiris et al: Lancet, May 24, 2003

Clinical SARS: % positive by RT-PCR

Faeces	9	10	10	17	21	29	64	42	19	34	67	38	12	6
NPA	39	57	62	41	42	26	34	27	9	9	18	13	11	



Peiris: personal communication

SARS Diagnostics

Specimen Selection and Timing

Specimen	<1 week post symptom onset	1 - 3 weeks post symptom onset	>3 weeks post symptom onset
Serum (separator tube)	++	++	++
Blood plasma (EDTA)	++	+	-
Respiratory (BAL, sputum, nasal aspirate & wash, np/op swabs)	+	++	+
Stool	+	++	++

SARS Diagnostics

Quality Assessment

- **QA CDC**
 - **Standardized test controls**
 - **Internal CDC confirmatory testing**
 - **External WHO quality assurance study**
- **QA LRN & APHL**
 - **Confirmatory testing**
 - **Proficiency testing**

SARS Diagnostics

Other Respiratory Pathogens – “rule-out testing”

Other respiratory pathogens, U.S. SARS surveillance, March-July, 2003.

M. pneumoniae	C. pneumoniae	L. pneumophila	Influenza A or B	hMPV	hPIV 1,2, 3	RSV	Adeno	Picornavirus (rhinovirus)
22/200 (11%)	2/197 (1%)	0/196 (0%)	15/140 (11%)	9/150 (6%)	10/150 (7%)	1/150 (0.7%)	7/150 (5%)	18/61 (30%)

Schrag SJ et al. SARS surveillance in the United States during the Emergency Public Health Response, March-July, 2003. EID (In press).

SARS Diagnostics

Other Respiratory Pathogens – “rule-out testing”

- **CDC can provide guidance on test selection**
 - **What other tests are available?**
 - **What are their performance characteristics?**
- **CDC can provide guidance on testing**
 - **Clinical presentation**
 - **Demographics (e.g., age)**
 - **Seasonality (NREVSS)**
- **CDC can provide RT-PCR protocols for other respiratory pathogens**

SARS Diagnostics

Key Messages

- **SARS diagnostic assays are sensitive and specific, but may not provide definitive diagnosis early in the illness**
- **Changes in the quantity, type, and timing of specimens collected may improve detection of SARS-CoV infection**
- **Rapid and accurate diagnosis of other respiratory pathogens associated with SARS-like illness may help rule out SARS-CoV infection and calm public fears**
- **Interpretation of test results must take into consideration possibility of false positives and negatives; a clear strategy to minimize such possibilities and to confirm test results are essential**