11

Landman and Bahter- Neurospora Lactase. I. Properties of Lactase Preparations
from a lactose utilizing and a lactose non-utilizing strain

1101

This paper is an introductory contribution of considerable merit on a subject of great interest and importance. In general, this reviewer has no doubt as to its acceptability. However, it has not been carefully edited by the writers, and should be revised in order to make it understandable and conformable to proper standards of scientific expression.

The reviewer finds two points in the argument inadequately supported. The evidence on pages 9 and 10 does not seem to have very critical bearing on the conclusion that "hydrolytic eleavage represents the major way in which lactose is utilized", for one might easily imagine that within the myselium the lactase system functioned differently than in extracts, or alternatively that the major pathway of lactose utilization was mediated by another enzyme whose adaptive responses parallel those of the hydrolytic lactase. Precedents for both types of action can be cited: e.g., transphesiphatase (Axelrod), and a-glucosidase in relation to maltase in yeast, respectively. On the other hand, the reviewer regards this issue as overemphasized in relation to the problem of gene-enzyme sentrol, and the writers' general conclusions are not greatly affected by this point.

The second issue is more important. Although some interesting details are given, this paper does not present an exhaustive study of the lactage as any enzyme. Its main interest is in the comparison of the lactages of the standard and a "lactoceless" mutant. From this point of view, the behavior of the mutant, especially its growth and adaptive enzyme responses to lactoce, are not adequated documented. One may infer that this is taken up in a companion paper "in press". Unless this paper, reference 16, and to some extent 21, is to adjoin the present paper in the pages of this journal, the reviewer questions the utility of splitting the information. At any rate, he is, to a large extent prevented from an adequate critical view of the present paper by a lack of information on the details of the other.

The discussion is rather long, but well-taken and should not be shortened. The recent papers of Cohm and Moned (Acta Biochim Biphys, 1951) and of Lederberg and Beadle (both in Genetics in the 20th Century, MacMillan, 1951) are all, howe extremelyppertinent and should be cited.

The following remarks concern the form rather than the content of the paper and are addressed to authors primarily for their own consideration:

- 1. Capitalization is inconsistent and often incorrect in expressions such as Ethanol, Minimal Medium, Standard Strain, Lactose. The entire paper should be carefully edited for orthographic errors. The same for abbreviations and contractions, especially mg/mg., ml/l., and so forth. Volume numbers are inconsistently treated in the References. Reference 18 cites Adv. in Enzym.; 19 shows Adv. Enzym.
  - 2. B", used repeatedly before "-galactosidase" should be read (beta)
  - 5. The datum "rpm", pp. 6, 11 is not useful. "ROF" should be given, or mentic of the type of centrifuge.

4. "lactoseless", p. 2, and throughout. This expression is bound to be confusing and should be discouraged if the authors agree. It is inconwith the usage in, for example, "methicalneless", which implies an organism lacking the ability to synthesize methicalne. Some organisms are known which require tryptophene; others cannot metabolize it. To use the "-less" terminology for both eases would be very unfortunate. Suggested alternatives: lactase-deficient, alactatic, or lactose-negative.

Specific (mis?)constructions:

- p. 3. 8: which/ that. result and resulted ?
  - L. 14 Is vitamin formation a mutant character?

The second sentence of this paragraph is melorard.

- p. 3 Ref. (13) should be moved framed back one phrase. Emerson's description of his strains' behavior on lactore should be cited (Fed. Proc. 1945-671)
- p. 4 P. 2 2% by weight etc./ 20 g of carbon source was added per 1. ef medium.
- p. 6 L. 8-9. Cation in buffer is not specified, but presumably K. pH 5 or pH 5.
  - L. 19 incomprehensible. Do writers mean: "Activities of different preparations are expressed in terms of constant dry weights of myselium per unit volume per unit time"?
  - 7 L. 5 5.5 for three and a third.
- L. 11 implies/ requires
- p. 8 L. 1 Concentrations/levels
  - 11 Confusing! Insert: "activity in myselia grown on"
- p. 9 L. 1 alternate/alternative
  - 10 "dealing" dangles.
  - 10 9 Two ideas in one sentence confuse!
  - 12 5 ff. One paragraph only
  - 12 14, and 15,7 calories/mole.
  - 12 7 The expression "ensyme-CNPG reaction" is vague. It might, but apparently does not, refer to the initial reaction of adsorption of substrate for which K<sub>s</sub> is given later. Since no indication of extrapolation for V<sub>max</sub> is given, it is difficult to determine whicher the temperature effects concern # K<sub>s</sub> or V<sub>max</sub>, or both, in view of the non-linearity.

- p. 14 L. 1 Same comment. (See Cohn and Monod for offects of cations on lactose/CNPG activity.
  - 14 4 E. coli: Escherichia or Entamosba?
  - 15 15 Why not document this? The inhibitions themselves, especially by xylose, would be of interest.

## Legende for figures:

This figure is a plot of 1/k against 1/8, not a Michaelis constant. The abcissa is incorrectly labelled. If the calculations for p.15 are correct, it should read  $1/8 \times 10^4$ . Most of the 8 concentrations are too low to be useful in the precise estimation of  $K_8$ . It must have been rather difficult to make an accurate determination of the first-order rate constants for an initial substrate concentration of 2.5  $\times$  10<sup>-6</sup> M (the rightmost points).

NOTE: If K is 4 x 10<sup>-4</sup>, and the assay system 2 x 10<sup>-5</sup> M, the statement on p. 7 that the reaction is first order under these conditions is not quite correct. When half the substrate has been used up, the rate would have decreased only about 15% from initial. Comments above on p. 12, 7, fellowed your conclusion on this, but the enzyme is actually about 80% saturated.

## Summeryi

Neurospora crassa (should be underlined). Note: the specific name is never given in the text- why in the summary!