

**NIST National Institute of Standards and Technology**  
...working with industry to develop and apply technology, measurements and standards

## Y-Chromosome and Mitochondrial DNA Work at the U.S. National Institute of Standards and Technology

**John M. Butler**

Amy E. Decker, Peter M. Vallone, Michael D. Coble, Janette W. Redman, Margaret C. Kline, and Richard Schoske\*

November 20, 2004  
IV. International Forensic Y-User Workshop

**NIST Human Identity Project Team**

Funding:  
Interagency Agreement between National Institute of Justice and NIST Office of Law Enforcement Standards

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### Presentation Outline

- General NIST Projects
  - STRBase, etc.
- Y-Chromosome Work
  - SWGDAIM Y-chromosome subcommittee
  - SRM 2395
  - New Y-STR loci under development
  - Y-SNPs
- Mitochondrial DNA Work
  - Coding region SNP assay development with AFDIL
  - LINEAR ARRAYS as an mtDNA screening tool
  - Standard Reference Material SRM 2392-I (Barbara Levin)
- Invitation to Participate in a New NIST Interlaboratory Study involving mixture interpretation

**National Institute of Justice**  
The Research, Development, and Evaluation Agency of the U.S. Department of Justice

### Current Areas of NIST Research Effort

- Standard Information Resources** (STRBase information, training materials/review articles, validation standardization, calibration datasets)
- Interlaboratory Studies** (Real-time PCR, mixture interpretation)
- Resources for "Challenging Samples"** (miniSTRs for degraded DNA)
- Information on New Loci** (Y-Chromosome, new STRs)

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## STRBase

### Short Tandem Repeat DNA Internet Database

**Recent Additions**

- Validation (summary sheets)  
[.../validation.htm](#)
- miniSTR information  
[.../miniSTR.htm](#)

**We Regularly Update**

- Reference List
- Variant Alleles
- Addresses for Scientists
- Links to Other Web Sites
- Y-STR Information

**NIST publications** and presentations as pdf files  
[.../NISTpub.htm](#)

<http://www.cstl.nist.gov/biotech/strbase>

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### Technology Development Efforts Centered around multiplex PCR...

Created Custom Primer Design Software

Standardized Assay Design Formats

Multiplex Design Schematic

Schoske et al. (2003) *Anal. Bioanal. Chem.* 375:333-343

Implemented Quality Control Methods for PCR Primers

Demonstrated Success with Multiple Projects and Collaborations

mtDNA 11plex SNP assay

Y-STR 20plex, cat STR 12plex

**AutoDimer Primer Screening Program**

**SHORT TECHNICAL REPORTS**  
Vallone, P.M. and Butler, J.M. (2004) *BioTechniques* 37:226-231

**AutoDimer: a screening tool for primer-dimer and hairpin structures**

Peter M. Vallone and John M. Butler  
National Institute of Standards and Technology, Gaithersburg, MD, USA

**BioTechniques 37:226-231 (August 2004)**

**Available for download from STRBase:**  
<http://www.cstl.nist.gov/biotech/strbase>

**Download Page**

**Home** **Download** **Case No.** **Forensics** **Referencing AutoDimer** **FAQ** **Support**

AutoDimer was packaged for installation using Visual Basic 6.0. I have tested the installation on PCs running Win98, 2000, XP and NT. However, I cannot guarantee installation success for each user's specific computer configuration.

By clicking the link below you will be downloading the file AutoDimer.zip. Once extracted ([www.winzip.com](http://www.winzip.com)), the files can be used to install the AutoDimer program (click setup.exe).

The end user is responsible for the installation and running of the program (this is done at your own risk). The author will not be held responsible for any subsequent computer/operating system issues due to conflicts with the AutoDimer software. AutoDimer is a general tool for screening sequences; we do not guarantee the success of your PCR assay.

**Please click here to download AutoDimer (~5 MB).**

**A web-based interface is in development (similar to Primer3)**

**Anal Bioanal Chem (2003) 375: 333–343**  
DOI 10.1007/s00216-002-1683-2

**ORIGINAL PAPER**

Richard Schoske · Pete M. Vallone  
Christian M. Rüttiger · John M. Butler

**Multiplex PCR design strategy used for the simultaneous amplification of 10 Y chromosome short tandem repeat (STR) loci**

Received: 3 July 2002 / Revised: 24 October 2002 / Accepted: 29 October 2002 / Published online: 14 January 2003  
© Springer-Verlag 2003

**Careful primer design**

- Uniform annealing temperatures
- Checking for all potential primer-primer interactions

**Potential Interaction**

$$\begin{array}{c} 3'-TAGTAGATGACAGAGGGATACA-5' \\ ||| \quad | \\ SCCCCCTCTCTGCTATCT-3' \end{array}$$

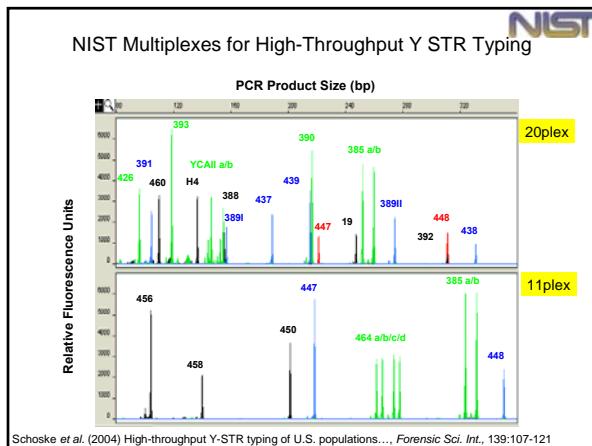
Butler et al. (2001) *Fresenius J. Anal. Chem.* 369:200-205

**Stringent primer quality control**

**Dye labeled oligos**

6FAM ("blue"), VIC ("green"), NED ("yellow")

Butler et al. (2001) *Forensic Sci. Int.* 119: 87-96



**NIST U.S. Population Samples**

As of 06/2003 **663 males** (anonymous; self-identified ethnicities)

260 Caucasians  
260 African Americans  
140 Hispanics  
3 Asians

Whole blood received from Interstate Blood Bank (Memphis, TN)

**Working tubes/plates 1 ng/uL**

To date: (~85,000 allele calls)

- Identifier (15 autosomal markers + Amelogenin) (10,608)
- Roche LINEAR ARRAYS (HVI/HV2 10 regions) (6,630)
- Y STRs 22 loci—27 amplicons (17,388)
- Y STRs 27 new loci (14,535), 6 new autosomal STRs (2,844)
- Y SNPs 50 markers on sub-set of samples (11,498)
- Orchid 70 autosomal SNPs on sub-set (13,230)
- miniSTR testing-new loci and CODIS concordance (9,228)
- mtDNA full control region sequences by AFDIL

**Stock tubes**

**Working tubes**

**Working plates**

**Samples supplied to collaborators for miniSTR typing and AFDIL for whole mtGenome sequencing**

**Standard U.S. Population Dataset**

<http://www.cstl.nist.gov/biotech/strbase/NISTpop.htm>

260 Caucasians, 260 African Americans, 140 Hispanics, 3 Asians = **663 males**

Genetic Markers	Loci Examined	Publications
Common STRs	Identifier kit 15 STRs (CODIS + D2S1338 & D19S433)	Butler et al. (2003) JFS
miniSTRs	All CODIS loci except D3S1358	Drabek et al. (2004) JFS
New autosomal STRs	New 6 loci for miniSTRs	Coble et al. (2005) JFS
Autosomal SNPs	70 C/T SNPs (Orchid panel)	Vallone et al. (2004) FSI
Common Y-STRs	22 loci (27 regions)	Schoske et al. (2004) FSI
Yfiler concordance study	<i>Data in ABI Yfiler database</i>	
<b>New Y-STRs</b>	<b>27 additional loci</b>	<b>Butler et al., submitted</b>
Y-SNPs	50 loci spanning haplogroups A-R	Vallone et al. (2004) JFS
mtDNA	LINEAR ARRAY and coding mtSNPs	Kline et al. (2005) JFS
	Full control regions by AFDIL	<i>inclusion in EMPOP</i>

**Y-Chromosome**

**SWGDAM, SRM 2395,**  
**New Y-STR Loci, and Y-SNPs**

<http://www.fbi.gov>

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Forensic Science Communications July 2004 – Volume 6 – Number 3  
Standards and Guidelines

**Report on the Current Activities of the Scientific Working Group on DNA Analysis Methods Y-STR Subcommittee**

Scientific Working Group on DNA Analysis Methods Y-STR Subcommittee

**Introduction**

Detecting DNA from a male perpetrator is the goal in the forensic investigation of most sexual assault cases. Y-chromosome-specific STR typing targets the male DNA and is a useful additional tool in cases that often involve a mixture of male and female DNA. Although many technical aspects of Y-STR testing are parallel to autosomal STR testing, the unilateral (patrilineal) inheritance of the Y-chromosome alleles creates a haplotype of linked loci, and the statistical evaluation and reporting of the results differ significantly. Therefore, the SWGDAM Y-STR Subcommittee was established to deal with all aspects of Y-chromosome-specific testing in forensic casework.

**Selection of U.S. Core Loci:**

- DYS19,
- DYS385 a/b,
- DYS389 I/II,
- DYS390,
- DYS391,
- DYS392,
- DYS393,
- DYS438,
- DYS439

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**SWGDAM Sub-Committee on the Y Chromosome**

- Formed in July 2002
- Members
  - Jack Ballantyne (UCF) – chair
  - Mecki Prinz (NYC) – co-chair
  - John Butler (NIST)
  - Ann Gross (MN)
  - John Hartmann (Orange Co.)
  - Sam Baechtel (FBI Lab)
  - Jill Smerick (FBI Lab)
  - Debra Figarelli (Phoenix)
  - Carl Ladd (CT)
  - Demris Lee (AFDIL)
  - Jonathan Newman (CFS-Toronto)
  - Phil Kinsey (OR)
  - Gary Sims (CA DOJ)
- U.S. CORE Y-STR LOCI selected in January 2003**
  - 60 sample set selected for screening markers and initial testing
  - Testing of Y-PLEX 6 and Y-PLEX 5 kits in all labs
    - All results completed agreed with NIST results sent to participating labs in Dec 2002
  - Jack Ballantyne's lab and John Butler's lab to examine additional Y-STR and Y-SNP markers in the same sample set

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**European and U.S. Core Y-STR Loci**

Marker Name	Allele Range (repeat numbers)	Repeat Motif
DYS19	10-19	TAGA
DYS385 a/b	7-28	GAAT
DYS389 I	I: 9-17	(TCTG) (TCTA)
DYS389 II	II: 24-34	(TCTG) (TCTA)
DYS390	17-28	(TCTA) (TCTG)
DYS391	6-14	TCTA
DYS392	6-18	TAT
DYS393	8-17	AGAT
YCAII a/b	11-25	CA
DYS438	6-14	TTTC
DYS439	8-15	AGAT

Minimal haplotype (Europe)

U.S. haplotype

Extended haplotype (Europe)

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**Y-Chromosome Standard NIST SRM 2395**

**Human Y-Chromosome DNA Profiling Standard**

- 5 male samples + 1 female sample (neg. control)
- 100 ng of each (50 µL at ~2 ng/µL) \$248
- 22 Y STR markers sequenced
- 9 additional Y STR markers typed
- 42 Y SNPs typed with Marligen kit

Certified for all loci in commercial Y-STR kits:

- Y-PLEX 6
- Y-PLEX 5
- Y-PLEX 12
- PowerPlex Y
- SWGDAM recommended loci: DYS19, DYS385 a/b, DYS389 I/II, DYS390, DYS391, DYS392, DYS393, DYS438, DYS439
- Y-filer - adds DYS635 (C4); now sequenced

Helps meet FBI Standard 9.5 (and ISO 17025)...traceability to a national standard

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**Sequence Summaries for SRM 2395**

DYS19	A	14	[TAGA] <sub>1</sub> tagg[TAGA] <sub>1</sub>	DYS438	A	12	[TTTC] <sub>12</sub>
B	14	[TAGA] <sub>1</sub> tagg[TAGA] <sub>1</sub>	B	9	[TTTC] <sub>9</sub>		
C	16	[TAGA] <sub>1</sub> tagg[TAGA] <sub>1</sub>	C	11	[TTTC] <sub>11</sub>		
D	15	[TAGA] <sub>1</sub> tagg[TAGA] <sub>1</sub>	D	11	[TTTC] <sub>11</sub>		
E	17	[TAGA] <sub>1</sub> tagg[TAGA] <sub>1</sub>	E	10	[TTTC] <sub>10</sub>		

DYS390	A	25	[TCTG] <sub>6</sub> [TCTA] <sub>12</sub> [TCTG] <sub>6</sub> [TCTA] <sub>6</sub>
B	23	[TCTG] <sub>6</sub> [TCTA] <sub>10</sub> [TCTG] <sub>6</sub> [TCTA] <sub>6</sub>	
C	21	[TCTG] <sub>6</sub> [TCTA] <sub>10</sub> ACTA [TCTA] <sub>6</sub> [TCTG] <sub>6</sub> [TCTA] <sub>6</sub>	
D	22	[TCTG] <sub>6</sub> [TCTA] <sub>10</sub> [TCTG] <sub>6</sub> [TCTA] <sub>6</sub>	
E	24	[TCTG] <sub>6</sub> [TCTA] <sub>10</sub> [TCTG] <sub>6</sub> [TCTA] <sub>6</sub>	

We will continue to add information on new Y-STR loci as they are adopted by the community and put into commercial kits

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**Y-SNP Results on SRM 2395**

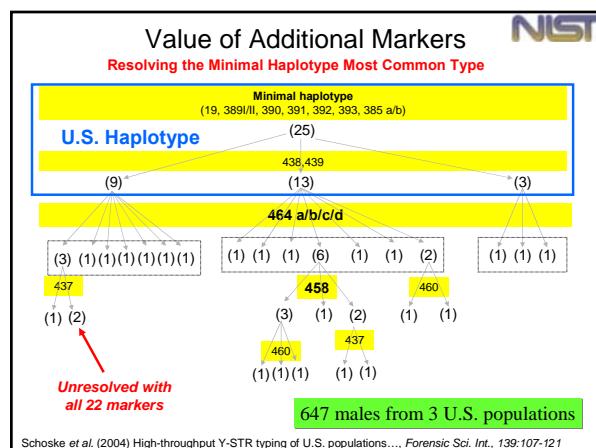
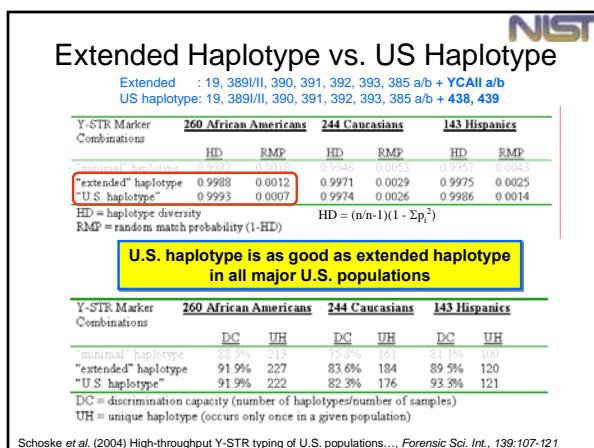
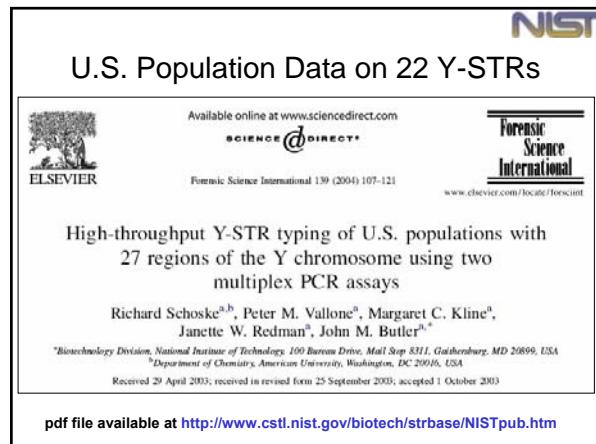
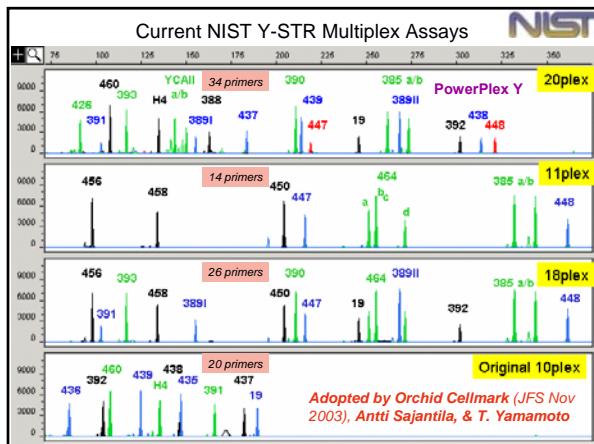
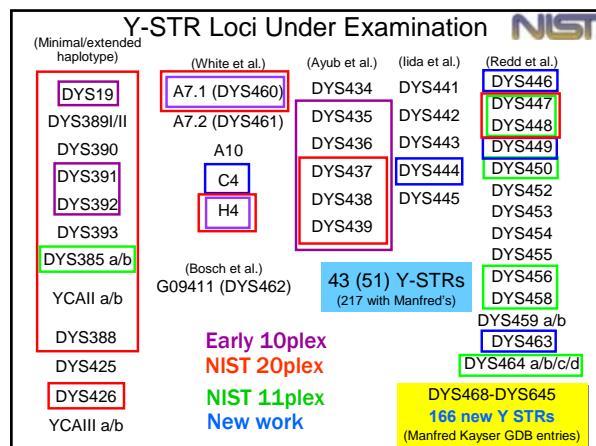
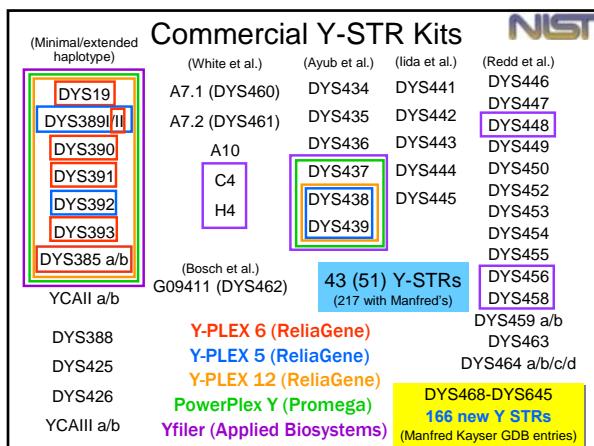
from Marligen Signet™ Multiplexes (Luminex bead assay)

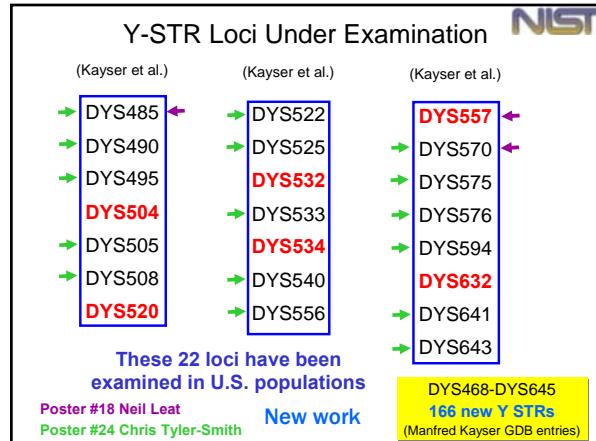
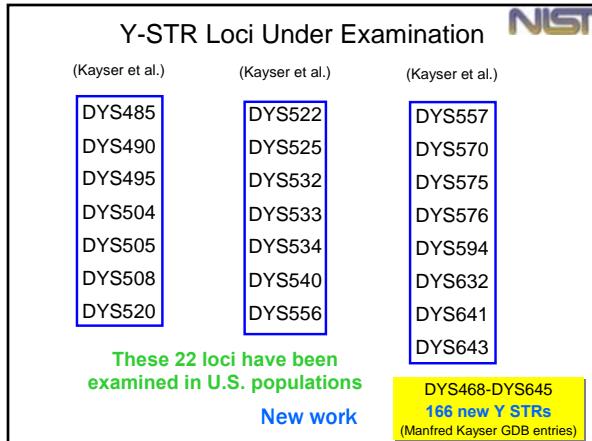
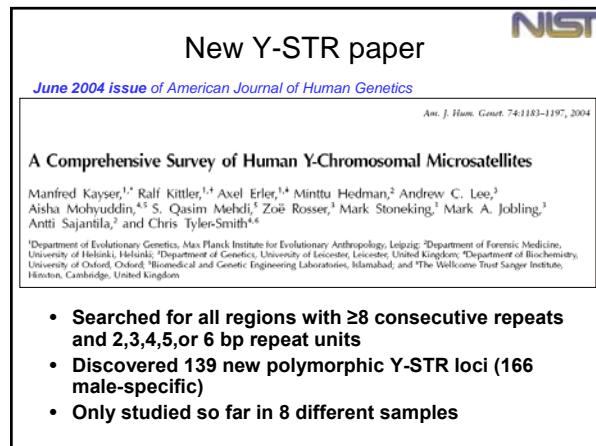
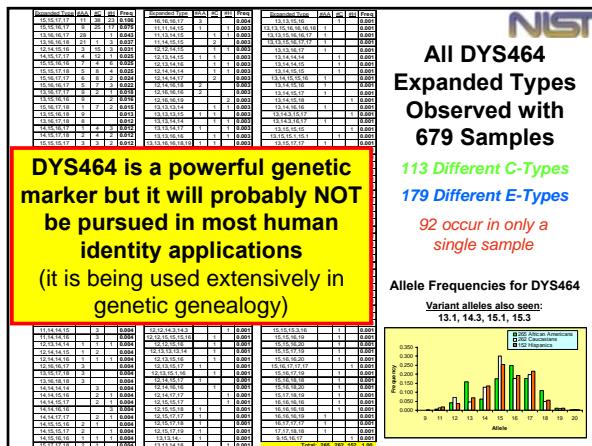
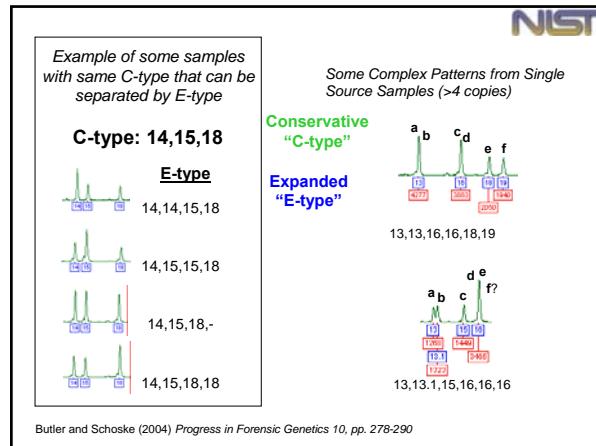
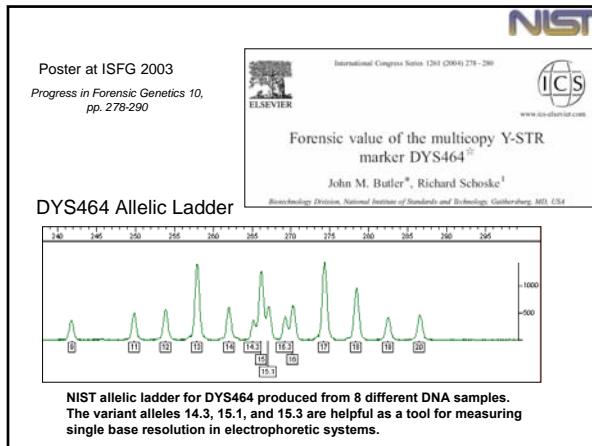
**42 Y-SNPs measured across all samples**

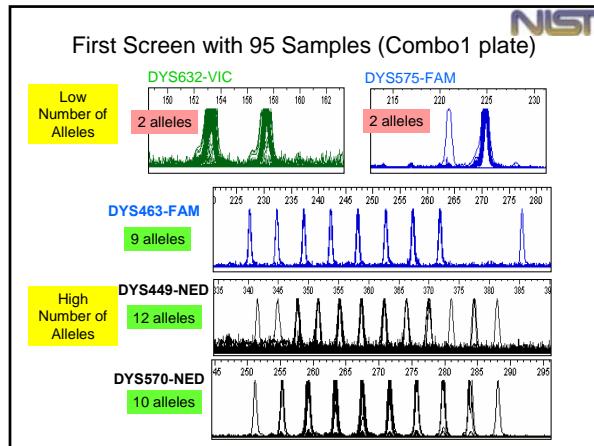
SRM 2395	AMEL	M207	M45	M89	DYS391	M2	M170	M172	M201
	(A/G)	(A/G)	(C/T)	(C/G)	(A/G)	(A/C)	(G/T)	(G/T)	
Component A	XY	<b>G</b>	<b>A</b>	T	C	A	A	T	G
Component B	XY	A	G	T	C	A	A	<b>G</b>	G
Component C	XY	A	G	<b>C</b>	<b>G</b>	A	T	G	
Component D	XY	A	G	T	C	A	A	T	<b>T</b>
Component E	XY	A	G	T	C	A	<b>C</b>	T	G
Component F	XX								

SRM components are all distinguishable from one another with these Y SNPs

5 male components in SRM 2395 have 5 different Y-SNP backgrounds: R1b, J2, E3a, G, and I







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Number of Alleles Seen with Various Y-STR Loci in Same Set of 95 U.S. Samples

Set	Locus ID	Dye	Length Range (bp)	# of alleles
B	DYS632	VIC	153-157	2
G	DYS575	FAM	221-225	2
E	DYS463	FAM	212-222	3
E	DYS449	NED	222-230	3
G	DYS540	NED	266-277	4
J	DYS522	VIC	359-370	4
A	DYS495	FAM	133-146	5
G	DYS594	VIC	264-289	5
C	DYS505	FAM	168-187	<b>6</b>
C	DYS508	VIC	180-200	<b>6</b>
C	DYS520	NED	181-201	<b>6</b>
E	DYS533	VIC	209-230	<b>6</b>
J	DYS446	FAM	298-325	<b>6</b>
H	DYS444	NED	299-325	<b>6</b>
B	DYS643	FAM	135-167	7
D	DYS534	NED	202-227	<b>7</b>
D	DYS557	VIC	196-219	<b>7</b>
H	DYS525	VIC	365-375	<b>7</b>
J	DYS526	NED	466-490	<b>7</b>
D	DYS576	FAM	178-206	<b>8</b>
A	DYS485	NED	138-164	<b>8</b>
H	DYS504	FAM	270-303	<b>9</b>
F	DYS463	FAM	227-277	<b>9</b>
F	DYS570	NED	251-288	<b>10</b>
A	DYS490	VIC	126-158	<b>10</b>
B	DYS449	NED	341-381	<b>12</b>

Loci Not Pursued Further...

Low Number of Alleles

DYS632  
DYS575

Primers Gave Artifacts in Female

DYS490 – duplicated and on chr X

DYS504

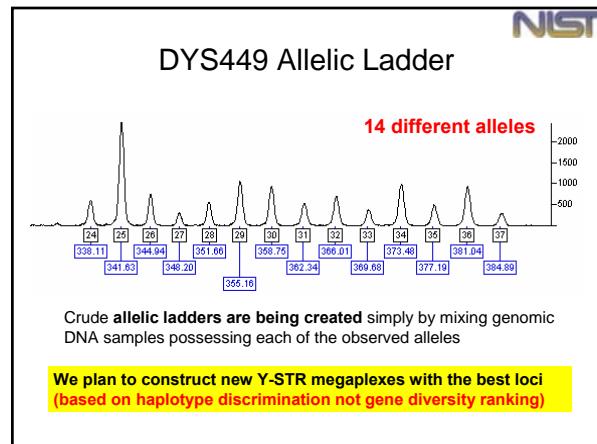
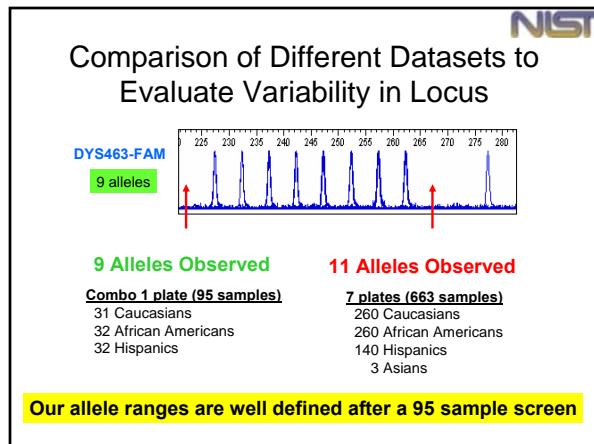
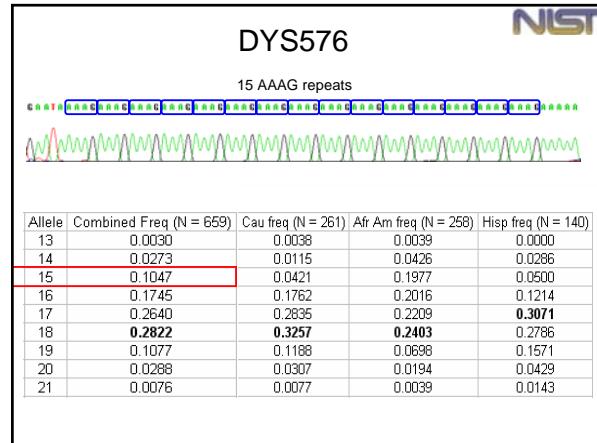
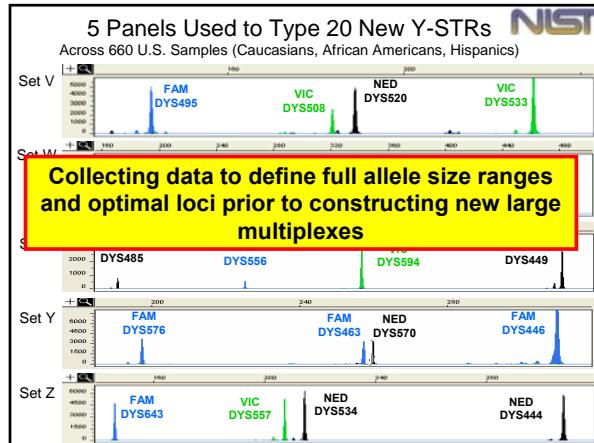
DYS525

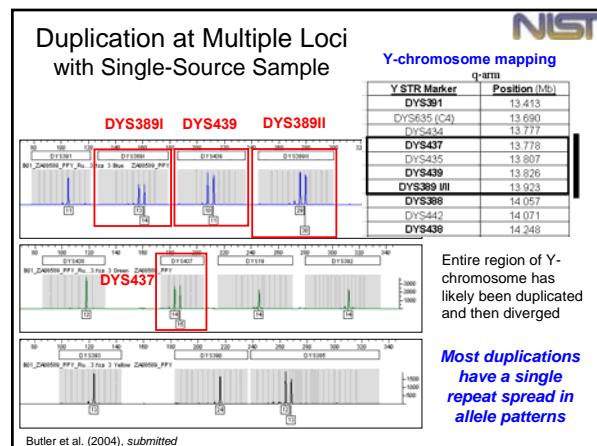
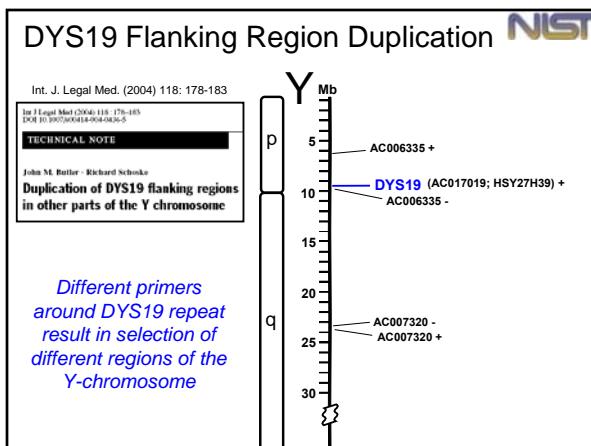
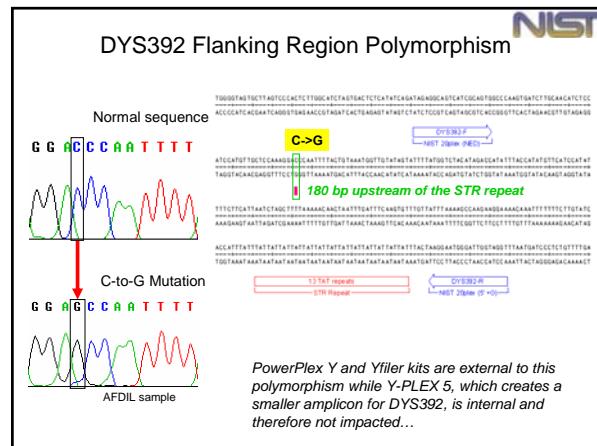
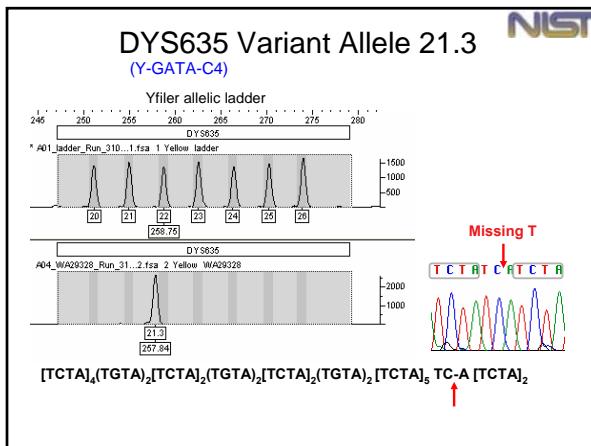
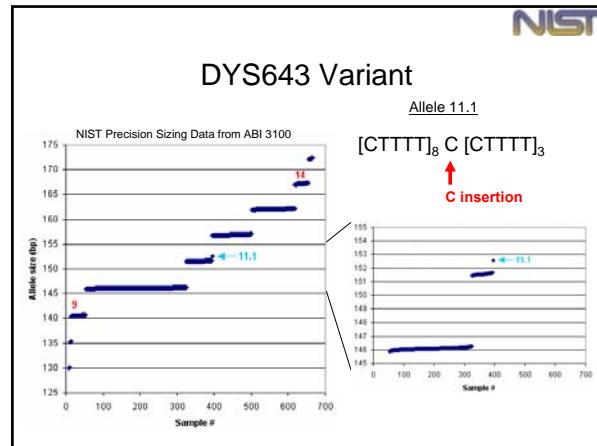
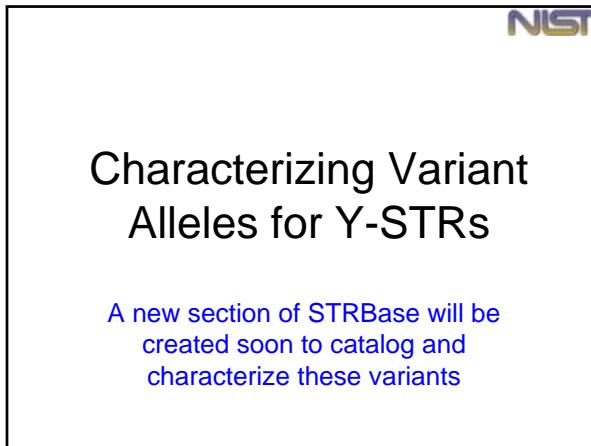
DYS557

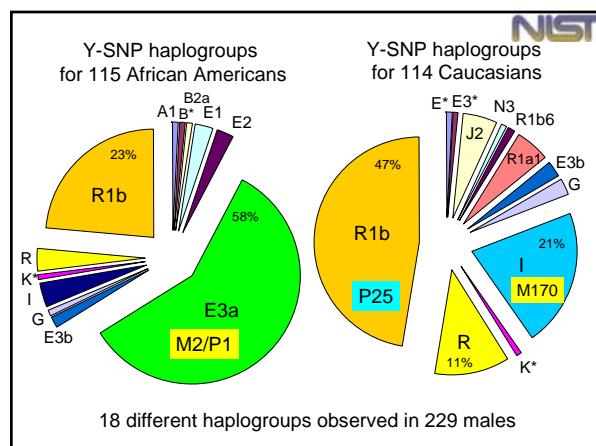
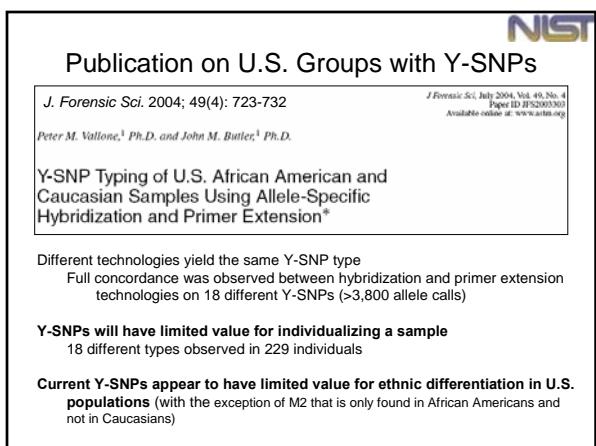
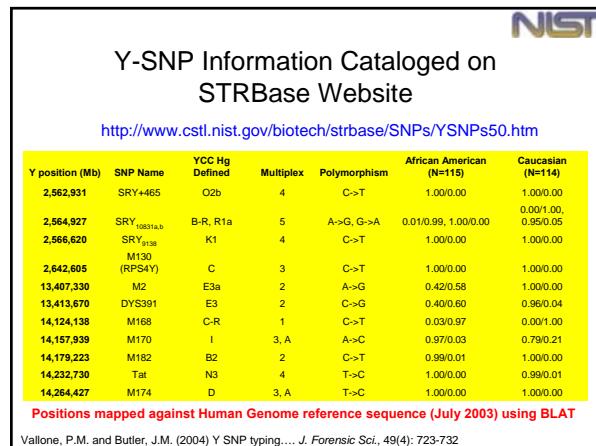
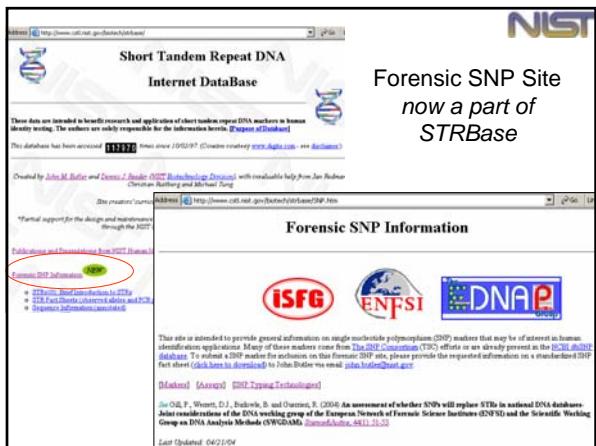
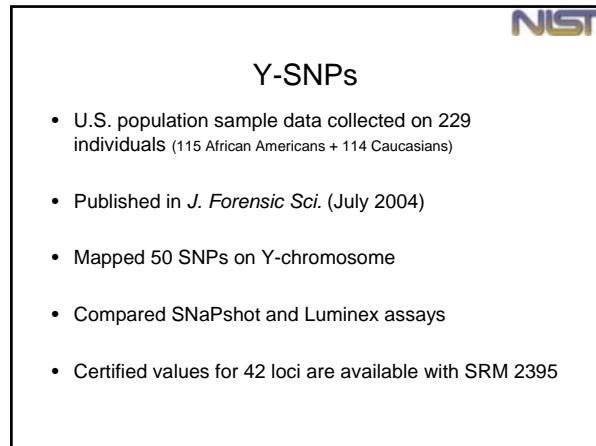
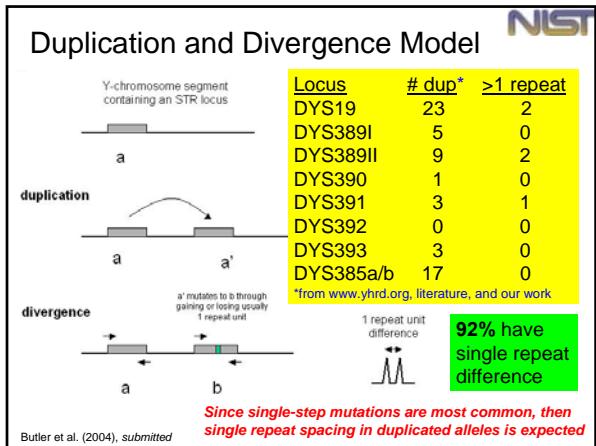
**14,535 types generated across 27 loci**

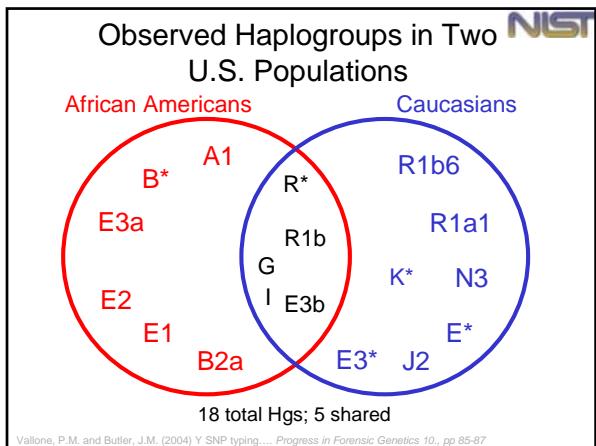
Combo 1 plate

31 Caucasians  
32 African Americans  
32 Hispanics





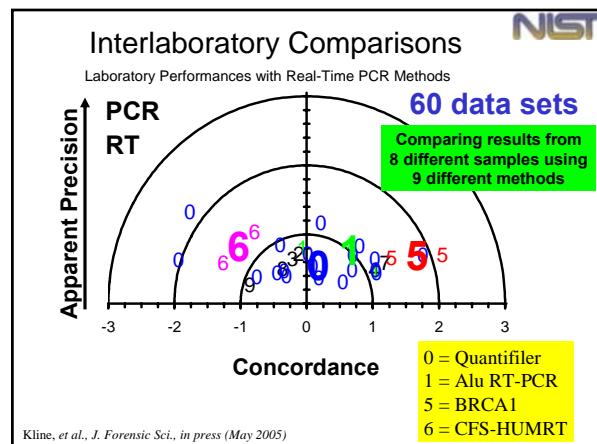
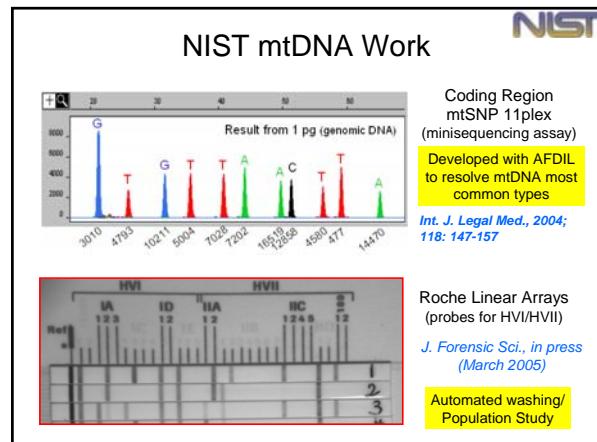
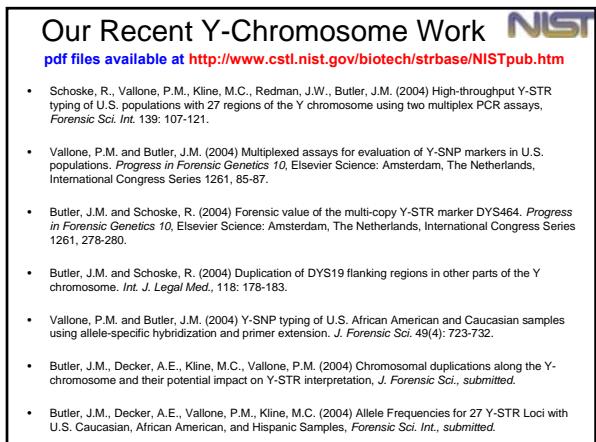




### NIST Summary of NIST Y-Chromosome Work

- Development of new Y-STR multiplex assays (**Y STR 20plex, 11plex, etc.**)
- Evaluation of new Y-STR loci in U.S. populations
- Evaluation of SNP typing methodologies and development of **Y-SNP assays** involving primer extension and the SNaPshot kit
- Creation of a Human Y-Chromosome Standard Reference Material (**SRM 2395**)
- Standardization of information on Y-chromosome markers with internet accessibility (**STRBase**)

>45,000 Y-chromosome allele calls generated to aid studies on optimal markers for U.S. populations



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## Acknowledgements

**NIST Project Team:**

John Butler  
Pete Vallone  
Margaret Kline  
Jan Redman  
Amy Decker  
Mike Coble  
Richard Schoske (former member)

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**National Institute of Justice** and  
**NIST Office of Law Enforcement  
Standards**

**Collaborators:**

Tom Parsons (AFDIL)  
Jodi Irwin (AFDIL)  
Sandy Calloway (Roche)

This presentation available as pdf file from  
<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>