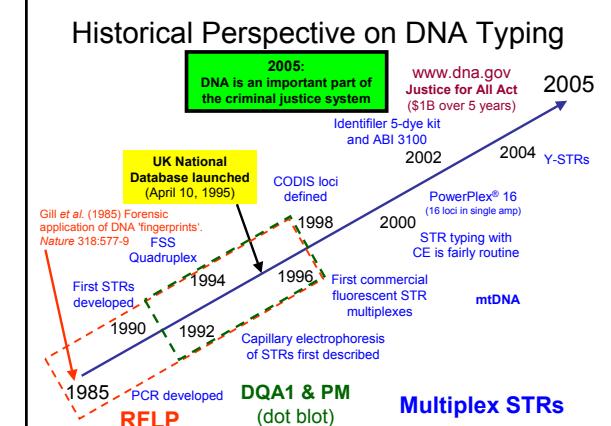


Setting Standards and Developing Technology to Aid the Human Identity Testing Community

John M. Butler
Peter M. Vallone, Michael D. Coble,
Janette W. Redman, Amy E. Decker, Carolyn R. Hill,
David L. Duewer, Margaret C. Kline

Human Identity Project Team, U.S. National Institute of Standards and Technology
ISFG Keynote Address – September 14, 2005

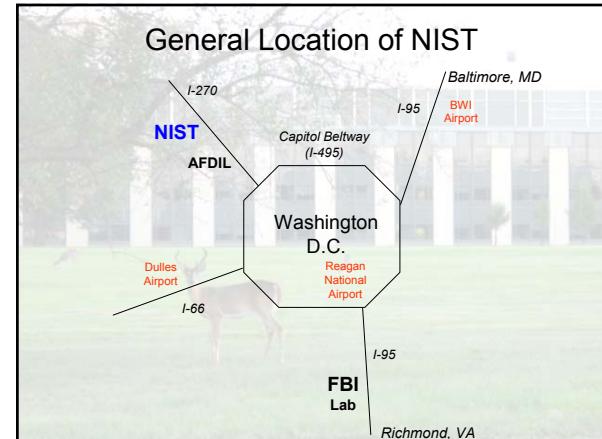
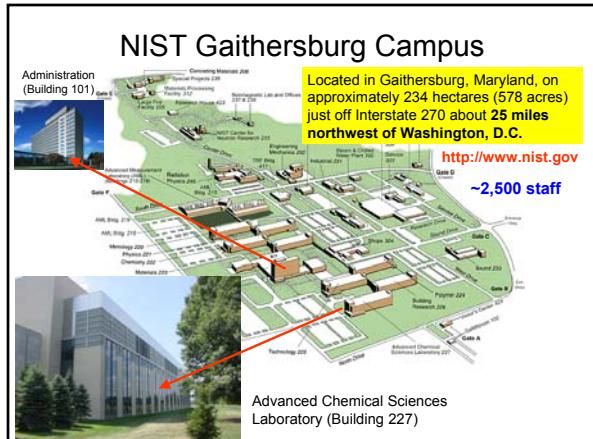


Presentation Overview

- Introduction to NIST and Project Team
- Standards
- Technology
- Training Materials

NIST Mission and History

- National Institute of Standards and Technology (NIST) was created in 1901 as the National Bureau of Standards (NBS). The name was changed to NIST in 1988.
- NIST is part of the U.S. Department of Commerce with a mission to develop and promote measurement, standards, and technology to enhance productivity, facilitate trade, and improve the quality of life.
- NIST supplies over 1,300 Standard Reference Materials (SRMs) for industry, academia, and government use in such areas as environmental analysis, health measurements, and industrial materials production and analysis.



NIST Human Identity Project Team Laboratory Space (within Building 227)

Equipment List

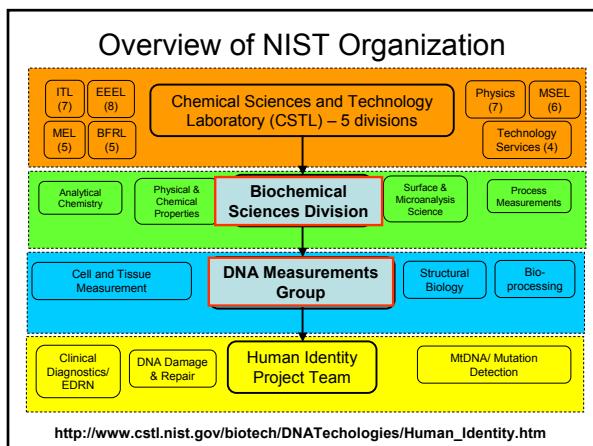
- GeneAmp 9700
- GeneAmp 9600
- ABI 310
- ABI 3100xl (on order)
- FMBIO III+
- ABI 7500 real-time PCR
- Agilent Bioanalyzer 2100
- Varian UV spec
- Varian HPLC
- Bruker TOF-MS
- MWG RoboAmp 4200 -80 °C freezers

Room B219 (DNA extraction) Room A230 (ABI 3130x)

NIST and Forensic DNA

- Dennis Reeder, Margaret Kline, and Kristy Richie started DNA work at NIST in late 1980s
- The FBI's DNA Advisory Board required one member be from NIST, which was Dennis Reeder
- NIST has been an invited guest to TWGDDM/SWGDAM since 1988
- A number of Standard Reference Materials have been developed since 1992: SRM 2390 (RFLP), SRM 2391 (PCR/STRs), SRM 2392 (mtDNA), SRM 2395 (Y-STRs), SRM 2372 (DNA quant)
- STRBase website developed in 1996-1997 while John Butler was a postdoc
- Human Identity Project Team formed in 2000 with John Butler as project leader (when Dennis Reeder left NIST for Applied Biosystems)
- Numerous research areas now funded by the National Institute of Justice

<http://www.cstl.nist.gov/biotech/strbase/NIJprojects.htm>



NIST Human Identity Project Team



Funding: Interagency Agreement 2003-IJ-R-029 between National Institute of Justice (NIJ) and NIST Office of Law Enforcement Standards (OLES)

Team Productivity

[Since last ISFG meeting \(Sept 2003\):](#)

- 43 publications
- 57 presentations
- Textbook: *Forensic DNA Typing, 2nd Edition*
- Widely-used website: hundreds of pages of new information added to STRBase

<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

Forensic DNA Typing, 2nd Edition: John Butler (not NIST)

Chapter 1 Overview & History of DNA Typing
Chapter 2 DNA Biology Review
Chapter 3 Sample Collection, Extraction, Quantitation
Chapter 4 PCR Amplification
Chapter 5 Common STRs and Commercial Kits
Chapter 6 Biology of STRs
Chapter 7 Forensic Issues
Chapter 8 Single Nucleotide Polymorphisms
Chapter 9 Y-Chromosomal DNA Tests
Chapter 10 Mitochondrial DNA Analysis
Chapter 11 Non-Human DNA and Microbial Forensics
Chapter 12 DNA Separation Methods
Chapter 13 DNA Detection Methods
Chapter 14 Instrumentation for STR Typing: ABI 310, 3100, FMBIO
Chapter 15 STR Genotyping Issues
Chapter 16 STR Data Analysis
Chapter 17 New Technologies, Automation, and Expert Systems
Chapter 18 CODIS and DNA Databases
Chapter 19 Basic Genetic Principles and Statistics
Chapter 20 STR Database Analysis
Chapter 21 Profile Frequency Estimates
Chapter 22 Statistical Analysis of Mixtures and Degraded DNA
Chapter 23 Kinship and Paternity Testing
Chapter 24 DNA Profiling and Victim Identification
Appendix I Reported STR Alleles
Appendix II U.S. Population Data-STR Allele Frequencies
Appendix III Suppliers of DNA Analysis Equipment
Appendix IV DAB QA Standards
Appendix V DAB Recommendations on Statistics
Appendix VI Application of NRC II to STR Typing
Appendix VII Example DNA Cases

double the size of the first edition
10 new chapters, >500 new references

National Institute of Justice
The Research, Development, and Evaluation Agency of the U.S. Department of Justice

Current Areas of NIST Effort with Forensic DNA

- Standards**
 - Standard Reference Materials
 - Standard Information Resources (STRBase website)
 - Interlaboratory Studies
- Technology**
 - Research programs in SNPs, miniSTRs, Y-STRs, mtDNA, qPCR
 - Assay and software development
- Training Materials**
 - Review articles and workshops on STRs, CE, validation
 - PowerPoint and pdf files available for download

Congress Passed **the DNA Identification Act of 1994** (Public Law 103-322)
Formalized the FBI's authority to establish a national DNA index for law enforcement purposes.

FBI's DNA Advisory Board
Quality Assurance Standards for Forensic DNA Testing Laboratories
(October 1, 1998)

STANDARD 9.5

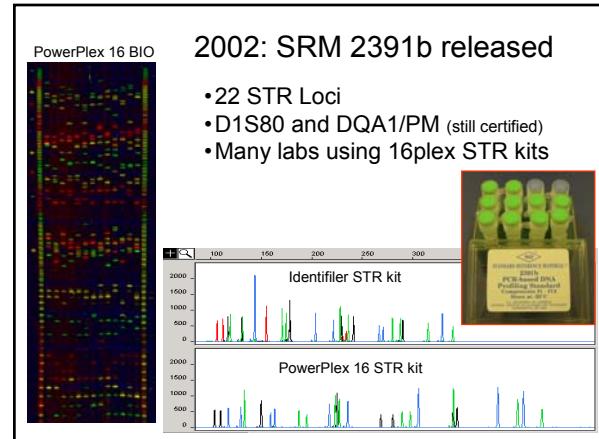
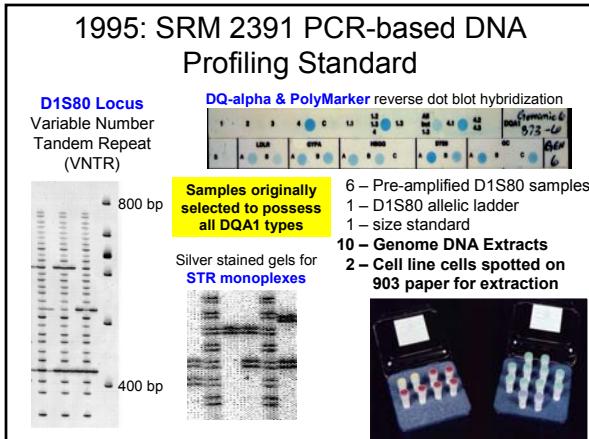
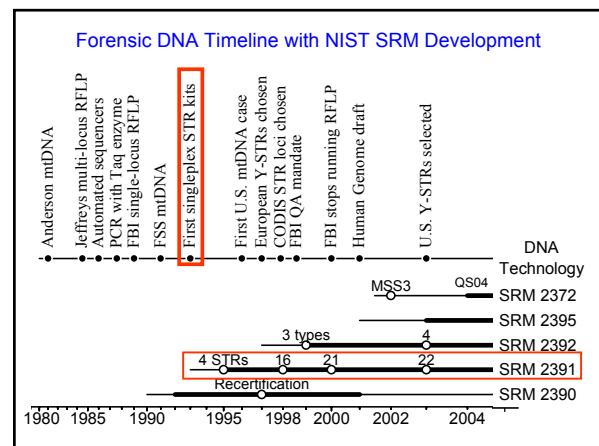
The laboratory shall check its DNA procedures annually or whenever substantial changes are made to the protocol(s) against an appropriate and available NIST standard reference material or standard traceable to a NIST standard.

Margaret Kline Jan Redman

Standard Reference Materials

- Relevant Forensic DNA SRMs
 - SRM 2391b (DNA profiling – STRs, D1S80, DQA1/PM)
 - SRM 2392-I (mtDNA)
 - SRM 2395 (Y-chromosome)
 - SRM 2372 (Human DNA quantitation); *in development*
- Provides national/international traceability and compatibility (aids in ISO 17025 compliance)

<http://www.nist.gov/srm>



NIST SRM 2391b

All current commercial kit STR loci are certified

	FES/IPS	LPL	Penta D	Penta E	D2S138	D19S433
0	12,12	10,11	10,15	7,12	17,23	13,16,2
1	10,11	9,11	7,12		17,26	14,16
2	11,12	11,12	11,12	13,14	20,24	12,14
3	10,13	10,12	8,9	5,12	17,23	11,13
4	11,13	10,12	10,13	7,13	17,19	12,2,14
5	11,11	10,12	9,12	12,14	25,25	12,14
6	11,11*	11,12	3,2,11	12,16	17,22	13,15,2
7	10,11					13,15
8	10,12					15
9	11,12					15
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Validation Summary Sheet for PowerPlex Y		
Study Completed (17 studies done)	Description of Samples Tested (performed in 7 labs and Promega)	# Run
Single Source (Concordance)	5 samples x 8 labs	40
Mixture Ratio (male:female)	6 labs x 2 MM mixture series x 11 ratios (1.0;1.1;1.0;1.100;1.300;1.1000;0.5;300;0.25;300;0.125;300, 0.0623;3.300;0.03;300) 132	
Mixture Ratio (male:male)	6 labs x 2 MM mixture series x 11 ratios (1.0; 19.1; 9.1; 5.1; 2.1; 1.1; 1.2; 1.5; 1.9; 1.19; 0.1) 132	
Sensitivity	7 labs x 2 series x 6 amounts (1.0/0.5/0.25/0.125/0.06/0.03)	84
Non-Human	24 animals	24
NIST SRM	6 components of SRM 2395	6
Precision (ABI 3100 and ABI 377)	10 ladder replicates + 10 sample replicated + [B ladders + 8 samples for 377]	36
Non-Probative Cases	65 cases with 102 samples	102
Stutter	412 males used	412
Peak Height Ratio	N/A (except for DYS385 but no studies were noted)	
Cycling Parameters	5 cycles (28/27/26/25/24) x 8 punch sizes x 2 samples	80
Annealing Temperature	5 labs x 5 temperatures (54/58/60/62/64) x 1 sample	25
Reaction volume	5 volumes (50/25/15/12.5/6.25) x [5 amounts + 5 concentrations]	50
Thermal cycler test	4 models (480/240/0960/0700) x 1 sample + [3 models x 3 sets x 12 samples]	76
Male-specificity	2 females x 1 titration series (0-50 ng female DNA) x 5 amounts each	10
TaqGold polymerase titration	5 amounts (1.38/0.62/0.38/0.19/0.13) U x 4 quantities (10.5/0.25/0.13 ng DNA)	20
Primer pair titration	5 amounts (0.5x/0.75x/1x/1.5x/2x) x 4 quantities (10.5/0.25/0.13 ng DNA)	20
Magnesium titration	5 amounts (1.25/1.5/1.75/2 mM Mg) x 4 quantities (10.5/0.25/0.13 ng DNA)	20

Krenke et al. (2005) *Forensic Sci. Int.* 148:1-14TOTAL SAMPLES EXAMINED **1269**

Laboratory Internal Validation Summaries																																																					
<p>We invite updates to this table. Please contact John Butler (julian.butler@fbi.gov) if you would like to add a summary of your laboratory validation studies with a particular forensic DNA test, instrument, or software program. Please submit information in a standard format summarizing the studies conducted, a description of samples run, and the number of samples examined using the downloadable Excel file [click here].</p> <p>Summaries of Validation Studies Conducted in Individual Laboratories (not published in the literature)</p> <table border="1"> <thead> <tr> <th>Kit, Assay or Instrument</th> <th>Laboratory</th> <th>Submitter</th> </tr> </thead> <tbody> <tr> <td>PowerPlex 16 Kit with ABI 310</td> <td>Pennsylvania State Police</td> <td>Christian Tomery</td> </tr> <tr> <td>Quantifiler with ABI 7000</td> <td>Alabama Department of Forensic Sciences</td> <td>Angelo Orsi Marzo</td> </tr> </tbody> </table>			Kit, Assay or Instrument	Laboratory	Submitter	PowerPlex 16 Kit with ABI 310	Pennsylvania State Police	Christian Tomery	Quantifiler with ABI 7000	Alabama Department of Forensic Sciences	Angelo Orsi Marzo																																										
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<p>Soliciting Information on Studies Performed by the Community</p> <table border="1"> <thead> <tr> <th>Study Category</th> <th>Description of samples used with a description of conditions</th> <th>Estimated Number of Samples</th> </tr> </thead> <tbody> <tr> <td>Single Source (Concordance)</td> <td>8 samples (Proenza concordance) = 200 samples (part of population concordance study)</td> <td>200 100</td> </tr> <tr> <td>Mixtures</td> <td>41</td> <td>45 10</td> </tr> <tr> <td>Peak Height Ratio</td> <td>1 sample x 11 ratios (1.0; 19.1; 9.1; 5.1; 2.1; 1.1; 1.2; 1.5; 1.9; 1.19; 0.1) x 2 replicates (30 seconds)</td> <td>22 39</td> </tr> <tr> <td>Sensitivity</td> <td>5 samples x 6 amounts (20/10; 5.6; 2.56; 1.50; 0.63; 0.32 ng) x 5 samples x 3 points (calibration standards)</td> <td>66 33</td> </tr> <tr> <td>Non-Human</td> <td>11 animals</td> <td>11 0</td> </tr> <tr> <td>NIST SRM 2391b</td> <td>12 components</td> <td>12 12</td> </tr> <tr> <td>Primer (ABI 310)</td> <td>(3 samples x 10 replicates) x 10 different locations of white laundry</td> <td>60 60</td> </tr> <tr> <td>Non-Probative Cases</td> <td>5 cases x 4 samples each (evidence from FBI Laboratory)</td> <td>20 20</td> </tr> <tr> <td>Stutter</td> <td>200 samples (data used from population samples)</td> <td>- -</td> </tr> <tr> <td>Peak Height Ratio</td> <td>(3 samples x 10 replicates) x 10 different locations of white laundry</td> <td>- -</td> </tr> <tr> <td>Cycling Parameters</td> <td>200 samples (data used from population samples)</td> <td>- -</td> </tr> <tr> <td>Annealing Temperature</td> <td>14 samples x 2 different cycle annealing temperatures x 2 replicates (30 seconds)</td> <td>66 -</td> </tr> <tr> <td>Proficiency</td> <td>3 samples x 4 concentrations (20; 10; 5.6; 2.56 ng) x 5 temperature (36/50/56/62/64)</td> <td>60 0</td> </tr> <tr> <td>Substrate</td> <td>9 common substrates x 4 samples per set</td> <td>36 12</td> </tr> <tr> <td>Environment</td> <td>9 common substrates x 1 sample each</td> <td>9 0</td> </tr> <tr> <td>Various tissues</td> <td>5 conditions (outside/inside/inside/outside/RT) x 6 time points (30/120/240 min)</td> <td>9 0</td> </tr> </tbody> </table> <p>Bone, hair, teeth, semen, paternity, urine, blood, semen, vaginal swab (mixtures of one sample each)</p> <p>TOTAL SAMPLES RUN: 632 200</p>			Study Category	Description of samples used with a description of conditions	Estimated Number of Samples	Single Source (Concordance)	8 samples (Proenza concordance) = 200 samples (part of population concordance study)	200 100	Mixtures	41	45 10	Peak Height Ratio	1 sample x 11 ratios (1.0; 19.1; 9.1; 5.1; 2.1; 1.1; 1.2; 1.5; 1.9; 1.19; 0.1) x 2 replicates (30 seconds)	22 39	Sensitivity	5 samples x 6 amounts (20/10; 5.6; 2.56; 1.50; 0.63; 0.32 ng) x 5 samples x 3 points (calibration standards)	66 33	Non-Human	11 animals	11 0	NIST SRM 2391b	12 components	12 12	Primer (ABI 310)	(3 samples x 10 replicates) x 10 different locations of white laundry	60 60	Non-Probative Cases	5 cases x 4 samples each (evidence from FBI Laboratory)	20 20	Stutter	200 samples (data used from population samples)	- -	Peak Height Ratio	(3 samples x 10 replicates) x 10 different locations of white laundry	- -	Cycling Parameters	200 samples (data used from population samples)	- -	Annealing Temperature	14 samples x 2 different cycle annealing temperatures x 2 replicates (30 seconds)	66 -	Proficiency	3 samples x 4 concentrations (20; 10; 5.6; 2.56 ng) x 5 temperature (36/50/56/62/64)	60 0	Substrate	9 common substrates x 4 samples per set	36 12	Environment	9 common substrates x 1 sample each	9 0	Various tissues	5 conditions (outside/inside/inside/outside/RT) x 6 time points (30/120/240 min)	9 0
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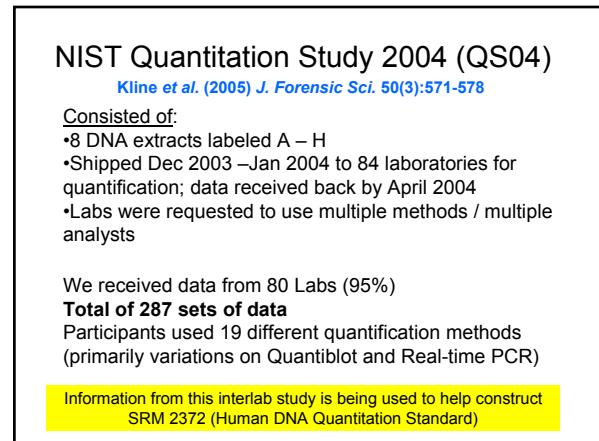
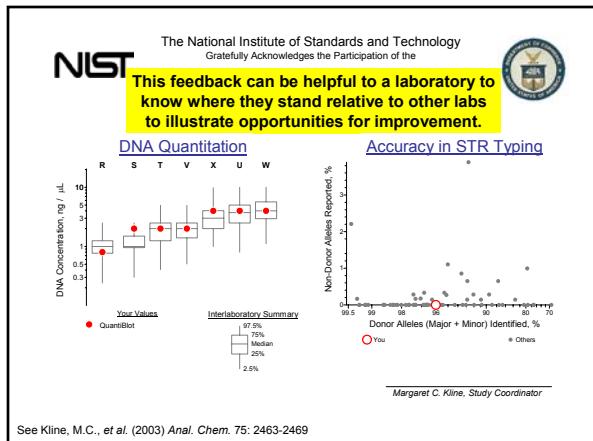

Interlaboratory Studies

Margaret Kline Dave Duewer Jan Redman John Butler

- DNA Quantitation Study (QS04)**
 - 8 DNA samples supplied
 - 84 laboratories signed up (80 labs returned results)
 - 287 data sets using 19 different methods
 - 60 data sets with real-time qPCR (37 Quantifiler data sets)
 - Publication in May 2005: *J. Forensic Sci.* 50(3): 571-578
- Mixture Interpretation Study (MIX05)**
 - 105 labs signed up (71 labs returned data)
 - Interpretation requested of provided e-grams for 4 mock sexual assault cases
 - Data analysis is still on-going...

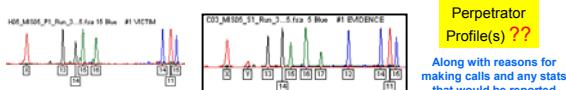
<http://www.cstl.nist.gov/biotech/strbase/interlab.htm>

NIST Initiated Interlaboratory Studies		
Studies involving STRs	# Labs	Publications
Evaluation of CSF1PO, TPOX, and TH01	34	Kline MC, Duewer DL, Newall P, Redman JW, Reeder DJ, Richard M. (1997) Interlaboratory evaluation of STR triplex CTT. <i>J. Forensic Sci.</i> 42: 897-906
Mixed Stain Studies #1 and #2 (Apr-Nov 1997 and Jan-May 1999)	45	Duewer DL, Kline MC, Redman JW, Newall PJ, Reeder DJ. (2001) NIST Mixed Stain Studies #1 and #2: interlaboratory comparison of DNA quantification practice and short tandem repeat multiplex performance with multiple-source samples. <i>J. Forensic Sci.</i> 46: 1199-1210
MSS3	74	Kline, M.C., Duewer, D.L., Redman, J.W., Butler, J.M. (2003) NIST mixed stain study 3: DNA quantitation accuracy and its influence on short tandem repeat multiplex signal intensity. <i>Anal. Chem.</i> 75: 2463-2469.
Mixed Stain Study #3 (Oct 2000-May 2001)	74	Duewer, D.L., Kline, M.C., Redman, J.W., Butler, J.M. (2004) NIST Mixed Stain Study #3: signal intensity balance in commercial short tandem repeat multiplexes. <i>Anal. Chem.</i> 76: 6928-6934.
DNA Quantitation Study (Jan-Mar 2004) QS04	80	Kline, M.C., Duewer, D.L., Redman, J.W., Butler, J.M. (2005) Results from the NIST 2004 DNA Quantitation Study. <i>J. Forensic Sci.</i> 50(3):571-578
MIX05	71	Data analysis currently on-going ... Will be presented at NIJ Grantees (June 2005), ISFG (Sept 2005), Promega (Sept 2005)



Mixture Interpretation Interlab Study (MIX05)

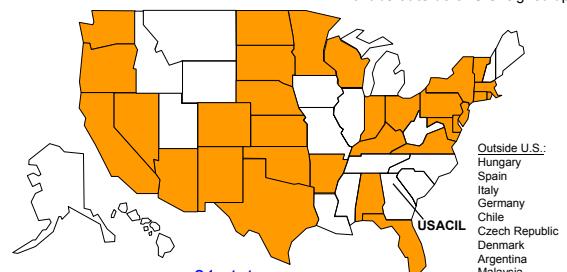
- Only involves interpretation of data
- 105 labs enrolled for participation (20 from overseas)
- 71 labs have returned results**
- Four mock cases supplied with "victim" and "evidence" electropherograms (GeneScan .fsa files – that can be converted for Mac or GeneMapper; gel files made available to FMBIO labs)
- Data available with Profiler Plus, COfiler, SGM Plus, PowerPlex 16, Identifier, PowerPlex 16 BIO (FMBIO) kits
- Summary of results will involve training materials to illustrate various approaches to solving mixtures



Participation in NIST Interlaboratory Study on Mixture Interpretation (MIX05)

Companies:
Myriad Genetics

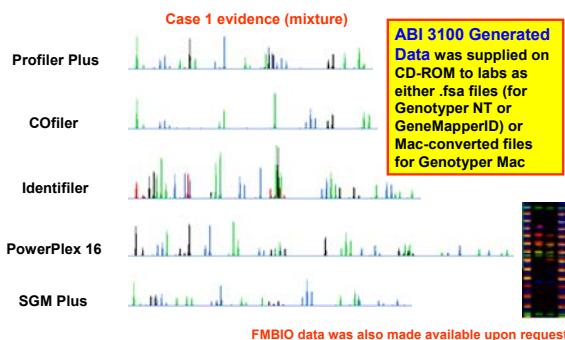
20 labs outside of U.S. signed up



105 laboratories signed up for study (71 returned results so far)

MIX05 Results on Multiple Kits

<http://www.cstl.nist.gov/biotech/strbase/interlab/MIX05.htm>



Purpose of MIX05 Study

- Goal is to understand the "lay of the land" regarding mixture analysis across the DNA typing community
- "If you show 10 colleagues a mixture, you will probably end up with 10 different answers"
 - Peter Gill, Human Identification E-Symposium, April 14, 2005
- One of the primary benefits we hope to gain from this study is **recommendations for a more uniform approach to mixture interpretation** and training tools to help educate the community

Value of the MIX05 Study

- Data sets exist with multiple mixture scenarios and a variety of STR kits that **can be used for training purposes**
- A wide variety of approaches to mixture interpretation have been applied on the **same data sets evaluated as part of a single study**
- Interpretation guidelines from many laboratories are being compared** to one another for the first time in an effort to determine challenges facing future efforts to develop "expert systems" for automated mixture interpretation
- We are exploring the challenges of supplying a common data set to a number of forensic laboratories (e.g., if a standard reference data set was ever desired for evaluating expert systems)

Technology: Research Programs

- miniSTRs
- Y-chromosome STRs
- mtDNA
- SNPs
- qPCR for DNA quantitation
- DNA stability studies
- Variant allele characterization and sequencing
- Software tools
- Assay development with collaborators

Standard U.S. Population Dataset

<http://www.cstl.nist.gov/biotech/strbase/NISTpop.htm>

260 Caucasians, 260 African Americans, 140 Hispanics, 3 Asians = **663 males**

DNA extracted from whole blood (anonymous; self-identified ethnicities) received from Interstate Blood Bank (Memphis, TN) and Millennium Biotech Inc. (Ft. Lauderdale, FL)

To date: (**>100,000 allele calls**)

Identifier (15 autosomal markers + Amelogenin) (10,608)
Roche Linear Arrays (HV1/HV2 10 regions) (6,630)
Y STRs 22 loci—27 amplicons (17,388)
Y STRs 27 new loci (14,535)
Yfiler kit 17 loci (11,237)
Y SNPs 50 markers on sub-set of samples (11,498)
Orchid 70 autosomal SNPs on sub-set (13,230)
miniSTR testing-new loci and CODIS concordance (9,228)
New miniSTR loci – for 11 loci, 7,293 genotypes
mtDNA full control region sequences by AFDIL

extracted genomic DNA
Stock tubes

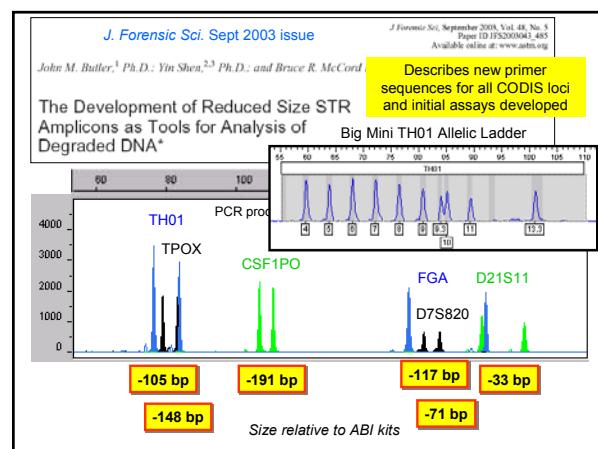
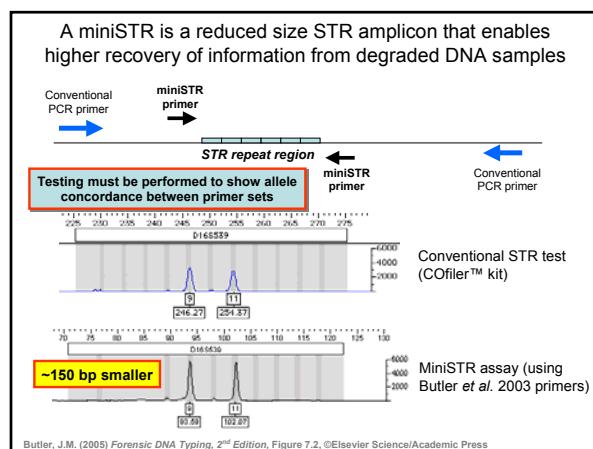
↓
Genotypes with various human identity testing markers

miniSTRs for Degraded DNA

Mike Coble Becky Hill John Butler

- Original miniSTR paper with CODIS loci, D2, D19, Penta D, Penta E
– *Butler et al. (2003) J. Forensic Sci.* 48: 1054-1064
- Many CODIS loci are too big and make poor miniSTRs
- New miniSTRs and assays: NC01, NC02
– *Coble, M.D. and Butler, J.M. (2005) J. Forensic Sci.* 50:43-53
- New miniSGM miniplex: AMEL, TH01, FGA, D18, D16, D2
- EDNAP/ENFSI degraded DNA study coordinated by Peter Gill
- Creation of miniSTR information on STRBase

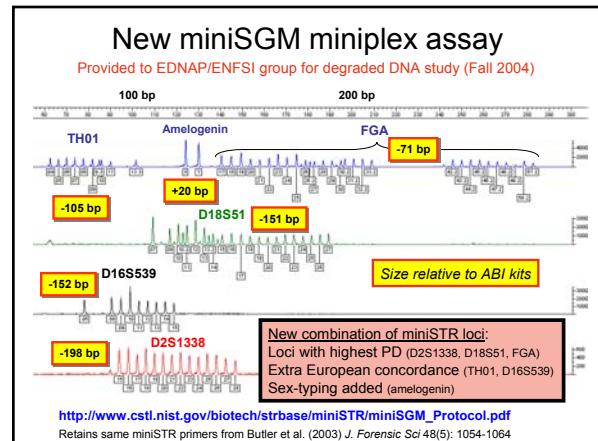
<http://www.cstl.nist.gov/biotech/strbase/miniSTR.htm>

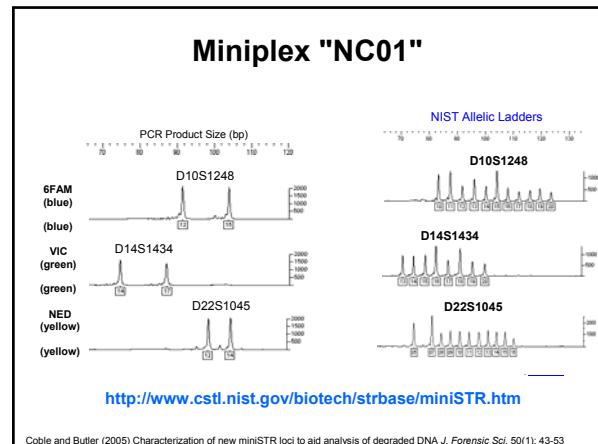
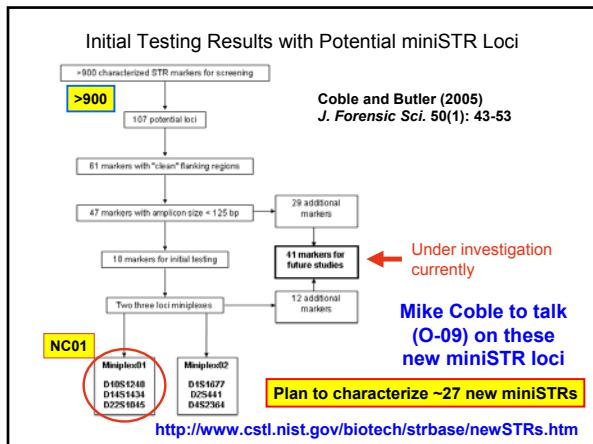


Recent Publications on miniSTRs

- Butler, J.M., Shen, Y., McCord, B.R. (2003) The development of reduced size STR amplicons as tools for analysis of degraded DNA. *J. Forensic Sci.* 48(5): 1054-1064.
- Chung, D.T., Drabek, J., Opel, K.L., Butler, J.M., McCord, B.R. (2004) A study on the effects of degradation and template concentration on the efficiency of the STR miniplex primer sets. *J. Forensic Sci.* 49(4): 733-740.
- Drabek, J., Chung, D.T., Butler, J.M., McCord, B.R. (2004) Concordance study between miniplex STR assays and a commercial STR typing kit. *J. Forensic Sci.* 49(4): 859-860.
- Coble, M.D. and Butler, J.M. (2005) Characterization of new miniSTR loci to aid analysis of degraded DNA. *J. Forensic Sci.*, 50: 43-53.

<http://www.cstl.nist.gov/biotech/strbase/miniSTR.htm>
<http://www.cstl.nist.gov/biotech/strbase/miniSTR/timeline.htm>





New Autosomal STR Loci

Mike Coble Becky Hill John Butler

- NC01 loci: **D10S1248, D14S1434, D22S1045**
- Peter Gill and the EDNAP/ENFSI group have recommended the NC01 loci as an extension of current European core loci following their demonstrated success in the recent EDNAP degraded DNA interlab study
- Being adopted/explored in multiple U.S. paternity testing labs (BRT Labs, Orchid Cellmark East Lansing, DNA Diagnostics Center)
- Population data, locus characterization, and allelic ladders for **27 new autosomal STRs under development** as new miniSTRs
- All new STR loci are physically unlinked to CODIS and ENFSI core loci

<http://www.cstl.nist.gov/biotech/strbase/newSTRs.htm>

Work with Y-STRs

John Butler Margaret Kline Pete Vallone Amy Decker

- Beta-testing of all commercial Y-STR kits
- U.S. population data supplied to Yfiler haplotype database
- **49 Y-STR loci evaluated with ~650 U.S. samples**
- New Y-chromosome information on STRBase linking to all available haplotype databases
- Human Y-Chromosome DNA Profiling Standard Reference Material (SRM 2395) – updates with DYS635 for Yfiler
- **Separation of two brothers with 47 Y-STRs**

http://www.cstl.nist.gov/biotech/strbase/y_strs.htm

New Y-Chromosome Information

Resources on STRBase

http://www.cstl.nist.gov/biotech/strbase/y_strs.htm

Commercial Y-STR Kits

- Europlex® Y (Eurogentec)
- PowerPlex® Y (Promega)
- PowerPlex® Y Multiplex (Promega)
- Europlex® Y-PLEX 5 Y-PLEX (Bioscience Technologies) – will not be sold after May 1, 2005
- DYSplex L, DYSplex L (Sevit, Bad Honnef, Germany)
- Menwyn® August Y-M1 (Menwyn, Dresden, Germany)

Haplotype Databases

- YHRD (Y Chromosome Haplotype Reference Database) (32,196 haplotypes with 9 loci): <http://www.yhrd.org/yhrd.html>
- YHRD with minimal haplotypes = D10S1248 and D14S1434 (7,097 haplotypes with 11 loci): http://www.yhrd.org/yhrd_min.html
- YHRD 10,000 haplotypes with 11 loci: http://www.ednaps.net/index.asp?menu_id=1&content_id=10
- Yfiler Haplotype Database (3,382 haplotypes with 17 loci): <http://www.appliedbiosystems.com/RealTimePCR/Products/YFILER/Products/Genotyping/Genotyping.aspx>
- Genetic Genealogy FamilyTree DNA Y-DNA STR (with 14, 22, or 27 loci): <http://www.ystr.org/>
- Genetic Genealogy Sorenson Molecular Genealogy Foundation (12,211 haplotypes with 16 loci): <http://www.smg.org/>
- Genetic Genealogy GedMatch (with 16 loci): <http://www.gedmatch.com/>

Y Chromosome Links

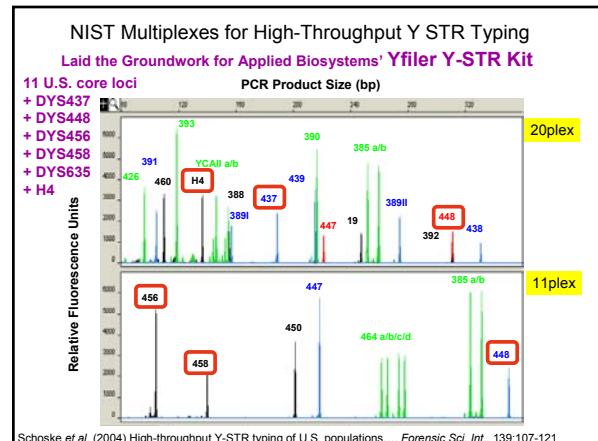
- Y-STR Haplotype Reference Database: <http://www.yhrd.org/yhrd.html>
- Y-STR Haplotype Reference Database: <http://www.ncbi.nlm.nih.gov/sra/>
- The Y Chromosome Consortium: <http://www.biotrust.ameba.jp/>
- Genetic Genealogy Family Tree DNA: <http://www.familytreedna.com>
- Genetic Genealogy DNA Heritage: <http://www.dnaheritage.com>
- Genetic Genealogy DNA Health: <http://www.dnahealth.com>
- Genetic Genealogy Oxford Ancestry: <http://www.oxfordancestry.com>
- Genetic Genealogy African Ancestry: <http://www.ashkenazi.com>
- Genetic Genealogy GedMatch (with 16 loci): <http://www.gedmatch.com/>
- Genetic Genealogy GedTree DNA Testing Center: <http://www.gedtree.com>

Locus boxes are hyperlinked to STR Fact Sheets

PowerPlex® Y

Largest Y-STR Database

YHRD has 7,897 haplotypes with SWGDAM recommended loci



Yfiler Haplotype Database

<http://www.appliedbiosystems.com/yfilerdatabase/>

Population	# Haplotypes
African American	985
Asian	330
Caucasian	1276
Filipino	105
Hispanic	597
Native American	106
Sub-saharan African	59
Vietnamese	103
All	3561

#Samples Contributed by NIST

Population	# Samples
259 African Americans	
3 Asians	
262 Caucasians	
139 Hispanics	

Data provided by NIST

663/3561 = 18.6%

NIST Work with New Y-STR Loci

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Available online at www.sciencedirect.com

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Forensic Science International xxx (2005) xxx-xxx

Announcement of population data

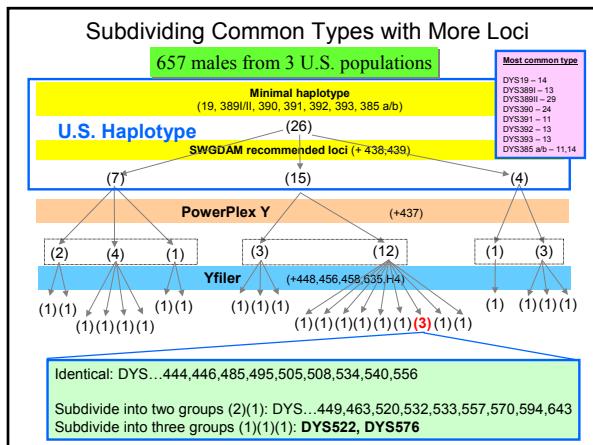
Allele frequencies for 27 Y-STR loci with U.S. Caucasian, African American, and Hispanic samples

John M. Butler*, Amy E. Decker, Peter M. Vallone, Margaret C. Kline
Biotechnology Division, National Institute of Standards and Technology, Gaithersburg, MD 20899-8711, USA
Received 26 January 2005; received in revised form 2 February 2005; accepted 22 February 2005

Abstract

A total of 263 U.S. Caucasians, 260 African Americans and 140 U.S. Hispanics, Americans, and 32 Hispanics were typed for 27 Y-chromosome short tandem repeat (Y-STR) markers: DYS444, DYS446, DYS449, DYS463, DYS485, DYS494, DYS504, DYS505, DYS508, DYS520, DYS522, DYS525, DYS532, DYS533, DYS534, DYS540, DYS556, DYS557, DYS570, DYS575, DYS576, DYS594, DYS632, DYS635, DYS641, and DYS643. Allele frequencies for each locus are reported along with nomenclature based on sequence analysis.

14,535 types generated across 27 new loci



Y-Chromosome Standard NIST SRM 2395

Available since July 2003

Human Y-Chromosome DNA Profiling Standard

STANDARD REFERENCE MATERIAL®
2395
Human Y Chromosome DNA Components A – F
Store at -20°C
www.nist.gov

• 5 male samples + 1 female sample (neg. control)
• 100 ng of each (50 µL at ~2 ng/µL)
• 22 Y STR markers sequenced
• 9 additional Y STR markers typed
• 42 Y SNPs typed with Marilgen kit

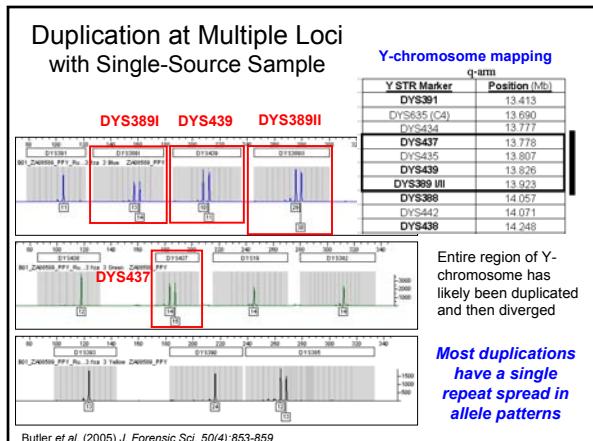
Certified for all loci in commercial Y-STR kits:

- Y-PLEX 6
- Y-PLEX 5
- Y-PLEX 12
- PowerPlex Y

SWGDAM recommended loci:
DYS19, DYS385 a/b, DYS389 I/II, DYS390, DYS391, DYS392, DYS393, DYS394, DYS395, DYS396, DYS397, DYS398, DYS399 III, DYS437, DYS438, DYS442, DYS443, DYS444, DYS446, DYS449, DYS463, DYS485, DYS494, DYS504, DYS505, DYS520, DYS522, DYS525, DYS532, DYS533, DYS534, DYS540, DYS556, DYS557, DYS570, DYS575, DYS576, DYS594, DYS632, DYS635, DYS641, and DYS643

Y-filer - adds DYS635 (C4); now sequenced

Helps meet FBI Standard 9.5 (and ISO 17025)...traceability to a national standard



Work with mtDNA

Mike Coble Pete Vallone Margaret Kline John Butler

- Collaboration with Armed Forces DNA Identification Laboratory to develop multiplex mtDNA SNP assays for coding region polymorphisms
- Beta-testing and automation of Roche LINEAR ARRAY HVI/HVII probes
- Population study performed with LINEAR ARRAY HVI/HVII probes
- Exploration of effective strategies for forensic analysis in the mitochondrial DNA coding region

<http://www.cstl.nist.gov/biotech/strbase/mtDNA.htm>

<http://www.cstl.nist.gov/biotech/strbase/>

Short Tandem Repeat DNA
Internet DataBase

These data are intended to benefit research and application of short tandem repeat DNA markers in forensic testing. The authors are solely responsible for the information levels. ©2002 NIST.

This database has been accessed 11,032 times since 1/05/97. (Create a counter www.cstl.nist.gov/biotech/strbase/)

Created by John M. Butler and Dennis Z. Butler, NIST Biotechnology Division, with invaluable help from Jim Redman, Christian Rothberg and Michael Long.

Site creators' names: <http://www.cstl.nist.gov/biotech/strbase/>

*Partial support for the design and maintenance through the FBI.

Publications & Presentations from 2002 (Search)

Forensic STR Information

ISFC ENFSI DNA[®] Group

This site is intended to provide general information on single nucleotide polymorphism (SNP) markers that may be of interest in human identification applications. Many of these markers come from the SNP Consortium (TDB) efforts or are already present in the NCBI SNP database. To submit a SNP marker for inclusion on this forensic SNP site, please provide the required information on a standard SNP sheet (John.Butler@nist.gov)

(Blanket) (Award) (SRM Typing Technology)

Will be updated with new SNP information from this meeting

Last Updated: 04/21/04

Allele Frequencies for 70 SNP Loci in U.S. Populations														
African American														
CC	0.405	1	AA	0.595	N = 74	CC	0.243	0.409	0.068	0.981	0.311	0.149	0.481	0.108
CT	0.477	2	AT	0.522		CT	0.243	0.136	0.514	0.135	0.189	0.338	0.122	0.378
TT	0.078	3	TC	0.922		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	4	GC	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GT	0.000	5	CT	1.000		GT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
AC	0.000	6	AT	1.000		AC	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
AT	0.000	7	TC	1.000		AT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
AG	0.000	8	CG	1.000		AG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TC	0.000	9	CT	1.000		TC	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CC	0.000	10	CC	1.000		CC	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CT	0.000	11	CT	1.000		CT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	12	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	13	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	14	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	15	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CC	0.000	16	CC	1.000		CC	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CT	0.000	17	CT	1.000		CT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	18	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	19	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	20	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	21	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CC	0.000	22	CC	1.000		CC	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CT	0.000	23	CT	1.000		CT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	24	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	25	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	26	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	27	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CC	0.000	28	CC	1.000		CC	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CT	0.000	29	CT	1.000		CT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	30	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	31	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	32	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	33	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CC	0.000	34	CC	1.000		CC	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CT	0.000	35	CT	1.000		CT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	36	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	37	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	38	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	39	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CC	0.000	40	CC	1.000		CC	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CT	0.000	41	CT	1.000		CT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	42	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	43	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	44	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	45	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CC	0.000	46	CC	1.000		CC	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
CT	0.000	47	CT	1.000		CT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	48	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	49	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	50	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	51	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	52	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	53	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	54	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	55	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	56	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	57	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	58	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	59	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	60	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	61	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	62	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	63	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	64	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	65	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	66	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	67	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	68	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
TT	0.000	69	TT	1.000		TT	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216
GG	0.000	70	GG	1.000		GG	0.243	0.446	0.243	0.162	0.473	0.365	0.108	0.216

Vallone et al. (2005) *Forensic Sci. Int.* 149:279-286

Publication on U.S. Groups with Y-SNPs

J. Forensic Sci. 2004; 49(4): 723-732

Peter M. Vallone,¹ Ph.D. and John M. Butler,¹ Ph.D.

Y-SNP Typing of U.S. African American and Caucasian Samples Using Allele-Specific Hybridization and Primer Extension*

Different technologies yield the same Y-SNP type
Full concordance was observed between hybridization and primer extension technologies on 18 different Y-SNPs (>3,800 allele calls)

Y-SNPs will have limited value for individualizing a sample
18 different types observed in 229 individuals

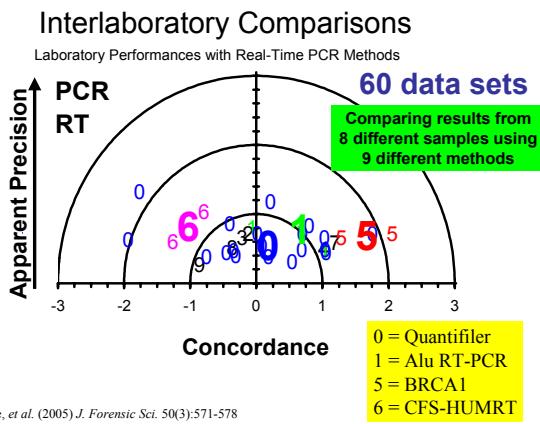
Current Y-SNPs appear to have limited value for ethnic differentiation in U.S. populations (with the exception of M2 that is only found in African Americans and not in Caucasians)

Evaluation of qPCR Assays

Margaret Kline, Pete Vallone, Amy Decker

- Evaluation of published assays on same samples
- Characterization of Quantifiler lot-to-lot performance
- Additional studies under way utilizing qPCR:
 - Examining the challenge of multiplexing qPCR assays
 - Studies to track DNA recovery from various types of tubes
 - Characterizing potential SRM 2372 components (Human DNA Quantitation Standard)

<http://www.cstl.nist.gov/biotech/strbase/DNAQuant.htm>



DNA Stability Studies

Margaret Kline

Dave Duewer

- Comparison of DNA recovery from FTA, IsoCode, S&S 903, Whatman papers with bloodstains from the same individuals
- Recovery of DNA from aged bloodstains on untreated paper stored at room temperature



Anal. Chem. (2002) 74:1863-1869
Polymerase Chain Reaction Amplification of DNA from Aged Blood Stains: Quantitative Evaluation of the "Suitability for Purpose" of Four Filter Papers as Archival Media

Margaret C. Kline,¹ David L. Deamer,² Jasmin W. Robinson,¹ and John M. Butler¹
¹Chemical Sciences and Technology Laboratory, National Institute of Standards and Technology
²Scripps Institution of Oceanography

David A. Boyce
Department of Defense DNA Registry, Armed Forces Institute of Pathology, Suite 530, 16000 Rockville Drive, Bethesda, Maryland 20889

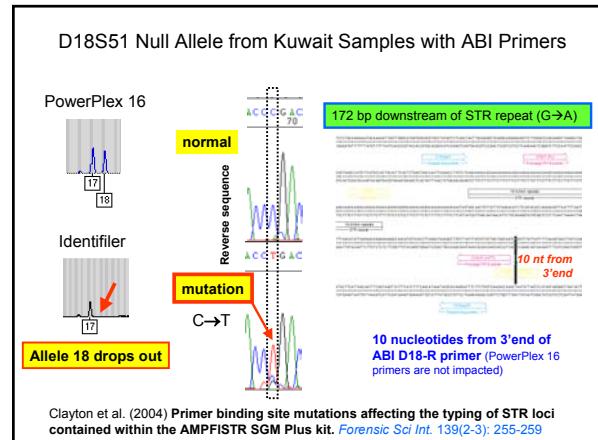
<http://www.cstl.nist.gov/biotech/strbase/NIJ/DNArecovery.htm>

STR Allele Sequencing and Characterization

Margaret Kline John Butler

- Variant characterization
 - TPOX 10.3 (Maryland State Police)
 - D18S51 null alleles** (FSS and Kuwait govt)
 - D18S51 allele 40 (Nebraska State Crime Lab)
 - D18S51 allele "5.3" (DNA Solutions)
 - DYS392 allele "10.3" (AFDIL)
 - VWA allele "15.1" (Peter de Knijff)
- Locus duplication or deletion
 - DYS390 deletion (CFS Toronto)
 - DYS392 deletion (MN BCA)
- Forensic labs are sending us unusual STR alleles for sequence characterization**

<http://www.cstl.nist.gov/biotech/strbase/STRseq.htm>



Software Tools

Pete Vallone Dave Duewer Chris DeAngelis

- AutoDimer – multiplex PCR primer screening tool
<http://www.cstl.nist.gov/biotech/strbase/AutoDimerHomepage/AutoDimerProgramHomepage.htm>
- mixSTR – mixture component resolution tool
- Multiplex_QA – quality assessment tool for monitoring instrument performance over time
- NIST U.S. population database (internal Access database)

<http://www.cstl.nist.gov/biotech/strbase/software.htm>

AutoDimer Overview

SHORT TECHNICAL REPORTS

AutoDimer: a screening tool for primer-dimer and hairpin structures

Peter M. Vallone and John M. Butler
National Institute of Standards and Technology, Gaithersburg, MD, USA

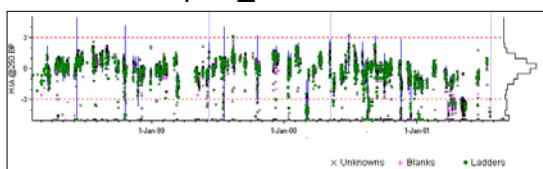
BioTechniques 37:226-231 (August 2004)

- Evaluates potential primer dimers and hairpins in batch mode.
- Requires input of primer list (already chosen in singleplex fashion with Primer3 or some other program). Runs as stand-alone Visual Basic program.
- Used in development of all NIST multiplex PCR/STR/SNP assays.

Available for download from STRBase:

<http://www.cstl.nist.gov/biotech/strbase/software.htm>

Multiplex_QA Overview

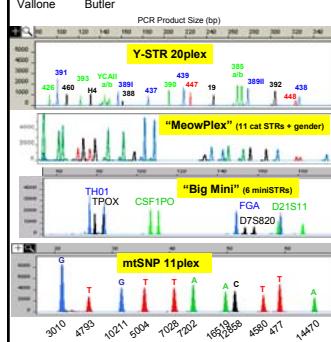


- **Research tool** that provides quality metrics to review instrument performance over time (e.g., examines resolution on internal size standard peaks)
- Runs with Microsoft Excel macros. Requires STR data to be converted with NCBI's BatchExtract program into numerical form.

Available for download from STRBase:

<http://www.cstl.nist.gov/biotech/strbase/software.htm>

Assay Development with Collaborators



- **Y-STR 20plex** (Mike Hammer and Alan Redd)
 - Butler et al. (2002) FSI
- **Cat STRs** (Marilyn Raymond and Victor David)
 - Butler et al. (2002) Profiles in DNA
 - Raymond et al. (2005) JFS
- **miniSTRs** (Bruce McCord)
 - Butler et al. (2003) JFS
- **mtDNA coding region SNPs (AFDIL)**
 - Vallone et al. (2004) IJLM
- **Others...**

Training Materials and Review Articles



- Workshops on STRs and CE (ABI 310/3100)
 - Taught with Bruce McCord (Florida Int. Univ.)
 - NEAFS (Sept 29-30, 2004)
 - U. Albany DNA Academy June 13-14, 2005)
- Validation Workshop
 - Taught with Robyn Ragsdale (FDLE) at NFSTC (August 24-26, 2005)
- PowerPoint slides from *Forensic DNA Typing, 2nd Edition*
 - >150 slides available now (~1,000 planned) for download
 - <http://www.cstl.nist.gov/biotech/strbase/FDT2e.htm>
- Review articles
 - ABI 310 and 3100 chemistry – Electrophoresis 2004, 25, 1397-1412

<http://www.cstl.nist.gov/biotech/strbase/training.htm>
<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

NEAFS Workshop Slide Handouts

Handouts available as downloadable pdf files from
<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm#NEAFSworkshop>

2 day workshop with **>500 slides** describing STRs and CE (ABI 310 and ABI 3100)

NEAFS CE-DNA Workshop (Butler and McCord)	Sept 29-30, 2004
Capillary Electrophoresis in DNA Analysis STR Analysis NEAFS Workshop Mystic, CT September 29-30, 2004 Dr. John M. Butler Dr. Bruce R. McCord	
Outline for Workshop <ul style="list-style-type: none"> • Introduction • STR Analysis • Introduction to CE and ABI 310 • Data Interpretation • Additional Topics – Real-time PCR and miniSTRs • Higher Throughput Approaches • Troubleshooting the ABI 310 (Participant Roundtable) • Additional Topics – Y-STRs, validation, accuracy • Review and Test 	

Review Article on STRs and CE

pdf available from <http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

Electrophoresis 2004, 25, 1397-1412

Review

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Forensic DNA using the ABI 310 for STR analysis

Contents	
1	Introduction
1.1	General aspects
1.2	Early work with CE
2	Sample preparation and injection
3	Sample separation
3.1	The polymer separation matrix
3.2	The buffer
3.3	The capillary
4	DNA typing with short tandem repeat applications including such as the ABI Prism for many laboratories
5	Sample detection
5.1	Sample interpretation
5.2	Software used
6	Assessing resolution of DNA separations
6.1	Applications of forensic DNA testing
6.2	Forensic casework
6.3	DNA databasing
7	Increasing sample throughput
7.1	Capillary array electrophoresis systems
7.2	Microchip CE systems
7.3	Future methods for DNA typing with STR markers
8	References

Analytical Chemistry Application Review

June 15, 2005 issue of *Analytical Chemistry*

Forensic Science

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250 articles referenced
covering forensic DNA analysis during 2003-2004

Review Contents

- Forensic DNA Analysis
- Collection, Characterization, Preservation, Extraction, and Quantitation of Biological Material
- Short Tandem Repeats
- Single-Nucleotide Polymorphisms
- YSTR, Y-DNA, and X-Chromosome Identification, and X-Chromosome Analysis
- Mitochondrial DNA Typing
- Nonshuman DNA Typing Systems and Microbial Profiling
- DNA Databases
- Interpretation and Statistical Weight of DNA Typing Results
- General Reviews

Acknowledgments

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John Butler Margaret Kline Pete Vallone Mike Coble Jan Redman Amy Decker Becky Hill Chris DeAngelis Dave Duewer

The many forensic scientists and their supervisors who took time out of their busy schedules to examine the MIX05 data provided as part of this interlaboratory study

Past and Present Collaborators (also funded by NIJ):

- Mike Hammer and Alan Redd (U. AZ) for Y-chromosome studies
- Tom Parsons, Rebecca Just, Jodi Irwin (AFDIL) for mtDNA coding SNP work
- Sandy Calloway (Roche) for mtDNA LINEAR ARRAYS
- Bruce McCorr and students (FL Int. U.) for miniSTR work
- Marilyn Raymond and Victor David (NCI-Frederick) for cat STR work
- Artie Eisenberg and John Planz (U. North Texas) for miniSTR testing on bones
- Murray Brilliant (U. AZ) for phenotype markers
- Ken Kidd (Yale U.) for SNP typing population samples

Thank you for your attention...

Questions?

Our publications and presentations are available at:
<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

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