

Session 5D

Quantitative PCR (qPCR) Tools for the DNA Analysis of Challenging Samples: the CAL DOJ Triplex Degradation Assay

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How To *Identify* Challenging Samples and Which Tool(s) to Use?

- **experience** (analyst, intra-lab, inter-lab, literature)
- **unsuccessful analysis** using routine methods
 - *i.e.*, partial or null typing results
 - ✓ inefficient use of analyst time
 - ✓ inefficient use of reagents and kits
 - ✓ inefficient use of (possibly limited) DNA extract
 - ✓ increased documentation and review

Goal

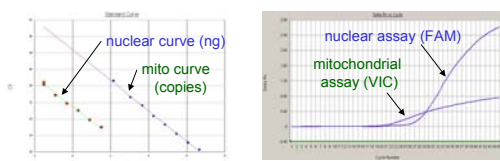
- to develop real-time qPCR tools to help identify challenging samples so that optimal strategies can be attempted at the outset of analysis
 - measure human DNA **quantities**
 - total nuclear DNA
 - mitochondrial genome copies
 - total male DNA (Y-chromosome)
 - assess DNA **quality**
 - degree of degradation
 - presence of PCR inhibitors

qPCR Assays at CA DOJ

- duplex nuclear-mitochondrial qPCR assay
 - developed for Missing Persons DNA Program (MPDP-Richmond), but also used for non-MPDP cases
 - on-line for ABI 7000; >20 qualified analysts
- triplex qPCR assay to assess nuclear DNA **quantity** and **quality** (DNA degradation and presence of PCR inhibitors)
 - developed for general casework (non-mito)
 - DOJ-wide (Richmond DNA lab and DOJ field labs)
 - validated on ABI 7500; analysts in training

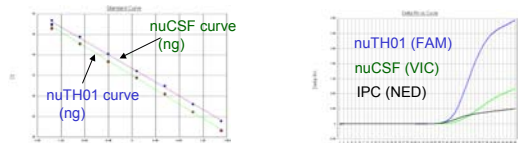
nuTH01-mtND1 Duplex qPCR Assay

- (development & validation: J.Forensic Sci. (2005) v.50, pp. 1044-60)
- on-line at Richmond DOJ DNA Lab ~1 year (~200 runs)
 - custom primers and TaqMan(MGB) probe oligonucleotides to co-amplify and detect nuclear and mitochondrial target sequences
 - validated on ABI 7000 SDS qPCR instrument
 - ABI Universal TaqMan Master Mix (+ BSA, 0.16 ug/uL)
 - 20 uL PCR with up to 4 uL of sample
 - DNA Standard: Promega Genomic Human DNA (male)



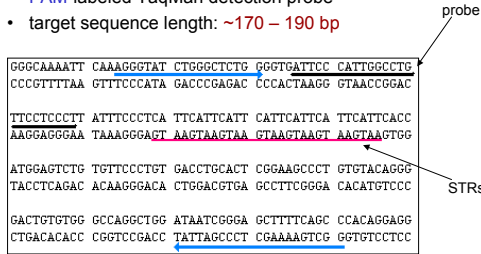
nuTH01-nuCSF-IPC Triplex qPCR Assay

- (development: Forensic Sci. International (2006), v. 158, pp. 14-26)
- **nuTH01 assay** (long nuclear target) **plus**:
 - nuCSF assay (short nuclear target) – for assessing degradation
 - IPC (internal PCR control) – for assessing PCR inhibition
 - validated on ABI 7500 qPCR instrument
 - 5-color instrument with improved NED detection and a Cy5 channel
 - ABI Universal TaqMan Master Mix (+ BSA, 0.16 ug/uL)
 - + 2.5 U AmpliTaq Gold
 - DNA Standard: Promega Genomic Human DNA (male)



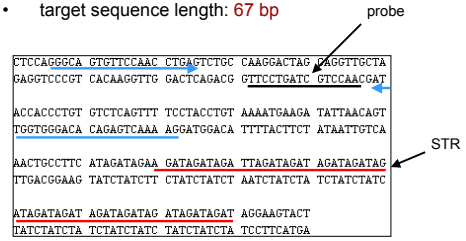
Nuclear nuTH01 qPCR Target

- target sequence spans TH01 CODIS STR locus (2 copies/diploid genome)
- FAM-labeled TaqMan detection probe
- target sequence length: ~170 – 190 bp



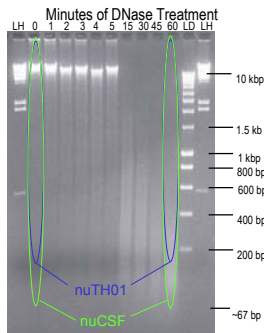
Nuclear nuCSF qPCR Target

- target sequence flanks the CODIS CSF STR region – (2 copies/diploid genome)
- VIC-labeled TaqManMGB detection probe
- target sequence length: 67 bp



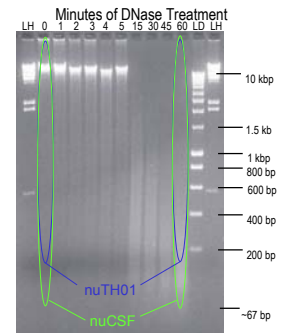
Using Short and Long Nuclear Targets to Assess DNA Fragmentation

- nuCSF assay – detects and quantifies DNA fragments larger than ~67bp
- nuTH01 assay – detects and quantifies DNA fragments larger than ~180bp

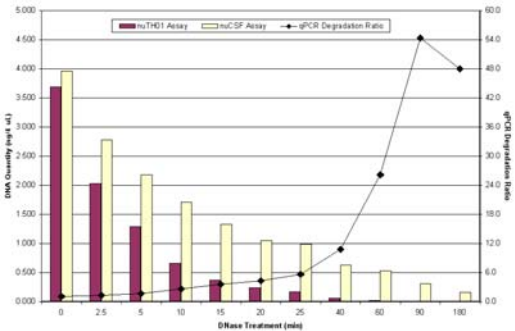


qPCR Degradation Ratio = nuCSF Quantity (ng) / nuTH01 Quantity (ng)

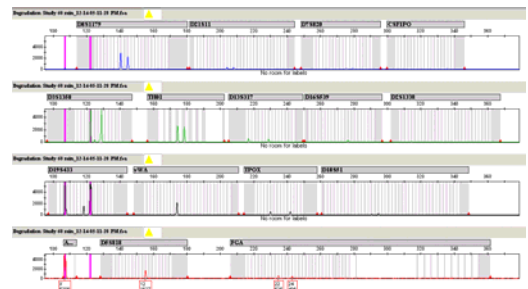
- For high-molecular weight DNA, expect the Degradation Ratio to be ~ 1.
- For highly-degraded DNA, expect the Degradation Ratio to be > 1.
- The bigger the qPCR Degradation Ratio, the more fragmented the DNA.



Triplex Quantifications and Degradation Ratios for DNaseI-Degraded Samples



qPCR Degradation Ratio ~ 25: "1 ng" (nuTH01) Identifier STR Results

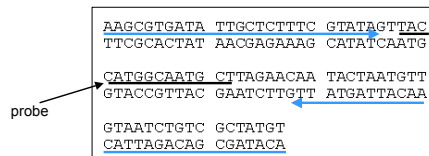


Interpreting the qPCR Degradation Ratio

Degradation Ratio	STR Implications
1 – 3	none
3 – 5	“wedge” effect, possible cross-dye pull-up
>5 (>10 ⇒ artifacts expected to be significant)	increasing “wedge” effect, pull-up, dropped-out alleles at larger loci, off-scale peaks, called stutter peaks, -A shouldering

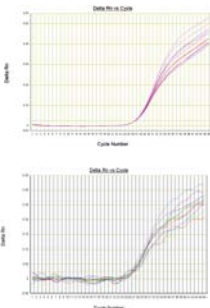
IPC qPCR Target

- template is a 77 nt synthetic oligonucleotide
- ‘artificial sequence’ - designed to be non-homologous to GenBank sequences
- oligo spiked into primer-probe mix (~90,000 copies/sample)
- NED-labeled TaqManMGB detection probe

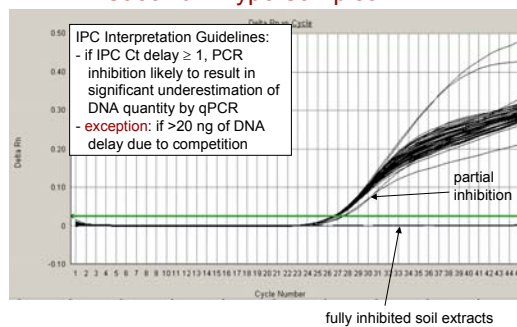


IPC for Non-Inhibited Samples Should Amplify to Give (Nearly) Identical Ct values

- ABI 7500 instrument
 - (assay **implementation**)
 - clean NED signal
 - identify PCR inhibition by
 - delay in Ct
- ABI 7000 instrument
 - (assay **development**)
 - poor NED signal-to-noise
 - easy to identify fully inhibited samples, hard to assess weak PCR inhibition



IPC qPCR Validation: Casework Type Samples



Semi-Automatic Calculation of Degradation Ratio and Delay in IPC Ct

- Excel Worksheet (Steven Myers, CA DOJ DNA Lab)
- for each qPCR run, Results are exported from
 - ABI 7500 analysis software and loaded into worksheet:
 - calculates the Degradation Ratio for each sample
 - “flags” samples with Degradation Ratio > 3 to alert for possibly degraded sample
 - calculates the delay in IPC Ct (“ΔIPC”) for each sample (= delay in Ct relative to the average IPC Ct for DNA standards < 32ng)
 - “flags” samples with ΔIPC > 0.75 cycles to alert for possibly inhibited sample
 - generates single-page report (values rounded to two significant figures)

Semi-Automatic Calculation of Degradation Ratio and Delay in IPC Ct

**Semi-Automatic Calculation
 of Degradation Ratio and Delay in IPC Ct**

	34 neat	34 1:10	34 1:100	34 1:1000
Und	Und	Und	1	-0.099
<LQ	<LQ	<LQ	<LQ	<LQ
<LQ	<LQ	<LQ	<LQ	<LQ

Sample Name	15
ΔIPC	-0.019
ΔqCSF	0.36
ΔqCSF	1.3
Retim	3.6

- Multiplex qPCR Analysis of
 Challenging Samples**
- nuTH01-nuCSF-IPC Triplex Assay
- IPC to detect PCR inhibition
 - re-quant with dilution series ?
 - achieve non-inhibited downstream analysis by:
 - » dilution
 - » augmented STR amp (extra Taq/BSA)
 - » sample clean-up

 - Degradation Ratio to detect DNA fragmentation
 - MiniSTRs (use nuCSF quant for input)
 - mitochondrial analysis