

Email: becky.hill@nist.gov

Phone: 301-975-4275

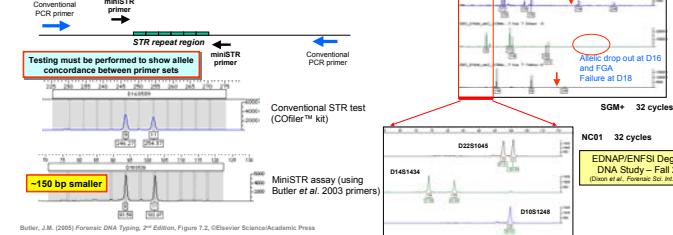
The development of new mini Short Tandem Repeat (miniSTR) loci for their use in forensic DNA typing is valuable in forensic casework involving DNA samples that are highly degraded due to exposure to environmental elements or contaminants (e.g. mass disasters such as the September 11, 2001 terrorist attack, the Indian Ocean Tsunami of December 2004, and Hurricane Katrina in August 2005). A number of studies have demonstrated that successful analysis of degraded DNA samples for human identification improve with smaller sized polymerase chain reaction (PCR) products (1). In addition, by moving PCR primers closer to the STR region, we have established that it is possible to decrease the incidence of allele or locus-dropout that may occur in degraded DNA samples.

The 27 new miniSTR loci were chosen based on their size and location on certain chromosomes. The candidate loci are all either located on chromosomes that differ from the 13 CODIS (COmbed DNA Index System) core loci or are at least ~ 50 Mb apart from an existing CODIS locus on the same chromosome, and therefore unlinked from that particular marker. New autosomal STR loci are being examined because many of the CODIS 13 core loci have large allele ranges (e.g., D21S11 and FGA) that make it impossible to create small PCR products. The first 6 loci (grouped into 2 miniplexes of 3 loci, NC01 and NC02) were previously characterized (2), and the process described for their development and characterization form the basis of the current work involving the remaining 21 new miniSTR loci. Each locus was evaluated across more than 600 samples representing the three major populations in the U.S.: Caucasian, African American, and Hispanic. All of the data was genotyped and allele frequencies and population statistics were determined.

Use of miniSTRs to Improve Analysis of Samples Containing Degraded DNA

In forensic casework, it is possible and quite likely that the DNA samples will be highly degraded due to exposure to environmental elements or natural contaminants that can result in loss of information at higher molecular weight STR loci. This loss of signal may be the result of the presence of PCR inhibitors or a DNA template that has been fragmented into smaller sizes, causing a partial DNA profile with allele or even complete locus dropout (1). Many of the commercial multiplex STR kits used in forensic DNA typing generate profiles up to 450 bp and can lead to loss of information at the higher molecular weight STR loci. In addition, many of the CODIS 13 core loci have large allele ranges that make it impossible to create small PCR products (1). Thus, by examining new STR loci that are the result of the size of the PCR products by moving the primers as close as possible to the STR repeat region, we have been able to observe a higher recovery of information from degraded DNA samples (2).

A miniSTR is a reduced size STR amplicon that enables higher recovery of information from degraded DNA samples

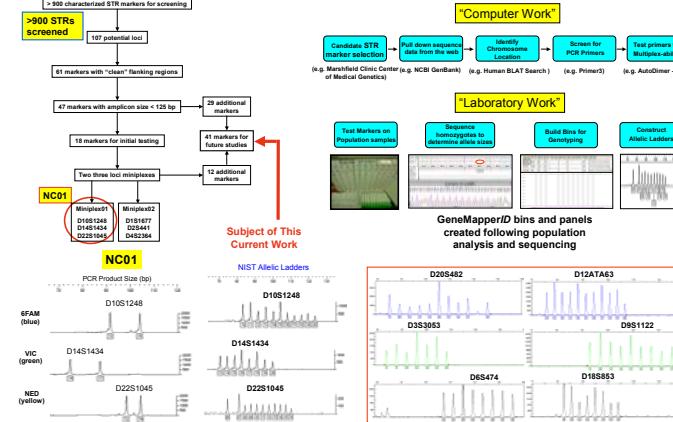


Butler, J.M. (2005) Forensic DNA Typing, 2nd Edition, Oxford Science/Academic Press

Selection and Characterization of New miniSTR Loci

The 27 new miniSTR loci were chosen based on their size and location on certain chromosomes. The candidate loci are all either located on chromosomes that differ from the 13 CODIS core loci or are at least ~ 50 Mb apart from an existing CODIS locus on the same chromosome, and therefore unlinked from that particular marker. The 27 new loci are separated into 9 groups of 3 loci, and are named "NC01 – NC09", for non-CODIS loci.

Coble and Butler (2005) J. Forensic Sci. 50(1): 43-53



Miniplex Sets Used in This Study

(choose to avoid linkage with CODIS 13 STRs to enable use of the product rule)

*Loci may be used in different combinations in the future**

NC01: D10S1249, D14S1434, D22S1045
NC02: D4S2364, D3S441, D16S1677
NC03: D3S3053, D6S474, D20S482
NC04: D16S1413, D12S1776, D5S2408
NC05: D151627, D5S200, D5S1115, D6S4529, D6S1017, P9S2517
NC07: P9S1122, D10S1453, D12A63, D22S1045
NC08: D17S1301, D18S853, D20S102
NC09: D6S1027, D17S974, D11S4463

Rejected Loci and Why

D9S234: allelic drop-out, complex repeats
D10S1430: complex repeats
D10S2327: tri-quad-allelic profiles
D14S297: poor heterozygosity
D15S817: tri-quad-allelic profiles

*32 loci x 663 samples = 21,216 total data points in this study

For more information, see STRbase:
<http://www.cstl.nist.gov/biotec/strbase/newSTRs.htm>
<http://www.cstl.nist.gov/biotec/strbase/miniSTR.htm>
<http://www.cstl.nist.gov/biotec/strbase/miniSTR/timeline.htm>

Positions determined along May 2004 Human Genome Reference Sequence (NCBI Build 35)



Development of 27 New MiniSTR Loci for Improved Analysis of Degraded DNA Samples

Carolyn R. Hill, Michael D. Coble, and John M. Butler

National Institute of Standards and Technology, Biochemical Science Division, 100 Bureau Drive MS 8311, Gaithersburg, MD 20899-8311

Copy of poster available:

<http://www.cstl.nist.gov/biotec/strbase/NISTpub.htm>

References

1. Butler, J.M., Shen, Y., McCord, B.R. (2003) The development of reduced size STR amplicons as tools for analysis of degraded DNA. *J. Forensic Sci.* 48(5): 1045-1064.
2. Coble, M.D. and Butler, J.M. (2005) Characterization of new miniSTR loci for the analysis of degraded DNA. *J. Forensic Sci.* 50: 43-53.
3. Gill, P., Ferday, L., Morling, N., Schneider, P.M. (2006) The evolution of DNA databases—recommendations for new European loci. *Forensic Sci. Int.* 158: 242-247.
4. Coble, M.D., Hill, C.R., Vellone, P.M., Butler, J.M. (2005) Characterization and performance of new miniSTR loci for typing degraded samples. *Progress in Forensic Genetics* 11, in press.
5. Dixon, L.A. et al. (2006) Analysis of artificially degraded DNA using STRs and SNPs—results of a collaborative European (EDNAP) exercise. *Forensic Sci. Int.*, in press.

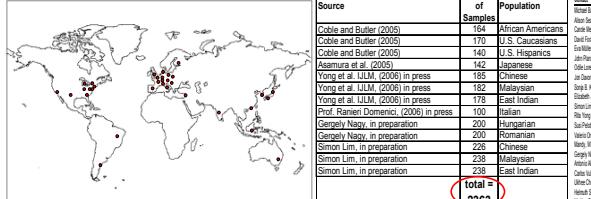
Population Data

A set of ~650 U.S. population samples (<http://www.cstl.nist.gov/biotec/strbase/NISTpub.htm>) were used to assess variation in African Americans, Caucasians, and Hispanics. The 3 loci boxed in blue are the new core European loci.



Population Data for NC01 and NC02 Loci is Being Collected by Other Labs

Global Impact of NC miniSTRs



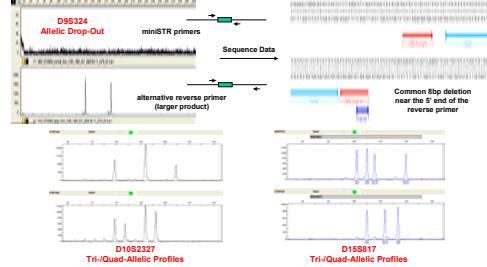
Materials and Methods

PCR primer sequences and full assay conditions will be included in a forthcoming publication

General protocols are available on the NIST STRBase website:

http://www.cstl.nist.gov/biotec/strbase/minISTR/updated_NC01_protocol.pdf
http://www.cstl.nist.gov/biotec/strbase/minISTR/Protocol_AlelicLadderRamp.pdf

Examples of Problematic Loci that were Rejected



Characteristics of New miniSTR Loci

Comparison of heterozygosity values for 27 non-CODIS miniSTRs (black) and 15 markers from the ABI Identifier loci (blue) across the U.S. samples examined in this study. The 3 loci boxed in blue are the new core European loci. The loci are arranged according to their Overall Rank, which is determined by the Heterozygosity Rank added to the Size Rank.

Loc	Report	Type	Chromosome	N	Heterozygosity Rank	African American	Caucasian	Hispanic	Observed Range (bp)	Size Rank	Overall Rank (Size Rank)
D20S1249 (NC01)	Tel	ATCA	1	661	0.844	4	0.864	0.779	71 - 101	4	1
D20S1045 (NC01)	Tel	TAA	Chr 22: 7.8 Mb	663	0.732	11	0.817	0.785	71 - 101	8	3
D20S441 (NC01)	Tel	TCTA	Chr 28: 21.6 Mb	660	0.774	12	0.798	0.721	78 - 110	9	4
D20S1082 (NC01)	Tel	GCG	Chr 10: 12.8 Mb	660	0.743	13	0.792	0.743	78 - 110	15	5
D10S1434 (NC02)	Tel	GATA	Chr 22: 2.3 Mb	663	0.766	15	0.798	0.770	82 - 139	16	6
D10S1777 (NC02)	Tel	ATT	Chr 10: 8.8 Mb	660	0.746	22	0.793	0.727	81 - 150	18	7
D10S1203 (NC02)	Tel	AGGG	Chr 1: 14.6 Mb	660	0.745	23	0.793	0.724	81 - 140	19	8
D10S167 (NC02)	Tel	GGAA	Chr 1: 16.5 Mb	660	0.746	23	0.743	0.748	81 - 117	13	9
D10S119 (NC02)	Tel	ATCC	Chr 21: 8.3 Mb	659	0.861	24	0.869	0.808	81 - 166	36	10
D21S151 (NC02)	Tel	TTCT	Chr 21: 4.8 Mb	659	0.844	25	0.829	0.871	185 - 244	34	12
D20S1249 (NC02)	Comp	[GATG]GACA	Chr 5: 58.7 Mb	664	0.747	26	0.757	0.729	85 - 125	19	12
D20S1045 (NC02)	Comp	[GATG]GATTC	Chr 1: 21.7 Mb	659	0.865	27	0.792	0.724	185 - 244	42	22
D20S441 (NC02)	Comp	[GATG]GATC	Chr 1: 8.4 Mb	660	0.761	28	0.787	0.757	97 - 145	24	18
D20S1082 (NC02)	Comp	[GATG]GATCA	Chr 3: 54.0 Mb	648	0.769	29	0.829	0.763	177 - 259	28	20
D10S1434 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.9 Mb	659	0.774	30	0.783	0.759	181 - 259	30	20
D10S1777 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.5 Mb	659	0.774	31	0.783	0.759	181 - 259	31	20
D10S1203 (NC02)	Comp	[GATG]GATCA	Chr 1: 14.6 Mb	660	0.746	32	0.793	0.727	181 - 259	32	20
D10S167 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.5 Mb	660	0.746	33	0.793	0.727	181 - 259	33	20
D10S119 (NC02)	Comp	[GATG]GATCA	Chr 1: 14.6 Mb	660	0.746	34	0.793	0.727	181 - 259	34	20
D20S1249 (NC02)	Comp	[GATG]GATCA	Chr 5: 58.7 Mb	664	0.741	35	0.785	0.724	181 - 259	35	20
D20S1045 (NC02)	Comp	[GATG]GATCA	Chr 1: 21.7 Mb	659	0.761	36	0.787	0.757	181 - 259	36	20
D20S441 (NC02)	Comp	[GATG]GATCA	Chr 1: 8.4 Mb	660	0.769	37	0.787	0.759	181 - 259	37	20
D20S1082 (NC02)	Comp	[GATG]GATCA	Chr 3: 54.0 Mb	648	0.769	38	0.793	0.727	181 - 259	38	20
D10S1434 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.9 Mb	659	0.745	39	0.793	0.727	181 - 259	39	20
D10S1777 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.5 Mb	659	0.745	40	0.793	0.727	181 - 259	40	20
D10S1203 (NC02)	Comp	[GATG]GATCA	Chr 1: 14.6 Mb	660	0.746	41	0.793	0.727	181 - 259	41	20
D10S167 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.5 Mb	660	0.746	42	0.793	0.727	181 - 259	42	20
D10S119 (NC02)	Comp	[GATG]GATCA	Chr 1: 14.6 Mb	660	0.746	43	0.793	0.727	181 - 259	43	20
D20S1249 (NC02)	Comp	[GATG]GATCA	Chr 5: 58.7 Mb	664	0.741	44	0.785	0.724	181 - 259	44	20
D20S1045 (NC02)	Comp	[GATG]GATCA	Chr 1: 21.7 Mb	659	0.761	45	0.787	0.757	181 - 259	45	20
D20S441 (NC02)	Comp	[GATG]GATCA	Chr 1: 8.4 Mb	660	0.769	46	0.787	0.759	181 - 259	46	20
D20S1082 (NC02)	Comp	[GATG]GATCA	Chr 3: 54.0 Mb	648	0.769	47	0.793	0.727	181 - 259	47	20
D10S1434 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.9 Mb	659	0.745	48	0.793	0.727	181 - 259	48	20
D10S1777 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.5 Mb	659	0.745	49	0.793	0.727	181 - 259	49	20
D10S1203 (NC02)	Comp	[GATG]GATCA	Chr 1: 14.6 Mb	660	0.746	50	0.793	0.727	181 - 259	50	20
D10S167 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.5 Mb	660	0.746	51	0.793	0.727	181 - 259	51	20
D10S119 (NC02)	Comp	[GATG]GATCA	Chr 1: 14.6 Mb	660	0.746	52	0.793	0.727	181 - 259	52	20
D20S1249 (NC02)	Comp	[GATG]GATCA	Chr 5: 58.7 Mb	664	0.741	53	0.785	0.724	181 - 259	53	20
D20S1045 (NC02)	Comp	[GATG]GATCA	Chr 1: 21.7 Mb	659	0.761	54	0.787	0.757	181 - 259	54	20
D20S441 (NC02)	Comp	[GATG]GATCA	Chr 1: 8.4 Mb	660	0.769	55	0.787	0.759	181 - 259	55	20
D20S1082 (NC02)	Comp	[GATG]GATCA	Chr 3: 54.0 Mb	648	0.769	56	0.793	0.727	181 - 259	56	20
D10S1434 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.9 Mb	659	0.745	57	0.793	0.727	181 - 259	57	20
D10S1777 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.5 Mb	659	0.745	58	0.793	0.727	181 - 259	58	20
D10S1203 (NC02)	Comp	[GATG]GATCA	Chr 1: 14.6 Mb	660	0.746	59	0.793	0.727	181 - 259	59	20
D10S167 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.5 Mb	660	0.746	60	0.793	0.727	181 - 259	60	20
D10S119 (NC02)	Comp	[GATG]GATCA	Chr 1: 14.6 Mb	660	0.746	61	0.793	0.727	181 - 259	61	20
D20S1249 (NC02)	Comp	[GATG]GATCA	Chr 5: 58.7 Mb	664	0.741	62	0.785	0.724	181 - 259	62	20
D20S1045 (NC02)	Comp	[GATG]GATCA	Chr 1: 21.7 Mb	659	0.761	63	0.787	0.757	181 - 259	63	20
D20S441 (NC02)	Comp	[GATG]GATCA	Chr 1: 8.4 Mb	660	0.769	64	0.787	0.759	181 - 259	64	20
D20S1082 (NC02)	Comp	[GATG]GATCA	Chr 3: 54.0 Mb	648	0.769	65	0.793	0.727	181 - 259	65	20
D10S1434 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.9 Mb	659	0.745	66	0.793	0.727	181 - 259	66	20
D10S1777 (NC02)	Comp	[GATG]GATCA	Chr 1: 16.5 Mb	659	0.745	67	0.793	0.727	181 - 259	67	20
D10S1203 (NC02)	Comp	[GATG]GATCA	Chr 1: 14.6 Mb	660	0.746						