



Capillary Electrophoresis in DNA Analysis

Higher Throughput Approaches

NEAFS Workshop
Mystic, CT
September 29-30, 2004
Dr. John M. Butler
Dr. Bruce R. McCord



NIST
National Institute of Standards and Technology
Technology Administration, U.S. Department of Commerce



FIU
FLORIDA INTERNATIONAL UNIVERSITY
Miami's public research university

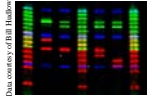
Outline for Workshop

- Introductions
- STR Analysis
- Introduction to CE and ABI 310
- Data Interpretation
- Additional Topics – Real-time PCR and miniSTRs
- **Higher Throughput Approaches**
- Troubleshooting the ABI 310 (Participant Roundtable)
- Additional Topics – Y-STRs, validation, accuracy
- Review and Test

STR Typing Technologies


<http://www.cstl.nist.gov/biotech/strbase/tech.htm>

Gels



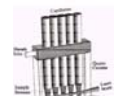
J. Forensic Sci. (1998) 43: 1168-1180

Capillary Electrophoresis



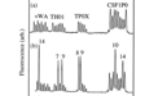
Electrophoresis. (1998) 19: 86-93

Capillary Arrays




Nucleic Acids Res. (1999) 27: e36

Microchip CE



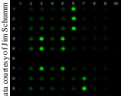
PNAS (1997) 94: 10273-10278

Mass Spectrometry



Int. J. Legal Med. (1998) 112: 45-49

Hybridization Arrays




Nucleic Acids Res. (2000) 28: e17

Ways to Increase Sample Throughput

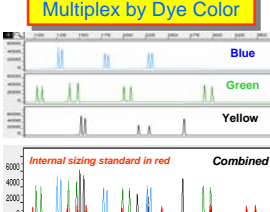
- Run more gels (FMBIO approach)
- Increase speed of single sample analysis (microchip CE systems)
- Multiplex fluorescent dyes of different colors (higher level PCR multiplexes)
- Parallel separations using capillary arrays
- New Detection Technologies (MALDI-TOF mass spectrometry)

Methods for Parallel Sample Processing

Multiplex by Size

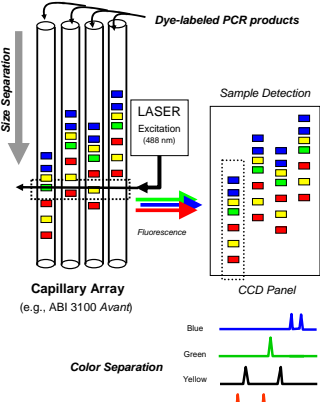


Multiplex by Dye Color



Multiplex by Number of Capillaries

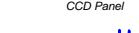
ABI 3100: 16 capillaries
ABI 3730: 96 capillaries
ABI 3100 Avant: 4 capillaries




Capillary Array
(e.g., ABI 3100 Avant)

Color Separation


Blue



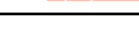
Green



Yellow



Red




From Butler, J.M. and Vallone, P.M. (2004) High-throughput genetic analysis through multiplexed PCR and multicapillary electrophoresis. PCR Technologies: Current Innovations (2nd edition), Weissensteiner, T., Griffin, H.G., Griffin, A. (Eds.), CRC Press: Boca Raton, Chapter 11, pp 111-120.

Capillary Array Electrophoresis

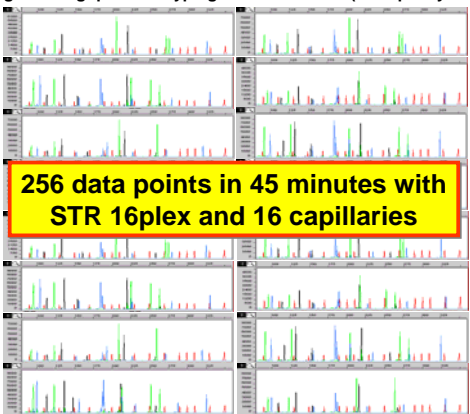
- Higher sample throughput
- Commercial 96 capillary systems were used to sequence the human genome
 - ABI 3700
 - MegaBACE
- Engineering and hardware challenges
- Software challenges

16 Capillary 3100



6 foot Table on wheels

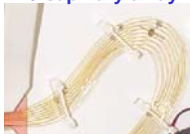
High-Throughput STR Typing on the ABI 3100 (16-capillary array)



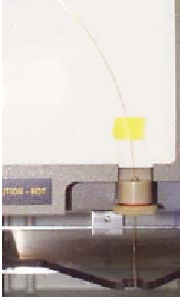
256 data points in 45 minutes with STR 16plex and 16 capillaries

Increasing Sample Throughput with Parallel Processing

ABI 3100
16-capillary array

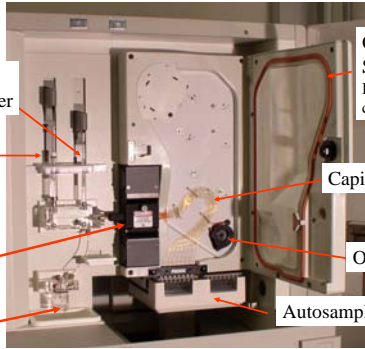


ABI 310
single capillary

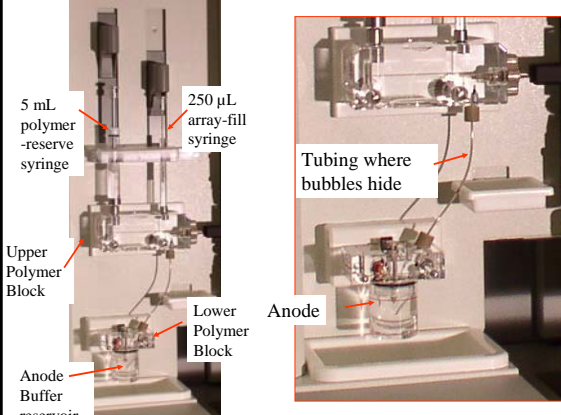


Subtle differences in matrix formation and sizing algorithms – NOT directly equivalent to 310

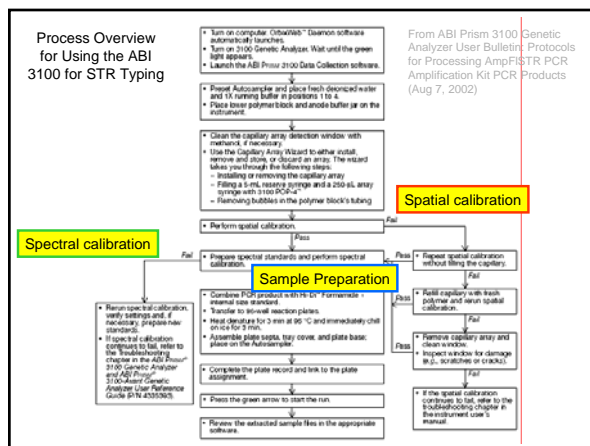
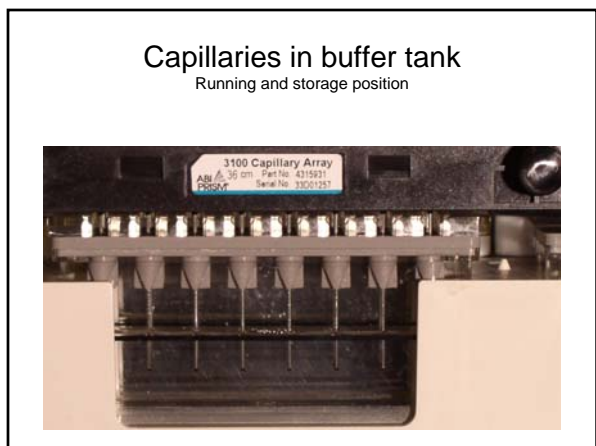
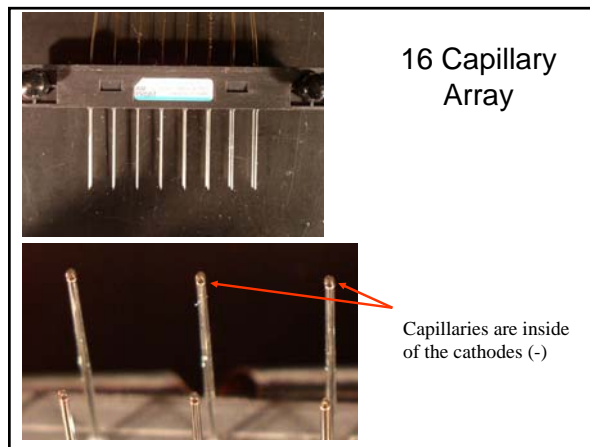
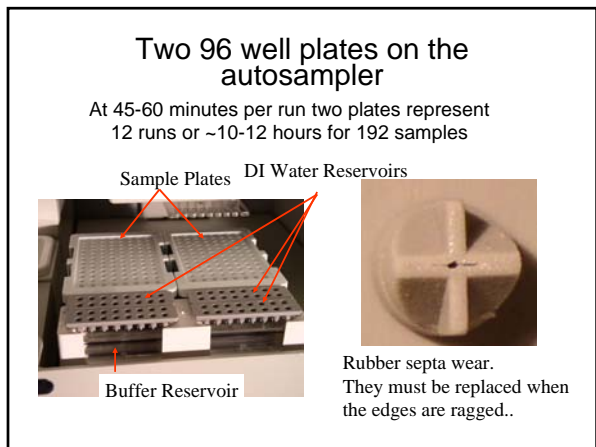
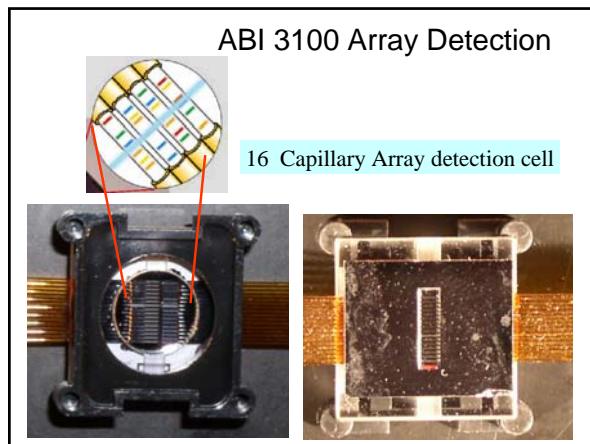
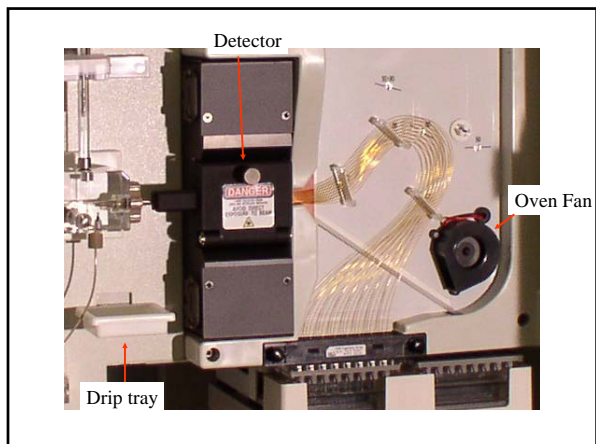
Inside the 3100



- 1 mL syringe Loads polymer
- 5 mL syringe Polymer reservoir
- Detection window
- Buffer reservoir
- Oven Seal Better temp control
- Capillary array
- Oven fan
- Autosampler



- 5 mL polymer-reserve syringe
- 250 µL array-fill syringe
- Upper Polymer Block
- Lower Polymer Block
- Anode Buffer reservoir
- Tubing where bubbles hide
- Anode



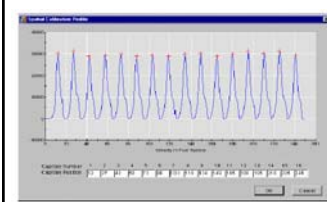
Spatial Calibration

Performed after:

- Installing or replacing a capillary array
- Removal of the array from the detection block, (Due to the design, to remove the upper polymer block for cleaning you must remove the Array from the detection window)

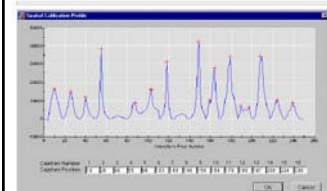
Information Provided:

Position of the fluorescence from each capillary on the CCD



Spatial Results

Good Results



Bad results
Try again

Maintenance of ABI 3100

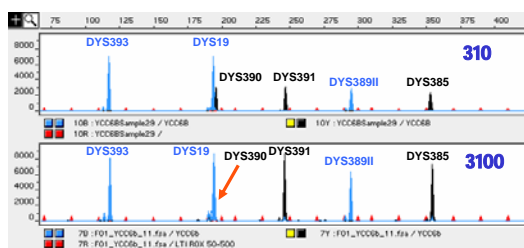
- Syringe – leaks cause capillary to not fill properly
- Capillary storage & wash – **it dries, it dies!**
- Pump block – cleaning helps insure good fill
- Change the running buffer regularly

YOU MUST BE CLEAN AROUND A CE!

Spectral Calibration

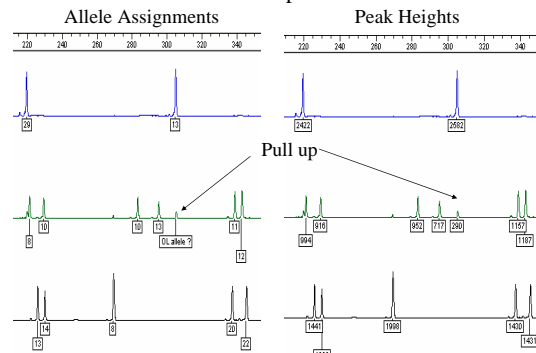
- Performed:
 - New dye set on the instrument
 - After Laser or CCD camera has been realigned
 - You begin to see a decrease in the spectral separation (pull-up, pull-down).
- You must have a valid separation matrix on the instrument prior to running samples.

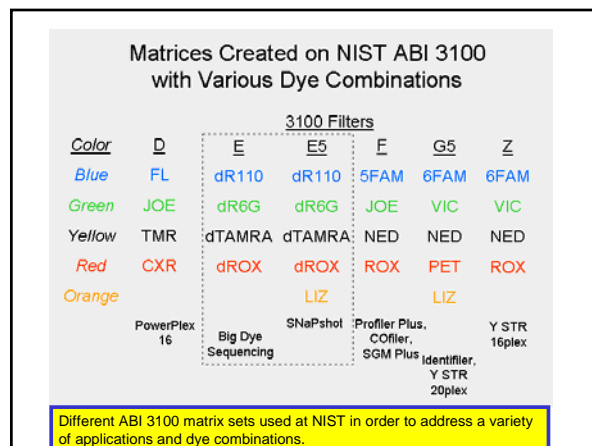
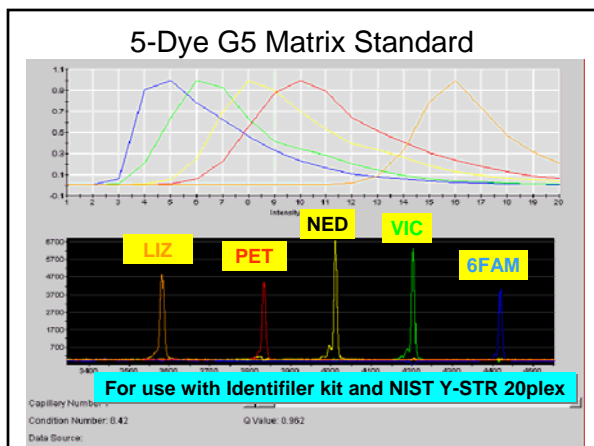
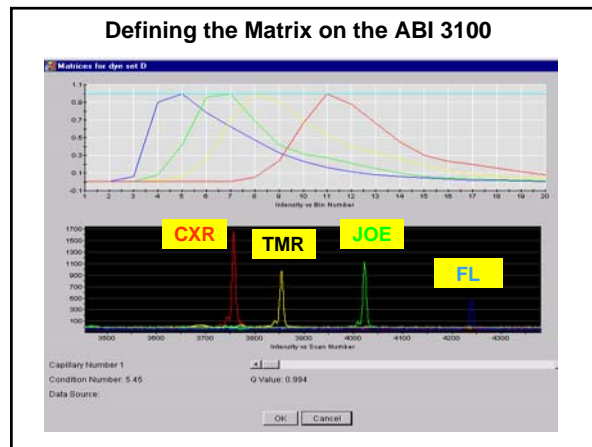
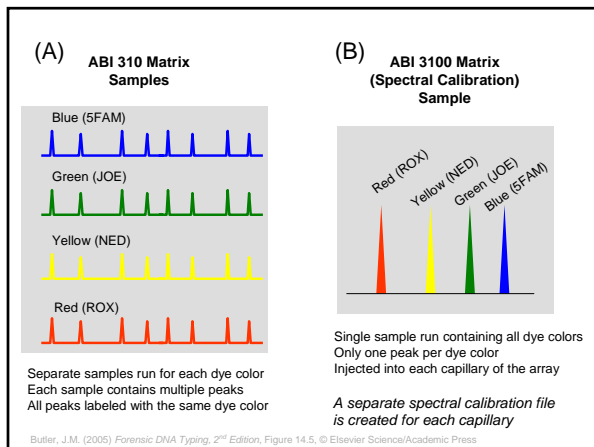
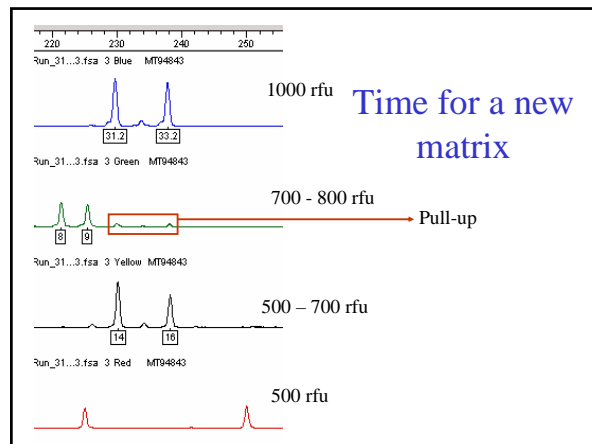
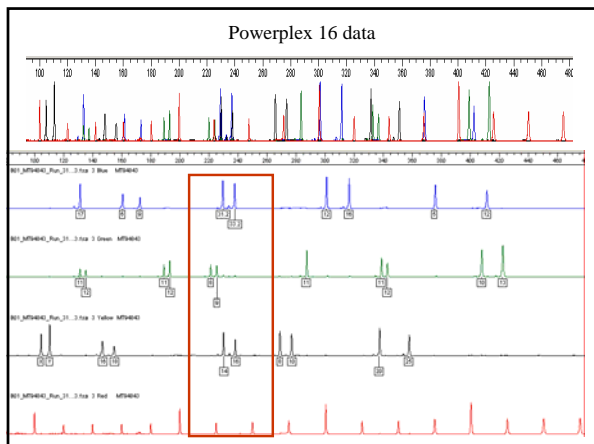
Use of the Correct Matrix is Critical

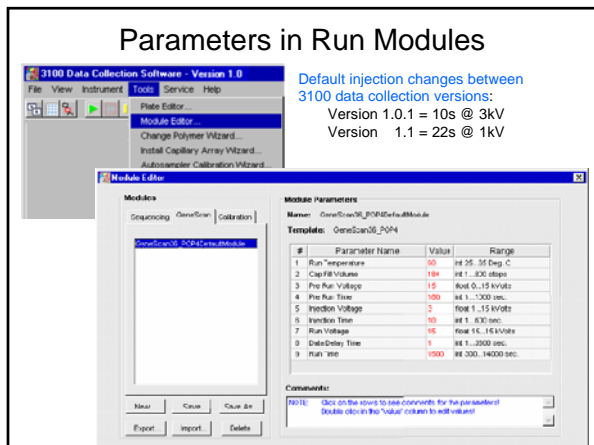
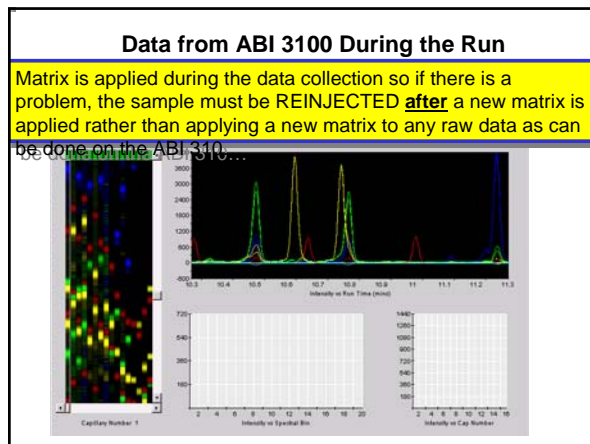
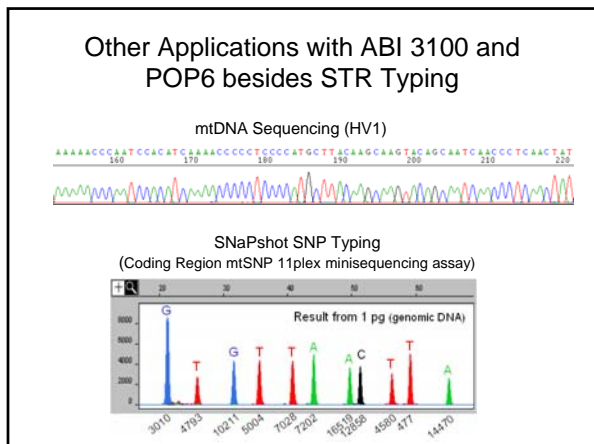


This figure shows the same sample, amplified with the Reliagene Y-Plex™ 6 kit, injected into both the 310 and the 3100. Note that the DYS390 allele, which is labeled with TAMRA as the yellow dye, does not show up in the 3100 result. The matrix used in this case contained NED rather than TAMRA. Thus, if the matrix for the particular dye combination is not established properly, peaks can disappear.

Pull-up issue







- ### Consumables
- ABI Optical Reaction Plates
 - \$2,200 / 500 plates = \$4.40 / plate
 - Phenix (mps-3590)
 - Plates \$291/100 plates = \$2.91 / plate
 - Hi – Di Formamide
 - \$28 / 25 mL
 - 36 cm 3100 Capillary Array (100 runs) \$695
 - 281 runs and still going (replace by resolution not # of injections)
 - 36 cm 3100 Avant Capillary Array (150 runs) \$560

- ### Consumables
- 10X Genetic Analyzer Buffer with EDTA
 - \$75/25 mL = \$0.30/mL 1X buffer (ABI)
 - Or A.C.E.™ Sequencing Buffer 10X
 - \$155/L = \$0.016/mL 1X buffer (Amresco)
 - 3100 POP-4 Polymer \$365 / 7 mL
 - 3100 POP-6 Polymer \$365 / 7 mL
 - 3700 POP-6 Polymer \$465 / 230 mL
 - What we have been using, runs take longer but you also get better resolution.

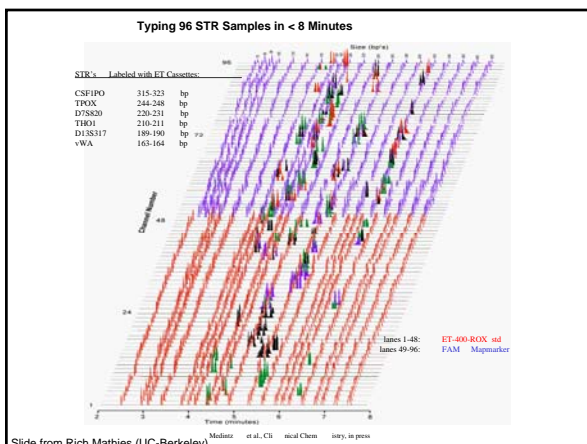
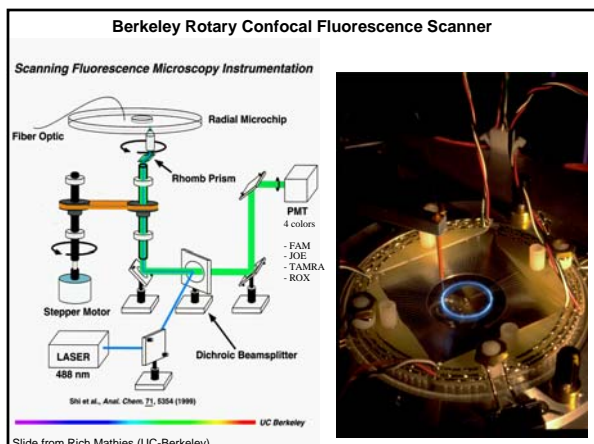
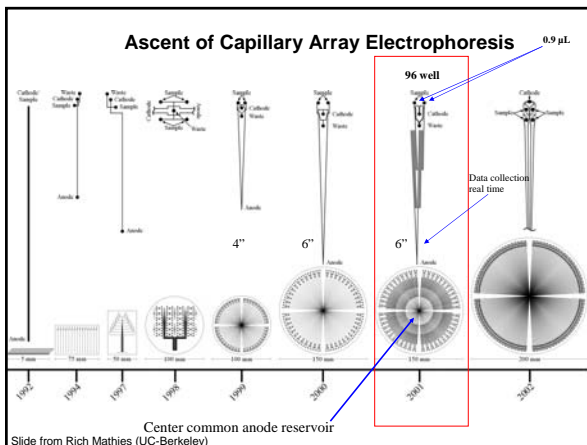
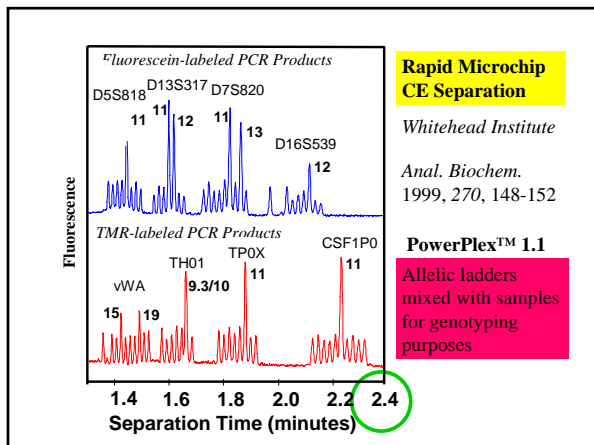
Microchip CE Systems

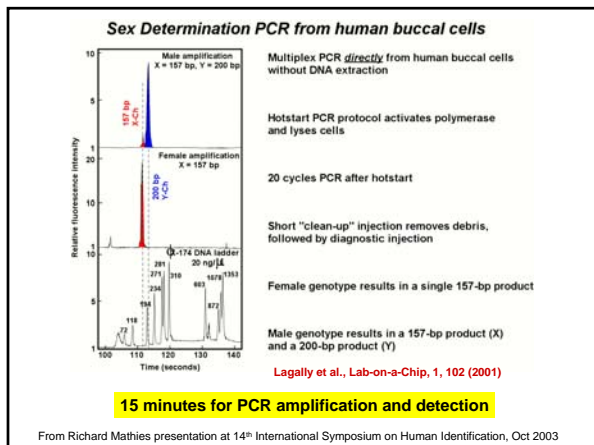
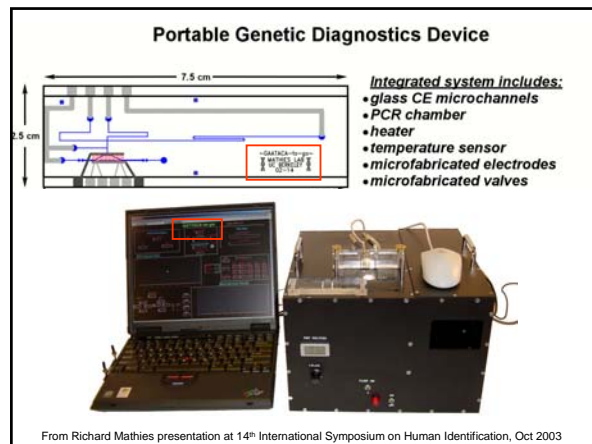
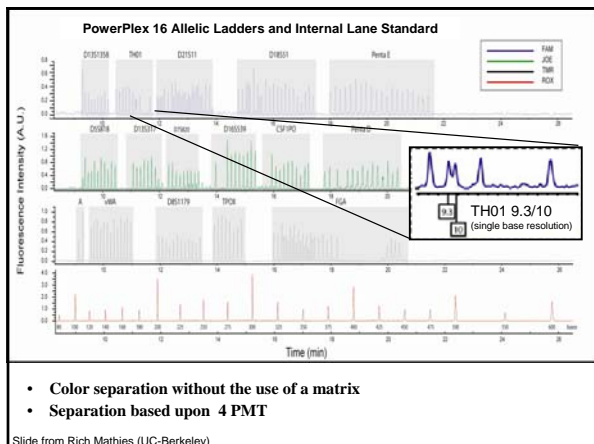
What is under development for STR typing?



CE Microchips

- Channels are etched in glass microscope slides to make miniature CE columns
- More rapid separations are possible due to the shorter separation length
- Possible to etch many channels CAE microchips





Virginia DNA Testing of Felon Arrestees

As of January 1, 2003, any individual arrested for a violent felony crime (Code of Virginia § 19.2-310.2:1) must provide a buccal sample for DNA analysis, with the resultant profile incorporated into the Virginia DNA Data Bank (Code of Virginia § 19.2-310.5).

Since January 2003

- Buccal swab collected upon arrest
- DNA sample processed within 72 hours
- DNA profile searched against state database (national database does not currently allow searches for individuals prior to conviction)
- If a match results, then arrestee is detained and later prosecuted
- From Jan 2003 – Dec 2003, VA processed 7,836 arrestee samples (not all analyzed) and scored 63 hits against their state database (Profiles in DNA, 2004, 7(1):3-5)

Time-of-Flight Mass Spectrometry

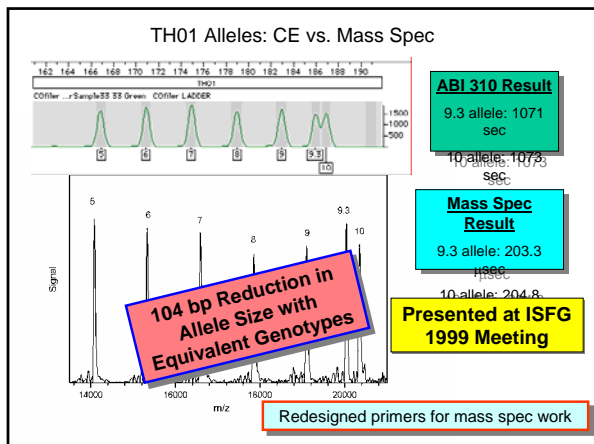
Why it will not become widely used...

Recent NIJ Publication

Final Report for NIJ Grant 97-LB-VX-0003 (work done at GeneTrace Systems Inc.)

- Describes new primer sets that are close to the STR repeat regions
- Many of these primers are being used in miniplex STR assays under development
- Y SNP multiplex primer sets are described
- 10plex mtSNP assay for HV1 and HV2 detailed

<http://www.ojp.usdoj.gov/nij/pubs-sum/188292.htm>



Timing for Data Collection

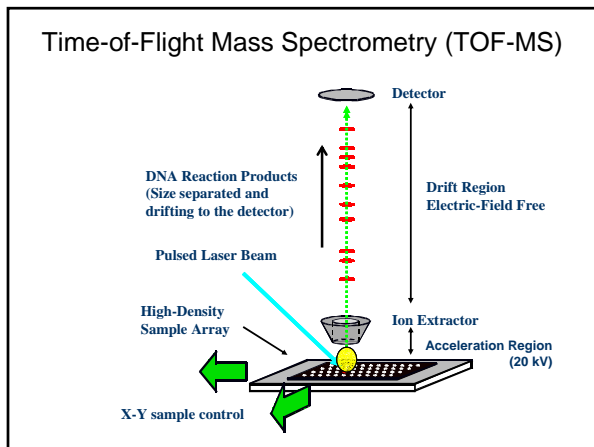
Laser pulse (10 nsec)	Wait (500 nsec)	Turn on voltages for ion optics (+20 kV)	Extract DNA ions	Turn off voltages
			Collect spectrum for ~300,000 nsec	

REPEAT process 100+ times

All this occurs in less than 5 seconds per sample

Sum multiple spectra into final sample spectrum

Data processing and genotype determination



Demonstrated Throughput at GeneTrace Systems

- 384 samples processed routinely in ~45 min (best was 96 samples in 2 min)
- ~4,000 samples in 11 hours on single mass spec and 3 robots
- averaged around 2,000 samples daily at GTS per instrument (Jan-Aug 1999)
- **most samples run as singleplex reactions** but demonstrated 10-plex SNP assay and 3-plex STR assay

High throughput mass spectrometer with rapid laser and automated sample acquisition

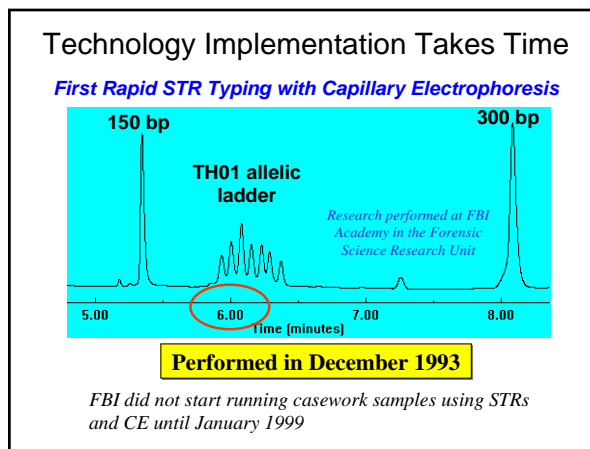
<http://www.atp.nist.gov/atp/success/genet.htm>

Improvements in Information Throughput with Multiplexed Markers and Multiple Capillaries

Time required to obtain each genotype...

	Single capillary (ABI 310)	16 capillary array (ABI 3100)	96 capillary array (ABI 3700)
	Each run: 30 min	Each run: 45 min	Run: 2 h 46 min
#Markers Multiplexed	1800 s (per capillary)	2700 s (per capillary)	9960 s (per capillary)
1	1800 s (30 min)	169 s (2.8 min)	104 s (1.7 min)
8	225 s	21 s	13 s
16	113 s	10.5 s	6.5 s

From Butler, J.M. and Vallone, P.M. (2004) High-throughput genetic analysis through multiplexed PCR and multicapillary electrophoresis. *PCR Technologies: Current Innovations* (2nd edition), Weissensteiner, T., Griffin, H.G., Griffin, A. (Eds.), CRC Press: Boca Raton, Chapter 11, pp 111-120.



Where is the Future Going?...
Miniaturization and Portability



Palm Pilot
(handheld computer)

http://www.nanogen.com/products/nanochip_cart.asp



NanoChip™ from Nanogen
(Miniature Bioassay Device)